

Research Article

Evaluation of pH, Calcium Ion Release, and Dimensional Stability of an Experimental Silver Nanoparticle-Incorporated Calcium Silicate-Based Cement

Teena Sheethal Dsouza ,¹ Aditya Shetty ,¹ and Neevan Dsouza ,²

¹Department of Conservative Dentistry & Endodontics, AB Shetty Memorial Institute of Dental Sciences, NITTE (Deemed to be University), Mangalore, India

²Department of Humanities, KS Hegde Medical Academy, NITTE (Deemed to be University), Mangalore, India

Correspondence should be addressed to Aditya Shetty; shetty_aditya1@yahoo.co.in

Received 3 August 2021; Revised 17 November 2021; Accepted 24 November 2021; Published 3 December 2021

Academic Editor: Brajesh Kumar

Copyright © 2021 Teena Sheethal Dsouza et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

An experimental calcium silicate-based root-end filling material incorporated with silver nanoparticles intended for use in periapical surgeries was developed with the purpose to overcome the drawbacks of existing materials and to satisfy the ideal requirements of root-end filling materials. This study was designed to evaluate the physicochemical properties, pH, calcium ion release, and dimensional stability of the experimental cement, and compare the results with commercially available ProRoot MTA (Dentsply). An independent sample test was used to analyze the data. Mean initial pH (immediately after mixing) of the experimental cement was 10.42 ± 0.04 which was higher than that of MTA. However, there was a significant increase in pH of MTA at 1 day, 2 days, and 7 days. Presence of calcium chloride favored the release of calcium ions which was significantly increased in the experimental group at 24 hours. At the end of 30 days, MTA showed a significant expansion when compared to the experimental cement ($p < 0.001$). In conclusion, the experimental nanoparticle-incorporated calcium silicate-based cement showed clinically acceptable physicochemical properties.

1. Introduction

The success rate of a conventional root canal treatment is very high and nearly approaching 95%. On occasional re-infections, nonsurgical retreatment is performed. However, when there is failure of nonsurgical root canal treatment or when retreatment is not advised, treatment by a surgical approach involving root-end resection, retrograde cavity preparation, and filling is indicated to obtain a good apical seal and resolve persistent infections [1, 2].

The introduction of calcium silicate-based materials in endodontics has been an explicit discovery for use in blood and moisture-field periapical surgeries. With the advent of mineral trioxide aggregate in 1993 by Torabinejad, several calcium silicate- and bioceramic-based materials have been marketed. Bioceramic sealers such as EndoSequence BC

Sealer, EndoSeal MTA, and MTA Fillapex have been appreciated for their favorable physicobiological properties [3, 4]. Calcium silicate-based cements have been developed as root-end filling materials mainly due to its hydraulic property which allows it to set even when in contact with tissue fluid and blood [5, 6]. Calcium silicate cements such as mineral trioxide aggregate (MTA) and other Portland-based cements have also shown promising results regarding biocompatibility and sealing ability [7]. Another notable feature and a booming interest in the potential clinical applications of hydraulic calcium silicate cements is their bioactivity that influences the surrounding environment [8, 9]. Shokouhinejad et al. proved that ProRoot MTA and EndoSequence root repair material were bioactive with increased effect over time [10]. A research study conducted by Gandolfi et al. in 2017 explored the ability of calcium silicate-based cements to

enable biominerilization, with an unmediated bond between the surface of these materials and the mineralized bone matrix [11].

However, due to the inherent drawbacks in these cements including high cost, slower setting, and limited antibacterial activity, new additives have been added to the existing commercially available materials. Selected accelerants such as calcium chloride, calcium nitrate, and calcium formate have been added to improve the physical properties of MTA [2, 12, 13]. In an attempt to increase the bioactivity and early formation of fluorapatite, Gandolfi et al., in 2011, introduced doping of fluorides in calcium silicate cements [14]. Zamparini et al. added tantalum pentoxide and zirconium oxide into bioceramics to enhance apatite-forming ability with fulfillment of the desired chemical and physical standards [15]. Antimicrobial agents such as silver and gold nanoparticles and titanium dioxide nanoparticles have been used to inhibit bacterial growth [16, 17]. Samiei et al. incorporated 1 wt% silver nanoparticles to MTA in an attempt to improve the antibacterial activity with desirable results [18].

