## Research Article

# Integrative Decomposition Procedure and Kappa Statistics for the Distinguished Single Molecular Network Construction and Analysis

## Lin Wang,<sup>1</sup> Ying Sun,<sup>1</sup> Minghu Jiang,<sup>2</sup> and Xiguang Zheng<sup>1</sup>

<sup>1</sup> Biomedical Center, School of Electronics Engineering, Beijing University of Posts and Telecommunications, Beijing 100876, China <sup>2</sup> Lab of Computational Linguistics, Tsinghua University, Beijing 100084, China

Correspondence should be addressed to Lin Wang, wanglin98@tsinghua.org.cn

Received 27 October 2008; Accepted 19 February 2009

Recommended by Zhenqiu Liu

Our method concentrates on and constructs the distinguished single gene network. An integrated method was proposed based on linear programming and a decomposition procedure with integrated analysis of the significant function cluster using Kappa statistics and fuzzy heuristic clustering. We tested this method to identify ATF2 regulatory network module using data of 45 samples from the same GEO dataset. The results demonstrate the effectiveness of such integrated way in terms of developing novel prognostic markers and therapeutic targets.

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## 1. Introduction

In the postgenomic era, with microarray technologies producing great deal of gene expression data, mining these data to get insight into biological processes at system-wide level has become a challenge for bioinformatics. On one hand, due to the complex and distribute nature of biological research, there is a great deal of methods for inferring gene regulatory networks. But all these methods focused on constructing the complicated entire network calculated from the given microarray data. The tremendous amounts of genes in those networks distribute analysts' attention, so it is hard to get any clear perception of valuable knowledge from such complicated networks, let alone further study of each single gene. On the other hand, the wide spread of knowledge over independent databases aggravates the hardness of integrating comprehensive annotation information for genes and lowers the study effectiveness. Thus, a novel method integrating both single molecular network construction and highly centralized gene-functional-annotation analysis is in demand for gene network and functional analysis.

This paper proposed an integrated method based on linear programming and a decomposition procedure with integrated analysis of the significant function cluster using Kappa statistics and fuzzy heuristic clustering. Our method concentrates on and constructs the distinguished single gene network integrated with function prediction analysis by DAVID. For the distinguished single molecular network, we did (1) control and experiment comparison, (2) identification of activation and inhibition networks, (3) construction of upstream and downstream feedback networks, and (4) functional module construction. We tested this method to identify ATF2 regulation network module using data of 45 samples from one and the same GEO dataset. The results demonstrate the effectiveness of such integrated way in terms of developing novel prognostic markers and therapeutic targets.

## 2. Methods

2.1. Distinguished Single Molecular Network Construction. The entire network was constructed using GRNInfer [1] and GVedit tools. GRNInfer is a novel mathematic method called gene network reconstruction (GNR) tool based on linear programming and a decomposition procedure that is used for inferring gene networks. The method theoretically ensures the derivation of the most consistent network structure with respect to all of the datasets, thereby not only significantly alleviating the problem of data scarcity but also remarkably improving the reconstruction reliability. The general solution for a single dataset is the following (1), which represents all of the possible networks:

$$J = (X' - A)U\Lambda^{-1}V^{T} + YV^{T} = \hat{J} + YV^{T},$$
(1)

where  $J = (J_{ij})_{n \times n} = \partial f(x)/\partial x$  is an  $n \times n$  Jacobian matrix or connectivity matrix,  $X = (x(t_1), \dots, x(t_m))$ ,  $A = (a(t_1), \dots, a(t_m))$ , and  $X' = (x'(t_1), \dots, x'(t_m))$  are all  $n \times m$  matrices with  $x'_i(t_j) = [x_i(t_{j+1}) - x_i(t_j)]/[t_{j+1} - t_j]$  for  $i = 1, \dots, n; j = 1, \dots, m$ .  $X(t) = (x_1(t), \dots, x_n(t))^T \in \mathbb{R}^n$ ,  $a = (a_1, \dots, a_n)^T \in \mathbb{R}^n$ ,  $x_i(t)$  is the expression level (mRNA concentrations) of gene *i* at time instance *t*.  $y = (y_{ij})$  is an  $n \times n$  matrix, where  $y_{ij}$  is zero if  $e_j \neq 0$  and is otherwise an arbitrary scalar coefficient.  $\wedge^{-1} = \text{diag}(1/e_i)$  and 1/e is set to be zero if  $e_i = 0$ . *U* is a unitary  $m \times n$  matrix of left eigenvectors,  $\wedge = \text{diag}(e_1, \dots, e_n)$  is a diagonal  $n \times n$  matrix containing the *n* eigenvalues, and  $V^T$  is the transpose of a unitary  $n \times n$  matrix of right eigenvectors.

