

## Research Article

# Organic Matter and Barium Absorption by Plant Species Grown in an Area Polluted with Scrap Metal Residue

Cleide Aparecida Abreu,<sup>1</sup> Mariana Cantoni,<sup>2</sup> Aline Renée Coscione,<sup>1</sup>  
and Jorge Paz-Ferreiro<sup>3</sup>

<sup>1</sup>Centro de Solo e Recursos Ambientais, IAC, Avenida Barão de Itapura, 1481, 13020-902 Campinas, SP, Brazil

<sup>2</sup>Programa de Pós-Graduação em Agricultura Tropical e Subtropical, IAC, 13020-902 Campinas, SP, Brazil

<sup>3</sup>Departamento de Edafología, Universidad Politécnica de Madrid, 28004 Madrid, Spain

Correspondence should be addressed to Jorge Paz-Ferreiro, jpaz@udc.es

Received 18 October 2011; Revised 27 December 2011; Accepted 8 January 2012

Academic Editor: Philip White

Copyright © 2012 Cleide Aparecida Abreu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The effect of organic matter addition on Ba availability to *Helianthus annuus* L., *Raphanus sativus* L., and *Ricinus communis* L. grown on a Neossolo Litólico Chernossólico fragmentário (pH 7.5), contaminated with scrap residue was evaluated. Four rates (0, 20, 40, and 80 Mg ha<sup>-1</sup>, organic carbon basis) of peat or sugar cane filter, with three replicates, were tested. Plant species were grown until the flowering stage. No effect of organic matter addition to soil on dry matter yield of oilseed radish shoots was observed, but there was an increase in sunflower and castor oil plant shoots when sugar cane filter cake was used. The average Ba transferred from roots to shoots was more than 89% for oilseed radish, 71% for castor oil plants, and 59% for sunflowers. Organic matter treatments were not efficient in reducing Ba availability due to soil liming.

## 1. Introduction

Accumulation of some chemical elements in the environment is of great concern because they can reach concentrations that may cause risks to human health and to the environment. Their concentration in soils depends on lithogenic and pedogenic processes, but also on anthropogenic activities. Soil pollution is a serious problem in many countries around the world. In São Paulo State, Brazil, since 2002, when the first survey was performed by the local environmental agency, more than 1600 contaminated areas have been identified [1].

The extensive industrial use of barium (Ba) adds up to the release of Ba in the environment and, as a result, Ba concentrations in air, water, and soil may be higher than naturally occurring concentrations on many locations [2–5]. Recently, it was observed that successive sewage sludge applications increased soil Ba concentration and accumulation in maize plants grown in the State of São Paulo [6]. Some research has shown probable Ba toxicity in plants, but such studies were short term and performed in nutrient solution

[7, 8]. Ba is an alkaline earth element which occurs as a trace metal in igneous and sedimentary rocks. In nature it occurs mainly as low soluble minerals such as barite (BaSO<sub>4</sub>) and witherite (BaCO<sub>3</sub>). Ba solubilization and, consequently, the release of Ba<sup>2+</sup> ions may occur under specific conditions. It has been shown to happen in acidic conditions [9], in the absence of oxygen, or even due to microbial action [10–13]. In contrast, Ba precipitates as a sulfate and/or carbonate salt in neutral or basic pH conditions. Therefore, the mobility of Ba is negligible in neutral or basic pH conditions, thus, reducing the risks of leaching and harmful health effects.

The application of lime and the addition of organic materials are considered the most efficient options to reduce heavy metal availability in soils [14–16]. The use of organic matter in chemically degraded areas can also be beneficial since plant development in such areas is frequently affected, exposing the soil to physical degradation.

Peat and humic materials concentrate reduced extractable Zn, Cu, Pb, and B in soil and mustard shoots [14] while liming reduced the available concentrations of Cd, Pb, Cu, and Zn in soils as well as its content in velvetbean shoots

TABLE 1: Chemical composition of the organic materials\*.

Source	pH	E.C. dS m <sup>-1</sup>	O.C. g kg <sup>-1</sup>	C/N	P	K	Ca g kg <sup>-1</sup>	Mg g kg <sup>-1</sup>	B	Cu	Fe	Mn Mg kg <sup>-1</sup>	Zn
Sugar cane filter cake	7.5	0.9	263.7	12	10.3	2.3	16.2	3.7	21	60	5900	557	141
Peat	5.5	0.2	163.1	24	0.8	1.4	1.7	1.7	16	45	6300	47	36

E.C.: electrical conductivity; O.C.: organic carbon.

\*Total elements concentration obtained by extraction with a mix of nitric and perchloric acids [17], results presented are the average of six replicates.

[16]. The bioavailability of heavy metals to soybean and black-oat cultures was close to zero, when 8 Mg ha<sup>-1</sup> sewage sludge, flue dust, and aqueous lime was applied to soil surface in no-till system [15]. However, lime and organic matter addition to the Ba contaminated soil and its availability to soil and its plants absorption have not received a great deal of attention and few information on the topic has been reported. The organic matter complexation of Ba ions can lead to insoluble species, decreasing the availability of Ba and enabling the growth of vegetation in highly contaminated areas [3]. Consequently, Ba effects on plant grown in soils containing Ba still needs to be further investigated.