Nonetheless, there are limited studies in the literature that reveal any adverse changes in the chemical properties of calcium silicate-based cements when these accelerants and nanoparticles are added. One characteristic whose change would be of concern is the dimensional stability because it is likely related to its ability to seal the root-end [12]. Another attribute to deal with is the alkalinity of the calcium silicate-based cements. Alteration in pH might cause changes in the antibacterial activity and the release of calcium ions. Calcium ions are desirable for hard tissue formation [12]. Neither scenario would be clinically acceptable. The purpose of this study was to evaluate the influence of adding calcium chloride and silver nanoparticles in terms of pH, dimensional stability, and calcium ion release on a low-temperature production-based experimental calcium silicate-based cement.

2. Materials and Methods

A series of experiments were initially performed to determine the most appropriate concentrations of the ingredients of Portland cement that best improved cohesiveness and workability. The components included 60% calcium oxide, 20% silicon dioxide, and 9% aluminium oxide. Other additives included bismuth oxide as a radiopacifier and 1 wt% silver nanoparticles (silver nanopowder, Type 2, APS: 20 nm, SRL Chemical) for antimicrobial action. 10% calcium chloride solution was used as the liquid component to accelerate the setting time. This was referred to as the experimental cement.

The powder components were mixed using the solid-state reaction technique using ethanol as a solvent. The resultant powder was further subjected to sintering using a hot air oven at 100°C for 24 hours. The final powder was passed through a 250-micron mesh to attain a desired homogenous powdered mixture. The experimental cement was subjected to further analysis. A pilot study was conducted to test the antibacterial activity using agar diffusion test on

E. faecalis which exhibited a zone of inhibition of 3-4 mm. The accelerated setting of the cement was evaluated by preliminary tests using a Gillmore needle apparatus which confirmed a shorter setting time of 6-7 min as reported by Dsouza et al. [19]. On comparison, ProRoot MTA showed a delayed setting time of 170 min. Elemental analysis was performed, and powder microstructure was evaluated to confirm the presence of oxides and silver nanoparticles in the final powder using energy-dispersive analysis by X-rays (EDAX) in the scanning electron microscope (Zeiss Gemini).

2.1. Groups

Group 1: experimental cement

Group 2: mineral trioxide aggregate (ProRoot MTA, Dentsply Sirona)

2.2. pH Measurement. Polyethylene tubes (3 mm in diameter and 1 mm length) were used. The powder (0.2 g) and liquid (0.14 mL) were mixed to get a desired putty consistency. After filling the tubes with the material, they were immersed in containers containing 10 mL of deionized water, and initial pH was immediately noted. Subsequent readings were made after 1 day, 2 days, and 7 days. A digital pH meter (Merck Life Science Pvt Ltd.) was used for pH measurement. The apparatus was previously calibrated with pH 7.0 ± 0.05 buffer capsules. Between each measurement, the electrode was washed with ultrapure water and blot-dried. Initial pH of deionized water was measured before the immersion of the materials. A similar procedure was followed to determine pH of ProRoot MTA (Dentsply).

2.3. Calcium Ion Release Analysis. Polyethylene tubes with 10 mm length and 1 mm internal diameter were used. The filled tubes with the experimental cement were immersed in 10 mL of deionized water. The calcium ion release profile from the cement samples was recorded at day 1 and day 7 using an atomic absorption spectrophotometer (GBC 932 Plus), equipped with a hollow calcium cathode lamp, according to the following operative conditions: lamp current 3 mA; fuel nitrous oxide; support oxygen; stoichiometry reducing; wavelength 422.7 nm; slit 02 nm. The results were estimated by using the equation of standard curve line, which was determined by the measurement of solutions with known concentrations. The calcium ion release of ProRoot MTA was also determined.