But the entire network is too complex to get any clear perception of such complicated relationships among those genes, let alone further study of each single gene. We constructed the distinguished single molecular network by selecting the centered gene and its directly related genes based on the entire network for further study. We take into account the effectiveness of biology study in order to concentrate on single molecular network rather than the intricate entire network. It is helpful to get intensive and deep insight of the whole network. For the distinguished single molecular network, we did (1) control and experiment comparison, (2) identification of activation and inhibition networks, (3) construction of upstream and downstream feedback networks, and (4) functional module construction.

2.2. Functional Annotation Clustering. For the function of genes that is neither determined by their sequence nor by the protein families they belong to [2], the function of those genes included in the same single molecular network should not be interpreted separately, but should be analyzed together according to the whole single molecular network. This method takes into account the network nature of biological annotation contents in order to concentrate on the larger biological picture rather than an individual gene. We used DAVID to do functional annotation clustering. It changes functional annotation analysis from term- or genecentric to biological module-centric [2] in accordance with our network analysis aim.

The DAVID gene functional clustering tool provides typical batch annotation and gene-GO term enrichment analysis for highly throughput genes by classifying them into gene groups based on their annotation term co-occurrence [3]. DAVID uses a novel algorithm to measure relationships among the annotation terms based on the degrees of their coassociation genes to group similar annotation contents from the same or different resources into annotation groups. The grouping algorithm is based on the hypothesis that similar annotations should have similar gene members. The functional annotation clustering integrates the same techniques of Kappa statistics to measure the degree of the common genes between two annotations, and fuzzy heuristic clustering to classify the groups of similar annotations according kappa values [4, 5]. The tool also allows observation of the internal relationships of the clustered terms by comparing it to the typical linear, redundant term report, over which similar annotation terms may be distributed among many other terms.

#### 3. Results and Discussion

We tested this method using microarrays containing 22215 genes in 40 MPM tumors and 5 normal pleural tissues from one and the same GEO datasets. We identified potential tumor molecular markers and chose the top 51 significant positive genes with normalization of log2, the minimum fold change = 3.5, delta = 1.59, and a false-discovery rate of 0% using SAM [6]. We selected activating transcription factor (ATF)-2 because it is one of the most distinguished genes in MPM. It is a member of the ATF/cyclic AMP-responsive element binding protein family of transcription factors.

3.1. Normal Tissues and Tumor Comparisons of Distinguished Single Molecular Network. We, respectively, constructed the interaction network of the above 51 genes in healthy tissues and that in tumor using GRNInfer [1] and GVedit tools and selected the ATF2-centered downstream subnetworks. With comparison of these ATF2-centered subnetworks, we can get a more clear perception of the notable differences between normal tissues and tumor, as shown in Figure 1. It appeared that ATF2 inhibits C11orf9, C18orf10, C20orf31, CALD1, CAMK2G, DDX3X, FALZ, GLS, GOLGA2, ID2, NME2, NMU, NONO, PAWR, PLOD2, PSMF1, RBMS1, RIC8A, RNF10, TEAD4, TIA1, TNPO1, unknown2, unknown3, WBSCR20C, and ZF in normal tissues, as shown in Figure 1(a). It appeared that ATF2 inhibits C11orf9, C15orf5, C18orf10, C20orf31, CAMK2G, CDR2, DDX3X, FALZ, FLJ10707, GLS, GOLGA2, ID2, KRT18, LRRC1, NME2, NMU, NONO, NSUN5, OBSL1\_2, PLOD2, PLXNA1, PTOV1, RBMS1, RIC8A, RNASEH1, RNF10, TEAD4, TIA1, UCK2, USP11, and ZF, while it activates CALD1 and TFAP2C in tumor, as shown in Figure 1(b).