The aim of the present work was to evaluate the effect of application rates of peat and sugar cane filter cake on Ba concentration in soil and its potential availability to sunflowers (*Helianthus annuus* L.), oilseed radish (*Raphanus sativus* L.), and castor oil plants (*Ricinus communis* L.) grown in a soil (pH 7.5) contaminated with scrap metal residue.

## 2. Material and Methods

In 2005, automobile scrap “shredder residue” was applied to the soil of an agricultural area of approximately 3 ha located in Piracicaba (22°42′30″ S, 47°38′01″ W), São Paulo State, Brazil. The residue’s metal content, obtained by the SW-846 3051 method [18] was, in mg kg<sup>-1</sup>: 170 of B, 7.4 of Cd, 2497 of Cu, 775 of Pb, 178 of Cr, 153 of Ni, 8157 of Zn, and 920 of Ba. Residue addition was performed based on the supposition that it may provide Zn and Cu to sugar cane crops, and the residue was incorporated into the soil at a depth of 30 cm. The local environmental agency (CETESB) later verified that the area was contaminated by heavy metals (copper and zinc) and boron. Lime (10 Mg ha<sup>-1</sup>) was added to the soil in order to reduce heavy metals mobility and potential leaching. The soil in this area is classified as Lithic Udorthent [19].

Soil samples were taken from the 0–20 cm depth layer, dried at room temperature, and sieved to 2.0 mm. The soil fertility attributes were measured as follows: pH<sub>CaCl2</sub> = 7.5; MO = 30.5 g dm<sup>-3</sup>; P<sub>resin</sub> = 43.3 mg dm<sup>-3</sup>; K<sub>resin</sub> = 2.6 mmol<sub>c</sub> dm<sup>-3</sup>; Ca<sub>resin</sub> = 294 mmol<sub>c</sub> dm<sup>-3</sup>; Mg<sub>resin</sub> = 59 mmol<sub>c</sub> dm<sup>-3</sup>; CEC = 364 mmol<sub>c</sub> dm<sup>-3</sup>; H + Al = 9.0 mmol<sub>c</sub> dm<sup>-3</sup>; V = 98% according to [20]. For the determination P, K, Ca a mixed (cationic and anionic) ion exchange resin method (Amberlite IRA 120 and Amberlite IRA 400) was used to simulate elements soil availability. It

employs a ratio of 2.5 of soil per 2.5 cm<sup>3</sup> of resin, which is kept in contact for 16 hours. The elements adsorbed by resin are washed away with 50 mL of a 0.8 mol L<sup>-1</sup> NH<sub>4</sub>Cl + 0.2 mol L<sup>-1</sup> HCl, producing an extract where the elements are determined. Some of total elements content in the soil were measured by SW-846 3051 method [20] as follows, in mg kg<sup>-1</sup>: 241 of Ba, 62 of B, 4.3 of Cd, 335 of Cu, 332 of Pb, 88.2 of Cr, 53.6 of Ni, and 2998 of Zn. This procedure consists of adding 10 mL of HNO<sub>3</sub> to 500 mg soil in a teflon capped vessel in a laboratory microwave system (CEM, Mars 5 model, Xpress vessels). The extraction is performed by raising the temperature to 170°C for 5 min and keeping it at this temperature during 10 minutes.

The experiment was carried out in a greenhouse at Campinas (São Paulo State, Brazil) in plastic pots (5 dm<sup>-3</sup>). The following plant species: sunflowers (*Helianthus annuus* L.), oilseed radish (*Raphanus sativus* L.), and castor oil plants (*Ricinus communis* L.) were selected for the experiment due to previous works showing them to be tolerant to high concentration of heavy metals and boron in soil [21–24].

The experimental design was in randomized complete blocks with four rates (0, 20, 40, and 80 Mg ha<sup>-1</sup>, organic carbon basis) of two organic matter sources (peat and sugar cane filter cake), with three replicates. The treatments were applied at (g pot<sup>-1</sup>): 0.0, 37.9, 75.8, and 151.6 g of sugar cane filter cake per pot, respectively and 0.0, 61.3, 122.6, and 245.2 g of peat per pot. The chemical compositions of the peat and sugar cane filter cake (Table 1) were obtained by determination of elements in a 0.5 mg of sample extracted with nitric perchloric acids (3 : 1 ratio) [17].

The soil/organic materials were carefully homogenized and incubated at room temperature for 20 days with soil moisture maintained at 60% water holding capacity (WHC). The pots received 200 mg kg<sup>-1</sup> of P as triple superphosphate (41% P<sub>2</sub>O<sub>5</sub>) and the samples were homogenized and incubated for an additional 15 days after the sowing of seeds.

