# Review Article Interferon Alpha in Systemic Lupus Erythematosus

# Timothy B. Niewold,<sup>1</sup> Daniel N. Clark,<sup>2</sup> Rafah Salloum,<sup>1</sup> and Brian D. Poole<sup>2</sup>

<sup>1</sup> Section of Rheumatology and Gwen Knapp Center for Lupus and Immunology Research, The University of Chicago, Chicago, IL 60637, USA

<sup>2</sup> Department of Microbiology and Molecular Biology, Brigham Young University, Provo, UT 84602, USA

Correspondence should be addressed to Brian D. Poole, bpoole@gmail.com

Received 22 January 2010; Accepted 20 May 2010

Academic Editor: Charles Via

Copyright © 2010 Timothy B. Niewold et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The pleiotropic cytokine interferon alpha is involved in multiple aspects of lupus etiology and pathogenesis. Interferon alpha is important under normal circumstances for antiviral responses and immune activation. However, heightened levels of serum interferon alpha and expression of interferon response genes are common in lupus patients. Lupus-associated autoantibodies can drive the production of interferon alpha and heightened levels of interferon interfere with immune regulation. Several genes in the pathways leading to interferon production or signaling are associated with risk for lupus. Clinical and cellular manifestations of excess interferon alpha in lupus combined with the genetic risk factors associated with interferon make this cytokine a rare bridge between genetic risk and phenotypic effects. Interferon alpha influences the clinical picture of lupus and may represent a therapeutic target. This paper provides an overview of the cellular, genetic, and clinical aspects of interferon alpha in lupus.

## 1. Introduction

In systemic lupus erythematosus, a finely tuned system of cells and signals is dysregulated, and the balance between tolerance and autoimmunity is disrupted. Cytokines, as a fundamental mechanism through which the immune system is kept in balance, play an important role in the etiology and pathogenesis of lupus. An example of an important cytokine involved in lupus etiology and pathogenesis is interferon alpha (IFN $\alpha$ ).

IFN $\alpha$  is a pleiotropic cytokine that can affect multiple cell types involved in lupus. Several genes in the interferon pathway are associated with risk for lupus, suggesting a role for this pathway in etiology. Additionally, increased IFN $\alpha$ levels and expression of IFN response genes are often found in lupus. IFN $\alpha$  may affect the clinical manifestations of lupus and is a promising target for therapeutic interventions.

### **2.** Cellular Aspects of IFNα in Lupus

Interferon alpha (IFN $\alpha$ ) is a key molecule in immune regulation. It is produced by multiple cell types in response

to viral infection. Plasmacytoid dendritic cells have a special role in the production of IFN $\alpha$  and are the main sources of serum interferon [1]. IFN $\alpha$  has the potential to dramatically influence the development, progression, and pathogenesis of SLE as it can influence the function and activation state of most major immune cell subsets and function as a bridge between innate and adaptive immunity.

2.1. Toll-Like Receptors and Interferon. One of the principal mechanisms through which IFN $\alpha$  is produced is through Toll-like receptor (TLR) signaling [2, 3]. TLR7 recognizes single-stranded RNA, culminating in interferon regulatory factor (IRF) 5 and IRF7 activation [4] and production of IFN [5–7]. Excessive TLR 7 signaling produces lupus-like autoimmunity in male Yaa mice, where an extra copy of the TLR7 gene is present on the Y chromosome [8–10]. The autoimmune phenotype conferred by the Yaa genotype is dependent on IFN  $\alpha$ , and addition of IFN $\alpha$  can partially duplicate the Yaa phenotype [11]. Additionally, knocking out the IRF7 gene or inhibiting its action with pharmacologic agents inhibits antibody production against RNA-containing

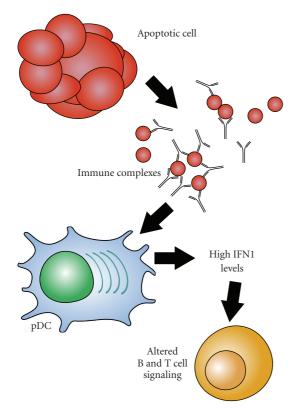


FIGURE 1: Putative source and effects of interferon alpha in lupus. RNA-containing complexes from apoptotic cells are bound by autoantibodies. These immune complexes are internalized after binding to FC receptors on plasmacytoid dendritic cells and stimulate toll-like receptors in the endosomes. Toll-like receptor ligation drives production of interferon alpha, leading to alteration of T-cell profiles, disruption of regulatory T-cell networks, and alteration of B-cell development.

nuclear components [12], suggesting that TLR7 is essential for this type of autoantibody production.

A characteristic of many cases of lupus is the production of antibodies against RNA-containing protein complexes such as Sm, nRNP, Ro, and La. In fact, antibodies against the spliceosomal protein Sm are so specific for lupus that they are used as a diagnostic criterion. The RNA found in these complexes is capable of promoting the production of IFN $\alpha$ through the stimulation of TLR7 [3, 13] (Figure 1).

Because TLR7 is located in the endosomes, RNAcontaining complexes must access the interior of the cell before they are able to act as activators. Autoantibodies specific for these lupus-associated riboproteins can bind with antigens derived from apoptotic cells and form antibodyprotein-RNA complexes. The Fc portions of the immune complexes are recognized and internalized by cells with Fc receptors, providing a route of entry for RNA to reach TLR7, resulting in interferon alpha production [3, 14]. This process is especially well established in PDCs [15, 16]. Interestingly, in addition to being produced as a result of TLR7 ligation, IFN $\alpha$  enhances TLR7 signaling in PDCs [17, 18], forming a positive feedback loop. Despite these data and the strong association between SLE-associated autoantibodies and serum IFN $\alpha$ , SLE-associated autoantibodies are not sufficient for high serum IFN $\alpha$  in humans *in vivo* [19]. Healthy subjects with anti-Ro antibodies do not have high serum IFN- $\alpha$ , while a significant proportion of anti-Ro positive patients with SLE or Sjogren's syndrome do have high serum IFN $\alpha$ , suggesting that these autoantibodies require other disease-associated factors to result in high serum IFN $\alpha$  in humans.