With comparison between the two results, notable differences can be shown clearly in order to get further perception of pathological changes in MPM. For example, ATF2 target genes appeared in ATF2 activation to CALD1, TFAP2C in MPM, as only shown in Figure 2(b). Caldesmon (CALD1) is a potential actomyosin regulatory protein found in smooth muscle and nonmuscle cells [7]. Transcription factor AP2gamma (TFAP2C) is alternatively titled AP2. Families of related transcription factors are often expressed in the same cell lineages but at different times or sites in the developing embryo. The AP2 family appears to regulate the expression of genes required for development of tissues of ectodermal origin such as neural crest and skin [8]. AP2 may also be Journal of Biomedicine and Biotechnology

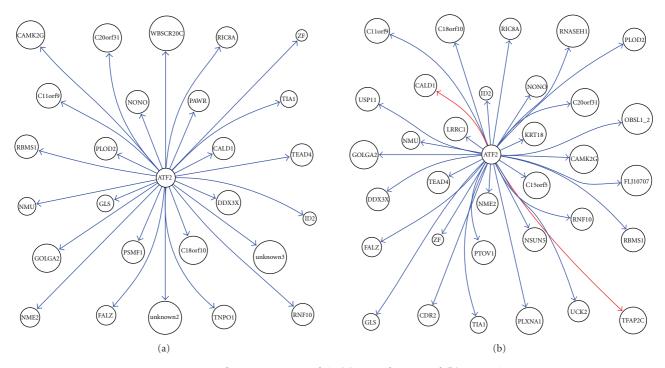


FIGURE 1: ATF2 downstream network in (a) normal tissue and (b) MPM tissue.

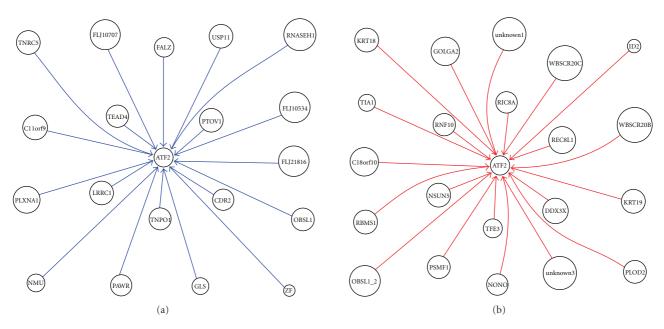


FIGURE 2: (a) ATF2 upstream inhibition network of MPM; (b) ATF2 upstream activation network of MPM.

involved in the overexpression of c-erbB-2 in human breast cancer cells [9].

3.2. Identification of Activation and Inhibition Networks for the Distinguished Single Molecule. We also identified the activation and inhibition networks, respectively, in order to simplify and intensify the analysis process. For example, in ATF2 upstream network of MPM, as shown in Figure 2, it appeared that C110rf9, CDR2, FALZ, FLJ10534, FLJ10707, FLJ21816, GLS, LRRC1, NMU, OBSL1, PAWR, PLXNA1, PTOV1, RNASEH1, TEAD4, TNPO1, TNRC5, USP11, and ZF inhibit ATF2, as shown in Figure 2(a), whereas C18orf10, DDX3X, GOLGA2, ID2, KRT18, KRT19, NONO, NSUN5, OBSL1\_2, PLOD2, PSMF1, RBMS1, REC8L1, RIC8A, RNF10, TFE3, TIA1, unknown1, unknown3, WBSCR20B, and WBSCR20C activate ATF2, as shown in Figure 2(b).