Three sunflowers and castor oil plants and ten oilseed radish were grown per pot. Deionized water was supplied by weighing the pots daily and adding the water needed to maintain 60% WHC. Nitrogen (30 mg N kg<sup>-1</sup> soil) was applied as ammonium nitrate (32% N) on emerging seedlings and again 15 days later.

Sunflower and oilseed radish were harvested 65 days after sowing, while castor oil plants were harvested 74 days after sowing. Shoots were separated from roots at harvest. The flowers were also separated when the oilseed radish and sunflowers were harvested. Roots were sieved, washed

and soaked for 90 min in a solution of 0.02 mmol L<sup>-1</sup> EDTA (disodium salt). After soaking, the oilseed radish roots were washed again with distilled water. Plant samples were washed, dried, and weighed and then digested using HNO<sub>3</sub>/H<sub>2</sub>O<sub>2</sub> in a CEM Mars 5 microwave oven and analyzed for macro- and micronutrients, and barium, lead, cadmium, chromium, and nickel.

Soil samples collected after incubation were air-dried and sieved through a 2 mm mesh screen and then characterized for total and available metal contents. Available Ba content was analyzed using Mehlich-3 method (CH<sub>3</sub>COOH 0.2 mol L<sup>-1</sup> + NH<sub>4</sub>NO<sub>3</sub> 0.25 mol L<sup>-1</sup> + NH<sub>4</sub>F 0.015 mol L<sup>-1</sup> + HNO<sub>3</sub> 0.015 mol L<sup>-1</sup> + EDTA 0.001 mol L<sup>-1</sup> at pH 2.5) by agitation of five cm<sup>3</sup> of soil and 20 mL the Mehlich-3 solution for 5 min [25]. The availability of several nutrients (P, K, Ca, and Mg) was evaluated by the ion exchange method [20].

Ba transported from soil to shoots was evaluated using the transfer factor (TF) as follows:  $TF = PC (mg\ kg^{-1})/SC (mg\ kg^{-1})$ , where CP is the Ba concentration in the whole plant (root and shoot), and CT is the concentration of Ba in the soil [26]. The ability of each species to translocate Ba from the roots to the shoots was calculated using the following translocation index (TI):  $TI (\%) = QPA (mg\ pot^{-1})/QAP (mg\ pot^{-1}) \times 100$ , where QPA is the element accumulation in the shoots, and QAP is the element accumulation in the whole plant (shoots and roots) [26].

The plant efficiency for the removal of elements (removal factor, *E*) was calculated using the following equation:  $E (\%) = QPA (mg\ pot^{-1})/QR (mg\ pot^{-1}) \times 100$ , where QR is the amount of Ba to be removed from the soil (mg pot<sup>-1</sup>) [27]. When considering a 75% reduction of Ba concentration in the soil as a target, the time (*T*, in years) needed for Ba removal was calculated as follows:  $T = (R/E)/NC$ , where *R* is the percentage of Ba reduction in the soil, *E* is the removal factor, and NC is the number of crop cycles/year (considered as 1 cycle/year).

The data were submitted to analyses of variance (ANOVA), and the mean values were compared according to Tukey's test ( $P \leq 0.05$ ). When significant, the results obtained with the different concentrations of organic material were also examined using regression analysis (linear and quadratic models tested).

### 3. Results and Discussion

The concentration of Ba found during the soil characterization (241 mg kg<sup>-1</sup>) was close to the intervention levels (300 mg kg<sup>-1</sup>) established by the Environmental Agency of the State of São Paulo [28]. Although the concentration of zinc, copper, and boron in this area is also worrisome, the use of plants that could help to remediate the soil was studied on previous works and was not significant in the present work as also discussed below [24, 29].

Mehlich-3 available Ba increased in soils amended with 40 Mg ha<sup>-1</sup> of peat and 80 Mg ha<sup>-1</sup> of sugar cane filter cake, with an average of 32.9 mg dm<sup>-3</sup> in the soils amended with sugar cane filter cake and 36.2 mg dm<sup>-3</sup> in the soils amended with peat (Table 2), which corresponded to a 12.3% and

TABLE 2: Ba extracted from soil with the Mehlich-3 method\*.

Rate	Sugar cane filter cake	Peat
Mg ha <sup>-1</sup>	mg dm <sup>-3</sup>	
0	31.5 a	34.9 a
20	32.9 a	34.1 a
40	32.9 a	36.7 ab
80	34.6 b	39.3 b
Average	32.9 A	36.2 A

\*Results presented are the average of 3 replicates. Means followed by the same letter are not significantly different by the Tukey's test at  $P \leq 0.05$ . Upper case letters, in columns, compare treatments and lower case letters, in lines, compare rate of amendments.

14.4% recovery, respectively. The recovery found in this study was lower than the one reported by others which was in a range from 50% to 78% [30]. The correlation between extractable Ba and soil organic carbon was 0.96  $P < 0.05$  (sugar cane filter cake) and 0.95  $P < 0.05$  (peat). However, no significant correlation was found between extractable Ba in the soil and the Ba accumulated in all of the plant tissues.