2.2. IFN $\alpha$  and Adaptive Immune Regulation. Excess serum IFN $\alpha$  and IFN-response gene expression are characteristics of lupus and are most likely the result of excessive PDC activation. Such high levels of interferon could contribute to lupus by promoting immune activation rather than tolerance. Dendritic cells are the primary activators of T cells, and affect both T-cell tolerance and activation, depending on the state of the dendritic cell. When treated with interferon alpha, dendritic cells mature and become more prone to activate T cells [20, 21]. Myeloid dendritic cells from lupus patients are able to phagocytose and present self-antigens to T cells in a stimulatory, rather than regulatory manner, a process which is interferon-dependent [22]. Such a process likely contributes to loss of T-cell tolerance to self-antigens and subsequent autoimmunity.

Exposure of the dendritic cell to IFN $\alpha$  contributes to T cell polarity. When CD4+ T cells are activated in the presence of IFN $\alpha$ -producing dendritic cells, their polarity is shifted towards IFN- $\gamma$  producing cells rather than IL-4 producing cells [23, 24], which may promote autoimmunity or immune dysregulation. The T-cells activated by IFN $\alpha$ -treated dendritic cells also are enriched for T-follicular helper cells, a recently described cell type which are adept at activating B cells and driving antibody production [25].

Regulatory T cells (T-reg) are attracting increased attention as a mechanism of immune regulation and suppression of autoimmunity. In lupus, T-regs are often, though not always, found in lower numbers than in controls [26– 31]. Those T-regs that are present in lupus are inefficient at suppressing inflammation and T-cell proliferation [27, 29, 30, 32]. T-reg development is suppressed by treatment of dendritic cells with IFN $\alpha$  [33]. In lupus patients, Treg activity is diminished, due at least in part to the action of IFN $\alpha$  [34] indicating that increased IFN $\alpha$  levels in lupus patients is likely contributing to the development or maintenance of autoimmunity through suppression of T-reg cells.

B cells are important in lupus, since humoral autoimmunity is a hallmark of the disease. IFN $\alpha$  can prevent apoptosis and enhance proliferation of primary B cells, even in the absence of mitogenic stimuli [35]. Interestingly, isolated B cells are inhibited from developing into antibodyproducing plasma cells by IFN $\alpha$  treatment [36]. However, this inhibition is reversed if the B cells are allowed to come into contact with monocytes, in which case IFN $\alpha$  actually stimulates B-cell development and antibody production [37]. The ability of IFN $\alpha$  to influence the activation and function of many major immune cell subsets is a testament to the wide and far-reaching effects of this cytokine. It is clear that interferon is dysregulated in lupus and that overexpression of IFN $\alpha$  can result from the autoantibodies present in lupus. Many components of the molecular pathways through which IFN $\alpha$  and TLRs drive immune activation include genetic risk factors for lupus, further implicating IFN $\alpha$  in lupus etiology and pathogenesis.

# 3. IFN and IFN-Related Genes Associated with SLE Risk

Lupus involves a combination of both environmental and genetic factors. Support for a genetic component includes a high sibling risk ratio [8–29], high heritability (greater than 66%), and higher concordance rates between monozygotic twins (20 to 40%) as compared to other full siblings and dizygotic twins (2 to 5%) [38, 39]. A large number of genetic risk factors are associated with increased susceptibility to the SLE. This genetically determined increased risk status has been referred to as a "threshold liability" [40], which is expected to be highly polygenic in nature and widely variable between individuals. Environmental factors also affect lupus susceptibility and likely interact with this "threshold liability", but as in the case of genetic factors, there is no single environmental cause. A person may have only a few of the genetic risk variations and never get SLE despite exposure to environmental triggers. In contrast, another person may have many of these variations and then develop SLE on first exposure to an environmental trigger.

3.1. Lupus-Associated Risk Loci. Research into the etiopathogenesis of SLE has recently been advanced by several large scale case-control genetic studies, including genome-wide association scans. There is now a pool of approximately 30 genes that have been associated with SLE susceptibility with a high degree of statistical certainty and many others with probable evidence for association (reviewed in [41-45]). With this large number of SLE-associated genes, we can begin to group the list of identified SLE associated genes which should provide insight into initial disease pathogenesis into functional categories. These categories include TLR and IFN signaling, apoptosis and clearance of immune complexes, and B- and T-cell signaling. Several genes affecting the interferon pathway have been associated with risk for lupus. The Interferon pathway normally serves an important function in defense against viral infection. Yet in people with genetic predisposition, environmental triggers such as viral infections may tip the scales in favor of autoimmunity.

Once a genetic variation is identified, functional inference then characterization is necessary to move from identification to an understanding of how the variation affects the etiology or pathogenesis of SLE. Since most of the genes involved in genetic susceptibility to SLE have been identified only recently, there remains much work to identify the functional differences in the genetic associations. However, work done thus far in human cohorts is promising, and the categories of genes and loci associated with risk of lupus already suggest pathways that are of high importance.

3.2. Interferon Regulatory Factors. Certain lupus-associated genetic variations have been shown to directly increase IFN $\alpha$  levels or response to IFN $\alpha$  signaling. Interferon regulatory factor 5 (IRF5) has been confirmed as a risk locus in several different ethnic groups [46–50]. Three main functional variants in IRF5 have been described, which combine to form a risk haplotype in individuals of European descent [51]. One of these loci, at rs2004640, creates an alternate splice site (exon 1B) in the untranslated first exon. Another is a copy number variation of a 30-bp insertion/deletion sequence in exon 6, and the final is rs10954213, which creates an alternate polyadenylation site, resulting in shorter, stabler mRNA [52].

