

Review Article

Biomaterials in Guided Bone and Tissue Regenerations: An Update

Reza Abdollahi Namanloo ¹, **Maedeh Ommani** ², **Kamyar Abbasi**,³ **Mostafa Alam**,⁴
Ashkan Badkoobeh,⁴ **Mahdi Rahbar** ⁵, **Hadi Kokabi Arasteh** ⁶,
Emran Hajmohammadi ⁷, **Reza Sayyad Soufdoost** ⁸, and **Seyed Ali Mosaddad** ⁹

¹Dentistry Department, Bogomolets National Medical University, Kyiv, Ukraine

²Student Research Committee, Department of Pediatric Dentistry, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran

³Department of Prosthodontics, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁴Department of Oral and Maxillofacial Surgery, School of Dentistry, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁵Department of Restorative Dentistry, School of Dentistry, Ardabil University of Medical Sciences, Ardabil, Iran

⁶Department of Periodontics, School of Dentistry, Hamadan University of Medical Sciences, Hamadan, Iran

⁷Department of Oral and Maxillofacial Surgery, School of Dentistry, Ardabil University of Medical Sciences, Ardabil, Iran

⁸Dentistry Research Institute, School of Dentistry, Shahed University, Tehran, Iran

⁹Student Research Committee, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran

Correspondence should be addressed to Seyed Ali Mosaddad; sepehr.mosaddad@gmail.com

Received 4 March 2022; Revised 1 April 2022; Accepted 4 April 2022; Published 5 May 2022

Academic Editor: Qian Chen

Copyright © 2022 Reza Abdollahi Namanloo et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Purpose. Guided tissue reconstruction can be performed to restore the supporting structure of a previously lost tooth, which, in addition to maintaining beauty, preserves the function of the tooth in the patient. *Materials and Methods.* In this review, Scopus, PubMed, and MEDLINE databases were searched using the keywords “biocompatible materials,” “membrane,” “bone regeneration,” “tissue reconstruction,” and “dental biomaterials.” Overall, 150 articles were reviewed, and finally, 107 articles published during 2000–2021 were included in the final paper. *Results.* Studies have been conducted on a variety of membranes in both clinical and experimental settings. The first half of this article explores the different kinds of membranes and diverse classes of biomaterials used in these procedures. Secondly, biomaterials are examined for their therapeutic uses such as growth factors, stem cells, and gene delivery vehicles. *Conclusion.* If a tooth has been extracted or if the gums have been infected with periodontal disease, guided bone regeneration procedures may be used to restore the lost bone. Recent years have seen a variety of approaches to regenerating these tissues. To prevent nonossifying cells from entering, membranes are heavily employed during guided rebuilding.

1. Introduction

Today, one of the most critical topics in periodontology is the repair of periodontal lesions and the reconstruction of lost jawbone through proliferating bone cells. A fundamental prerequisite for successful implant treatments is the adequate volume of hard bone tissue [1]. On the other hand, for the successful repair of damaged tissue, it is necessary to prevent the invasion of cells with high-speed migration to the damaged tissue and guide sedentary cells to the lesion site.

In recent years, tissue engineering using biocompatible polymers has introduced new methods for repairing damaged tissues [2]. Guided bone regeneration (GBR) and guided tissue regeneration (GTR) are the most important conventional reconstruction methods. The GBR is used to regenerate alveolar bone in toothless areas, and GTR is utilized to repair damaged periodontal tissue [3, 4]. In general, GTR and GBR are surgical techniques that use a porous polymer membrane to physically prevent the migration of undesirable tissues and cells to the lesion site [4]. As a result, a suitable space and substrate are provided for

repairing damaged tissue by proliferating the cells in question [5]. The GBR can maintain and strengthen the alveolar bulge, regenerate alveolar bone, correct contractions around the implant or fenestration, and regenerate the bone around the implant [6]. In contrast, GTR refers to periodontal ligament (PDL) regeneration, bone regeneration, and cementum around the tooth [7].

Although porous membranes in the GBR technique seem necessary, repairing and growing damaged bone require osteogenic cells, as well as osteoconductive and bone-inducing (osteoinductive) materials [8]. The ideal membrane used in guided bone tissue repair should have unique properties, such as good biocompatibility and functional stability over the required time. In addition, the membrane must maintain the space and biomechanical stability of the lesion area by filtering out the disturbing cells and tissues and protecting the newly formed tissue [9].

Generally, membranes used in GTR and GBR techniques may fall into the following categories: (1) bioabsorbable based on natural polymers, including collagen, chitosan, and gelatin, or synthetic polymers; (2) nonresorbable membranes, including expanded polytetrafluoroethylene, titanium-reinforced e-PTFE, and dense-PTFE [10]; and (3) metals and inorganic compounds considered as another group of guided membranes, which will be discussed in detail.

2. Materials and Methods

Based on the institutional regulations, this research was granted an exemption regarding approval since it was a literature review. A comprehensive search of the Scopus, PubMed, and MEDLINE databases was performed. All relevant articles published during 2000–2021 were obtained using the keywords “biocompatible materials,” “membrane,” “bone regeneration,” “tissue reconstruction,” and “dental biomaterials.” Afterwards, articles were reviewed by titles and abstracts. The papers that were less relevant to the subject of study were excluded. The remaining full-text articles were evaluated, and those unrelated to the subject were removed. The filtered papers were further analyzed by the team of authors, and this review was structured.

3. Bioabsorbable

A significant advantage of this type of membrane is that it does not have to be removed by secondary surgery, that it has a better repair and healing ability, that it is biocompatible, and that it reduces the risk of inflammation and infection [11]. However, there are several limitations to these structures, the most important of which is the low ability to make a suitable space for new tissue growth and the high destruction rate. These structures are destroyed in the body much faster than expected and leave the damaged area before completing the tissue formation process [12, 13]. The two kinds of adsorbable membranes consist of natural polymers and synthetic polymers [14].

3.1. Natural Adsorbable Membranes. The application of natural polymers has expanded due to their properties in the GTR and GBR processes. In other words, inherent bioactivity and the ability to provide active sites for cell attachment are among the most important advantages of natural polymers over synthetic polymers [15]. However, some problems related to the intrinsic bioactivity of polymers, including strong immune responses, complications associated with the purification of these polymers, and the possibility of disease transmission, restrict the use of these polymers [16]. Table 1 summarizes the disadvantages and advantages of various types of membranes.

3.1.1. Collagen Membranes. Collagen is the hardest filament in the connective tissue. This membrane has significant advantages, such as good tissue integrity, fast vascularization, biodegradability without external reactions, weak immunogenicity, hemostatic property, biocompatibility, and wound healing capacity [42]. In addition, collagen is a chemotactic factor for fibroblasts and accelerates cell migration. Collagen membranes may secondarily increase tissue thickness as a result of enzymatic degradation and replacement by surrounding connective tissue [43]. Such properties have led to collagen-based membranes in GTR and GBR research. This type of membrane is degradable and adsorbable by enzymatic degradation, carried out by collagenases and proteases [44, 45]. Porcine collagen fiber types I and II are incorporated into Bio-Gide, one of the most commonly used commercial collagen membranes. Soft tissue invasion and development into a defect may be limited by the smooth surface of Bio-Gide, which may also act as a scaffold for the attachment of fibroblasts [46–50]. The rough, porous side of Bio-Gide functions as a framework for the migration and proliferation of blood vessels and bone cells [43, 46].

Disadvantages of collagen membranes have rapid biodegradability and reduced membrane capability in maintaining the space and biomechanical stability of the lesion area in wet conditions. The destruction time of these membranes is about 4–8 weeks, which is not enough for the full regeneration of bone tissue [51, 52]. To improve the mechanical properties and decrease the rate of deterioration of collagen membranes, several approaches have been investigated [42, 53]. There are a number of possibilities for treatment with UV light or chemical therapies with genipin (Gp) [54, 55].

Moreover, various physical, chemical, and biological crosslinking methods effectively strengthen the mechanical characteristics and augment the stability of collagen membranes against biodegradability. The most common chemical crosslinking agents are glutaraldehyde, 3-dimethyl-amino-propyl, carbodiimide, and polyoxy [45, 55]. Crosslinking is accompanied by pros and cons. Its advantage is improving the tensile strength of collagen and delaying the breakdown time [56]. Drawbacks entail limiting the use of membranes by creating potential toxic effects by the remaining crosslinking agent or the formation of byproducts during collagen degradation, which can lead to severe inflammation at the

TABLE 1: Advantages and disadvantages of various membranes.

Membrane type	Membrane materials	Advantage	Disadvantage	References
Natural polymers	Collagen, gelatin, chitosan	<ul style="list-style-type: none"> • High biocompatibility • Improved cellular Interaction • Hydrophilicity • Antibacterial effect • Cell/drug containing • Enhancement of wound healing • No surgical removal 	<ul style="list-style-type: none"> • Lack of bioactivity • Rapid degradation rate • Low mechanical strength • Residual crosslinking agents • Possible disease transmission • Questionable barrier function • Hard to control biodegradation • Few studies 	[17–27, 51, 52, 57, 60]
Synthetic polymers	PLA/PLGA, PCL, PEG	<ul style="list-style-type: none"> • Mechanical strength can be processed • Able to seed mesenchymal cells/growth factors • Favorable biocompatibility • Favorable barrier function • High reproducibility • Manageable biodegradability and mechanical properties • No surgical removal needed 	<ul style="list-style-type: none"> • Slow degradation rate • Hydrophobic (PCL) • Low cell affinity • Poor cellular response • Not suitable for a drug delivery system • Acidic byproducts 	[28–34]
Nonresorbable	e-PTFE, d-PTFE, TR-ePTFE	<ul style="list-style-type: none"> • Many studies demonstrate their success • Could be titanium-reinforced • Remain intact until removal • Easily fixed with titanium or resorbable tacks • Greater bone fill if the membrane is not exposed • Minimal tissue response if the membrane is not exposed • High chemical stability • High biocompatibility • High barrier function 	<ul style="list-style-type: none"> • Require a second surgery for removal • Increase patient morbidity • If exposed, must be removed in most cases • Could be technique sensitive • Membrane exposure 	[12, 35–38, 87]
Metals	Titanium, titanium alloy	<ul style="list-style-type: none"> • High biocompatibility • High barrier function • Mechanical strength, durability 	<ul style="list-style-type: none"> • Surgical removal required • Expensive 	[39–41, 91]

implant site [51, 57]. Crosslinking, in addition to reducing the rate of degradation, greatly diminishes tissue integration and angiogenesis and also affects the biocompatibility of the final structure. Increasing the percentage of crosslinker decreases the adhesion and proliferation of PDL fibroblasts and osteoblasts [58, 59]. Therefore, the success of the operation by collagen membrane, in addition to the source of the collagen, depends on the preparation and processing stages, including decellularization, sterilization, and crosslinking methods. The Gp and D-ribose, as two natural safe and nontoxic compounds, have been proposed to raise the mechanical strength of collagen and diminish its degradation rate [51, 60, 61].