2.4. Test of Dimensional Stability. This test was based on ISO 6876:7.6 with split molds (height 6 mm and width 3 mm). After mixing, the samples were transferred to the molds until slightly overfilled. Filled molds were flattened on each end, wrapped in a water-moistened gauze, and placed in a 37°C incubator with humidity maintained at 95%. After 48 hours, the samples were drawn across fresh 600-grit sandpaper to flatten their ends. The samples were removed from the molds, their lengths were measured using a digital caliper

with a resolution of 0.01 mm, and they were stored in distilled water at 37°C. After 30 days, their lengths were measured again. The change in length during a period of 30 days divided by the original length was our measure of dimensional stability. The test was repeated 3 times. In accordance with the ISO and ANSI/ADA, the results must not exceed 1.0% of contraction or 0.1% of expansion.

3. Results

3.1. EDAX SEM Analysis. Elemental analysis confirmed the presence of oxides of the powders with pronounced peaks for calcium, silicon, and oxygen. Powder microstructure of the unhydrated cement showed coarse and irregular particles under energy-dispersive analysis by X-rays (EDAX) in the scanning electron microscope (Figure 1).

3.2. Evaluation of pH and Calcium Ion Release. Throughout the experimental period, the results of pH measurements of both cements were alkaline. Independent sample test showed a statistically significant difference which was observed in immediate pH as well as pH at 1 day, 2 days, and 7 days between the two groups. Mean pH of the experimental group in the immediate period was 10.42 ± 0.04 which was higher than MTA (group 2). However, there was a significant increase in pH of group 2 (MTA) at 1 day, 2 days, and 7 days compared to group 1 ($p < 0.05$) (Table 1).

Presence of calcium chloride favored the release of calcium ions which was significantly increased in group 1 at 24 hours ($p < 0.001$). However, no significant difference was noted at 7 days between group 1 and group 2 ($p = 0.698$) (Table 2).

3.3. Evaluation of Dimensional Stability. At the end of 30 days, group 2 showed a significant expansion when compared to group 1 ($p < 0.001$) (Table 3).

4. Discussion

Hydraulic calcium silicate cements have been a boon to the clinicians as alternative biomaterials for dentine replacement, in vital pulp therapy, apexogenesis and apexification, root perforation repair and resorptive defects, root canal, and root-end restorations [8, 15].

This study evaluated the physicochemical properties of a low-temperature fabricated calcium silicate-based cement. There were pronounced peaks of calcium and silicon in the EDAX analysis which were similar to the study results conducted by Gandolfi et al. They confirmed that calcium silicate materials, i.e., ProRoot MTA, MTA Plus, and Bio-dentine, also showed an increase in calcium and phosphorus that was necessary for biomineralization [11]. Concordance to our study, Shokouhinejad et al. analyzed the elemental composition of calcium silicate-based EndoSequence root repair material, BioAggregate, and ProRoot MTA with display of high peaks for calcium [10].

On physicochemical evaluation, the addition of calcium chloride and silver nanoparticles to the experimental

