

Supplementary Material

MHC I Stabilizing Potential of Computer-Designed Octapeptides

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Supplementary 1: Descriptor list and specifications. Five defined ‘descriptor spaces’ (sets a.-e.):

- 40-dimensional (8×5): hydrophobicity [1]; hydrophilicity [2]; bulkiness [3]; refractivity [4]; H-2K^b stabilization values [5].
- 40-dimensional (8×5): flexibility [6]; number of possible hydrogen bonds [7]; molecular weight [8]; isoelectricity [9]; hydrophobicity [10].
- 48-dimensional (8×6): six pharmacophoric features of the 20 standard amino acids [11].
- 264-dimensional (8×33): 33 descriptors selected from a set of 184 descriptors that were computed by the software package MOE (Molecular Operation Environment, v2006.08, Chemical Computing Group, Inc., Montreal, Canada). The 33 descriptors were selected based on Kolmogorov-Smirnov statistics [12], so that they separate stabilizing from non-stabilizing octapeptides in at least five residue positions at the 5% significance level. The descriptor space d. consists of following 33 descriptors: diameter, VDistEq, VDistMa, einerPath, weinerPol, BCUT_PEOE_2, CUT_SLOGP_2, BCUT_SMR_1, BCUT_SMR_2, BCUT_SMR_3, GCUT_PEOE_3, GCUT_SLOGP_3, a_count, a_IC, a_ICM, b_count, chi0v, chi1v, a_heavy, a_nC, b_heavy, chi0, chi1_C, VAdjEq, zagreb, PEOE_RPC, PEOE_VSA-0, PEOE_VSA_POS, Kier2, KierA1, KierA2, KierA3, vsa_hyd.
- 904-dimensional (8×113): 113 descriptors selected from a set of more than 1600 descriptors of the software package DragonX (DRAGON for Linux (software for Molecular Descriptor Calculations) Version 1.2, 2006) using Kolmogorov-Smirnov statistics which separate stabilizing from non-stabilizing octapeptides in at least six residue positions. The descriptor space e. includes following 113 descriptors: MW, Sv, Se, Sp, Ss, nAT, nBT, ZM1V, ZMV, Dz, SMTIV, GMTIV, WhetZ, Whetm, Whetv, Whete, Whetp, Jhetp, DEL, S0K, S1K, S2K, X0v, X1v, XMO, ISIZ, IAC, TIC0, BIC0, SIC1, CIC1, BIC1, IC3, IC4, TIC4, IC5, TIC5, SIC5, BIC5, ATS1m, ATS2m, ATS3m, ATS1e, ATS2e, ATS3e, ATS4e, MATS1m, MATS2m, MATS1v, MATS2v, MATS1e, MATS1p, MATS2p, EEig02d, EEig03d, EEig04d, EEig06d, EEig09d, ESpm02d, ESpm03d, ESpm04d, ESpm05d, ESpm06d, ESpm03r, ESpm04r, ESpm05r, BEHm3, BEHm4, BEHm5, BEHm6, BELm1, BELm2, BELm4, BELm6, BEHv2, BEHv3, BEHv4, BEHv5, BEHv6, BEHv7, BELv5, BEHe2, BEHe3, BEHe4, BEHe5, BEHe6, BELe2, BELe4, BEHp2, BEHp4, BEHp5, BEHp6, BEHp7, BELp5, Eig1Z, Eig1m, Eig1v, Eig1e, Eig1p, AEigZ, AEigm, AEigv, AEige, AEigp, VEZ1, VEZ2, VEm1, VEm2, VRv1, VEe1, VEe2, VRp1, AMR.

Supplementary 2: Experimentally tested octapeptides. Sequences are given in single-letter code, motif anchor positions fulfilling the SYFPEITHI motif are underlined. *S*-score: SYFPEITHI score [13]. IEDB-ANN score: predicted IC_{50} values by IEDB-ANN method [14]. < 500 nM: recommended cutoff for IEDB-ANN score [14] for binders predicted with intermediate affinity; *active*: IEDB-ANN score < 500 nM; *0*: IEDB-ANN score > 500 nM. EC_{50} values correspond to the peptide concentration required for 50% of maximal MHC I protein stabilization; values in brackets are standard deviations ($N = 3$). n.d.: not detectable.