In most plants, the concentration of Ba ranges from 4 to 50 mg kg<sup>-1</sup> [31], and concentrations of 200 and 500 mg kg<sup>-1</sup> are considered to be slightly toxic or toxic, respectively [32]. The average Ba concentrations in the shoots after addition of sugar cane filter cake or peat were as follows: 44.47 or 50.97 mg kg<sup>-1</sup>, respectively, in sunflowers; 29.68 or 30.03 mg kg<sup>-1</sup>, respectively, in castor oil plants; and 77.23 or 74.46 mg kg<sup>-1</sup>, respectively, in oilseed radish (Table 3). Similar results have been reported for the same plant species grown in Rhodic Hapludox using BaSO<sub>4</sub> additions of 0, 150, and 300 mg kg<sup>-1</sup>. The plant tissue Ba concentrations found in the present study were higher than previously reported (21.3 mg kg<sup>-1</sup> for sunflowers, 19.4 mg kg<sup>-1</sup> for mustard plants, and 10.6 mg kg<sup>-1</sup> for castor oil plants [30]).

However, Ba concentrations in this study were less than those reported by Suwa et al., 2008 [8] who observed that high Ba concentrations affected soybeans and resulted in reduced development, stomatal closing, and reduced photosynthetic activity. In contrast, Ba accumulation in maize plants grown in soil with much lower Ba concentrations (soil pH in the range of 5.1 to 5.7) has also been reported and no phytotoxic symptoms or nutritional imbalance correlations were observed [6].

In this study no effects or symptoms of phytotoxicity were found in the plants. Moreover, no nutritional imbalance was observed in the soil samples. In the presence of high Ca concentrations, such as those of the area studied, Ba can precipitate [9]. The absence of phytotoxic in this study might be explained by the high levels of available Ca (294 mmol, dm<sup>-3</sup>).

Shoot dry matter yields varied depending on the treatment and plant species (Table 4). Among the species tested, oilseed radish was the least affected by the treatments, and the peat addition promoted a higher dry matter yield in the oilseed radish roots. Sunflowers and castor oil plants showed similar results regarding shoot and root dry matter production, which were both higher when the sugar cane

TABLE 3: Barium in plant parts of the tested species according to the rate of organic material *treatment*\*.

Rate	Oilseed radish				Sunflower				Castor oil plant		
	Root	Straw (S)	Pod (P)	S + P	Root	Straw (S)	Flower (F)	S + F	Roots	Shoots	
	Mg ha <sup>-1</sup>				mg kg <sup>-1</sup>						
Sugar cane filter cake	0	111.0 a	56.6 a	19.2 a	75.8 a	76.7 a	31.1 a	18.5 a	49.56 a	39.8 b	30.1 ba
	20	109.7 a	56.0 a	21.3 a	77.3 a	76.0 a	32.2 a	15.2 b	47.4 ba	53.1 a	31.7 a
	40	105.0 a	58.5 a	20.7 a	79.2 a	75.0 a	27.8 ba	15.5 ba	43.4 bc	46.5 ba	30.9 a
	80	103.8 a	56.8 a	19.7 a	76.6 a	75.0 a	25.6 b	11.9 c	37.5 c	50.2 b	26.0 b
Average	107.4 A	57.0 A	20.2 A	77.2 A	75.7 A	29.2 A	15.3 B	44.5 B	47.4 A	29.7 A	
Peat	0	107.4 a	58.1 a	17.0 a	75.1 a	73.3 a	31.2 a	21.4 a	52.6 a	44.9 a	533.7 a
	20	108.3 a	59.4 a	17.6 a	77.0 a	76.0 a	32.0 a	21.4 a	53.4 a	44.3 a	627.9 a
	40	105.5 a	57.8 a	13.8 a	71.6 a	72.0 a	30.9 a	22.8 a	53.7 a	41.2 a	560.2 a
	80	109.7 a	58.8 a	15.4 a	74.2 a	76.7 a	25.9 b	18.3 b	44.2 b	42.7 a	618.3 a
Average	107.7 A	58.5 A	16.0 A	74.5 B	74.5 A	30.0 A	21.0 A	51.0 A	43.3 B	585.0 A	

\*Result presented are the average of 3 replicates. Means followed by the same letter are not significantly different by the Tukey's test at  $P \leq 0.05$ . Upper case letters, in columns, compare plant tissues and lower case letters, in columns, compare rate of amendments.

TABLE 4: Dry matter yield for different plant parts of the species tested according to the rate of organic material *treatment*\*.