3.1.2. Gelatin-Based Membrane. Gelatin is a protein that is soluble and is made from collagen. Improved cell adhesion, suitable biocompatibility, reasonable price, and flexibility of this protein have made it a favorable biomaterial for tissue engineering, GTR, and GBR [62, 63]. Protein membranes have poor mechanical properties, and they degrade rapidly. A method for improving stability and mechanical properties is crosslinking glutaraldehyde and N-hydroxyl succinamide using heat treatment [64]. However, crosslinking can reduce the modulus of elasticity under humid conditions, even though it improves the tensile properties of gelatin fiber membranes. Because of this, gelatin can sometimes be used alone in GTR and GBR [65, 66].

3.1.3. Chitosan-Based Membranes. Studies showed that chitosan is well adapted to cells *in vitro* and facilitates bone regeneration at the site of rat skull lesions [67]. Chitosan is also a suitable membrane for GTR and GBR because of its reasonable price, high biocompatibility, good degradation rate, antibacterial properties, wound healing potential, and flexibility in humid environments. Chitosan membranes deteriorate at different rates depending on their molecular weight and manufacture method [68]. One of the ways to boost mechanical strength and reduce the rate of chitosan degradation is chemical crosslinking. Gp-crosslinked chitosan membranes produce fewer inflammatory reactions than glutaraldehyde-crosslinked membranes, accelerating lesion healing [69]. Chitosan-based electrospinning membranes crosslinked with Gp have a much lower degradation rate than nonlattice membranes. Due to the high cost of Gp and the toxicity of glutaraldehyde, ionic crosslinking with sodium tripolyphosphate has been proposed as an alternative for crosslinking [70].

3.2. Synthetic Adsorbable Membranes

3.2.1. PLA and PLGA. The PLA is among the most important and common synthetic polymers applied in the GTR and GBR processes. This polymer has high mechanical and biocompatible properties. To control the hydrophilicity of PLA and its degradation rate, its copolymers are synthesized based on lactide, caprolactone, and glycolide [71]. Polyglycolic acid (PLGA), in orthopedic applications, is an

excellent alternative to PLA. Despite the biodegradability and nontoxicity of PLA and PLGA-based membranes, there are limitations to using these membranes. One of these problems is the possibility of inflammatory reactions and reactions to an external object *in vivo* when oligomers and acidic byproducts are released during degradation [72]. Their hardness is another problem that limits the medical applications of these membranes. One effective solution for the hardness is to add emollients, such as N-methyl-2-pyrrolidone (NMP). The NMP softens PLGA membranes, thereby accelerating bone regeneration and osteoblast growth [73].

3.2.2. Poly-Caprolactone (PCL). A low-cost polymer is also biocompatible and possesses excellent mechanical strength. Therefore, it has received much attention in bone tissue engineering. There are still few studies on PCL-based GTR membranes. One of the advantages of this polymer over PLA and PLGA polymers is that PCL degradation does not increase environment acidity [74]. The complete bioabsorption of the PCL membrane in the body takes about 3 years, which is long for use in the GBR and GTR methods. On the other hand, the hydrophobicity of its pure membranes reduces adhesion and cell proliferation. Consequently, it is always combined with other polymers or as a copolymer for medical applications [75, 76].

3.2.3. Polyethylene Glycol (PEG). This polymer is remarkably biodegradable and biocompatible, making it a suitable option for use in GTR and GBR membranes [77]. The main positive points of this polymer membrane are regulated biodegradation, manageability, processability, and the ability to encapsulate the medicine [75, 78]. In contrast, poor instability and high degradation rates produce a robust inflammatory response that affects the outcome of bone regeneration and leads to regenerated bone resorption [79, 80].

4. Nonresorbable

The first used membranes were nonresorbable and able to keep the bone lesion separate from other tissue cells for a long time. In these membranes, reoperation is often required to remove the membrane. The repair will not occur spontaneously if these membranes are exposed. In addition, there is a possibility of bacterial contamination of the exposed membrane resulting in infection [81].

4.1. Expanded Polytetrafluoroethylene (e-PTFE). The e-PTFE membranes are the first synthetic polymer used for GBR with cavities of 0.02–20 μm and are an excellent candidate to cover the defect opening around the implant and repair or maintain the bone around the implant in the area [82–85]. Due to its high chemical stability, this membrane is biologically considered the most stable polymer, which resists degradation caused by host tissues and does not lead to immunological reactions [80]. They also prevent bone and

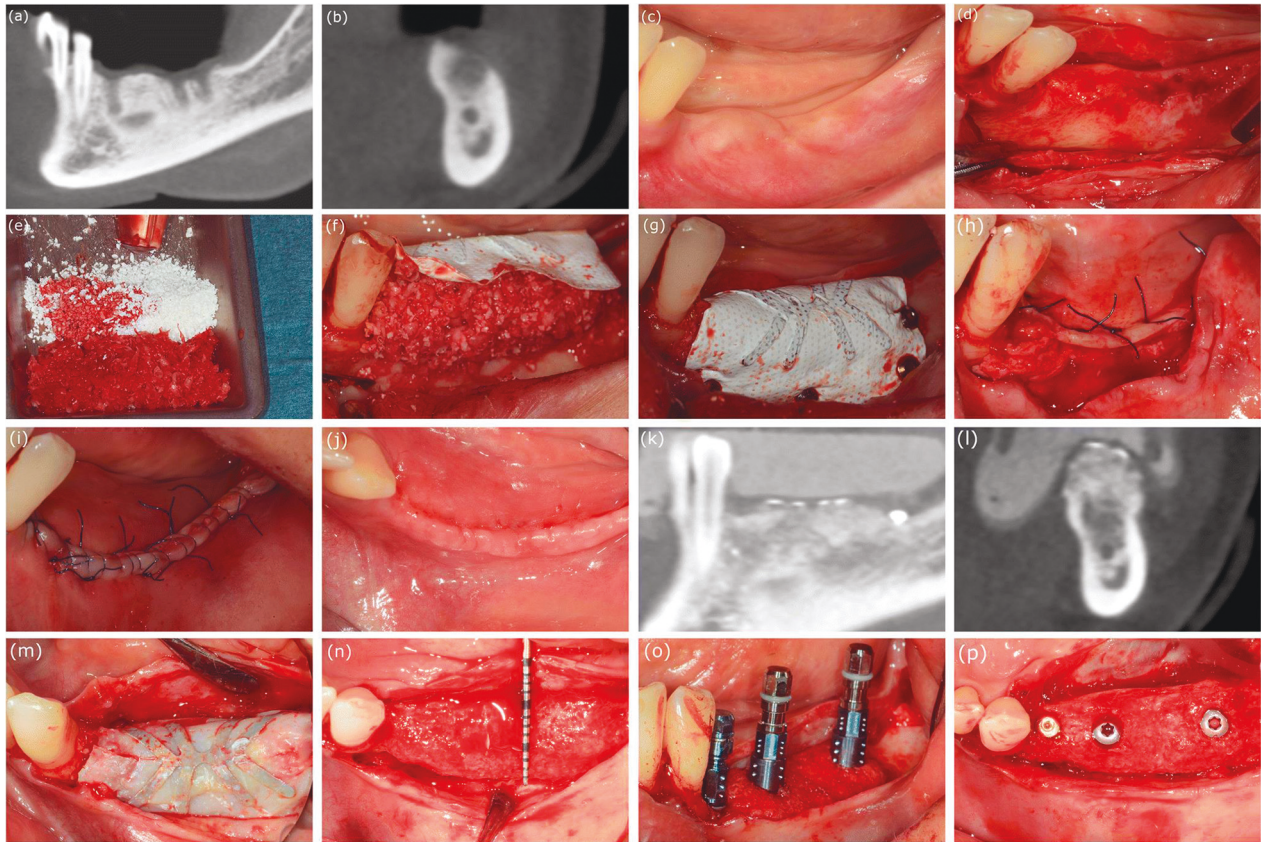


FIGURE 1: Bone augmentation with a titanium-reinforced d-PTFE membrane. (a) Primary CBCT scan, parasagittal section showing the horizontal and vertical bone defect. (b) Primary CBCT scan, frontal section showing the horizontal and vertical bone defect. (c) Clinical photograph of the edentulous mandible. (d) Flap preparation with the split-thickness method. (e) Bovine-derived xenograft (Geistlich Bio-Oss) and particulate autogenous bone graft. (f) Adaptation of nonresorbable titanium-reinforced d-PTFE membrane (Osteogenics Cytoplast) over the graft. (g) Fixing the membrane with titanium pins. (h) Suturing. (i) Closure of the mucosal layer with horizontal mattress and noninterrupted sutures. (j) Healing two weeks post-op. (k) Postoperative CBCT scan after nine months: parasagittal section. (l) Postoperative CBCT scan after nine months: frontal section. (m) Reentry for removing the membrane. (n) Good quantity of bone tissue for implant placement. (o) Guided implant surgery (Straumann bone level). (p) Final positioning of implants through a prosthetically driven technique [92].

other connective tissue cells from migrating to the lesion [86]. As a result, bone marrow cells, which migrate less rapidly, are more likely to be present at the lesion site. Despite the expansion of tetrafluoroethylene membranes, osteogenesis without interfering with other tissues leads to repairing damaged bone tissue in an average period of three months. In contrast, osteogenesis occurs incompletely in control samples without membrane because of connective tissue interference [87].

4.2. Dense Polytetrafluoroethylene (D-PTFE). The porosity of the e-PTFE membrane can allow germs to migrate. This problem was addressed through the creation of d-PTFE membranes with pores smaller than 0.3 μ m. Clinical studies have found that d-PTFE does not generate bacterial colonies as rapidly as e-PTFE [88, 89]. The d-PTFE membranes used in the mouth cavity may prevent bacteria from passing through, but they do allow oxygen to enter and tiny molecules to pass through. On the other hand, e-PTFE membranes have larger holes and can interact more strongly with

soft tissue. This results in membrane separation being significantly more challenging and requiring deeper incisions following hard tissue regeneration. As there is no development or attachment to soft tissues, d-PTFE membrane separation is straightforward [90].