	Peptide number	Sequence	<i>S</i> -score	IEDB-ANN score [μ M]	< 500 nM	Experimental EC_{50} [μ M]
stabilizing peptides	1	FRYPYK <u>T</u> L	27	0.5	active	24.0 (\pm 11)
	2	FRYI <u>Y</u> H <u>T</u> L	27	0.2	active	20.0 (\pm 4.0)
	3	FHW <u>D</u> Y <u>R</u> G <u>L</u>	22	0.4	active	25.0 (\pm 11)
	4	WRFKY <u>D</u> N <u>L</u>	22	2.3	0	7.0 (\pm 4.0)
	5	WRFV <u>Y</u> W <u>R</u> L	20	1.2	0	8.0 (\pm 4.0)
	6	WRFY <u>F</u> N <u>L</u>	21	0.4	active	3.6 (\pm 1.2)

	7	WDFK <u>F</u> DS <u>V</u>	17	4.7	0	9.0 (\pm 4.0)
	8	WK <u>F</u> I <u>F</u> DP <u>V</u>	16	3.0	0	0.4 (\pm 0.1)
	9	FHH <u>A</u> HRT <u>V</u>	8	10.5	0	9.0 (\pm 4.0)
non-stabilizing peptides of category i	10	FR <u>Y</u> E <u>Y</u> RS <u>L</u>	28	0.1	active	n.d.
	11	WR <u>Y</u> I <u>Y</u> HS <u>I</u>	22	2.5	0	n.d.
	12	HR <u>Y</u> V <u>Y</u> RNI	24	0.4	active	n.d.
	13	FH <u>Y</u> A <u>Y</u> RS <u>V</u>	23	0.1	active	n.d.
	14	YR <u>Y</u> K <u>Y</u> DRL	27	0.5	0	n.d.
	15	WR <u>Y</u> Q <u>Y</u> DNL	27	0.7	0	n.d.
	16	WR <u>Y</u> R <u>Y</u> WS <u>L</u>	27	0.6	0	n.d.
	17	WR <u>Y</u> N <u>Y</u> DPL	26	1.7	0	n.d.
	18	WR <u>Y</u> H <u>Y</u> DPL	26	2.4	0	n.d.
	19	WK <u>Y</u> Q <u>Y</u> DNL	27	0.3	active	n.d.
	20	WK <u>Y</u> I <u>F</u> DP <u>V</u>	22	1.2	0	n.d.
21	WK <u>Y</u> P <u>F</u> DP <u>V</u>	22	2.0	0	n.d.	
non-stabilizing peptides of category ii	22	FR <u>Y</u> L <u>Y</u> KNA	17	1.2	0	n.d.
	23	FR <u>Y</u> V <u>W</u> RTL	18	1.7	0	n.d.
	24	FH <u>Y</u> L <u>Y</u> HTA	17	0.9	0	n.d.
	25	FR <u>Y</u> P <u>Y</u> HTP	17	17.8	0	n.d.
	26	FR <u>W</u> E <u>Y</u> RGL	22	0.7	0	n.d.
	27	FR <u>H</u> I <u>Y</u> RTI	18	9.3	0	n.d.
	28	FR <u>H</u> G <u>Y</u> RQI	18	12.6	0	n.d.
	29	FH <u>W</u> A <u>Y</u> HTV	17	1.2	0	n.d.
	30	WR <u>W</u> L <u>Y</u> KGV	16	4.3	0	n.d.