Since IRF5 activates IFN $\alpha$  production, these more stable variants may pose a risk due to their ability to produce excess IFN $\alpha$ . In fact, studies of this gene in human SLE cohorts have shown that the risk variant predisposes to greater serum IFN- $\alpha$ , supporting the idea that the risk haplotype is a gain-of-function variant [53]. IRF5 itself is activated by IFN $\alpha$  signaling, producing a potential positive feedback loop. Another IRF, IRF7, has been highlighted by the association of the IRF7/KIAA1542 locus with lupus in recent studies [54, 55]. Several SNPs in this area were shown to correlate with IFN $\alpha$  levels and alter autoantibody profiles in certain ethnicities [56].

IRF5 and IRF7 are activated by signaling through the endosomal toll-like receptors (TLRs) 7, 8, and 9. Interestingly, both of the IRF variants which are implicated in SLE predispose to higher serum IFN- $\alpha$ , but only in the presence of SLE-associated autoantibodies [53, 56] suggesting that these autoantibodies may provide chronic stimulation of the endosomal TLR pathway of IFN- $\alpha$  generation that when combined with gain-of-function polymorphisms in the IRFs results in dysregulation of the pathway in vivo. Additionally, TLRs 8 and 9 were identified in recent studies as containing susceptibility loci to SLE [57, 58]. The role of TLRs in the interferon production was discussed above.

3.3. Interferon-Associated Genes. Another confirmed locus of susceptibility is in the gene encoding IL1 receptorassociated kinase 1 (IRAK1). This kinase is part of the signal transduction which follows TLR ligation. In a mouse model of lupus, IRAK deficiency eliminated most lupus symptoms, showing the importance of this key intermediate [59]. Since this gene is on the X chromosome, gene dosage may contribute to the risk and the prevalence of the disease in women [59].

Two interacting proteins involved in inflammation, TNF $\alpha$ -induced protein 3 (TNFAIP3) and TNFAIP3-interacting protein 1 (TNIP1), have been identified as risk loci [60–64]. TNFAIP3 encodes the protein A20, which helps turn off signaling through NF $\kappa$ B after an inflammatory response [65, 66]. TNIP1 interacts with TNFAIP3 and is involved in several signal transduction pathways.

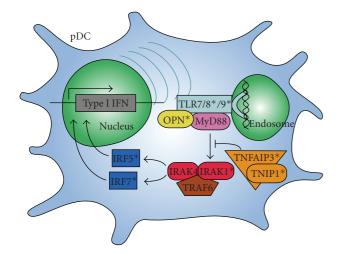


FIGURE 2: Multiple genes involved in interferon production and regulation are associated with risk for lupus. Shown are components of the signal transduction pathway from TLR stimulation by nucleic acids to IFN production. Genes that have been associated with risk for lupus are marked (\*). IFN: interferon, IRAK: interleukin-1 receptor-associated kinase, IRF: interferon regulatory factor, MyD88: myeloid differentiation primary response gene 88, OPN: osteopontin, pDC: plasmacytoid dendritic cell, TLR: toll-like receptor, TNFAIP3: tumor necrosis factor alpha induced protein 3, TNIP1: TNFAIP3 interacting protein 1, and TRAF6: tumor necrosis factor receptor-associated factor 6.

Signal transducer and activator of transcription 4 (STAT4) is another risk locus with direct links to the interferon pathway. It is involved in proliferation, differentiation, and apoptosis. STAT4 has 2 risk loci, one at rs7574865 which has been shown to increase sensitivity to IFN $\alpha$  [67], and another at rs3821236 which increases STAT4 transcription and interacts with IRF5 susceptibility loci [68]. The presence of both of these risk alleles gives an additive effect, increasing risk to SLE [69]. Osteopontin (OPN) is a key molecule for IFN $\alpha$  production in pDCs [70]. Presence of a lupus risk-associated form of this gene was recently tied to high IFN levels in males and young-onset female lupus patients [71].

Possible interactions of the IFN-associated genes that have been linked to lupus are shown in Figure 2. The risk variants of these genes influence the production of and response to IFN $\alpha$ , likely driving the increased levels seen in lupus patients and affecting the clinical manifestations of the disease.

#### 4. Clinical Aspects of IFNα in Lupus

Lupus primarily affects women in the reproductive years; however people of all ages, genders, and ancestral backgrounds are susceptible. Disease features range from mild manifestations such as rash or arthritis to life-threatening end-organ manifestations such as glomerulonephritis or thrombosis, and it is difficult to predict which manifestations will affect a given patient. 4.1. IFN- $\alpha$  as a Causal Factor in Human Lupus. A number of patients treated with IFN $\alpha$  have developed lupus or lupus-like syndrome [72-74]. In these reports, many specific manifestations of idiopathic lupus such as malar or discoid rash, oral ulcers, photosensitivity, renal involvement, and anti-Sm and anti-dsDNA antibodies were represented, suggesting that these cases were not "drug-induced" SLE but instead resembled idiopathic SLE [73]. Discontinuation of IFN $\alpha$  typically resulted in remission of SLE symptoms [73], supporting a causal relationship with IFN- $\alpha$ . While only a minority of patients treated with IFN $\alpha$  develop SLE (<1% of patients) [75], these data support the idea that IFN $\alpha$  can be sufficient to induce SLE in some individuals. Many more IFNa-treated individuals develop a "lupus-like" syndrome [74], with some SLE symptoms which are insufficient to meet formal diagnostic criteria for SLE [76]. IFN-induced SLE can be severe, and there are reports of life-threatening multiorgan SLE involvement including glomerulonephritis, serositis, discoid rash, myopericarditis, and vasculitis [77, 78].

Another finding which supports the hypothesis of IFN $\alpha$ as a primary causal factor in human SLE is the clustering of high serum IFN $\alpha$  in lupus families [79]. Patients with lupus and their healthy relatives have higher serum IFN $\alpha$ activity as compared to healthy unrelated individuals [79]. Strong familial correlations in serum IFN $\alpha$  were observed regardless of disease status (affecteds versus unaffecteds), and SLE probands in the same family tended to have similar IFN $\alpha$ levels [79]. Spouses of SLE patients did not have high serum IFN $\alpha$  activity, and taken together these data suggest that high serum IFN $\alpha$  is a heritable risk factor for SLE. Interestingly, age-related patterns of serum IFN $\alpha$  were also observed in SLE families in which the ages of highest IFN $\alpha$  mirrored the ages of peak SLE incidence [80, 81]. The discovery of several lupus risk loci in IFN-related genes provides further support for the above observation that serum IFN- $\alpha$  is heritable, and the SLE risk variants of each of these genes result in a gain of function increase in IFN $\alpha$  signaling as detailed above.