ATF2 upstream genes TFE3, REC8L1 showed activation to ATF2. TFE3 is a member of the helix-loop-helix family

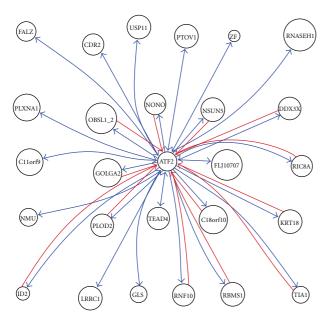


FIGURE 3: ATF2 feedback subnetwork of MPM.

of transcription factors and binds to the mu-E3 motif of the immunoglobulin heavy-chain enhancer and is expressed in many cell types [10]. Nakagawa et al. [11] identified TFE3 as a transactivator of metabolic genes that are regulated through an E box in their promoters which led to metabolic consequences such as activation of glycogen and protein synthesis, but not lipogenesis, in liver [11]. REC8L1 is the human homolog of yeast Rec8, a meiosis-specific phosphoprotein involved in recombination events [12]. Brar et al. (2006) showed that phosphorylation of the cohesin subunit REC8 contributes to stepwise cohesin removal [13].

3.3. Constructing Feedback Network of the Distinguished Single Upstream and Downstream Gene. We took into account the feedback relationship and setup ATF2 feedback network, as shown in Figure 3. ATF2 target genes appeared in ATF2 inhibition to CDR2, GLS, and USP11, consistently, its upstream genes also appeared in CDR2, GLS, and USP11 inhibition to ATF2. CDR2 is also called CDR62, where CDR means cerebellar degeneration-related. On Western blot analysis of Purkinje cells and tumor tissue, the anti-Yo sera react with at least 2 antigens, a major species of 62 kD called CDR62 and a minor species of 34 kD called CDR34 [14]. Sahai (1983) demonstrated phosphate-activated glutaminase (GLS) in human platelets [15]. It is the major enzyme yielding glutamate from glutamine. Significance of the enzyme derives from its possible implication in behavior disturbances in which glutamate acts as a neurotransmitter [16]. USP11 is also called UHX1. Swanson et al. (1996) cited evidence indicating that ubiquitin hydrolases play a role in oncogenesis (oncogenes and tumor suppressor gene products are degraded in ubiquitin-dependent pathways) [17]. The relationship of ATF2 with CDR2, GLS, and USP11 represents a negative feedback loop.

	Ribonuclease h1
	RNA binding motif, single stranded interacting protei
	Prostate tumor overexpressed gene 1
	Non-pou domain containing, octamer-binding
	Chromosome 11 open reading frame 9
	Proteasome (prosome, macropain) inhibitor subunit 1 (PI31)
	TIA1 cytotoxic granule-associated RNA binding prot
	TEA domain family member 4
	Glutaminase
	Inhibitor of DNA binding 2, dominant negative helix-loop-helix pro.
	Ubiquitin specific peptidase 11
	Transportin 1
	PRKC, apoptosis, WT1, regulator
	Procollagen-lysine, 2-oxoglutarate 5-dioxygenas
	Transcription factor binding to IGHM enhance
	Activating transcription fact
Primary metabolic proccess Cellular metabolic proccess	

FIGURE 4: One ATF2 upstream gene metabolic network including RBMS1, RNASEH1, PTOV1, NONO, C11orf9, PSMF1, TIA1, TEAD4, GLS, ID2, USP11, TNPO1, PAWR, PLOD2, and TFE3.

Metabolic proccess

3.4. Functional Module Construction of the Distinguished Single Gene. According to ATF2 upstream network, we did DAVID analysis of function cluster, respectively. The DAVID functional annotation clustering results appeared that one ATF2 regulation network was identified as consisting of the ATF2 upstream genes including RBMS1, RNASEH1, PTOV1, NONO, C110rf9, PSMF1, TIA1, TEAD4, GLS, ID2, USP11, TNPO1, PAWR, PLOD2, and TFE3, as shown in Figure 4.

According to Figure 2, it appeared that RBMS1, NONO, PSMF1, TIA1, ID2, PLOD2, TFE3 activate ATF2; whereas RNASEH1, PTOV1, C11orf9, TEAD4, GLS, USP11, TNPO1, and PAWR inhibit ATF2.

RBMS1, NONO, TIA1, ID2, and TFE3 enhance nucleoside, nucleotide, and nucleic acid metabolism because RBMS1, NONO, TIA1, ID2, and TFE3 are involved in these metabolism; PSMF1 activation to ATF2 means the increase of Acyl-CoA metabolism and porphyrin metabolism; PLOD2 activation to ATF2 indicates the progress of cholesterol metabolism and other protein metabolism, as shown in Figure 5.