	31	WR <u>F</u> P <u>Y</u> DQL	21	6.6	0	n.d.
	32	WR <u>F</u> K <u>Y</u> DPL	21	3.3	0	n.d.
	33	WR <u>F</u> P <u>Y</u> DKL	21	6.8	0	n.d.
	34	WR <u>F</u> V <u>Y</u> DNL	21	1.9	0	n.d.
	35	WK <u>F</u> K <u>F</u> DP <u>V</u>	17	2.8	0	n.d.
36	WK <u>I</u> N <u>F</u> DP <u>V</u>	17	8.2	0	n.d.	
37	TTE <u>W</u> Y <u>T</u> KI	18	3.4	0	n.d.	
5 of 23 non-stabilizing peptides of	38	RGE <u>V</u> F <u>T</u> AT	13	24.0	0	n.d.
	39	FH <u>Y</u> D <u>H</u> RNA	9	6.5	0	n.d.
	40	HR <u>W</u> V <u>F</u> WQP	11	26.0	0	n.d.
	41	SIK <u>N</u> F <u>H</u> YY	12	13.6	0	n.d.
	42	FR <u>H</u> D <u>Y</u> HSP	11	29.2	0	n.d.
7 of 120 non-stabilizing peptides of category iv	43	TVE <u>Q</u> G <u>V</u> TQ	1	37.5	0	n.d.
	44	FHH <u>G</u> H <u>N</u> VP	0	36.8	0	n.d.
	45	QDG <u>H</u> E <u>I</u> HR	1	38.4	0	n.d.
	46	TVE <u>Q</u> G <u>V</u> TQ	1	37.5	0	n.d.
	47	GVD <u>Q</u> S <u>Y</u> LK	0	38.1	0	n.d.
	48	SVEN <u>P</u> ILR	2	33.7	0	n.d.
	49	NEG <u>W</u> T <u>I</u> HR	1	38.8	0	n.d.
	50	SVD <u>H</u> S <u>I</u> FK	2	36.1	0	n.d.
18 of 23 non-stabilizing peptides of category iii	51	FH <u>Y</u> G <u>H</u> KTA	7	16.2	0	n.d.
	52	FH <u>Y</u> I <u>H</u> RSA	7	0.42	0	n.d.
	53	WK <u>F</u> R <u>V</u> DR <u>V</u>	7	15.9	0	n.d.
	54	HGE <u>F</u> Y <u>P</u> LP	11	30.0	0	n.d.
	55	HGE <u>W</u> Y <u>P</u> FD	11	32.2	0	n.d.
	56	TIK <u>N</u> DD <u>S</u> I	7	31.8	0	n.d.
	57	GIK <u>Q</u> Y <u>R</u> YK	12	28.4	0	n.d.
	58	RGD <u>Y</u> Y <u>A</u> TE	13	23.3	0	n.d.
	59	GIE <u>N</u> F <u>H</u> KD	12	34.5	0	n.d.
	60	VVD <u>H</u> F <u>R</u> SP	11	23.2	0	n.d.
	61	VVE <u>H</u> F <u>R</u> LW	11	17.3	0	n.d.
	62	VIEN <u>F</u> SAY	12	9.5	0	n.d.
	63	HGE <u>Y</u> Y <u>R</u> QH	13	32.9	0	n.d.
64	HGD <u>F</u> Y <u>I</u> VS	12	29.4	0	n.d.	
65	RGE <u>P</u> NS <u>L</u> I	9	33.0	0	n.d.	