4.3. Titanium-Reinforced Expanded Polytetrafluoroethylene (TR-ePTFE). Flexible titanium mesh-reinforced polytetrafluoroethylene membranes allow the membrane to be ductile to fit the shape and size of the defect in the desired area and provide adequate structural stability in bone defects around the implant [91]. Figure 1 demonstrates an implant surgical procedure using a titanium-reinforced d-PTFE membrane [92]. Clinical and experimental studies on nonabsorbable membranes have shown good therapeutic results in GTR and GBR applications. However, one of the most important disadvantages of these membranes is the need for secondary surgery to remove them after bone growth. In addition to raising the cost to patients, this sometimes leads to the loss of a part of the regenerated tissue. Stiffness and rigidity are

other weaknesses of nonabsorbable membranes that screws are used in most cases to make them more stable [56].

4.4. Metals. A frequent medical material used in maxillofacial surgery and orthopedics, titanium, is highly biocompatible, strong, corrosion-resistant, dense, and light [93]. In addition to bone grafts, titanium mesh has been shown to improve the localized deficiency of the alveolar ridge prior to or after implant placement [94, 95]. Studies have shown that titanium causes less lasting inflammation compared with PTFE [96]. When considering chronic inflammation connected to titanium micro/nanoparticles, there is a well-established mechanism by which metal nanoparticles cause inflammation via their immunomodulatory properties, which mostly act at the macrophage level [97, 98]. A number of factors increase with increasing oxidative stress, including DNA damage, protein carbonylation, lipid peroxidation, and superoxide dismutase activity, while catalase and total glutathione are all declining. In addition, they result in macrophage activation that is aberrant, which is associated with an increase in inflammation and a decrease in innate immunity [99, 100]. Ti nanoparticles were investigated *in vitro* to determine their effects on MSC physiology, ROS generation, and phenotyping of osteogenic and adipogenic cells. Activating PKC beta, which is implicated in MSC commitment to adipocyte lineage, also causes ROS production, and metal particles are not degradable, so ROS production causes abnormal neutrophil recruitment. The bone regeneration function of VEGF is reduced by a genetic abnormality. This imbalance affects the osteogenic commitment [101]. Another used metal is cobalt-chromium (CoCr), which has suitable mechanical properties, but less biocompatibility than titanium. One study found that placing a CoCr membrane on the tibial defect of a rabbit created enough space for bone regeneration [102].

5. Inorganic Components

Calcium sulfate (CaS) is a substance used to form significant GBR membranes because it is osteoconductive, biocompatible, and bioresorbable [103, 104]. It is possible to make solid substances with relatively stable and less absorbable crystals by hydrating CaS hemihydrate powder [105, 106]. It is a calcium phosphate of the hydroxyapatite type. HA is frequently used in bone applications due to its resemblance to bone minerals and biocompatibility. The biocompatibility and osteoconductivity of HA make it a common material used in bone treatments. Membranes with strong mechanical properties can withstand soft tissue static pressure while allowing bone to regenerate more readily [107]. When combined with nonabsorbable and absorbable membranes, membranes that are mixed with HA have been shown to enhance osteoblast-like cell activity *in vitro* [108–113].

6. Biomaterial-Based Delivery

6.1. Growth Factor Delivery. Signal molecules such as growth factors control how cells grow and develop. Several cell functions are affected by growth factors, including

proliferation, migration, and extracellular matrix (ECM) formation. Some of them play a role in cell differentiation [114]. Growth factors include VEGFs, FGFs, PTHs, PDGFs, and IGFs. When cells begin to repair themselves, PDGF is released, which is chemotactic and mitogenic. Angiogenesis is induced by FGF, and cells such as fibroblasts, periodontal ligaments, osteoblasts, and endothelium are proliferated and differentiated by this growth factor [115]. Similarly, FGF signaling contributes to the development of the cranial skeleton [116]. Figure 2 illustrates a method developed to deliver growth factors to scaffolds [117]. The combined use of PDGF and grafts results in bone regeneration of up to 5 mm, whereas grafts alone result in bone regeneration of about 1–2 mm. Therefore, it is expected that growth factors will enhance the results of grafting materials [114]. To maintain skeletal health and growth, IGFs are necessary [118]. The mesenchyme also plays a role in vascularization, proliferation, and bone formation in the skull and maxilla [119]. Periodontal lesions are treated with plasma-rich plasma (PRP) derived from one's own centrifuged blood. This can be used alone or with grafting materials. Although there is insufficient evidence to suggest a relation between PRP and maxillary sinus floor elevation, it appears to have benefits in treating periodontal lesions [120, 121]. The PDGFs are known to contribute to bone repair, wound healing, and regeneration following trauma or infection by stimulating osteoblastic progenitor cells to multiply [122]. There is histological evidence that the enamel matrix derivative (EMD), Emdogain, can regenerate periodontal lesions due to its high amelogenin content and small amounts of enamel and other proteins. Additionally, angular periodontal lesions can be treated with it [123, 124]. In addition, EMD contains growth factors that stimulate the production of growth factors, such as bone morphogenetic proteins (BMPs), and enhance angiogenesis. In addition, EMD contains growth factors that encourage angiogenesis and enhance bone morphogenetic proteins (BMPs). In several studies, the combination of demineralized freeze-dried bone allograft (DFDBA) with EMD has been studied and found to be successful in regenerating periodontal lesions [125, 126].

Bone matrix proteins, or BMPs, play an important role in bone development at all stages. In addition to their contribution to the formation of the neural crest, rhBMP-7 and rhBMP-2 are also involved in the development of the teeth, lips, palates, and facial primordia, as well as creating soft and hard calluses [116, 127]. In the process of creating it, orthopedics, periodontics, and dentistry were considered. By interacting with the ECM, growth factors are stabilized and maintained *in vivo*. To achieve steady and local release of one or several GFs, selecting the right biomaterial delivery method is crucial [128]. To capture GFs physically or chemically, sponges, micro/nanoparticles, nanofiber membranes, and hydrogels are being used as delivery vehicles [129]. In addition to maintaining release kinetics, the GFs in the biomaterial provide a porous scaffold that facilitates internal bone growth [118, 130]. The following examples demonstrate two effective strategies. According to Jung, recombinant BMP-2 has the potential to enhance and expedite gingival resorption in humans, as well as for a wide

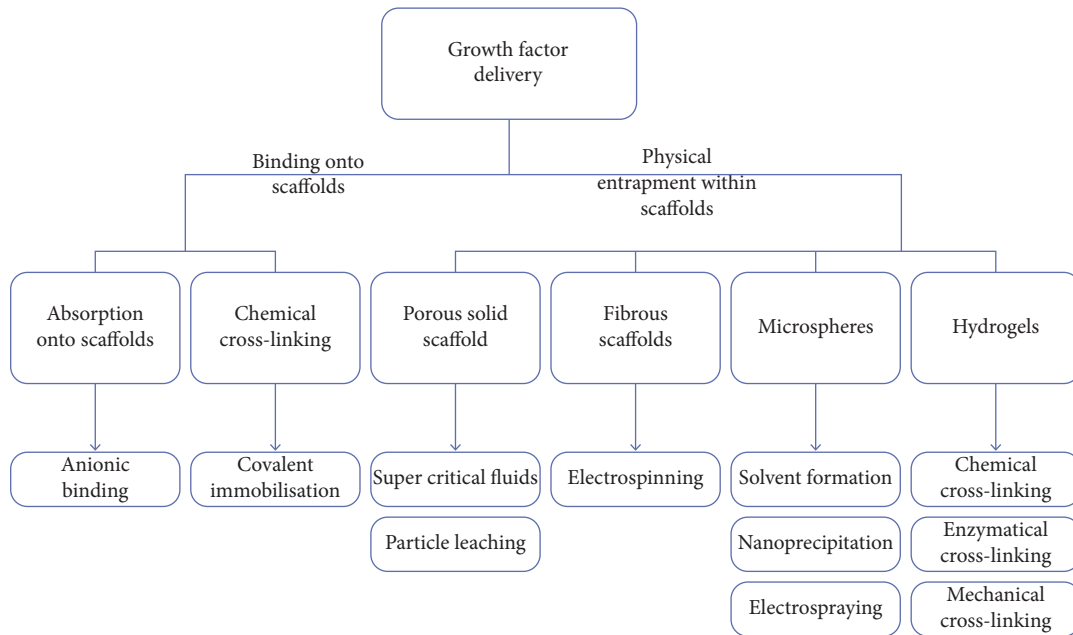


FIGURE 2: Current methods for fabrication of growth factor-loaded scaffolds [117].

range of bone defects. These include (1) rhBMP-2 injected into absorbable collagen for sinus lift and local augmentation of alveolar ridges, (2) OP-1 putty and rhBMP-7 for nonbonded fractures, and (3) TCP injected with rhPDGF for periodontal tissue [131]. Another study revealed that human periodontal ligament cells developed early osteoblasts when recombinant BMP-7 was added [132]. In bone development, bone production, and homeostasis, BMPs function in tandem with transforming growth factor-beta (TGF-beta) by activating signaling pathways [133].

6.2. Stem Cell Delivery Vehicles. Biomaterials can also be used for the delivery of stem cells. Embedded cells can be connected and developed using biomaterials rather than native ECM, avoiding ankylosis. In many cases, biomaterials can be altered to activate the cellular processes required for tissue regeneration and to prevent a host immune response [134, 135]. Hydrogels allow stem cells to be delivered minimally invasively to the face and jaw to repair deformities and disorders of the skull. Adipose tissue, dental pulp placenta, bone marrow, and limbus adult progenitor and stem cells were utilized in clinical studies [136]. Only a few commercially accessible materials based on mesenchymal stem cells are available for clinical uses such as DBM, Trinity Evolution Matrix™, Map3™, Osteocel Plus®, and AlloStem® [137]. A report by Soe and others indicated that mesenchymal stem cells existed in the periodontal ligament [138]. Overall, stem cell administration via biomaterials appears to help regenerate and restructure the oral cavity. Nevertheless, further research is required to evaluate the safety of the medication and its effectiveness in the long run [139].