Rate	Oilseed radish				Sunflower				Castor oil plant		
	Root	Straw (S)	Pod (P)	S + P	Roots	Straw (S)	Flower (F)	S + F	Roots	Shoots	
	Mg ha <sup>-1</sup>				mg kg <sup>-1</sup>						
Sugar cane filter cake	0	0.8 a	10.8 a	5.0 a	15.7 a	2.2 a	12.7 b	4.9 a	17.6 b	4.7 a	18.5 b
	20	0.7 a	11.7 a	6.3 a	17.9 a	2.7 a	15.9 ba	3.4 b	19.3 ba	5.8 a	19.5 ba
	40	0.8 a	12.2 a	5.6 a	17.7 a	3.2 a	18.3 a	3.9 ba	22.2 a	5.6 a	21.3 c
	80	0.7 a	12.5 a	5.7 a	18.2 a	2.4 a	15.0 ba	5.2 a	20.2 ba	5.6 a	20.7 bc
Average	0.8 B	11.8 A	5.6 A	17.4 A	2.6 A	15.5 A	4.4 A	19.8 A	5.4 A	20.0 A	
Peat	0	0.9 a	12.1 a	4.8 a	16.9 a	1.6 a	12.7 a	3.2 a	15.8 a	4.4 a	18.5 b
	20	1.0 a	11.5 a	6.0 a	17.5 a	2.4 a	14.4 a	3.1 a	17.5 a	4.8 a	19.7 ba
	40	0.9 a	12.6 a	5.0 a	17.6 a	2.0 a	13.3 a	3.3 a	16.7 a	5.1 a	18.9 b
	80	1.0 a	12.0 a	5.9 a	17.9 a	2.5 a	15.5 a	3.0 a	18.5 a	5.4 a	20.8 a
Average	0.9 A	12.0 A	5.5 A	17.5 A	2.1 B	14.0 B	3.2 B	17.1 B	4.9 B	19.5 B	

\*Results presented are the average of 3 replicates. Means followed by the same letter, are not significantly different by the Tukey's test at  $P \leq 0.05$ . Upper case letters, in columns, compare plant tissues and lower case letters, in columns, compare rate of amendments.

filter cake was used. Sugar cane filter cake has a low C/N ratio (Table 1), which may explain the trend to be a more useful source of nutrients than peat, as sugar cane is more easily decomposed than peat. Figure 1 shows that the increased organic material positively affected the shoot dry matter yield in the castor oil plants. However, despite the statistical significance of the regression models, from the agronomic or ecological point of view no marked quantitative difference in dry matter production among treatments would be enough to recommend one amendment over the other since the increase in dry matter production was, overall, discrete.

The addition of organic material to the soil affected differently Ba concentration among the three plant species (Table 3). Oilseed radish did not show a significant effect, but an increase in Ba in the castor oil plant roots was observed after the addition of sugar cane filter cake, from 39.8 mg/kg (no addition) to 50.2 mg/kg (80 Mg ha<sup>-1</sup>), which corresponded to an increase of 26%. In castor bean shoots,

Ba increased from 533.7 mg/kg in the control to 618 mg/kg in soils amended with 80 Mg ha<sup>-1</sup> peat respectively, which represented an increase of 16%. In sunflowers, the Ba concentrated in the flowers and straw+flower tissues was higher when the sugar cane filter cake was used. Moreover, the increase in the organic material rate (sugar cane filter cake and peat) resulted in a linear decrease in the Ba concentration in the flowers of the sunflower plants (Figure 2), up to 15% with sugar cake filter cake and 16% with peat addition.

When peat was used a negative correlation was observed for Ba and P in the castor oil plants ( $r = -0.55$ ) and sunflower tissues ( $r = -0.48$ ) (Table 5). The same trend was observed for K in the oilseed radish ( $r = -0.83$ ) and Ca in the castor oil plants ( $r = -0.67$ ) and sunflowers ( $r = 0.52$ ) with the use of sugar cane filter cake (Table 5). A nutritional imbalance of Ca, K, and S in the presence of Ba has been reported by several authors. These reports suggest that the imbalance is related to the plant species [7, 8, 31].

TABLE 5: Correlation between Ba and P, K and Ca concentrations in shoots.

Element	Oilseed radish		Sunflower		Castor oil plant	
	Sugar cane filter cake	Peat	Sugar cane filter cake	Peat	Sugar cane filter cake	Peat
P	0.51*	0.07NS	0.76*	-0.48NS	-0.78*	0.55*
K	-0.52*	-0.83*	0.12NS	-0.00NS	0.78*	0.42NS
Ca	-0.05NS	0.13NS	-0.52*	-0.04NS	0.73*	-0.67

\*Significant at  $P < 0.05$  and NS: not significant.

TABLE 6: Transfer factor (TF) and translocation index (TI) of Ba in the tested species.