calcium silicate-based cement showed favorable results. In terms of pH, alkaline pH was maintained throughout the study period. The rich concentration of calcium oxide is converted into calcium hydroxide in an aqueous solution. Calcium hydroxide dissociates into calcium and hydroxyl ions, thus increasing pH of the solution and thus contributing to antibacterial action [20]. Alkaline pH is required as it has proven to offer osteogenic potential, biocompatibility, and antibacterial ability [4]. Initial pH after mixing was similar to that of MTA. However, variations in pH were seen at different time intervals between the experimental cement and MTA. At the end of 7 days, pH was reduced in the experimental group, but alkalinity was still maintained. The results of MTA were in accordance to a long-term study conducted by Fridland and Rosado who reported that usually reported pH of 12-13 may slightly decrease over time [21]. Thus, an additional ingredient of imparting an antibacterial action would be necessary to maintain a bacteria-free environment. This purpose was fulfilled by adding silver nanoparticles, the results of which were validated during the pilot study. Samiei et al. assessed the antimicrobial activity of MTA with silver nanoparticles with favorable results [18]. Silver nanoparticles (Ag NPs) are one of the most widely used nanoparticles, most notably serving as an antimicrobial agent for medical applications [22, 23]. Small-sized Ag NPs can inhibit the growth of nitrifying bacteria more than that by silver ions at the same total silver concentrations [24, 25]. The size of the particle was also related to antimicrobial activity; the smaller particles give more bactericidal effects compared to larger particles [26-28]. Gomes-Filho et al. reported that Ag NP dispersion was biocompatible, mainly at low concentrations [29].

Additional calcium incorporated into the experimental cement via the calcium chloride liquid boosted the calcium ion release. These findings further led to the assumption with the study results by Takita et al. that the continuous release of calcium ions is required for the proliferation of human dental pulp cells in calcium silicate-based cements [30]. A known fact is the formation of an interfacial calcium phosphate layer that forms a chemical bond between calcium silicate-based materials and walls of dentin [3]. Also, these are placed directly in contact with periradicular tissues; therefore, an adequate response of bioactivity is much desired [14]. The amount of calcium ions released from the experimental cement was enhanced in 24 hours and almost similar to that of MTA after 7 days. Calcium chloride alone has low pH of 4.4, and its addition to experimental cement did alter pH. However, it did not seem to alter significantly the calcium ion release of the cement. This is in accordance with the findings reported by Antunes Bortoluzzi et al. which revealed that the addition of calcium chloride improved the calcium release properties of commercially available MTA cements [31]. Contradictory to the pH results of this study, Vazquez-Garcia et al. evaluated the radiopacity, setting time, pH, compressive strength, and solubility of Portland cement incorporated with silver nanoparticles and concluded that the addition of Ag NPs to PC/ZrO₂ maintained pH, lowered the solubility, and increased the setting time and compressive strength [32].

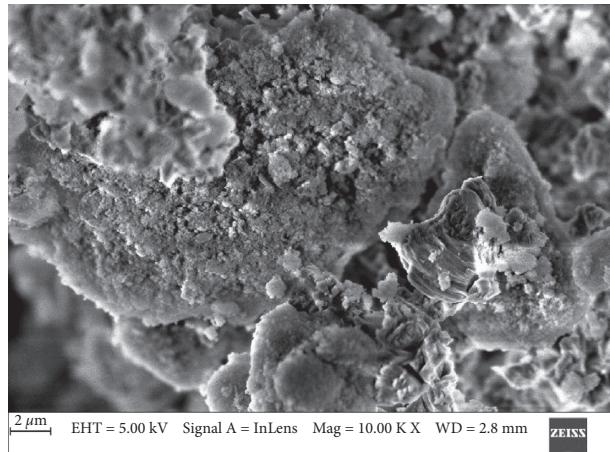


FIGURE 1: Powder microstructure under the EDAX SEM.

TABLE 1: Comparison of pH analysis between the groups.

	Mean ± SD		<i>p</i> value	95% confidence interval of the difference	
	Group 1	Group 2		Lower	Upper
Immediate	10.42 ± 0.04	10.27 ± 0.06	0.001	0.12	0.19
24 hours	10.29 ± 0.06	11.88 ± 0.12	<0.001	-1.66	-1.52
48 hours	9.78 ± 0.11	11.47 ± 0.45	<0.001	-1.93	-1.44
7 days	8.90 ± 0.18	9.82 ± 0.05	<0.001	-1.01	-0.81

TABLE 2: Comparison of calcium ion release between the groups.