6.3. Gene Delivery. The short half-lives of many growth factors used in tissue engineering restrict their availability at the right time and in the right amount. Recently, genes have

been used to stimulate cells to create growth factors. Biomaterials have been preferred over other virus vector systems for their safety and ease of modification and mutagenesis [118]. A study by Giannobile et al. successfully transferred BMP-7 and PDGF genes to fibroblast, cementoblasts, and other periodontal cells [140]. Transplantation of cells expressing these genes into periodontal wounds stimulated bone and cement regeneration in rats [141]. Using this technology, periodontal repair simulations can be simulated, although additional studies on the efficiency and safety of the method are necessary [142].

6.4. Scaffold and Cell-Free Technologies. To diagnose prognostic human illness, scientists continually develop new models for diseases, detect early indicators, and experiment with new treatments. Humans are capable of curing a variety of illnesses through multipotent stem cells. Cells of the mesenchymal stem cell line may differentiate into a variety of cell types and function as paracrine glands that secrete endogenous chemicals that affect the immune response and aid tissue repair [143]. It is difficult to safely use MSC banking for rapid regenerative applications due to the unique challenges specific to each MSC type, including technical, legal, and ethical issues. EVs are a group of small vesicles that are released from a different cell types and heterogeneous cultures by budding from the plasma membrane. There are many different vesicles that are seen in EVs, such as exosomes, shed vesicles, nanoparticles, and apoptotic bodies [144]. The terms ectosome, microparticle, and nanoparticle refer to single vesicles released directly from the plasma membrane. The origin of exosomes has been implicated in numerous studies; however, reliable information regarding EV origins is often lacking. Nano- or microvesicles are classified according to their size. The secretome of a cell includes extracellular vesicles. The major

components of exosomes are nucleic acids, proteins, cytokines, enzymes, and misfolded proteins [143]. In light of their many features to be utilized for diagnosing, prognosticating, and therapeutic purposes, EVs are regarded as new and clever theranostic instruments [144]. Studies have examined how MSC-derived exosomes can be used in the regeneration of kidney, liver, hearts, and neurological damage in a variety of model illnesses and conditions. Scientists have recently focused on ESC-MSCs. Several methods were employed for examining the components in the conditioned medium, including multidimensional protein identification technologies, gene microarrays, and cytokine antibody arrays [144]. Based on computational analysis of the acquired data, computational analysis of the gene products involved in immune response and tissue differentiation was predicted [145]. According to Zhang et al., exosomes produced from human embryonic MSCs can be used to heal cartilage and subchondral bones and can therefore be considered a “cell-free” therapeutic method for osteochondral disorders [146].

6.5. Other Potential Biomaterials. These days, smart material research focuses on allotropic materials that have unique properties including medicinal applications that make them valuable components. Smart two-dimensional materials are being studied mainly due to their unique allotropic properties, which have shown potential in a number of applications. Two-dimensional materials that are nanometer scalable can be used to enhance the interaction between cells and human tissue. Biocompatible and bioactive interfaces are needed between cells and biomaterials. Recent studies have demonstrated the effectiveness of MoS₂, WSe₂, and h-BN as biomedical device fabrication materials [147]. It has been consistently found that two-dimensional materials exhibit distinct physical, chemical, electrical, and optical properties. The antimicrobial and physical properties of graphene have been widely reported in recent years, but the material also has certain limitations. In the coming years, black phosphorene (BP) could replace graphene as a potential material for biomedical applications due to its material structure and biological properties. In the same way as other 2D materials, BP may be used to make colorimetric and fluorescence detectors and biosensors. Furthermore, BP produces nontoxic byproducts in the body after *in vivo* biodegradation. The property of this material could be beneficial for pharmaceuticals, prosthetic coatings, and scaffolds. Because BP is a low cytotoxic agent, there are no local effects [147].

Inzana and others focused their 2014 research on creating calcium phosphate scaffolds using low-temperature 3D printing. Biolinking these scaffolds to bone tissue chemicals is possible because these scaffolds showed good cytocompatibility and osteoconductive properties [148]. There is already BP in bone, but in tiny amounts, comprising 1% (slightly more than 660 grams) of the total body weight [149]. A biolinkage may use this characteristic to combine with chemicals that enhance osteoconduction [148]. The role of calcium and phosphate in bone healing is well known in

tissue engineering. The researchers focused on bioglass-based scaffolds made from BP nanosheets. A variety of scaffolds can be created using bio-printing. A 3D printing technique that uses biomaterials that have been doped or coated with BP might be a viable way of improving osteosarcoma therapy [150]. Wang et al. created experimental scaffolds that mimicked medullary bone by creating reticular damage to promote and enhance cellular attachment and colonization. Nanosheets of BP (200–400 nm) were applied to the scaffolds to bind them safely and effectively. Based on results from *in vitro* testing, the coated scaffolds were very capable of promoting bone formation due to increased cell proliferation on their surfaces, which may have been explained by their unique shape. The BP-BG scaffolds have been demonstrated to be more effective at treating post-oncological bone abnormalities in a mouse model of osteosarcoma [150]. The BP coating was also applied to hydrogels. In particular, gels were created by photo-reticulation of gelatin containing methacrylamide with ultraviolet light (GelMA). GelMA and BP were coated with arginine and poly(ester amide). A functionalized hydrogel facilitated bone development. The hydrogel's compression modulus and biodegradability time were measured *in vitro*, where BP submerged in mimicked physiological fluids produced a positive response. In addition, BP-based hydrogels promoted the proliferation of human dental pulp stem cells (hDPSCs) when differentiation into osteoblasts occurred. Based on the results of this study, BP-coated hydrogels may be suitable for use in dentistry if they provide the optimal environment for hDPSCs [151]. Then, 2D boron sheets have been grown on Ag substrates to produce two-dimensional triangular structures known as borophene (BO). As with graphene, borophene regularly exhibits anisotropic properties. Because of its simple 3D structure, borons are classified as metalloids because neither metals nor nonmetals can be formed from their structure. Several semiconductors are fabricated using this material. The metal properties of boron are more apparent when it is arranged in a two-dimensional structure. This is comparable to allotropes such as graphene [152]. A borophene ridge's form and size are determined by how strongly boron atoms bond together. Because of this, the surfaces of graphene and borophene are significantly different. Borophene is a polymorphous and anisotropic compound due to its structural feature [153]. The properties of borophene make it a fascinating material for biomedical applications. This material is known as the “chameleon” of biomedical materials because it exhibits a variety of chemistry and physical properties that are suitable for both medical devices and customized biomedicine, exhibiting a variety of behavior and existence in many phases.

7. Conclusion

To place a suitable implant, doctors have examined the tissue and bone repair to put a crown/root ratio that is ideal, as well as long-term stability of soft tissue. The epithelium is separated from the damaged tissue with the help of different kinds of degradable septic membranes in GTR and GBR

treatments. Because e-PTFE membranes are indestructible, additional surgery is required to remove them. They have the greatest flaw of all. Despite their biodegradability and cell adhesion, natural polymer-based membranes have poor mechanical strength and short degradation cycles. Synthetic polymer-based membranes can be controlled for their biodegradability and mechanical strength. They, however, have a lower biological activity than natural polymers. Additionally, their decomposition products might trigger inflammatory responses outside the body. Despite some drawbacks, the importance and irreplaceability of biodegradable polymers cannot be overstated in GTR and GBR procedures, while periodontal biomaterials have become increasingly popular over the past several decades, owing to their many benefits ranging from testing membranes to everyday use. Biomaterials will be determined by factors found in genes and stem cells that control cell growth. Furthermore, membrane ossification activity needs to be studied to maintain a balance between their mechanical and biological features.

Abbreviations

GTR:	Guided tissue reconstruction/regeneration
PDL:	Periodontal ligament
GBR:	Guided bone regeneration
PLA:	Poly(lactic acid)
PLGA:	Poly(lactide-co-glycolide)
NMP:	N-methyl-2-pyrrolidone
PCL:	Poly(ϵ -caprolactone)
FGF:	Fibroblast growth factor
PEG:	Poly(ethylene glycol)
PTFE:	Poly(tetrafluoroethylene)
e-PTFE:	Expanded poly(tetrafluoroethylene)
d-PTFE:	Dense poly(tetrafluoroethylene)
TR-ePTFE:	Titanium-reinforced expanded poly(tetrafluoroethylene)
CoCr:	Cobalt-chromium
CaS:	Calcium sulfate
HA:	Hydroxyapatite
PDGF:	Platelet-derived growth factor
PTH:	Parathyroid hormone
BMP:	Bone morphogenetic proteins
PRP:	Platelet-rich plasma
IGF:	Insulin-like growth factor
EMD:	Enamel matrix derivative
ECM:	Extracellular matrix
VEGF:	Vascular endothelial growth factor
DFDBA:	Demineralized freeze-dried bone allograft
GFs:	Growth factors
DBM:	Demineralized bone matrix
Gp:	Genipin
TGF- β :	Transforming growth factor-beta
MSCs:	Mesenchymal stem cells
EVs:	Extracellular vesicles
ESC- MSCs:	Human embryonic stem cell-derived MSCs
2D:	Two-dimensional
BP:	Black phosphorene

GelMA:	Methacrylamide gelatin
U-Arg- PEAs:	Arginine and poly (ester amide)
hDPSCs:	Human dental pulp stem cells.

Data Availability

The datasets used and/or analyzed during this study are available from the corresponding author on reasonable request.

Conflicts of Interest

The authors declare no conflicts of interest.