Treatment	Mg ha <sup>-1</sup>	TF			TI (%)		
		Oilseed radish	Sunflower	Castor oil plant	Oilseed radish	Castor oil plant	Sunflower
Sugar cane filter cake	0	0.70	0.47	0.26	89.23	74.64	59.22
	20	0.70	0.46	0.32	90.63	66.71	60.92
	40	0.69	0.45	0.29	90.32	71.77	60.45
	80	0.65	0.41	0.28	90.52	65.70	60.13
Average		0.69	0.45	0.29	90.20	69.57	60.21
Peat	0	0.75	0.52	0.30	89.01	72.86	59.59
	20	0.71	0.49	0.29	88.24	74.76	59.87
	40	0.68	0.48	0.27	89.52	72.69	59.37
	80	0.71	0.47	0.28	88.10	72.98	59.82
Average		0.71	0.49	0.29	88.71	73.35	59.66

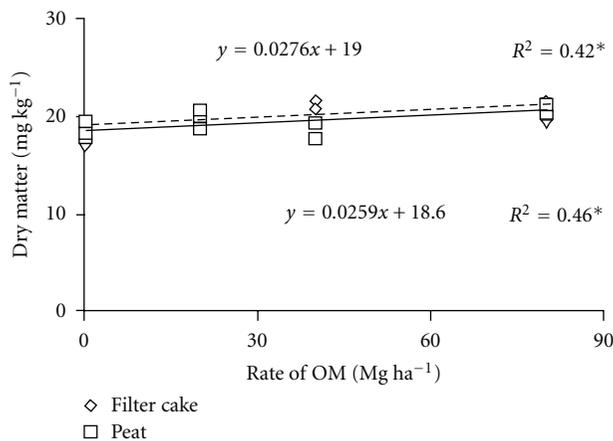


FIGURE 1: Effect of increasing concentrations of organic materials on shoot dry matter yield in castor oil plants shoots (d.w.). Significant at  $P < 0.05$ .

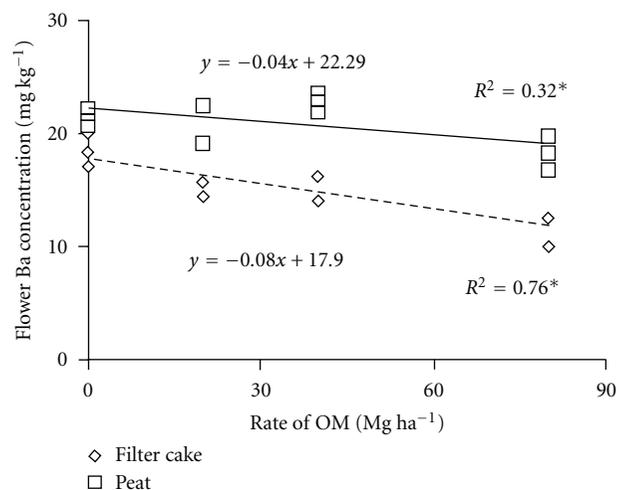


FIGURE 2: Effect of increasing concentrations of organic materials on Ba concentration in the flowers of sunflowers (d.w.). Significant at  $P < 0.05$ .

The transfer index (calculated as the Ba shoot concentration divided by the total Ba in the soil) decreased as follows: oilseed radish (0.70) > sunflowers (0.47) > castor oil plants (0.29). The average Ba transfer from the roots to the shoots in the oilseed radish, castor oil plants, and sunflowers was found to be 89%, 71%, and 59%, respectively. These values indicated that the Ba was highly mobile in the xylem of the oilseed radish and castor oil plants. From the total Ba transfer values, at least 50% of the Ba shoot concentration

was found in the flowers of the sunflowers, and 35% of the Ba shoot concentration was found in the pods of the oilseed radish (Table 3). Ba concentrations in flowers with the addition of sugar cane filter cake and peat were as follows: 20.2 and 16.0 mg kg<sup>-1</sup>, respectively, for oilseed radish; 15.3 and 21.0 mg kg<sup>-1</sup>, respectively, for sunflowers. A high Ba mobility has also been observed in cotton and white beets

during development and flowering, whereas a large amount of Ba accumulation has been reported in the leaves of corn ( $105.7 \text{ mg kg}^{-1}$ ) when compared with the grains of corn ( $1.05 \text{ mg kg}^{-1}$ ) [6, 33].

The obtained transfer factors (lower than 1) suggested that the tested species were inadequate in accumulating or extracting Ba from soil (Table 6). Similarly, low transfer factors have also been reported for castor oil plants, sunflowers, and mustard plants [30]. Indeed, these authors [30] reported that none of the plants grown in soils containing Ba ranging from  $132.3$  to  $1,130 \text{ mg kg}^{-1}$  were able to accumulate measurable concentrations of Ba, thus, highlighting the low transfer of this element from soil to plants [34].

A decreasing trend for Ba transfer to oilseed radish and sunflowers was found when the sugar cane filter cake concentration increased (Table 6). In addition to improving the physical and chemical conditions of the soil, organic ligands are promising in the mitigation of heavy metal contaminated soils. Peat and a concentrate containing humic substances from coal favor mustard development in a contaminated soil due to the mitigation of Zn, Cu, Mn, Pb, and B by the organic ligands [14].