	Mean ± SD		<i>p</i> value	95% confidence interval of the difference	
	Group 1	Group 2		Lower	Upper
24 hours	15.92 ± 2.37	11.02 ± 0.92	<0.001	3.56	6.23
7 days	17.71 ± 5.91	18.30 ± 0.35	0.698	-3.73	2.56

TABLE 3: The dimensional stability (%) of the tested sealers at 30 days (mean ± standard deviation).

	Mean ± SD	
	Group 1	Group 2
Baseline (0 th day)	6.00 ± 0.00	6.00 ± 0.00
30 days	5.99 ± 0.06	6.00 ± 0.04

The dimensional stability of the experimental cement was favorable. In accordance with the ISO and ANSI/ADA, the results must not exceed 1.0% of contraction or 0.1% of expansion. The experimental cement did not exceed 1% of contraction than the original dimension; however, MTA showed slight expansion which was also acceptable. In an *in vitro* dye leakage study conducted by Bortoluzzi et al., calcium chloride actually improved the sealing ability of MTA placed as a root-end filling material when compared with the control [33]. The dimensional stability of samples set with calcium chloride in this study supports the study results of Bortoluzzi et al. in that they exhibited no contraction. In another bacterial leakage study conducted by Dsouza et al. in 2016, incorporation of calcium chloride to

MTA as a substitute for distilled water showed better sealing ability than when mixed with distilled water, chlorhexidine, and doxycycline [2]. The favorable dimensional change in this experiment may be related to the lower porosity of the cement and due to the occupancy of the silver nanoparticles into the empty spaces of the cement matrix [32].

5. Conclusion

In conclusion, the experimental calcium silicate-based cement with the incorporation of silver nanoparticles and calcium chloride had favorable physicochemical properties that can be useful to maintain a bacteria-free environment and for improved healing, which are necessary in root-end

surgeries. Further investigations are indicated to assess the biological properties and the long-term sealing ability.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors wish to acknowledge the Department of Oral Pathology, AB Shetty Memorial Institute of Dental Sciences, Nitte (Deemed-to-be-University), for the assistance with the pH measurements. They also acknowledge the Department of Chemical Engineering, National Institute of Technology, Karnataka, for assisting in the calcium ion release analysis and Mr. Murari, Mangalore University, for carrying out the EDAX SEM analysis.