4.2. Clinical Correlations with IFN Alpha. A very strong correlation is consistently observed between the presence of SLE-associated autoantibodies, such as anti-Ro, anti-La, anti-Sm, anti-RNP, and anti-dsDNA [79, 82]. Lupus patients with high serum IFN $\alpha$  had a significantly higher prevalence of cutaneous and renal disease in most studies [82–84]. It is interesting that both of these clinical manifestations share an association with a particular serology (rash with anti-Ro and nephritis with anti-dsDNA), and whether these clinical manifestations are associated independent of serology has not been shown to our knowledge.

A number of studies have shown that IFN $\alpha$  correlates with disease activity when assessed cross sectionally [82–85]. Results are conflicting regarding the potential fluctuation of IFN $\alpha$  with disease activity in SLE, and there are a number of studies which did not find a longitudinal correlation [86, 87]. In these studies, a cross-sectional relationship between IFN $\alpha$  and disease activity is still observed, suggesting that IFN $\alpha$  may indicate those patients who generally have higher disease activity as compared to other patients. A recent prospective study evaluated the utility of serum interferonregulated chemokine levels as potential biomarkers of SLE disease activity [88]. In this study, IFN $\alpha$ -induced chemokines correlated with disease activity cross sectionally, rose at the time of a flare, and decreased as the disease remitted [88]. In this same study, high chemokine levels were predictive of SLE flare over the next year in a subset of patients.

4.3. Anti-IFN $\alpha$  Therapies in Lupus. Given all of the studies presented above, there has been considerable interest in therapies which block IFN $\alpha$ . To date there is one published study describing a phase I trial of a fully human monoclonal antibody that binds to the majority of the subtypes of human interferon alpha [89]. Treatment with this anti-IFN antibody resulted in a dose-dependent inhibition of interferon-induced gene expression in peripheral blood cells as well as skin lesions in patients with mild to moderate SLE [89]. No obvious safety signals were reported during the phase I trial of anti-IFN therapy, and the proof-ofprinciple analyses supported a biological effect blocking the IFN pathway in humans. Phase two trials to assess efficacy of these agents in treating SLE are currently underway. There are many known predictors of high serum IFN $\alpha$  in SLE patients, including both serologic and genetic markers outlined in this paper. We anticipate that incorporation of these variables into clinical trial design would enhance efficacy and potentially minimize side effects by targeting the most relevant patient group. Long-term safety data will be important, since IFN $\alpha$  is such a highly conserved and important immunological mediator of viral defense.

#### 5. Conclusions

IFN- $\alpha$  is associated with SLE through multiple lines of evidence. These include genetic, immunological/serological, and clinical associations, as described in this review. It is likely that IFN- $\alpha$  plays a key role in SLE etiology, pathogenesis, and/or disease persistence. Despite this large body of evidence associating IFN-alpha with lupus, the association between interferon alpha and SLE is largely inferential. The exact cellular and immunological mechanisms through which IFN affects lupus also remain undiscovered for the most part. These mechanisms and pathways are potentially fertile areas for future investigation. Such studies will likely lead to new therapeutic targets as well as a greater understanding of lupus as a disease.

### References

- P. Fitzgerald-Bocarsly, J. Dai, and S. Singh, "Plasmacytoid dendritic cells and type I IFN: 50 years of convergent history," *Cytokine and Growth Factor Reviews*, vol. 19, no. 1, pp. 3–19, 2008.
- [2] T. Ito, Y.-H. Wang, and Y.-J. Liu, "Plasmacytoid dendritic cell precursors/type I interferon-producing cells sense viral infection by Toll-like receptor (TLR) 7 and TLR9," *Springer Seminars in Immunopathology*, vol. 26, no. 3, pp. 221–229, 2005.