References

- [1] V. Venugopalan, R. Vamsi, S. Shenoy, K. Ashok, and B. Thomas, "Guided bone regeneration-A comprehensive review," *Journal of Clinical and Diagnostic Research*, vol. 15, no. 4, 2021.
- [2] B. Lenka, R. Mohanty, R. Nayak, G. Mohanty, A. Satpathy, and S. Panda, "Current research directions in regenerative therapy," *Indian Journal of Forensic Medicine & Toxicology*, vol. 14, no. 4, 2020.
- [3] Y.-D. Cho, K.-H. Kim, Y.-M. Lee, Y. Ku, and Y.-J. Seol, "Periodontal wound healing and tissue regeneration: a narrative review," *Pharmaceuticals*, vol. 14, no. 5, p. 456, 2021.
- [4] A. Simonelli, L. Minenna, T. Leonardo, and F. Roberto, "Single flap approach with or without enamel matrix derivative in the treatment of severe supraosseous defects: a retrospective study," *Clinical Oral Investigations*, vol. 25, no. 11, pp. 6385–6392, 2021.
- [5] F. Diomedede, G. D. Marconi, L. Fonticoli et al., "Functional relationship between osteogenesis and angiogenesis in tissue regeneration," *International Journal of Molecular Sciences*, vol. 21, no. 9, p. 3242, 2020.
- [6] A. Hoornaert and P. Layrolle, *Bone Regenerative Issues Related to Bone Grafting Biomaterials*, pp. 207–215, Dental Implants and Bone Grafts, Woodhead Publishing, Sawston, UK, 2020.
- [7] N. Doan, P. Reher, Q. T. Duong, G. Wang, and L. Truong, "Application of blood stem cells (CD34+ and CD45)/concentrated growth factors (CGF) in guided bone regeneration (GBR) and guided tissue regeneration (GTR) in conjunction with mls laser and piezoelectric surgery," *International Journal of Oral and Maxillofacial Surgery*, vol. 48, p. 62, 2019.
- [8] S. Zahid, A. S. Khan, A. A. Chaudhry et al., "Fabrication, in vitro and in vivo studies of bilayer composite membrane for periodontal guided tissue regeneration," *Journal of Biomaterials Applications*, vol. 33, no. 7, pp. 967–978, 2019.
- [9] S. Beigi-Broujeni and S. Babanzadeh, "Polymeric membranes used for guided periodontal tissue regeneration: a review-Part I," *Basparesh*, vol. 8, no. 2, pp. 83–90, 2018.
- [10] M. Toledano Pérez, Á. Carrasco-Carmona, A. L. Medina-Castillo, M. Toledano-Osorio, and R. Osorio, "Protein adsorption and bioactivity of functionalized electrospun membranes for bone regeneration," *Journal of Dentistry*, vol. 102, p. 103473, 2020.
- [11] I. Sanz-Sánchez, A. Carrillo de Albornoz, E. Figuero, F. Schwarz, R. Jung, and M. Sanz, "Effects of lateral bone augmentation procedures on peri-implant health or disease:

- a systematic review and meta-analysis," *Clinical Oral Implants Research*, vol. 29, pp. 18–31, 2018.
- [12] M. Chiapasco and M. Zaniboni, "Clinical outcomes of GBR procedures to correct peri-implant dehiscences and fenestrations: a systematic review," *Clinical Oral Implants Research*, vol. 20, pp. 113–123, 2009.
- [13] A. V. Imbronito, J. H. Todescan, C. V. Carvalho, and V. E. Arana-Chavez, "Healing of alveolar bone in resorbable and non-resorbable membrane-protected defects. A histologic pilot study in dogs," *Biomaterials*, vol. 23, no. 20, pp. 4079–4086, 2002.
- [14] F. Donnalaja, E. Jacchetti, M. Soncini, and M. T. Raimondi, "Natural and synthetic polymers for bone scaffolds optimization," *Polymers*, vol. 12, no. 4, p. 905, 2020.
- [15] N. Iqbal, A. S. Khan, A. Asif, M. Yar, J. W. Haycock, and I. U. Rehman, "Recent concepts in biodegradable polymers for tissue engineering paradigms: a critical review," *International Materials Reviews*, vol. 64, no. 2, pp. 91–126, 2019.
- [16] L. S. Nair and C. T. Laurencin, "Biodegradable polymers as biomaterials," *Progress in Polymer Science*, vol. 32, no. 8–9, pp. 762–798, 2007.
- [17] F. Döri, T. Huszár, D. Nikolidakis, N. B. Arweiler, I. Gera, and A. Sculean, "Effect of platelet-rich plasma on the healing of intra-bony defects treated with a natural bone mineral and a collagen membrane," *Journal of Clinical Periodontology*, vol. 34, no. 3, pp. 254–261, 2007.
- [18] A. Kozlovsky, G. Aboodi, O. Moses et al., "Bio-degradation of a resorbable collagen membrane (Bio-Gide) applied in a double-layer technique in rats," *Clinical Oral Implants Research*, vol. 20, no. 10, pp. 1116–1123, 2009.
- [19] A. Turri, I. Elgali, F. Vaziriani et al., "Guided bone regeneration is promoted by the molecular events in the membrane compartment," *Biomaterials*, vol. 84, pp. 167–183, 2016.
- [20] T. Gueldenpfennig, A. Houshmand, S. Najman et al., "The condensation of collagen leads to an extended standing time and a decreased pro-inflammatory tissue response to a newly developed pericardium-based barrier membrane for guided bone regeneration," *In Vivo*, vol. 34, no. 3, pp. 985–1000, 2020.
- [21] Y. Ueyama, K. Ishikawa, T. Mano et al., "Usefulness as guided bone regeneration membrane of the alginate membrane," *Biomaterials*, vol. 23, no. 9, pp. 2027–2033, 2002.
- [22] H. He, J. Huang, F. Ping, G. Sun, and G. Chen, "Calcium alginate film used for guided bone regeneration in mandible defects in a rabbit model," *Cranio: The Journal of Cranio-mandibular & Sleep Practice*, vol. 26, no. 1, pp. 65–70, 2008.
- [23] S. Ma, A. Adayi, Z. Liu, M. Li, M. Wu, L. Xiao et al., "Asymmetric collagen/chitosan membrane containing minocycline-loaded chitosan nanoparticles for guided bone regeneration," *Scientific Reports*, vol. 6, no. 1, pp. 1–10, 2016.
- [24] T. Zhou, X. Liu, B. Sui, C. Liu, X. Mo, and J. Sun, "Development of fish collagen/bioactive glass/chitosan composite nanofibers as a GTR/GBR membrane for inducing periodontal tissue regeneration," *Biomedical Materials*, vol. 12, no. 5, p. 055004, 2017.
- [25] C. Wu, H. Su, A. Karydis et al., "Mechanically stable surface-hydrophobilized chitosan nanofibrous barrier membranes for guided bone regeneration," *Biomedical Materials*, vol. 13, no. 1, p. 015004, 2017.
- [26] D. Huang, L. Niu, J. Li et al., "Reinforced chitosan membranes by microspheres for guided bone regeneration," *Journal of the Mechanical Behavior of Biomedical Materials*, vol. 81, pp. 195–201, 2018.
- [27] A. T. Shah, S. Zahid, F. Ikram et al., "Tri-layered functionally graded membrane for potential application in periodontal regeneration," *Materials Science and Engineering: C*, vol. 103, p. 109812, 2019.
- [28] N. C. Geurs, J. M. Korostoff, P. J. Vassilopoulos et al., "Clinical and histologic assessment of lateral alveolar ridge augmentation using a synthetic long-term bioabsorbable membrane and an allograft," *Journal of Periodontology*, vol. 79, no. 7, pp. 1133–1140, 2008.
- [29] M. Annunziata, L. Nastro, G. Cecoro, and L. Guida, "The use of poly-D,L-lactic acid (pdlla) devices for bone augmentation techniques: a systematic review," *Molecules*, vol. 22, no. 12, p. 2214, 2017.
- [30] I. Yoshimoto, J.-I. Sasaki, R. Tsuboi, S. Yamaguchi, H. Kitagawa, and S. Imazato, "Development of layered PLGA membranes for periodontal tissue regeneration," *Dental Materials*, vol. 34, no. 3, pp. 538–550, 2018.
- [31] A. Haghghat, S. Shakeri, M. Mehdikhani, S. S. Dehnavi, and A. Talebi, "Histologic, histomorphometric, and osteogenesis comparative study of a novel fabricated nanocomposite membrane versus cytoplasm membrane," *Journal of Oral and Maxillofacial Surgery*, vol. 77, no. 10, pp. 2027–2039, 2019.
- [32] H. Y. Zhang, H. B. Jiang, J.-H. Ryu, H. Kang, K.-M. Kim, and J.-S. Kwon, "Comparing properties of variable pore-sized 3D-printed PLA membrane with conventional PLA membrane for guided bone/tissue regeneration," *Materials*, vol. 12, no. 10, p. 1718, 2019.
- [33] G. L. Abe, J.-I. Sasaki, C. Katata et al., "Fabrication of novel poly(lactic acid/caprolactone) bilayer membrane for GBR application," *Dental Materials*, vol. 36, no. 5, pp. 626–634, 2020.
- [34] M. Chi, M. Qi, L. A et al., "Novel bioactive and therapeutic dental polymeric materials to inhibit periodontal pathogens and biofilms," *International Journal of Molecular Sciences*, vol. 20, no. 2, p. 278, 2019.
- [35] T. Korzinskas, O. Jung, R. Smeets et al., "In vivo analysis of the biocompatibility and macrophage response of a non-resorbable PTFE membrane for guided bone regeneration," *International Journal of Molecular Sciences*, vol. 19, no. 10, p. 2952, 2018.
- [36] J. Garcia, A. Dodge, P. Luepke, H.-L. Wang, Y. Kapila, and G.-H. Lin, "Effect of membrane exposure on guided bone regeneration: a systematic review and meta-analysis," *Clinical Oral Implants Research*, vol. 29, no. 3, pp. 328–338, 2018.
- [37] P. Gallo and D. Diaz-Báez, "Management of 80 complications in vertical and horizontal ridge augmentation with nonresorbable membrane (d-PTFE): a cross-sectional study," *The International Journal of Oral & Maxillofacial Implants*, vol. 34, no. 4, pp. 927–935, 2019.
- [38] M. Ronda, A. Rebaudi, L. Torelli, and C. Stacchi, "Expanded vs. dense polytetrafluoroethylene membranes in vertical ridge augmentation around dental implants: a prospective randomized controlled clinical trial," *Clinical Oral Implants Research*, vol. 25, no. 7, pp. 859–866, 2014.
- [39] Y. Sumi, O. Miyaishi, I. Tohna, and M. Ueda, "Alveolar ridge augmentation with titanium mesh and autogenous bone," *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology & Endodontics*, vol. 89, no. 3, pp. 268–270, 2000.
- [40] Y. D. Rakhmatia, Y. Ayukawa, A. Furuhashi, and K. Koyano, "Current barrier membranes: titanium mesh and other membranes for guided bone regeneration in dental applications," *Journal of prosthodontic research*, vol. 57, no. 1, pp. 3–14, 2013.