RNASEH1, PTOV1, and TEAD4 inhibition to ATF2 decreases nucleoside, nucleotide, and nucleic acid metabolism mediated by the three genes; C110rf9 inhibition to ATF2 means the decline of polysaccharide metabolism, whereas GLS represents the weakness of amino acid and cyclic nucleotides metabolism; USP11 inhibition to ATF2 indicates the fall-off in protein metabolism and modification, whereas PAWR in glycogen metabolism, as shown in Figure 5.

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RBMS1	rna binding motif, single stranded interacting protein 1 R	lelated genes	Homo sapiens
ANTHER_MF_ALL	MF00039: Other transcription factor, MF00042: Nucleic acid binding, MF00053: Other RNA-binding protein, MF00057: DNA topoisomoera	ase, MF00068: mRNA splicin	g factor, MF0007:
	Chromatin/Aromatin-binding protein, MF00076: Other nucleic acid binding, MF00085: Cation transporter, MF00101: Guanyl-nucleotide e histocompatibility complex antigen, MF00202: Other miscellaneous function protein, MF00208: Molecular function unclassified, MF00213: KRAB box transcription factor, MF00232: Interleukin, MF00250: Serine protease inhibitor, MF00259: Cadherin,		
RNASEH1	ribonuclease h1 R	elated genes	Homo sapiens
ANTHER_MF_ALL	MF00042: Nucleic acid binding, MF00053: Other RNA-binding protein, MF00072: Translation initiation factor, MF00212: Other G-protein	modulator,	
PTOV1		lelated genes	Homo sapiens
ANTHER_MF_ALL	MF00031: Voltage-gated ion channel, MF00033: Voltage-gated calcuim channel, MF00036: Transcription factor, MF00075: Ribosomal protei Guany1-nucleotide exchange factor, MF00146: Deacetylase, MF00175: Major histocompatibility complex antigen, MF00220: Other miscellan modulator, MF00222: Zinc finger transcription factor, MF00224: KRAB box transcription factor, MF00283: Ubiquitin-protein ligase,	neous function protein, MF0	0212: Other G-protein
NONO ANTHER_MF_ALL	non-pou domain containing, octamer-binding R MF00042: Nucleic acid binding, MF00065: mRNA processing factor, MF00068: mRNA splicing factor, MF00084: ATP-binding cassette (ABC	lelated genes	Homo sapiens
– – C11orf9		elated genes	Homo sapiens
ANTHER_MF_ALL	MF00072: Translation initiation factor, MF00086: Other transporter, MF00101: Guanyl-nucleotide exchange factor, MF00135: Transaldolase MF00174: Complement component, MF00189: Other select calcium binding proteins, MF00208: MF00208: Molescular function unclassifier MF00224: KRAB box transcription factor, MF00250: Serine protease inhibitor, MF00279: Tumor necrosis factor receptor,	ed, MF00213: Non-receptor s	erine/threonine protein ki
PSMF1 ANTHER_MF_ALL	proteasome (prosome, macropain) inhibitor subunit 1 (pi31) R MF00002: G-protein coupled receptor, MF00006: Interleukin receptor, MF00008: mRNA splicing factor, MF00072: Translation initiation fac inhibitor, MF00175 : Major histocompatibility complex antigen, MF00208: Molecular function unclassified, MF00227: Basic helix-loop-heli: protein, MF00240: Inmunoglobulin, MF00243: DNA helicase, MF00291: Other enzyme activator,		
TIA1		telated genes	Homo sapiens
NTHER_MF_ALL	Mf00042: Nucleic acid binding, MF00053: Other RNA-binding protein, MF00055: Single-stranded DNA-binding protein, MF00212: Other ( motor protein, MF00243: DNA helicase,	G-protein modulator, MF002	231: Microtubule binding
TEAD4		telated genes	Homo sapiens
ANTHER_MF_ALL	MF00036: Transcription factor, MF00039: Other transcription factor, MF00067: mRNA polyadenylation factor, MF00068: mRAN polyadeny Apolipoprotein ,MF00224: KRAB box transcription factor, MF00242: RNA helicase, MF00243: DNA helicase,		NA splicing factor, MF000
GLS		telated genes	Homo sapiens