References

- [1] S. Shahi and S. Rahimi, "Sealing ability of white and gray mineral trioxide aggregate mixed with distilled water and 0.12% chlorhexidine gluconate when used a root-end filling materials," *Journal of Endodontics*, vol. 33, no. 12, pp. 1429–1432, 2007.
- [2] T. Dsouza, S. Bhandary, and R. Srinivasan, "An invitro comparative evaluation of the sealing ability of Mineral Trioxide Aggregate with different mixing reagents," *World Journal of Dentistry*, vol. 7, no. 2, pp. 54–58, 2016.
- [3] G. T. D. M. Candeiro, F. C. Correia, M. A. H. Duarte, D. C. Ribeiro-Siqueira, and G. Gavini, "Evaluation of radiopacity, pH, release of calcium ions, and flow of a bioceramic root canal sealer," *Journal of Endodontics*, vol. 38, no. 6, pp. 842–845, 2012.
- [4] J. K. Lee, S. W. Kwak, J. H. Ha, W. Lee, and H. C. Kim, "Physicochemical properties of epoxy resin-based and bioceramic-based root canal sealers," *Bioinorganic Chemistry and Applications*, vol. 2017, Article ID 2582849, 8 pages, 2017.
- [5] J. Camilleri, "Characterization and hydration kinetics of tricalcium silicate cement for use as a dental biomaterial," *Dental Materials*, vol. 27, no. 8, pp. 836–844, 2011.
- [6] J. Camilleri, L. Formosa and D. Damidot, "The setting characteristics of MTA Plus in different environmental conditions," *International Endodontic Journal*, vol. 46, no. 9, pp. 1–10, 2013.
- [7] P. Yan, B. Peng, B. Fan, M. Fan, and Z. Bian, "The effects of sodium hypochlorite (5.25%), chlorhexidine (2%), and glyde file prep on the bond strength of MTA-Dentin," *Journal of Endodontics*, vol. 32, no. 1, pp. 58–60, 2006.
- [8] L.-N. Niu, K. Jiao, T.-D. Wang et al., "A review of the bioactivity of hydraulic calcium silicate cements," *Journal of Dentistry*, vol. 42, no. 5, pp. 517–533, 2014.
- [9] N. J. Coleman, K. Awosanya, and J. W. Nicholson, "Aspects of the in vitro bioactivity of hydraulic calcium (alumino)silicate cement," *Journal of Biomedical Materials Research Part A*, vol. 90A, no. 1, pp. 166–174, 2009.
- [10] N. Shokouhinejad, M. H. Nekoofar, H. Razmi et al., "Bioactivity of EndoSequence root repair material and bioaggregate," *International Endodontic Journal*, vol. 45, no. 12, pp. 1127–1134, 2012.
- [11] M. G. Gandolfi, G. Iezzi, A. Piattelli, C. Prati, and A. Scarano, "Osteoinductive potential and bone-bonding ability of Pro-Root MTA, MTA Plus and Biodentine in rabbit intra-medullary model: microchemical characterization and histological analysis," *Dental Materials*, vol. 33, no. 5, pp. e221–e238, 2017.
- [12] K. B. Wiltbank, S. A. Schwartz, and W. G. Schindler, "Effect of selected accelerants on the physical properties of Mineral Trioxide Aggregate and Portland cement," *Journal of Endodontics*, vol. 33, no. 10, pp. 1235–1238, 2007.
- [13] B.-N. Lee, Y.-C. Hwang, J.-H. Jang et al., "Improvement of the properties of Mineral Trioxide Aggregate by mixing with hydration accelerators," *Journal of Endodontics*, vol. 37, no. 10, pp. 1433–1436, 2011.
- [14] M. G. Gandolfi, P. Taddei, F. Siboni, E. Modena, M. P. Ginebra, and C. Prati, "Fluoride-containing nanoporous calcium-silicate MTA cements for endodontics and oral surgery: early fluorapatite formation in a phosphate-containing solution," *International Endodontic Journal*, vol. 44, no. 10, pp. 938–949, 2011.
- [15] F. Zamparini, F. Siboni, C. Prati, P. Taddei, and M. G. Gandolfi, "Properties of calcium silicate-monobasic calcium phosphate materials for endodontics containing tantalum pentoxide and zirconium oxide," *Clinical Oral Investigations*, vol. 23, no. 1, pp. 445–457, 2019.
- [16] T. Dsouza, A. Shetty, M. N. Hegde, J. E. Packayam, V. Radhakrishna, and A. D. Monteiro, "Biocompatibility of calcium silicate based cement incorporated with silver or gold nanoparticles- an invitro study," *Annals of Dental Specialty*, vol. 8, no. 2, pp. 62–66, 2020.
- [17] T. Dsouza, M. N. Hegde, V. Radhakrishna, N. Dsouza, and S. Kumari, "In vitro cytotoxic evaluation of Mineral Trioxide Aggregate with silver and titanium dioxide nanoparticles," *World Journal of Dentistry*, vol. 10, no. 6, 2019.
- [18] M. Samiei, M. Aghazadeh, M. Lotfi, S. Shakoei, Z. Aghazadeh, and S. M. Vahid Pakdel, "Antimicrobial efficacy of mineral trioxide aggregate with and without silver nanoparticles," *Iranian Endodontic Journal*, vol. 8, no. 4, pp. 166–170, 2013.
- [19] T. S. Dsouza, A. Shetty, M. N. Hegde, J. E. Packayam, and A. D. Monteiro, "EDAX and FTIR characterization and setting time of an experimental nanoparticle incorporated root-end filling material," *E-Journal of Surface Science and Nanotechnology*, vol. 18, pp. 289–292, 2020.
- [20] J. L. Gonsalves, R. Viapiana, C. E. Miranda, A. H. Borges, and A. M. Filho, "Evaluation of physicochemical properties of Portland cements and MTA," *Brazilian Oral Research*, vol. 24, no. 3, pp. 277–283, 2010.
- [21] M. Fridland and R. Rosado, "MTA solubility: a long term study," *Journal of Endodontics*, vol. 31, no. 5, pp. 376–379, 2005.
- [22] I. Sondi and B. Salopek-Sondi, "Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria," *Journal of Colloid and Interface Science*, vol. 275, no. 1, pp. 177–182, 2004.
- [23] E. Navarro, F. Piccapietra, B. Wagner et al., "Toxicity of silver nanoparticles to *Chlamydomonas reinhardtii*," *Environmental Science & Technology*, vol. 42, no. 23, pp. 8959–8964, 2008.
- [24] O. Choi, K. K. Deng, N.-J. Kim, L. Ross Jr., R. Y. Surampalli, and Z. Hu, "The inhibitory effects of silver nanoparticles, silver ions, and silver chloride colloids on microbial growth," *Water Research*, vol. 42, no. 12, pp. 3066–3074, 2008.