- [41] H. Hasegawa, S. Masui, and H. Ishihata, "New micro-perforated pure titanium membrane created by laser processing for guided regeneration of bone," *British Journal of Oral and Maxillofacial Surgery*, vol. 56, no. 7, pp. 642-643, 2018.
- [42] A. Bouguezzi, A. Debibi, A. Chokri, S. Sioud, H. Hentati, and J. Selmi, "Cross-linked versus natural collagen membrane for guided bone regeneration? A literature review," *American Journal of Medical and Biological Research*, vol. 8, no. 1, pp. 12-16, 2020.
- [43] F. Schwarz, D. Rothamel, M. Hertzen et al., "Immunohistochemical characterization of guided bone regeneration at a dehiscence-type defect using different barrier membranes: an experimental study in dogs," *Clinical Oral Implants Research*, vol. 19, no. 4, pp. 402-415, 2008.
- [44] J. L. Rudolf, C. Moser, A. Sculean, and S. Eick, "In-vitro antibiofilm activity of chlorhexidine digluconate on polylactide-based and collagen-based membranes," *BMC Oral Health*, vol. 19, no. 1, p. 291, 2019.
- [45] J. Jiménez García, S. Berghazan, J. M. M. Caramès, M. M. Dard, and D. N. S. Marques, "Effect of cross-linked vs non-cross-linked collagen membranes on bone: a systematic review," *Journal of Periodontal Research*, vol. 52, no. 6, pp. 955-964, 2017.
- [46] F. Schwarz, D. Rothamel, M. Hertzen, M. Sager, and J. Becker, "Angiogenesis pattern of native and cross-linked collagen membranes: an immunohistochemical study in the rat," *Clinical Oral Implants Research*, vol. 17, no. 4, pp. 403-409, 2006.
- [47] D. Rothamel, F. Schwarz, M. Sager, M. Hertzen, A. Sculean, and J. Becker, "Biodegradation of differently cross-linked collagen membranes: an experimental study in the rat," *Clinical Oral Implants Research*, vol. 16, no. 3, pp. 369-378, 2005.
- [48] D. Rothamel, F. Schwarz, A. Sculean, M. Hertzen, W. Scherbaum, and J. Becker, "Biocompatibility of various collagen membranes in cultures of human PDL fibroblasts and human osteoblast-like cells," *Clinical Oral Implants Research*, vol. 15, no. 4, pp. 443-449, 2004.
- [49] H. Tal, A. Kozlovsky, Z. Artzi, C. E. Nemcovsky, and O. Moses, "Long-term bio-degradation of cross-linked and non-cross-linked collagen barriers in human guided bone regeneration," *Clinical Oral Implants Research*, vol. 19, no. 3, pp. 295-302, 2008.
- [50] N. U. Zitzmann, R. Naef, and P. Schärer, "Resorbable versus nonresorbable membranes in combination with Bio-Oss for guided bone regeneration," *The International Journal of Oral & Maxillofacial Implants*, vol. 12, no. 6, pp. 844-852, 1997.
- [51] L. Sbricoli, R. Guazzo, M. Annunziata, L. Gobato, E. Bressan, and L. Natri, "Selection of collagen membranes for bone regeneration: a literature review," *Materials*, vol. 13, no. 3, p. 786, 2020.
- [52] P. Bunyaratavej and H.-L. Wang, "Collagen membranes: a review," *Journal of Periodontology*, vol. 72, no. 2, pp. 215-229, 2001.
- [53] E. A. Grebenik, L. P. Istranov, E. V. Istranova, S. N. Churbanov, B. S. Shavkuta, R. I. Dmitriev et al., "Chemical cross-linking of xenopericardial biomeshes: a bottom-up study of structural and functional correlations," *Xenotransplantation*, vol. 26, no. 3, p. e12506, 2019.
- [54] O. Uğur and O. Karaman, "Development of synthetic barrier dental membrane for guided bone regeneration," in *Proceedings of the 2018 Medical Technologies National Congress (TIPTEKNO)*, IEEE, Magusa, Cyprus, 2018.
- [55] G. Sam and B. R. M. Pillai, "Evolution of barrier membranes in periodontal regeneration—are the third generation membranes really here?" *Journal of Clinical and Diagnostic Research: Journal of Clinical and Diagnostic Research*, vol. 8, no. 12, p. ZE14, 2014.
- [56] I. Elgali, O. Omar, C. Dahlin, and P. Thomsen, "Guided bone regeneration: materials and biological mechanisms revisited," *European Journal of Oral Sciences*, vol. 125, no. 5, pp. 315-337, 2017.
- [57] B. Wessing, S. Lettner, and W. Zechner, "Guided bone regeneration with collagen membranes and particulate graft materials: a systematic review and meta-analysis," *The International Journal of Oral & Maxillofacial Implants*, vol. 33, no. 1, pp. 87-100, 2018.
- [58] S. Dowlatshahi, C. Y. Chen, H. Zigdon-Giladi et al., "Volumetric assessment of changes in the alveolar ridge dimension following GBR using a combination FDBA with collagen membrane or novel resorbable scaffold: a prospective two-center clinical trial," *Journal of Periodontology*, vol. 93, no. 3, pp. 343-353, 2021.
- [59] S. Vahabi, Z. Yadegary, and M. Karamshahi, "Evaluating the adhesion of human gingival fibroblasts and MG-63 osteoblast-like cells to activated PRP-coated membranes," *Cell and Tissue Banking*, vol. 20, no. 3, pp. 339-349, 2019.
- [60] M. C. Bottino, V. Thomas, G. Schmidt et al., "Recent advances in the development of GTR/GBR membranes for periodontal regeneration-A materials perspective," *Dental Materials*, vol. 28, no. 7, pp. 703-721, 2012.
- [61] O. Omar, A. Dahlin, A. Gasser, and C. Dahlin, "Tissue dynamics and regenerative outcome in two resorbable non-cross-linked collagen membranes for guided bone regeneration: a preclinical molecular and histological study in vivo," *Clinical Oral Implants Research*, vol. 29, no. 1, pp. 7-19, 2018.
- [62] G. D. Mogoşanu and A. M. Grumezescu, "Natural and synthetic polymers for wounds and burns dressing," *International Journal of Pharmaceutics*, vol. 463, no. 2, pp. 127-136, 2014.
- [63] T. Jiang, E. J. Carbone, K. W.-H. Lo, and C. T. Laurencin, "Electrospinning of polymer nanofibers for tissue regeneration," *Progress in Polymer Science*, vol. 46, pp. 1-24, 2015.
- [64] K. Noritake, S. Kuroda, M. Nyan, K. Ohya, Y. Tabata, and S. Kasugai, "Development of a new barrier membrane for guided bone regeneration: an in vitro and in vivo study," *Journal of Oral Tissue Engineering*, vol. 9, no. 2, pp. 53-63, 2011.
- [65] F. R. da Silva, R. O. Silva, H. M. de Castro Oliveira et al., "Gelatin-based membrane containing usnic acid-loaded liposomes: a new treatment strategy for corneal healing," *Biomedicine & Pharmacotherapy*, vol. 130, p. 110391, 2020.
- [66] T. Ahmadi, A. Monshi, V. Mortazavi et al., "Fabrication and characterization of polycaprolactone fumarate/gelatin-based nanocomposite incorporated with silicon and magnesium co-doped fluorapatite nanoparticles using electrospinning method," *Materials Science and Engineering: C*, vol. 106, p. 110172, 2020.
- [67] J. Shao, N. Yu, E. Kolwijck et al., "Biological evaluation of silver nanoparticles incorporated into chitosan-based membranes," *Nanomedicine*, vol. 12, no. 22, pp. 2771-2785, 2017.
- [68] N. Ghadri, K. M. Anderson, P. Adatrow et al., "Evaluation of bone regeneration of simvastatin loaded chitosan nanofiber membranes in rodent calvarial defects," *Journal of*