## 4. Conclusions

Under the conditions studied the elevated soil pH reached due to liming overcame the organic matter addition effect and determined the barium availability and its absorption by the plant species grown in the area polluted with scrap metal residue. This is suggested by the absence of phytotoxic effects on plants, the moderate Ba accumulation in shoots compared to the usual content of Ba in plants, the small effect of organic matter treatments on plants dry matter yields, and finally the levels of Ba extracted by Mehlich-3.

## Acknowledgment

The authors would like to thank Fundação de Apoio a Pesquisa do Estado de São Paulo (FAPESP) for the financial support with the 2006/60987-0 Research Grant.

## References

- [1] Cetesb, "Relatório de Totalização de áreas contaminadas e reabilitadas-Dezembro de 2010," 2011, <http://www.cetesb.sp.gov.br/areas-contaminadas/relacoes-de-areas-contaminadas/15-publicacoes>.
- [2] J. A. Ippolito and K. A. Barbarick, "Biosolids affect soil barium in a dryland wheat agroecosystem," *Journal of Environmental Quality*, vol. 35, no. 6, pp. 2333–2341, 2006.
- [3] L. C. S. Merlino, W. Melo, F. G. Macedo et al., "Barium, cadmium, chromium and lead in maize plants and in an oxisol after eleven years of sewage sludge applications," *Revista Brasileira de Ciência do Solo*, vol. 34, pp. 2031–2039, 2010.
- [4] E. Simon, M. Braun, A. Vidic, D. Bogoy, I. Fabian, and B. Tothneresz, "Air pollution assessment ion elemental concentration of leaves tissue and foliage dust along an urbanization gradient in Vienna," *Environmental Pollution*, vol. 159, pp. 1229–1233, 2011.
- [5] E. Nogaj, J. Kwapulinski, and H. Misiolek, "Pharyngeal Tonsil as new biomarker of pollution on example of barium," *Polish Journal of Environmental Studies*, vol. 20, pp. 161–172, 2011.
- [6] T. A. R. Nogueira, W. J. deMelo, I. M. Fonseca, M. O. Marques, and Z. He, "Barium uptake by maize plants as affected by sewage sludge in a long-term field study," *Journal of Hazardous Materials*, vol. 181, no. 1-3, pp. 1148–1157, 2010.
- [7] M. Llugany, C. Poschenrieder, and J. Barceló, "Assessment of barium toxicity in bush beans," *Archives of Environmental Contamination and Toxicology*, vol. 39, no. 4, pp. 440–444, 2000.
- [8] R. Suwa, K. Jayachandran, N. T. Nguyen, A. Boulouvar, K. Fujita, and H. Saneoka, "Barium toxicity effects in soybean plants," *Archives of Environmental Contamination and Toxicology*, vol. 55, no. 3, pp. 397–403, 2008.
- [9] C. A. Menzie, B. Southworth, G. Stephenson, and N. Feisthauer, "The importance of understanding the chemical form of a metal in the environment: the case of barium sulfate (barite)," *Human and Ecological Risk Assessment*, vol. 14, no. 5, pp. 974–991, 2008.
- [10] F. Baldi, M. Pepi, D. Burrini, G. Kniewald, D. Scali, and E. Lanciotti, "Dissolution of barium from barite in sewage sludges and cultures of *Desulfovibrio desulfuricans*," *Applied and Environmental Microbiology*, vol. 62, no. 7, pp. 2398–2404, 1996.
- [11] A. A. Carbonell, R. Pulido, R. D. DeLaune, and W. H. Patrick, "Soluble barium in barite and phosphogypsum amended Mississippi River alluvial sediment," *Journal of Environmental Quality*, vol. 28, no. 1, pp. 316–321, 1999.
- [12] G. A. Ulrich, G. N. Breit, I. M. Cozzarelli, and J. M. Suffita, "Sources of sulfate supporting anaerobic metabolism in a contaminated aquifer," *Environmental Science and Technology*, vol. 37, no. 6, pp. 1093–1099, 2003.
- [13] C. M. Davidson, M. D. Gibson, E. Hamilton, B. H. MacGillivray, J. Reglinski, and E. Rezabal, "The long-term environmental behaviour of strontium and barium released from former mine workings in the granites of the Sunart region of Scotland, UK," *Chemosphere*, vol. 58, no. 6, pp. 793–798, 2005.
- [14] G. C. G. Dos Santos and A. A. Rodella, "Effect of sources of organic matter in the alleviation of the toxic effects of B, Zn, Cu, Mn and Pb to Brassica Juncea," *Revista Brasileira de Ciência do Solo*, vol. 31, no. 4, pp. 793–804, 2007.
- [15] J. C. Corrêa, L. T. Büll, W. D. S. Paganini, and I. A. Guerrini, "Heavy metal exchangeable in an Oxisol with surface application of flue dust, aqueous lime, sewage sludge and limestone," *Pesquisa Agropecuária Brasileira*, vol. 43, no. 3, pp. 411–419, 2008.
- [16] E. E. C. de Melo, C. W. A. do Nascimento, A. C. Q. Santos, and A. S. da Silva, "Availability and fractionation of Cd, Pb, Cu, and Zn in soil as a function of incubation time and pH," *Ciência e Agrotecnologia*, vol. 32, no. 3, pp. 776–784, 2008.
- [17] O. C. Bataglia, A. M. C. Furlani, J. P. F. Teixeira, and J. R. Gallo, *Métodos de Análise Química de Plantas*, Boletim Técnico, 78, Instituto Agrônomo, Campinas, Brazil, 1983.
- [18] United States Environmental Protection Agency, "Method 3051: microwave assisted acid digestion of sediments, sludges, soil and soils," 2009, [http://www.epa.gov/epaoswer/hazwaste/test/3\\_series.htm](http://www.epa.gov/epaoswer/hazwaste/test/3_series.htm).
- [19] Empresa Brasileira de Pesquisa Agropecuária, *Brazilian System of Soil Classification*, vol. 306, Embrapa-Centro Nacional de Pesquisa de Solos, Rio de Janeiro, Brazil, 2nd edition, 2006.
- [20] B. van Raij, J. C. Andrade, H. Cantarella, and J. A. Quaggio, *Análise Química Para Avaliação da Fertilidade de Solos Tropicais*, Instituto Agrônomo, Campinas, Brazil, 2001.