- [3] T. Lövgren, M.-L. Eloranta, B. Kastner, M. Wahren-Herlenius, G. V. Alm, and L. Rönnblom, "Induction of interferonalpha by immune complexes or liposomes containing systemic lupus erythematosus autoantigen- and Sjogren's syndrome autoantigen-associated RNA," *Arthritis and Rheumatism*, vol. 54, no. 6, pp. 1917–1927, 2006.
- [4] A. Schoenemeyer, B. J. Barnes, M. E. Mancl et al., "The interferon regulatory factor, IRF5, is a central mediator of tolllike receptor 7 signaling," *Journal of Biological Chemistry*, vol. 280, no. 17, pp. 17005–17012, 2005.
- [5] K. Hoshino, T. Sugiyama, M. Matsumoto et al., "I $\kappa$ B kinase- $\alpha$  is critical for interferon- $\alpha$  production induced by Toll-like receptors 7 and 9," *Nature*, vol. 440, no. 7086, pp. 949–953, 2006.
- [6] T. Kawai, S. Sato, K. J. Ishii et al., "Interferon-α induction through Toll-like receptors involves a direct interaction of IRF7 with MyD88 and TRAF6," *Nature Immunology*, vol. 5, no. 10, pp. 1061–1068, 2004.
- [7] S. Löseke, E. Grage-Griebenow, H. Heine et al., "In vitro-generated viral double-stranded RNA in contrast to polyinosinic: polycytidylic acid induces interferon-α in human plasmacytoid dendritic cells," *Scandinavian Journal of Immunology*, vol. 63, no. 4, pp. 264–274, 2006.
- [8] A.-M. Fairhurst, S.-H. Hwang, A. Wang et al., "Yaa autoimmune phenotypes are conferred by overexpression of TLR7," *European Journal of Immunology*, vol. 38, no. 7, pp. 1971–1978, 2008.
- [9] P. Pisitkun, J. A. Deane, M. J. Difilippantonio, T. Tarasenko, A. B. Satterthwaite, and S. Bolland, "Autoreactive B cell responses to RNA-related antigens due to TLR7 gene duplication," *Science*, vol. 312, no. 5780, pp. 1669–1672, 2006.
- [10] S. Subramanian, K. Tus, Q.-Z. Li et al., "A Tlr7 translocation accelerates systemic autoimmunity in murine lupus," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 26, pp. 9970–9975, 2006.
- [11] M. Ramanujam, P. Kahn, W. Huang et al., "Interferon-α treatment of Female (NZW × BXSB)F1 mice mimics some but not all features associated with the Yaa mutation," *Arthritis and Rheumatism*, vol. 60, no. 4, pp. 1096–1101, 2009.
- [12] F. J. Barrat, T. Meeker, J. H. Chan, C. Guiducci, and R. L. Coffmann, "Treatment of lupus-prone mice with a dual inhibitor of TLR7 and TLR9 leads to reduction of autoan-tibody production and amelioration of disease symptoms," *European Journal of Immunology*, vol. 37, no. 12, pp. 3582–3586, 2007.
- [13] F. J. Barrat, T. Meeker, J. Gregorio et al., "Nucleic acids of mammalian origin can act as endogenous ligands for Toll-like receptors and may promote systemic lupus erythematosus," *Journal of Experimental Medicine*, vol. 202, no. 8, pp. 1131– 1139, 2005.
- [14] F. Batteux, P. Palmer, M. Daëron, B. Weill, and P. Lebon, "FCyRII (CD32)-dependent induction of interferon-alpha by serum from patients with lupus erythematosus," *European Cytokine Network*, vol. 10, no. 4, pp. 509–514, 1999.
- [15] U. Båve, M. Magnusson, M.-L. Eloranta, A. Perers, G. V. Alm, and L. Rönnblom, "FcyRIIa is expressed on natural IFN-αproducing cells (plasmacytoid dendritic cells) and is required for the IFN-α production induced by apoptotic cells combined with Lupus IgG," *Journal of Immunology*, vol. 171, no. 6, pp. 3296–3302, 2003.
- [16] E. Savarese, O.-W. Chae, S. Trowitzsch et al., "U1 small nuclear ribonucleoprotein immune complexes induce type I interferon in plasmacytoid dendritic cells through TLR7," *Blood*, vol. 107, no. 8, pp. 3229–3234, 2006.

- [17] D. Rajagopal, C. Paturel, Y. Morel, S. Uematsu, S. Akira, and S. S. Diebold, "Plasmacytoid dendritic cell-derived type I interferon is crucial for the adjuvant activity of Toll-like receptor 7 agonists," *Blood*, vol. 115, no. 10, pp. 1949–1957, 2010.
- [18] C. Asselin-Paturel, G. Brizard, K. Chemin et al., "Type I interferon dependence of plasmacytoid dendritic cell activation and migration," *Journal of Experimental Medicine*, vol. 201, no. 7, pp. 1157–1167, 2005.
- [19] T. B. Niewold, T. L. Rivera, J. P. Buyon, and M. K. Crow, "Serum type I interferon activity is dependent on maternal diagnosis in anti-SSA/Ro-positive mothers of children with neonatal lupus," *Arthritis and Rheumatism*, vol. 58, no. 2, pp. 541–546, 2008.
- [20] A. Farkas, G. Tonel, and F. O. Nestle, "Interferon-α and viral triggers promote functional maturation of human monocytederived dendritic cells," *British Journal of Dermatology*, vol. 158, no. 5, pp. 921–929, 2008.
- [21] M. Sakakibara, T. Kanto, M. Inoue et al., "Quick generation of fully mature dendritic cells from monocytes with OK432, low-dose prostanoid, and interferon-α as potent immune enhancers," *Journal of Immunotherapy*, vol. 29, no. 1, pp. 67– 77, 2006.
- [22] P. Blanco, A. K. Palucka, M. Gill, V. Pascual, and J. Banchereau, "Induction of dendritic cell differentiation by IFN-α in systemic lupus erythematosus," *Science*, vol. 294, no. 5546, pp. 1540–1543, 2001.
- [23] T. Ito, R. Amakawa, M. Inaba et al., "Plasmacytoid dendritic cells regulate Th cell responses through OX40 ligand and type I IFNs," *Journal of Immunology*, vol. 172, no. 7, pp. 4253–4259, 2004.
- [24] M. Mohty, A. Vialle-Castellano, J. A. Nunes, D. Isnardon, D. Olive, and B. Gaugler, "IFN-α skews monocyte differentiation into toll-like receptor 7-expressing dendritic cells with potent functional activities," *Journal of Immunology*, vol. 171, no. 7, pp. 3385–3393, 2003.
- [25] H. Cucak, U. Yrlid, B. Reizis, U. Kalinke, and B. Johansson-Lindbom, "Type I interferon signaling in dendritic cells stimulates the development of lymph-node-resident T follicular helper cells," *Immunity*, vol. 31, no. 3, pp. 491–501, 2009.
- [26] S. Baráth, P. Soltész, E. Kiss et al., "The severity of systemic lupus erythematosus negatively correlates with the increasing number of CD4<sup>+</sup>CD25<sup>high</sup>FoxP3<sup>+</sup> regulatory T cells during repeated plasmapheresis treatments of patients," *Autoimmunity*, vol. 40, no. 7, pp. 521–528, 2007.
- [27] M. Bonelli, A. Savitskaya, K. Von Dalwigk et al., "Quantitative and qualitative deficiencies of regulatory T cells in patients with systemic lupus erythematosus (SLE)," *International Immunology*, vol. 20, no. 7, pp. 861–868, 2008.
- [28] S. Hu, W. Xiao, F. Kong, D. Ke, R. Qin, and M. Su, "Regulatory T cells and their molecular markers in peripheral blood of the patients with systemic lupus erythematosus," *Journal* of Huazhong University of Science and Technology—Medical Science, vol. 28, no. 5, pp. 549–552, 2008.
- [29] J.-L. Suen, H.-T. Li, Y.-J. Jong, B.-L. Chiang, and J.-H. Yen, "Altered homeostasis of CD4<sup>+</sup>FoxP3<sup>+</sup> regulatory T-cell subpopulations in systemic lupus erythematosus," *Immunology*, vol. 127, no. 2, pp. 196–205, 2009.
- [30] R. K. C. Venigalla, T. Tretter, S. Krienke et al., "Reduced CD4<sup>+</sup>,CD25-T cell sensitivity to the suppressive function of CD4<sup>+</sup>, CD25<sup>high</sup>, CD127<sup>-/low</sup> regulatory T cells in patients with active systemic lupus erythematosus," *Arthritis and Rheumatism*, vol. 58, no. 7, pp. 2120–2130, 2008.