66	HGEWYHFA	11	25.7	0	n.d.
67	HGEWYPPG	11	29.6	0	n.d.
68	RGKWEKRI	8	30.3	0	n.d.
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69	SDGHQPHR	1	36.0	0	n.d.
70	QEGWDGRR	0	38.5	0	n.d.
71	NEGHGIHR	1	39.0	0	n.d.
72	NGGWGVHR	1	36.6	0	n.d.
73	NEGYDPHR	0	38.8	0	n.d.
74	NEGHTIHR	1	39.2	0	n.d.
75	QDGHQAHR	0	37.7	0	n.d.
76	NEGWQAKR	1	39.0	0	n.d.
77	NDGWDPHR	0	38.9	0	n.d.
78	QDGHSVHR	0	38.1	0	n.d.
79	QDGYNPHR	0	37.0	0	n.d.
80	SDGHQPKR	2	36.9	0	n.d.
81	QDGWGPFR	0	37.1	0	n.d.
82	NEVHGARN	0	38.2	0	n.d.
83	NDPHEIHN	2	38.1	0	n.d.
84	NEVYGPHQ	0	37.4	0	n.d.
85	QGIYGVHN	2	33.0	0	n.d.
86	QDEHGIRN	1	39.0	0	n.d.
87	QDIHGARQ	1	35.8	0	n.d.
88	NEPFTIKN	3	37.8	0	n.d.
89	NDAWGLHQ	0	39.2	0	n.d.
90	QEVHGIHQ	1	37.5	0	n.d.
91	GVETDADN	0	39.6	0	n.d.
92	GVEWKADN	0	39.6	0	n.d.
93	GVEADRDN	1	40.0	0	n.d.
94	GAEGERDN	2	40.0	0	n.d.
95	GAEADAWN	1	39.6	0	n.d.
96	GVEEKADN	1	39.7	0	n.d.
97	GVEGEADN	0	39.7	0	n.d.
98	GAEDERDN	3	39.9	0	n.d.
99	SVEGEADN	1	38.6	0	n.d.
100	GVEDEPDN	1	39.6	0	n.d.
101	GVEGDPDN	0	39.7	0	n.d.
102	GVEGDRVN	1	40.0	0	n.d.
103	GVEGDADN	0	39.8	0	n.d.
104	GAEEERVN	3	39.6	0	n.d.
105	SVDGERTK	3	38.6	0	n.d.
106	GADAKADK	1	39.3	0	n.d.
107	GVDGEADK	0	39.3	0	n.d.
108	GADDEPDK	2	39.5	0	n.d.
109	GVDFEADK	0	38.9	0	n.d.
110	GVDSDAWK	0	39.1	0	n.d.
111	GVKGKADK	0	29.6	0	n.d.
112	GVDDEADK	1	30.5	0	n.d.
113	GVDQSWKG	1	38.4	0	n.d.
114	SIKQSNYY	2	33.5	0	n.d.
115	SVDNSVVF	1	33.4	0	n.d.
116	TPENPTYW	1	35.1	0	n.d.
117	SVEKGVVH	2	37.6	0	n.d.
118	SVETVTSP	2	35.7	0	n.d.
119	SIDNGVW	2	30.8	0	n.d.
120	SVENGVLR	1	33.9	0	n.d.
121	TVDNVILR	1	33.2	0	n.d.
122	TPENSVVR	0	36.9	0	n.d.
123	VVEWPVHN	0	34.1	0	n.d.
124	SVEQSVVH	1	38.0	0	n.d.
125	GIFQQNLP	1	32.0	0	n.d.
126	SVELSIHP	2	35.5	0	n.d.
127	GIDQGVFH	1	38.3	0	n.d.
128	SVEQGILR	2	34.3	0	n.d.
129	TVENSIHK	1	36.0	0	n.d.
130	SIWQVTKD	4	24.2	0	n.d.
131	SVDQGLLQ	1	36.5	0	n.d.