ANTHER_MF_ALL	MF00002: G-protein coupled receptor, MF00023: Other signaling molecule, MF00034: Voltage-gated potassium channel, MF00083: Cation 1 Guany-hucleotide exchange factor, MF0013 8: Transminase, MF00141: Hydrolase, MF00148: Phosphodiesterase, MF00173: Defense/immu MF00231: Microtubule binding motor protein, MF00262: Non-motor actin binding protein,	anity protein, MF00180: Extr	
ID2		telated genes	Homo sapiens
ANTHER_MF_ALL	MF00021: Neuropeptide, MF00036: Transcription factor, MF00039 : Other transcription factor, MF00068: mRNA splicing factor, MF00074: adhesion molecule,		
USP11 ANTHER_MF_ALL	ubiguitin specific peptidase 11 R MF00034: Voltage-gated potassium channel, MF00101: Guanyl-nucleotide exchange factor, MF00153: Protease, MF00215: Cysteine protease	lelated genes e. MF00225: Other zinc finge	Homo sapiens
	MF00242: RNA helicase,		
TNPO1 ANTHER_MF_ALL	transportin 1 R MF00087: Transfer/carrier protein, MF00230: Actin binding motor protein, MF00231: Microtubule binding motor protein, MF00261: Actin	lelated genes	Homo sapiens
	family cytoskeletal protein,		
PAWR		lelated genes	Homo sapiens
ANTHER_MF_ALL	MF00042: Nucleic acid binding, MF00096: Phosphatase modulator, MF00138: Transaminase, MF00208: Molecular function unclassified, MI transcription factor binding to ighm enhancer 3 R		
TFE3 ANTHER_MF_ALL	MF00036: Transcription factor, MF00042: Nucleic acid binding, MF00227: Basic helix-loop-helix transcription factor,	lelated genes	Homo sapiens
PLOD2		lelated genes	Homo sapiens
ANTHER_MF_ALL	MF00117: Other phosphatase, MF00123: Oxidoreductase, MF00124: Oxygenase, MF00130: Other oxidoreductase, MF00143: Phospholipase MF00208: Molecular function unclassified, MF00212: Other G-protein modulator, MF00213: Non-receptor serine/threonine protein kinase,	e, MF00202: Other miscellane e, MF00265: Tubulin,	eous function protein,
RBMS1 ANTHER_BP_ALL	rna binding motif, single stranded interacting protein 1 Re BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00040: mRNA transcription, BP00044: mRNA transcription regulation, B1 BP00071: Proteolysis, BP00077: Oxidative phosphorylation BP00142: Ion transport, BP00143: Cation transport, BP00149: T-cell mediated in BP00151: MHCII-mediated immunity, BP00193: Developmental processes, BP00216: Biological process unclassified, BP00273: Chromatin p	mmunity, BP00150: MHCI-r	nediated immunity, P00287: Cell motility,
RNASEH1 ANTHER_BP_ALL		elated genes	Homo sapiens
PTOV1	BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00143: Cation transport, BP00197: Spermatogenesisand motility, BP00256 prostate tumor overexpressed gene 1 R	elated genes	Homo sapiens
ANTHER_BP_ALL	BP00014: Amino acid biosynthesis, BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00044: mRNA transcription regulatior BP00077: Oxisative phosphorylation, BP00104: G-protein mediated signaling, BP00142: Ion transport, BP00143: Cation transport, BP00149 immunity, BP00289: Other metabolism,	9: T-cell mediated immunity,	BP00150: MHCI-mediat
NONO ANTHER BP ALL	non-pou domain containing, octamer-binding Re BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00047: Pre-mRNA processing, BP00048: mRNA splicing, BP00216: Biolog	elated genes	Homo sapiens
C11orf9		elated genes	Homo sapiens
ANTHER_BP_ALL	BP0009: Other polysaccharide metabolism, BP00036: DNA repair, BP00044: mRNA transcription regulation, BP00071, Proteolysis, BP0007 mediated signaling, BP00111: Intracellular signaling cascade, BP00112: Calcium mediated signaling, BP00153: Complement-mediated imm BP00273: Chromatin packaging and remodeling, BP00286: Cell structure,		
PSMF1		elated genes	Homo sapiens