- [25] O. Choi and Z. Hu, "Size dependent and reactive oxygen species related nanosilver toxicity to nitrifying bacteria," *Environmental Science & Technology*, vol. 42, no. 12, pp. 4583–4588, 2008.
- [26] D. R. Monteiro, L. F. Gorup, A. S. Takamiya, A. C. Ruvollo-Filho, E. R. D. Camargo, and D. B. Barbosa, "The growing importance of materials that prevent microbial adhesion: antimicrobial effect of medical devices containing silver," *International Journal of Antimicrobial Agents*, vol. 34, no. 2, pp. 103–110, 2009.
- [27] C. Baker, A. Pradhan, L. Pakstis, D. Pochan, and S. I. Shah, "Synthesis and antibacterial properties of silver nanoparticles," *Journal of Nanoscience and Nanotechnology*, vol. 5, no. 2, pp. 244–249, 2005.
- [28] A. Panacek, L. Kvítek, R. Prucek et al., "Silver colloid nanoparticles: synthesis, characterization, and their antibacterial activity," *The Journal of Physical Chemistry B*, vol. 110, no. 33, pp. 16248–16253, 2006.
- [29] J. E. Gomes-Filho, F. O. Silva, S. Watanabe et al., "Tissue reaction to silver nanoparticles dispersion as an alternative irrigating solution," *Journal of Endodontics*, vol. 36, no. 10, pp. 1698–1702, 2010.
- [30] T. Takita, M. Hayashi, O. Takeichi et al., "Effect of mineral trioxide aggregate on proliferation of cultured human dental pulp cells," *International Endodontic Journal*, vol. 39, no. 5, pp. 415–422, 2006.
- [31] E. Antunes Bortoluzzi, N. Juarez Broon, M. Antonio Hungaro Duarte, A. de Oliveira Demarchi, and C. Monteiro Bramante, "The use of a setting accelerator and its effect on pH and calcium ion release of mineral trioxide aggregate and white Portland cement," *Journal of Endodontics*, vol. 32, no. 12, pp. 1194–1197, 2006.
- [32] F. Vazquez-Garcia, M. Tanomaru-Filho, M. Tanomaru-Filho et al., "Effect of silver nanoparticles on physicochemical and antibacterial properties of calcium silicate cements," *Brazilian Dental Journal*, vol. 27, no. 5, pp. 508–514, 2016.
- [33] E. A. Bortoluzzi, N. J. Broon, C. M. Bramante, R. B. Garcia, I. G. de Moraes, and N. Bernardineli, "Sealing ability of MTA and radiopaque Portland cement with or without calcium chloride for root-end filling," *Journal of Endodontics*, vol. 32, no. 9, pp. 897–900, 2006.