- [31] J. Yates, A. Whittington, P. Mitchell, R. I. Lechler, L. Lightstone, and G. Lombardi, "Natural regulatory T cells: number and function are normal in the majority of patients with lupus nephritis," *Clinical and Experimental Immunology*, vol. 153, no. 1, pp. 44–55, 2008.
- [32] J. Gómez, C. Prado, P. López, A. Suárez, and C. Gutiérrez, "Conserved anti-proliferative effect and poor inhibition of TNFα secretion by regulatory CD4<sup>+</sup>CD25<sup>+</sup> T cells in patients with systemic lupus erythematosus," *Clinical Immunology*, vol. 132, no. 3, pp. 385–392, 2009.
- [33] M. Gigante, M. Mandic, A. K. Wesa et al., "Interferon-alpha (IFN-α)-conditioned DC preferentially stimulate type-1 and limit treg-type in vitro T-cell responses from RCC patients," *Journal of Immunotherapy*, vol. 31, no. 3, pp. 254–262, 2008.
- [34] B. Yan, S. Ye, G. Chen, M. Kuang, N. Shen, and S. Chen, "Dysfunctional CD4<sup>+</sup>,CD25<sup>+</sup> regulatory T cells in untreated active systemic lupus erythematosus secondary to interferonα-producing antigen-presenting cells," *Arthritis and Rheumatism*, vol. 58, no. 3, pp. 801–812, 2008.
- [35] K. Ruuth, L. Carlsson, B. Hallberg, and E. Lundgren, "Interferon-α promotes survival of human primary Blymphocytes via phosphatidylinositol 3-kinase," *Biochemical and Biophysical Research Communications*, vol. 284, no. 3, pp. 583–586, 2001.
- [36] H. Oka, S. Hirohata, T. Inoue, and K. Ito, "Effects of interferon-α on human B cell responsiveness: biphasic effects in cultures stimulated with Staphylococcus aureus," *Cellular Immunology*, vol. 139, no. 2, pp. 478–492, 1992.
- [37] H. Oka and S. Hirohata, "Regulation of human B cell responsiveness by interferon-α: interferon-α-mediated suppression of B cell function is reversed through direct interactions between monocytes and B cells," *Cellular Immunology*, vol. 146, no. 2, pp. 238–248, 1993.
- [38] D. Alarcón-Segovia, M. E. Alarcón-Riquelme, M. H. Cardiel et al., "Familial aggregation of systemic lupus erythematosus, rheumatoid arthritis, and other autoimmune diseases in 1,177 lupus patients from the GLADEL cohort," *Arthritis and Rheumatism*, vol. 52, no. 4, pp. 1138–1147, 2005.
- [39] D. Deapen, A. Escalante, L. Weinrib et al., "A revised estimate of twin concordance in systemic lupus erythematosus," *Arthritis and Rheumatism*, vol. 35, no. 3, pp. 311–318, 1992.
- [40] A. Wandstrat and E. Wakeland, "The genetics of complex autoimmune diseases: non-MHC susceptibility genes," *Nature Immunology*, vol. 2, no. 9, pp. 802–809, 2001.
- [41] G. D. Sebastiani and M. Galeazzi, "Immunogenetic studies on systemic lupus erythematosus," *Lupus*, vol. 18, no. 10, pp. 878– 883, 2009.
- [42] K. L. Moser, J. A. Kelly, C. J. Lessard, and J. B. Harley, "Recent insights into the genetic basis of systemic lupus erythematosus," *Genes and Immunity*, vol. 10, no. 5, pp. 373– 379, 2009.
- [43] I. T. W. Harley, K. M. Kaufman, C. D. Langefeld, J. B. Harley, and J. A. Kelly, "Genetic susceptibility to SLE: new insights from fine mapping and genome-wide association studies," *Nature Reviews Genetics*, vol. 10, no. 5, pp. 285–290, 2009.
- [44] R. R. Graham, G. Hom, W. Ortmann, and T. W. Behrens, "Review of recent genome-wide association scans in lupus," *Journal of Internal Medicine*, vol. 265, no. 6, pp. 680–688, 2009.
- [45] B. Rhodes and T. J. Vyse, "The genetics of SLE: an update in the light of genome-wide association studies," *Rheumatology*, vol. 47, no. 11, pp. 1603–1611, 2008.