- Biomaterials and Nanobiotechnology*, vol. 09, no. 02, pp. 210–231, 2018.
- [69] A. J. Bavariya, P. Andrew Norowski, K. Mark Anderson et al., “Evaluation of biocompatibility and degradation of chitosan nanofiber membrane crosslinked with genipin,” *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, vol. 102, no. 5, pp. 1084–1092, 2014.
- [70] S. Ma, Z. Chen, F. Qiao et al., “Guided bone regeneration with tripolyphosphate cross-linked asymmetric chitosan membrane,” *Journal of Dentistry*, vol. 42, no. 12, pp. 1603–1612, 2014.
- [71] H. Zhou, J. G. Lawrence, and S. B. Bhaduri, “Fabrication aspects of PLA-CaP/PLGA-CaP composites for orthopedic applications: a review,” *Acta Biomaterialia*, vol. 8, no. 6, pp. 1999–2016, 2012.
- [72] F. Ebrahimi and H. Ramezani Dana, “Poly lactic acid (PLA) polymers: from properties to biomedical applications,” *International Journal of Polymeric Materials and Polymeric Biomaterials*, pp. 1–14, 2021.
- [73] L. S. Karfeld-Sulzer, C. Ghayor, B. Siegenthaler, B. Gjoksi, T. H. Pohjonen, and F. E. Weber, “Comparative study of NMP-preloaded and dip-loaded membranes for guided bone regeneration of rabbit cranial defects,” *Journal of tissue engineering and regenerative medicine*, vol. 11, no. 2, pp. 425–433, 2017.
- [74] R. De Santis, A. Russo, A. Gloria et al., “Towards the design of 3D fiber-deposited poly(-caprolactone)/Iron-Doped hydroxyapatite nanocomposite magnetic scaffolds for bone regeneration,” *Journal of Biomedical Nanotechnology*, vol. 11, no. 7, pp. 1236–1246, 2015.
- [75] P. Gentile, V. Chiono, C. Tonda-Turo, A. M. Ferreira, and G. Ciardelli, “Polymeric membranes for guided bone regeneration,” *Biotechnology Journal*, vol. 6, no. 10, pp. 1187–1197, 2011.
- [76] N. K. de Moura, E. F. Martins, R. L. M. S. Oliveira et al., “Synergistic effect of adding bioglass and carbon nanotubes on poly (lactic acid) porous membranes for guided bone regeneration,” *Materials Science and Engineering: C*, vol. 117, Article ID 111327, 2020.
- [77] S. Soltani Dehnavi, M. Mehdikhani, M. Rafenia, and S. Bonakdar, “Preparation and in vitro evaluation of polycaprolactone/PEG/bioactive glass nanopowders nanocomposite membranes for GTR/GBR applications,” *Materials Science and Engineering: C*, vol. 90, pp. 236–247, 2018.
- [78] K. Budak, O. Sogut, and U. A. Sezer, “A review on synthesis and biomedical applications of polyglycolic acid,” *Journal of Polymer Research*, vol. 27, no. 8, pp. 1–19, 2020.
- [79] P. Rider, Ž. P. Kačarević, A. Elad, D. Tadic, D. Rothamel, G. Sauer et al., “Biodegradable magnesium barrier membrane used for guided bone regeneration in dental surgery,” *Bioactive Materials*, vol. 14, pp. 152–168, 2022.
- [80] A. M. Greenberg, “Localized ridge augmentation using guided bone regeneration,” in *Craniofacial Reconstructive and Corrective Bone Surgery*, pp. 155–163, Springer, New York, NY, USA, 2019.
- [81] N. Kohli, J. C. Stoddart, and R. J. van Arkel, “The limit of tolerable micromotion for implant osseointegration: a systematic review,” *Scientific Reports*, vol. 11, no. 1, p. 10797, 2021.
- [82] H. Gluckman and J. Du Toit, “The management of recession midfacial to immediately placed implants in the aesthetic zone,” *Int Dent Afr Ed*, vol. 5, no. 1, pp. 6–15, 2015.
- [83] F. Jabari, B. Houshmand, and S. Hesarakhi, “The role of barrier membranes in guided bone regeneration: a review,” *Journal of Dental Medicine*, vol. 31, no. 3, pp. 198–207, 2018.
- [84] Ö. Solakoglu, W. Götz, M. C. Kiessling, C. Alt, C. Schmitz, and E. U. Alt, “Improved guided bone regeneration by combined application of unmodified, fresh autologous adipose derived regenerative cells and plasma rich in growth factors: a first-in-human case report and literature review,” *World Journal of Stem Cells*, vol. 11, no. 2, pp. 124–146, 2019.
- [85] Ö. Solakoglu, G. Heydecke, N. Amiri, and E. Anitua, “The use of plasma rich in growth factors (PRGF) in guided tissue regeneration and guided bone regeneration. A review of histological, immunohistochemical, histomorphometrical, radiological and clinical results in humans,” *Annals of Anatomy - Anatomischer Anzeiger*, vol. 231, Article ID 151528, 2020.
- [86] J. P. Matinlinna, *Handbook of Oral Biomaterials*, Pan Stanford, Jenny Stanford Publishing, Singapore, 2019.
- [87] M. Trobos, A. Juhlin, F. A. Shah, M. Hoffman, H. Sahlin, and C. Dahlin, “In vitro evaluation of barrier function against oral bacteria of dense and expanded polytetrafluoroethylene (PTFE) membranes for guided bone regeneration,” *Clinical Implant Dentistry and Related Research*, vol. 20, no. 5, pp. 738–748, 2018.
- [88] F. Mandelli, T. Traini, and P. Ghensi, “Customized-3D zirconia barriers for guided bone regeneration (GBR): clinical and histological findings from a proof-of-concept case series,” *Journal of Dentistry*, vol. 114, Article ID 103780, 2021.
- [89] M. Duskova, E. Leamerova, B. Sosna, and O. Gojis, “Guided tissue regeneration, barrier membranes and reconstruction of the cleft maxillary alveolus,” *Journal of Craniofacial Surgery*, vol. 17, no. 6, pp. 1153–1160, 2006.
- [90] P. Ghensi, W. Stablum, E. Bettio, M. C. Soldini, T. R. Tripi, and C. Soldini, “Management of the exposure of a dense PTFE (d-PTFE) membrane in guided bone regeneration (GBR): a case report,” *Oral Implantology*, vol. 10, no. 3, p. 335, 2017.
- [91] M. Degidi, A. Scarano, and A. Piattelli, “Regeneration of the alveolar crest using titanium micromesh with autologous bone and a resorbable membrane,” *Journal of Oral Implantology*, vol. 29, no. 2, pp. 86–90, 2003.
- [92] P. Windisch, K. Orban, G. E. Salvi, A. Sculean, and B. Molnar, “Vertical-guided bone regeneration with a titanium-reinforced d-PTFE membrane utilizing a novel split-thickness flap design: a prospective case series,” *Clinical Oral Investigations*, vol. 25, no. 5, pp. 2969–2980, 2021.
- [93] D. M. Brunette, P. Tengvall, M. Textor, and P. Thomsen, *Titanium in Medicine: Material Science, Surface Science, Engineering, Biological Responses and Medical Applications*, Springer Science & Business Media, Berlin, Germany, 2012.
- [94] F. Zeynalzadeh and A. Zahedpasha, “Vertical ridge augmentation by titanium mesh,” in *Innovative Perspectives in Oral and Maxillofacial Surgery*, pp. 117–124, Springer, Cham, Switzerland, 2021.
- [95] L. Ricci, V. Perrotti, L. Ravera, A. Scarano, A. Piattelli, and G. Iezzi, “Rehabilitation of deficient alveolar ridges using titanium grids before and simultaneously with implant placement: a systematic review,” *Journal of Periodontology*, vol. 84, no. 9, pp. 1234–1242, 2013.
- [96] C. Chakar Carole, N. Mokbel, S. Khalil, and A. R. Kassir, “Current Knowledge and Future Perspectives of Barrier Membranes: a Biomaterials Perspective = Connaissances Actuelles et Perspectives d’Avenir des Membranes

- Barrières,” *International Arab Journal of Dentistry*, vol. 11, no. 1, pp. 43–50, 2020.
- [97] M. Noronha Oliveira, W. V. H. Schunemann, M. T. Mathew et al., “Can degradation products released from dental implants affect peri-implant tissues?” *Journal of Periodontal Research*, vol. 53, no. 1, pp. 1–11, 2018.
- [98] C. M. Lappas, “The immunomodulatory effects of titanium dioxide and silver nanoparticles,” *Food and Chemical Toxicology*, vol. 85, pp. 78–83, 2015.
- [99] A. Dubey, M. Goswami, K. Yadav, and D. Chaudhary, “Oxidative stress and nano-toxicity induced by TiO₂ and ZnO on WAG cell line,” *PLoS One*, vol. 10, no. 5, Article ID e0127493, 2015.
- [100] E. Bressan, L. Ferroni, C. Gardin et al., “Metal nanoparticles released from dental implant surfaces: potential contribution to chronic inflammation and peri-implant bone loss,” *Materials*, vol. 12, no. 12, p. 2036, 2019.
- [101] O. Decco, A. Cura, V. Beltrán, M. Lezcano, and W. Engelke, “Bone augmentation in rabbit tibia using microfixed cobalt-chromium membranes with whole blood, tricalcium phosphate and bone marrow cells,” *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 1, pp. 135–44, 2015.
- [102] L. Di Alberti, F. Tamborrino, L. Lo Muzio, A. D’Agostino, L. Trevisiol, D. de santis et al., “Calcium sulfate barrier for regeneration of human bone defects. 3 years randomized controlled study,” *Minerva Stomatologica*, vol. 62, pp. 9–13, 2013.
- [103] M. Debel, S. Toma, B. Vandenberghe, M. C. Brecx, and J. F. Lasserre, “Alveolar ridge dimensional changes after two socket sealing techniques. A pilot randomized clinical trial,” *Clinical Oral Investigations*, vol. 25, no. 3, pp. 1235–1243, 2021.
- [104] M. Arnav, “Calcium sulfate: an unconventional bone graft in the management of furcation involvement - a case series,” *Journal of the International Clinical Dental Research Organization*, vol. 11, no. 1, p. 36, 2019.
- [105] J. Takamoli, A. Pascual, J. Martinez-Amargant, B. Garcia-Mur, J. Nart, and C. Valles, “Implant failure and associated risk indicators: a retrospective study,” *Clinical Oral Implants Research*, vol. 32, no. 5, pp. 619–628, 2021.
- [106] R. Malik, A. Gupta, P. Bansal, R. Sharma, and S. Sharma, “Evaluation of alveolar ridge height gained by vertical ridge augmentation using titanium mesh and novabone putty in posterior mandible,” *Journal of maxillofacial and oral surgery*, vol. 19, no. 1, pp. 32–39, 2020.
- [107] S.-L. Bee and Z. A. A. Hamid, “Characterization of chicken bone waste-derived hydroxyapatite and its functionality on chitosan membrane for guided bone regeneration,” *Composites Part B: Engineering*, vol. 163, pp. 562–573, 2019.
- [108] J. M. Song, S. H. Shin, Y. D. Kim et al., “Comparative study of chitosan/fibroin-hydroxyapatite and collagen membranes for guided bone regeneration in rat calvarial defects: micro-computed tomography analysis,” *International Journal of Oral Science*, vol. 6, no. 2, pp. 87–93, 2014.
- [109] H. Guo, D. Xia, Y. Zheng, Y. Zhu, Y. Liu, and Y. Zhou, “A pure zinc membrane with degradability and osteogenesis promotion for guided bone regeneration: in vitro and in vivo studies,” *Acta Biomaterialia*, vol. 106, pp. 396–409, 2020.
- [110] M. A. Basile, G. G. d’Ayala, M. Malinconico et al., “Functionalized PCL/HA nanocomposites as microporous membranes for bone regeneration,” *Materials Science and Engineering: C*, vol. 48, pp. 457–468, 2015.
- [111] N. Ribeiro, S. R. Sousa, C. A. van Blitterswijk, L. Moroni, and F. J. Monteiro, “A biocomposite of collagen nanofibers and nanohydroxyapatite for bone regeneration,” *Biofabrication*, vol. 6, no. 3, p. 035015, 2014.
- [112] D. M. Verissimo, R. F. Leitão, S. D. Figueiró et al., “Guided bone regeneration produced by new mineralized and reticulated collagen membranes in critical-sized rat calvarial defects,” *Experimental Biology and Medicine*, vol. 240, no. 2, pp. 175–184, 2015.
- [113] D. K. Singh and S. Kumar, “Tissue engineering: the biologic modifiers in periodontal regeneration,” *Journal of Emerging Technologies and Innovative Research*, vol. 8, no. 1, pp. 1082–1094, 2021.
- [114] A. Wiedlocha, E. M. Haugsten, and M. Zakrzewska, *Roles of the FGF-FGFR signaling system in cancer development and inflammation*, *Cell*, vol. 10, no. 9, p. 2231, 2021.
- [115] M. Pakvasa, P. Haravu, M. Boachie-Mensah et al., “Notch signaling: its essential roles in bone and craniofacial development,” *Genes & Diseases*, vol. 8, no. 1, pp. 8–24, 2021.
- [116] K. A. Blackwood, N. Bock, T. R. Dargaville, and M. Ann Woodruff, “Scaffolds for growth factor delivery as applied to bone tissue engineering,” *International Journal of Polymer Science*, vol. 2012, pp. 174942–25, 2012.
- [117] G. Thirivikraman, A. Athirasala, C. Twohig, S. K. Boda, and L. E. Bertassoni, “Biomaterials for craniofacial bone regeneration,” *Dental Clinics of North America*, vol. 61, no. 4, pp. 835–856, 2017.
- [118] X. Duan, S. R. Bradbury, B. R. Olsen, and A. D. Berendsen, “VEGF stimulates intramembranous bone formation during craniofacial skeletal development,” *Matrix Biology*, vol. 52–54, pp. 127–140, 2016.
- [119] M. Zhang, F. Jiang, X. Zhang et al., “The effects of platelet-derived growth factor-BB on human dental pulp stem cells mediated dentin-pulp complex regeneration,” *Stem Cells Translational Medicine*, vol. 6, no. 12, pp. 2126–2134, 2017.
- [120] A. Stähli, F. J. Strauss, and R. Gruber, “The use of platelet-rich plasma to enhance the outcomes of implant therapy: a systematic review,” *Clinical Oral Implants Research*, vol. 29, pp. 20–36, 2018.
- [121] A. I. Caplan and D. Correa, “PDGF in bone formation and regeneration: new insights into a novel mechanism involving MSCs,” *Journal of Orthopaedic Research*, vol. 29, no. 12, pp. 1795–1803, 2011.
- [122] R. Cimpões, L. M. Santiago, A. de Franca Caldas Júnior, B. de Carvalho Farias Vajgel, J. Perussolo, and N. Donos, “Treatment of intrabony periodontal defects in controlled diabetic patients with an enamel matrix derivative: a split-mouth randomized clinical trial,” *Clinical Oral Investigations*, vol. 26, no. 3, pp. 2479–2489, 2021.
- [123] F. Seshima, T. Kigure, and A. Saito, “Periodontal regenerative therapy using enamel matrix derivative for treatment of generalized severe chronic periodontitis: a 2-year case report,” *The Bulletin of Tokyo Dental College*, vol. 60, no. 2, pp. 97–104, 2019.
- [124] S. D. Aspriello, L. Ferrante, C. Rubini, and M. Piemontese, “Comparative study of DFDBA in combination with enamel matrix derivative versus DFDBA alone for treatment of periodontal intrabony defects at 12 months post-surgery,” *Clinical Oral Investigations*, vol. 15, no. 2, pp. 225–232, 2011.
- [125] S. P. Bienz, *Primary Bone Augmentation Leads to Equally Stable Marginal Tissue Conditions Comparing the Use of Xenograft Blocks Infused with BMP-2 and Autogenous Bone Blocks: A 3D Analysis after 3 Years*, *Clinical Oral Implants Research*, vol. 32, no. 12, pp. 4433–1443, 2021.