ANTHER_BP_ALL	BP00024: Acyl- CoA metabolism, BP00040: mRNA transcription, BP00044: mRNA transcription regulation, BP00071: Proteolysis, BP00087 mediated signal transduction, BP00104: G-protein mediated signaling, BP00119: Other intracellular signaling cascade, BP00122: Ligand-me BP00150: MHCI-mediated immunity, BP00151: MHCII-mediated immunity, BP00152: B-cell and antibody-mediated immunity, BP00216: communication,	ediated signaling, BP00149: T	-cell mediated immunity
TIA1 ANTHER_BP_ALL	tial cytotoxic granule-associated rna binding protein Re BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00047: Pre-mRNA processig, BP00048: mRNA splicing,	elated genes	Homo sapiens
TEAD4	tea domain family member 4 Re	elated genes	Homo sapiens
ANTHER_BP_ALL GLS	BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00040: mRNA transcription, BP00044: m	RNA transcription regulatior related genes	n, Homo sapiens
ANTHER_BP_ALL	BP00013: Amino acid metabolism, BP00014: Amino acid biosynthesis, BP00036: DNA repair, BP00042: mRNA transcription initiation, BP0 polyadenylation, BP00056: Metabolism of cyclic nucleotides, BP00071: Proteolysis, BP00090: Nitrogen metabolism, BP00102: Signal transdu BP00289: Other metabolism,	uction, BP00142: Ion transpo	rt, BP00143: Cation trans
ID2 ANTHER_BP_ALL	BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00040: mRNA transcription, BP00044: mRNA transcription regulation, B G-protein mediated signaling, BP00128: Constitutive exocytosis, BP00148: Immunity and defense, BP00273: Chromatin packaging and rem	iodeling,	
USP11 ANTHER_BP_ALL	ubiguitin specific peptidase 11 R BP00060: Protein metabolism and modification, BP00071: Proteolysis, BP0014: G-protein mediated signaling, BP00143: Cation transport,	elated genes BP00179: Apoptosis, BP0025	Homo sapiens 50: Muscle development,
TNPO1 ANTHER_BP_ALL		elated genes	Homo sapiens
PAWR		lelated genes	Homo sapiens
ANTHER_BP_ALL TFE3	BP00040: mRNA transcription, BP00043: mRNA transcription elongation, BP00044: mRNA transcription regulation, BP00179: Apoptosis, B transcription factor binding to ighm enhancer 3	3P00298 : Glycogen metaboli: telated genes	sm, Homo sapiens
ANTHER_BP_ALL	BP00031: Nucleoside, nucleotide and nucleic acid metabolism, BP00040: mRNA transcription, BP00044: mRNA transcription regulation,	ance genes	- Homo sapiens
PLOD2 ANTHER_BP_ALL	procpllagen-lysine, 2-oxoglutarate 5-dioxygenase 2 BP00026: Cholesterol metabolism, BP00041: General mRNA transcription activities, BP00060: Protein metabolism and modification, BP00	elated genes	Homo sapiens
Dr_nee	BY00020: Choosetero metadonism, browen: General mix/s transcription activities, provokor rrotein metadolism and mountation, provo metabolism, BP00104: G-protein mediated signaling, BP00142: Ion transport, BP00150: MHCI-mediated immunity, BP00216: Biological pr radical removal,		

FIGURE 5: Molecular function and biological process from DAVID.

## 4. Conclusions

Our method concentrates on and constructs the distinguished single gene network integrated with function prediction analysis by DAVID. For the distinguished single molecular network, we did (1) control and experiment comparison, (2) identification of activation and inhibition networks, (3) construction of upstream and downstream feedback networks, and (4) functional module construction. We tested this method to identify ATF2 regulation network module using data of 45 samples from one and the same GEO dataset. The results demonstrate the effectiveness of such integrated way in terms of developing novel prognostic markers and therapeutic targets.

#### Acknowledgments

This work was supported by the National Natural Science Foundation in China (no. 60673109 and no. 60871100) and the Teaching and Scientific Research Foundation for the Returned Overseas Chinese Scholars, State Education Ministry, State Key Lab of Pattern Recognition Open Foundation.

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