- [46] Y. H. Lee and G. G. Song, "Association between the rs2004640 functional polymorphism of interferon regulatory factor 5 and systemic lupus erythematosus: a meta-analysis," *Rheumatology International*, vol. 29, no. 10, pp. 1137–1142, 2009.
- [47] A. Kawasaki, C. Kyogoku, J. Ohashi et al., "Association of IRF5 polymorphisms with systemic lupus erythematosus in a Japanese population: support for a crucial role of intron 1 polymorphisms," *Arthritis and Rheumatism*, vol. 58, no. 3, pp. 826–834, 2008.
- [48] J. A. Kelly, J. M. Kelley, K. M. Kaufman et al., "Interferon regulatory factor-5 is genetically associated with systemic lupus erythematosus in African Americans," *Genes and Immunity*, vol. 9, no. 3, pp. 187–194, 2008.
- [49] M. V. P. L. Reddy, R. Velázquez-Cruz, V. Baca et al., "Genetic association of IRF5 with SLE in Mexicans: higher frequency of the risk haplotype and its homozygozity than Europeans," *Human Genetics*, vol. 121, no. 6, pp. 721–727, 2007.
- [50] K. Shimane, Y. Kochi, R. Yamada et al., "A single nucleotide polymorphism in the IRF5 promoter region is associated with susceptibility to rheumatoid arthritis in the Japanese population," *Annals of the Rheumatic Diseases*, vol. 68, no. 3, pp. 377–383, 2009.
- [51] R. R. Graham, C. Kyogoku, S. Sigurdsson et al., "Three functional variants of IFN regulatory factor 5 (IRF5) define risk and protective haplotypes for human lupus," *Proceedings* of the National Academy of Sciences of the United States of America, vol. 104, no. 16, pp. 6758–6763, 2007.
- [52] R. R. Graham, C. Kyogoku, S. Sigurdsson et al., "Three functional variants of IFN regulatory factor 5 (IRF5) define risk and protective haplotypes for human lupus," *Proceedings* of the National Academy of Sciences of the United States of America, vol. 104, no. 16, pp. 6758–6763, 2007.
- [53] T. B. Niewold, J. A. Kelly, M. H. Flesch, L. R. Espinoza, J. B. Harley, and M. K. Crow, "Association of the IRF5 risk haplotype with high serum interferon-α activity in systemic lupus erythematosus patients," *Arthritis and Rheumatism*, vol. 58, no. 8, pp. 2481–2487, 2008.
- [54] J. B. Harley, M. E. Alarcón-Riquelme, L. A. Criswell et al., "Genome-wide association scan in women with systemic lupus erythematosus identifies susceptibility variants in ITGAM, PXK, KIAA1542 and other loci," *Nature Genetics*, vol. 40, no. 2, pp. 204–210, 2008.
- [55] M. Suarez-Gestal, M. Calaza, E. Endreffy et al., "Replication of recently identified systemic lupus erythematosus genetic associations: a case-control study," *Arthritis Research and Therapy*, vol. 11, no. 3, article 69, 2009.
- [56] R. Salloum, B. Franek, S. Kariuki, T. Utset, and T. T. Niewold, "Genetic variation at the IRF7/KIAA1542 locus is associated with autoantibody profile and serum interferon alpha levels in lupus patients," *Clinical Immunology*, vol. 131, supplement 1, p. S54, 2009.
- [57] D. L. Armstrong, A. Reiff, B. L. Myones et al., "Identification of new SLE-associated genes with a two-step Bayesian study design," *Genes and Immunity*, vol. 10, no. 5, pp. 446–456, 2009.
- [58] C.-J. Xu, W.-H. Zhang, H.-F. Pan, X.-P. Li, J.-H. Xu, and D.-Q. Ye, "Association study of a single nucleotide polymorphism in the exon 2 region of toll-like receptor 9 (TLR9) gene with susceptibility to systemic lupus erythematosus among Chinese," *Molecular Biology Reports*, vol. 36, no. 8, pp. 2245– 2248, 2009.
- [59] C. O. Jacob, J. Zhu, D. L. Armstrong et al., "Identification of IRAK1 as a risk gene with critical role in the pathogenesis

of systemic lupus erythematosus," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 15, pp. 6256–6261, 2009.

- [60] J. S. Bates, C. J. Lessard, J. M. Leon et al., "Meta-analysis and imputation identifies a 109kb risk haplotype spanning TNFAIP3 associated with lupus nephritis and hematologic manifestations," *Genes and Immunity*, vol. 10, no. 5, pp. 470– 477, 2009.
- [61] V. Gateva, J. K. Sandling, G. Hom et al., "A large-scale replication study identifies TNIP1, PRDM1, JAZF1, UHRF1BP1 and IL10 as risk loci for systemic lupus erythematosus," *Nature Genetics*, vol. 41, no. 11, pp. 1228–1233, 2009.
- [62] R. R. Graham, C. Cotsapas, L. Davies et al., "Genetic variants near TNFAIP3 on 6q23 are associated with systemic lupus erythematosus," *Nature Genetics*, vol. 40, no. 9, pp. 1059–1061, 2008.
- [63] J.-W. Han, H.-F. Zheng, Y. Cui et al., "Genome-wide association study in a Chinese Han population identifies nine new susceptibility loci for systemic lupus erythematosus," *Nature Genetics*, vol. 41, no. 11, pp. 1234–1237, 2009.
- [64] S. L. Musone, K. E. Taylor, T. T. Lu et al., "Multiple polymorphisms in the TNFAIP3 region are independently associated with systemic lupus erythematosus," *Nature Genetics*, vol. 40, no. 9, pp. 1062–1064, 2008.
- [65] K. Honma, S. Tsuzuki, M. Nakagawa et al., "TNFAIP3/A20 functions as a novel tumor suppressor gene in several subtypes of non-Hodgkin lymphomas," *Blood*, vol. 114, no. 12, pp. 2467–2475, 2009.
- [66] M. Kato, M. Sanada, I. Kato et al., "Frequent inactivation of A20 in B-cell lymphomas," *Nature*, vol. 459, no. 7247, pp. 712– 716, 2009.
- [67] S. N. Kariuki, K. A. Kirou, E. J. MacDermott, L. Barillas-Arias, M. K. Crow, and T. B. Niewold, "Cutting edge: autoimmune disease risk variant of STAT4 confers increased sensitivity to IFN-alpha in lupus patients in vivo," *Journal of Immunology*, vol. 182, no. 1, pp. 34–38, 2009.
- [68] A. K. Abelson, A. M. Delgado-Vega, S. V. Kozyrev et al., "STAT4 associates with systemic lupus erythematosus through two independent effects that correlate with gene expression and act additively with IRF5 to increase risk," *Annals of the Rheumatic Diseases*, vol. 68, no. 11, pp. 1746–1753, 2009.
- [69] S. Sigurdsson, G. Nordmark, S. Garnier et al., "A risk haplotype of STAT4 for systemic lupus erythematosus is overexpressed, correlates with anti-dsDNA and shows additive effects with two risk alleles of IRF5," *Human Molecular Genetics*, vol. 17, no. 18, pp. 2868–2876, 2008.
- [70] W. Cao and Y.-J. Liu, "Opn: key regulator of pDC interferon production," *Nature Immunology*, vol. 7, no. 5, pp. 441–443, 2006.
- [71] S. N. Kariuki, J. G. Moore, K. A. Kirou, M. K. Crow, T. O. Utset, and T. B. Niewold, "Age- and gender-specific modulation of serum osteopontin and interferon-*α* by osteopontin genotype in systemic lupus erythematosus," *Genes and Immunity*, vol. 10, no. 5, pp. 487–494, 2009.
- [72] L. E. Ronnblom, G. V. Alm, and K. E. Oberg, "Possible induction of systemic lupus erythematosus by interferon αtreatment in a patient with a malignant carcinoid tumour," *Journal of Internal Medicine*, vol. 227, no. 3, pp. 207–210, 1990.
- [73] T. B. Niewold and W. I. Swedler, "Systemic lupus erythematosus arising during interferon-alpha therapy for cryoglobulinemic vasculitis associated with hepatitis C," *Clinical Rheumatology*, vol. 24, no. 2, pp. 178–181, 2005.