- [21] C. De Freitas Zeitouni, R. S. Berton, and C. A. De Abreu, "Phytoextraction of cadmium and zinc from an oxisol contaminated with heavy metals," *Bragantia*, vol. 66, no. 4, pp. 649–657, 2007.
- [22] A. M. M. Pires, C. A. Abreu, A. R. Coscione, V. A. Silva, and N. P. Ramos, "Initial growth of sunflower in soils with high concentrations of boron and heavy metals," in *Proceedings of the 17th International Sunflower Conference*, vol. 2, pp. 315–318, Córdoba, Spain, 2008.
- [23] G. C. G. dos Santos, A. A. Rodella, C. A. de Abreu, and A. R. Coscione, "Vegetable species for phytoextraction of boron, copper, lead, manganese and zinc from contaminated soil," *Scientia Agricola*, vol. 67, no. 6, pp. 713–719, 2010.
- [24] R. A. B. Jorge, C. A. de Abreu, C. A. de Andrade, and O. A. de Camargo, "Filter cake and peat as amendments of contaminated soil with residue of scrap rich in boron," *Bragantia*, vol. 69, no. 2, pp. 467–476, 2010.
- [25] A. Mehlich, "Mehlich 3 soil test extractant: a modification of Mehlich 2 extractant," *Communications in Soil Science and Plant Analysis*, vol. 15, no. 12, pp. 1409–1416, 1984.
- [26] A. D. Abichequer and H. Bohnen, "Efficiency of phosphorus uptake, translocation and utilization in wheat varieties," *Revista Brasileira de Ciência do Solo*, vol. 22, pp. 21–26, 1998.
- [27] S. Lubben and D. Sauerbeck, "The uptake and distribution of heavy metals by spring wheat," *Water, Air, and Soil Pollution*, vol. 57-58, pp. 239–247, 1991.
- [28] Companhia de Tecnologia de Saneamento Ambiental, "Guiding values for soils and groundwater in the State of Sao Paulo," 2011, [http://www.cetesb.sp.gov.br/Solo/relatorios/tabela\\_valores.2005.pdf](http://www.cetesb.sp.gov.br/Solo/relatorios/tabela_valores.2005.pdf).
- [29] M. B. Gabos, G. Casagrande, C. A. Abreu, and J. Paz-Ferreiro, "Uso da matéria orgânica como mitigadora de solo multicontaminado e do girassol como fitoextratora," *Revista Brasileira de Engenharia Agrícola e Ambiental*, vol. 15, no. 12, pp. 1298–1306, 2011.
- [30] A. R. Coscione and R. S. Berton, "Barium extraction potential by mustard, sunflower and castor bean," *Scientia Agricola*, vol. 66, no. 1, pp. 59–63, 2009.
- [31] F. M. Chaudhry, A. Wallace, and R. T. Mueller, "Barium toxicity in plants," *Communications in Soil Science and Plant Analysis*, vol. 8, no. 9, pp. 795–797, 1977.
- [32] I. Pais and J. R. Jones, *The Handbook of Trace Elements*, St. Lucie Press, Boca Ratón, Fla, USA, 1998.
- [33] Z. En, A. Vasidov, V. V. Tsipin, T. Tillaev, and G. I. Jumaniyazova, "Study of element uptake in plants from the soil to assess environmental contamination by toxic elements," *Nuclear Instruments and Methods in Physics Research A*, vol. 505, no. 1-2, pp. 462–465, 2003.
- [34] J. Pichtel, K. Kuroiwa, and H. T. Sawyerr, "Distribution of Pb, Cd and Ba in soils and plants of two contaminated sites," *Environmental Pollution*, vol. 110, no. 1, pp. 171–178, 2000.



**Hindawi**

Submit your manuscripts at  
<http://www.hindawi.com>