- [126] A. Oryan, S. Alidadi, A. Moshiri, and A. Bigham-Sadegh, "Bone morphogenetic proteins: a powerful osteoinductive compound with non-negligible side effects and limitations," *BioFactors*, vol. 40, no. 5, pp. 459–481, 2014.
- [127] H. S. Azevedo and I. Pashkuleva, "Biomimetic supramolecular designs for the controlled release of growth factors in bone regeneration," *Advanced Drug Delivery Reviews*, vol. 94, pp. 63–76, 2015.
- [128] H. D. N. Tran, K. D. Park, Y. C. Ching, C. Huynh, and D. H. Nguyen, "A comprehensive review on polymeric hydrogel and its composite: matrices of choice for bone and cartilage tissue engineering," *Journal of Industrial and Engineering Chemistry*, vol. 89, pp. 58–82, 2020.
- [129] E. P. Ramly, A. R. Alfonso, R. S. Kantar et al., "Safety and efficacy of recombinant human bone morphogenetic protein-2 (rhBMP-2) in craniofacial surgery," *Plastic and reconstructive surgery. Global open*, vol. 7, no. 8, Article ID e2347, 2019.
- [130] R. E. Jung, S. I. Windisch, A. M. Eggenschwiler, D. S. Thoma, F. E. Weber, and C. H. Hämmerle, "A randomized-controlled clinical trial evaluating clinical and radiological outcomes after 3 and 5 years of dental implants placed in bone regenerated by means of GBR techniques with or without the addition of BMP-2," *Clinical Oral Implants Research*, vol. 20, no. 7, pp. 660–666, 2010.
- [131] X. Dereka, C. E. Markopoulou, A. Mamalis, and I. A. Vrotsos, "Effect of rhBMP-7 combined with two bone grafts on human periodontal ligament cell differentiation," *Growth Factors*, vol. 27, no. 5, pp. 274–279, 2010.
- [132] M. Wu, G. Chen, and Y.-P. Li, "TGF- β and BMP signaling in osteoblast, skeletal development, and bone formation, homeostasis and disease," *Bone Research*, vol. 4, no. 1, Article ID 16009, 2016.
- [133] C. Qi, X. Yan, C. Huang, A. Melerzanov, and Y. Du, "Biomaterials as carrier, barrier and reactor for cell-based regenerative medicine," *Protein & cell*, vol. 6, no. 9, pp. 638–653, 2015.
- [134] X. Fu, G. Liu, A. Halim, Y. Ju, Q. Luo, and G. Song, "Mesenchymal stem cell migration and tissue repair," *Cells*, vol. 8, no. 8, p. 784, 2019.
- [135] A. Trounson and C. McDonald, "Stem cell therapies in clinical trials: progress and challenges," *Cell Stem Cell*, vol. 17, no. 1, pp. 11–22, 2015.
- [136] H. T. Temple and T. I. Malinin, "Orthobiologics in the foot and ankle," *Foot and Ankle Clinics*, vol. 21, no. 4, pp. 809–823, 2016.
- [137] B.-M. Seo, M. Miura, S. Gronthos et al., "Investigation of multipotent postnatal stem cells from human periodontal ligament," *The Lancet*, vol. 364, no. 9429, pp. 149–155, 2004.
- [138] A. Queiroz, E. Albuquerque-Souza, L. M. Gasparoni et al., "Therapeutic potential of periodontal ligament stem cells," *World Journal of Stem Cells*, vol. 13, no. 6, pp. 605–618, 2021.
- [139] W. V. Giannobile, C. S. Lee, M. P. Tomala, K. M. Tejada, and Z. Zhu, "Platelet-derived growth factor (PDGF) gene delivery for application in periodontal tissue engineering," *Journal of Periodontology*, vol. 72, no. 6, pp. 815–823, 2001.
- [140] Q. Jin, O. Anusaksathien, S. A. Webb, M. A. Printz, and W. V. Giannobile, "Engineering of tooth-supporting structures by delivery of PDGF gene therapy vectors," *Molecular Therapy*, vol. 9, no. 4, pp. 519–526, 2004.
- [141] Q.-M. Jin, O. Anusaksathien, S. A. Webb, R. B. Rutherford, and W. V. Giannobile, "Gene therapy of bone morphogenetic protein for periodontal tissue engineering," *Journal of Periodontology*, vol. 74, no. 2, pp. 202–213, 2003.
- [142] M. Tatullo, B. Marrelli, M. J. Zullo et al., "Exosomes from human periapical cyst-MSCs: theranostic application in Parkinson's disease," *International Journal of Medical Sciences*, vol. 17, no. 5, pp. 657–663, 2020.
- [143] B. Codispoti, M. Marrelli, F. Paduano, and M. Tatullo, "NANOmetric BIO-banked MSC-derived exosome (NANOBIOME) as a novel approach to regenerative medicine," *Journal of Clinical Medicine*, vol. 7, no. 10, p. 357, 2018.
- [144] S. K. Sze, D. P. V. de Kleijn, R. C. Lai et al., "Elucidating the secretion proteome of human embryonic stem cell-derived mesenchymal stem cells," *Molecular & Cellular Proteomics*, vol. 6, no. 10, pp. 1680–1689, 2007.
- [145] S. Zhang, W. C. Chu, R. C. Lai, S. K. Lim, J. H. P. Hui, and W. S. Toh, "Exosomes derived from human embryonic mesenchymal stem cells promote osteochondral regeneration," *Osteoarthritis and Cartilage*, vol. 24, no. 12, pp. 2135–2140, 2016.
- [146] M. Tatullo, F. Genovese, E. Aiello et al., "Phosphorene is the new graphene in biomedical applications," *Materials*, vol. 12, no. 14, p. 2301, 2019.
- [147] J. A. Inzana, D. Olvera, S. M. Fuller et al., "3D printing of composite calcium phosphate and collagen scaffolds for bone regeneration," *Biomaterials*, vol. 35, no. 13, pp. 4026–4034, 2014.
- [148] S. Comber, M. Gardner, K. Georges, D. Blackwood, and D. Gilmour, "Domestic source of phosphorus to sewage treatment works," *Environmental Technology*, vol. 34, no. 10, pp. 1349–1358, 2013.
- [149] B. Yang, J. Yin, Y. Chen et al., "2D-Black-Phosphorus-Reinforced 3D-printed scaffolds: A stepwise countermeasure for osteosarcoma," *Advanced Materials*, vol. 30, no. 10, Article ID 1705611, 2018.
- [150] K. Huang, J. Wu, and Z. Gu, "Black phosphorus hydrogel scaffolds enhance bone regeneration via a sustained supply of calcium-free phosphorus," *ACS Applied Materials & Interfaces*, vol. 11, no. 3, pp. 2908–2916, 2018.
- [151] M. Tatullo, B. Zavan, F. Genovese et al., "Borophene is a promising 2D allotropic material for biomedical devices," *Applied Sciences*, vol. 9, no. 17, p. 3446, 2019.
- [152] D. Li, J. He, G. Ding et al., "Stretch-driven increase in ultrahigh thermal conductance of hydrogenated borophene and dimensionality crossover in phonon transmission," *Advanced Functional Materials*, vol. 28, no. 31, Article ID 1801685, 2018.
- [153] J. Jung, J. S. Park, M. Dard, B. Al-Nawas, and Y.-D. Kwon, "Effect of enamel matrix derivative liquid combined with synthetic bone substitute on bone regeneration in a rabbit calvarial model," *Clinical Oral Investigations*, vol. 25, no. 2, pp. 547–554, 2021.