- [74] Y. Ioannou and D. A. Isenberg, "Current evidence for the induction of autoimmune rheumatic manifestations by cytokine therapy," *Arthritis and Rheumatism*, vol. 43, no. 7, pp. 1431–1442, 2000.
- [75] C. Gota and L. Calabrese, "Induction of clinical autoimmune disease by therapeutic interferon-α," *Autoimmunity*, vol. 36, no. 8, pp. 511–518, 2003.
- [76] E. M. Tan, A. S. Cohen, and J. F. Fries, "The 1982 revised criteria for the classification of systemic lupus erythrematosus," *Arthritis and Rheumatism*, vol. 25, no. 11, pp. 1271–1277, 1982.
- [77] V. Ho, A. McLean, and S. Terry, "Severe systemic lupus erythematosus induced by antiviral treatment for hepatitis C," *Journal of Clinical Rheumatology*, vol. 14, no. 3, pp. 166–168, 2008.
- [78] T. B. Niewold, "Interferon alpha-induced lupus: proof of principle," *Journal of Clinical Rheumatology*, vol. 14, no. 3, pp. 131–132, 2008.
- [79] T. B. Niewold, J. Hua, T. J. A. Lehman, J. B. Harley, and M. K. Crow, "High serum IFN-α activity is a heritable risk factor for systemic lupus erythematosus," *Genes and Immunity*, vol. 8, no. 6, pp. 492–502, 2007.
- [80] T. B. Niewold, J. E. Adler, S. B. Glenn, T. J. A. Lehman, J. B. Harley, and M. K. Crow, "Age- and sex-related patterns of serum interferon-α activity in lupus families," *Arthritis and Rheumatism*, vol. 58, no. 7, pp. 2113–2119, 2008.
- [81] C. E. Weckerle and T. B. Niewold, "The unexplained female predominance of systemic lupus erythematosus: clues from genetic and cytokine studies," *Clinical Reviews in Allergy and Immunology*, 2010. In press.
- [82] K. A. Kirou, C. Lee, S. George, K. Louca, M. G. E. Peterson, and M. K. Crow, "Activation of the interferon-α pathway identifies a subgroup of systemic lupus erythematosus patients with distinct serologic features and active disease," *Arthritis and Rheumatism*, vol. 52, no. 5, pp. 1491–1503, 2005.
- [83] M. C. Dall'Era, P. M. Cardarelli, B. T. Preston, A. Witte, and J. C. Davis Jr., "Type I interferon correlates with serological and clinical manifestations of SLE," *Annals of the Rheumatic Diseases*, vol. 64, no. 12, pp. 1692–1697, 2005.
- [84] X. Feng, H. Wu, J. M. Grossman et al., "Association of increased interferon-inducible gene expression with disease activity and lupus nephritis in patients with systemic lupus erythematosus," *Arthritis and Rheumatism*, vol. 54, no. 9, pp. 2951–2962, 2006.
- [85] H. Zhuang, S. Narain, E. Sobel et al., "Association of antinucleoprotein autoantibodies with upregulation of type I interferon-inducible gene transcripts and dendritic cell maturation in systemic lupus erythematosus," *Clinical Immunology*, vol. 117, no. 3, pp. 238–250, 2005.
- [86] C. Landolt-Marticorena, G. Bonventi, A. Lubovich et al., "Lack of association between the interferon-α signature and longitudinal changes in disease activity in systemic lupus erythematosus," *Annals of the Rheumatic Diseases*, vol. 68, no. 9, pp. 1440–1446, 2009.
- [87] M. Petri, S. Singh, H. Tesfasyone et al., "Longitudinal expression of type I interferon responsive genes in systemic lupus erythematosus," *Lupus*, vol. 18, no. 11, pp. 980–989, 2009.
- [88] J. W. Bauer, M. Petri, F. M. Batliwalla et al., "Interferonregulated chemokines as biomarkers of systemic lupus erythematosus disease activity: a validation study," *Arthritis and Rheumatism*, vol. 60, no. 10, pp. 3098–3107, 2009.

[89] Y. Yao, L. Richman, B. W. Higgs et al., "Neutralization of interferon-α/β-inducible genes and downstream effect in a phase I trial of an anti-interferon-α monoclonal antibody in systemic lupus erythematosus," *Arthritis and Rheumatism*, vol. 60, no. 6, pp. 1785–1796, 2009.

8



The Scientific World Journal



Gastroenterology Research and Practice





Journal of Diabetes Research



Disease Markers



Journal of Immunology Research





Submit your manuscripts at http://www.hindawi.com





BioMed Research International



Journal of Ophthalmology

Computational and Mathematical Methods in Medicine



Stem Cells International



Behavioural Neurology

CAM

Evidence-Based Complementary and Alternative Medicine







Research and Treatment





Oxidative Medicine and Cellular Longevity