113 of 120 non-stabilizing peptides of category iv

132	TVESVSSP	1	36.4	0	n.d.
133	GIRQSLKE	2	37.5	0	n.d.
134	TPDNGLFR	0	36.6	0	n.d.
135	SVDQGLKE	2	36.9	0	n.d.
136	TVENSVPQ	0	35.3	0	n.d.
137	TVENLSSY	1	33.0	0	n.d.
138	SVEPESHG	2	37.2	0	n.d.
139	SVENGILW	2	32.0	0	n.d.
140	SIDQGVAE	2	32.7	0	n.d.
141	SIEWGLFK	2	34.3	0	n.d.
142	SVEPDSHG	2	37.6	0	n.d.
143	TVENGLVQ	0	38.2	0	n.d.
144	SVENGVPQ	1	34.1	0	n.d.
145	SIKQSNPE	2	36.9	0	n.d.
146	STEPDVAW	1	34.9	0	n.d.
147	TVDTTVSW	0	33.1	0	n.d.
148	IVDHGTAP	2	37.0	0	n.d.
149	TVEQGILH	1	37.9	0	n.d.
150	TVEPWRSW	1	32.9	0	n.d.
151	GLDPDRHW	1	38.7	0	n.d.
152	WTEWVSPP	1	34.8	0	n.d.
153	KVEQSSPE	1	36.2	0	n.d.
154	IVEWGVHW	1	23.8	0	n.d.
155	IVEHSDVP	1	38.7	0	n.d.
156	IVKQGTKW	3	28.2	0	n.d.
157	VVEQSIRY	3	30.6	0	n.d.
158	HVEQGSLY	1	38.2	0	n.d.
159	IVEWITAG	2	35.2	0	n.d.
160	HPEQGV RW	0	36.6	0	n.d.
161	TVDNSVAY	0	35.8	0	n.d.
162	VVDPVLHY	0	29.3	0	n.d.
163	HPEHGV RW	0	37.1	0	n.d.
164	SVEQPLDP	1	37.4	0	n.d.
165	TLDWGV RW	0	31.2	0	n.d.
166	TLKNSKNW	1	31.6	0	n.d.
167	TFAHGV RW	0	24.7	0	n.d.
168	IVELTSAY	2	30.8	0	n.d.
169	SVENPIAQ	2	35.0	0	n.d.
170	RGTYHR TK	4	34.4	0	n.d.
171	RGKYGP FE	2	34.8	0	n.d.
172	RGDREPPA	3	33.7	0	n.d.
173	VVEWVEHY	1	31.8	0	n.d.
174	RGDNPSYT	3	34.7	0	n.d.
175	RGEPLALA	2	34.0	0	n.d.
176	RGEHWIDQ	3	36.2	0	n.d.
177	RGEIWIFW	3	27.1	0	n.d.
178	TVDISTVH	1	39.0	0	n.d.
179	RGEIWHLQ	2	36.4	0	n.d.
180	SVEISIAQ	2	35.8	0	n.d.

Supplementary References:

- [1] D. M. Engelman, T. A. Steitz, A. Goldman, *Annu. Rev. Biophys. Biophys. Chem.* 1986, 15, 321–353.
- [2] T. P. Hopp, K. R. Woods, *Proc. Natl. Acad. Sci. USA* 1981, 8, 3824–3828.
- [3] D. D. Jones, *J. Theor. Biol.* 1975, 50, 167–184.
- [4] K. Udaka, K. H. Wiesmuller, S. Kienle, G. Jung, P. Walden, *J. Biol. Chem.* 1995, 270, 24130–24134.
- [5] R. Bhaskaran, P. K. Ponnuswamy, *Int. J. Pept. Prot. Res.* 1988, 32, 241–255.
- [6] D. S. Dwyer, *J. Biomol. Struct. Dyn.* 2001, 18, 881–892.
- [7] G. D. Fasman, *Handbook of Biochemistry and Molecular Biology, Proteins* CRC Press, Cleveland, 1976.
- [8] G. D. Fasman, *Handbook of Biochemistry and Molecular Biology, Proteins* CRC Press, Cleveland, 1976.
- [9] J. M. Zimmerman, N. Eliezer, R. Simha, *J. Theor. Biol.* 1968, 21, 170–201.
- [10] S. D. Black, D. R. Mould, *Anal. Biochem.* 1991, 193, 72–82.

- [11] G. Schneider, K.-H. Baringhaus, *Molecular Design – Concepts and Applications*, Wiley-VCH, Weinheim, 2008.
- [12] D. N. Rassokhin, D. K. Agrafiotis, *J. Mol. Graph. Model.* 2000, 18,368–382.
- [13] H. G. Rammensee, J. Bachmann, N. N. Emmerich, O. A. Bachor and S. Stevanovic, "SYFPEITHI: database for MHC ligands and peptide motifs", *Immunogenetics*, vol. 50, pp. 213-219, 1999.
- [14] M. Nielsen, C. Lundegaard, P. Worning, S. L. Lauemøller, K. Lamberth, S. Buus, S. Brunak, O. Lund, "Reliable prediction of T-cell epitopes using neural networks with novel sequence representations", *Protein Science*, vol. 12, pp. 1007-1017, 2003.