

Review Article

Role of miRNA *Let-7* and Its Major Targets in Prostate Cancer

Siegfried Wagner,^{1,2} Anaclet Ngezahayo,² Hugo Murua Escobar,^{1,3} and Ingo Nolte¹

¹ *Small Animal Clinic, University of Veterinary Medicine Hannover, 30559 Hannover, Germany*

² *Institute of Biophysics, University Hannover, 30419 Hannover, Germany*

³ *Division of Medicine, Department of Haematology/Oncology, University of Rostock, 18057 Rostock, Germany*

Correspondence should be addressed to Ingo Nolte; ingo.nolte@tiho-hannover.de

Received 16 April 2014; Revised 11 August 2014; Accepted 18 August 2014; Published 3 September 2014

Academic Editor: Andreas Doll

Copyright © 2014 Siegfried Wagner et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Prostate cancer is worldwide the sixth leading cause of cancer related death in men thus early detection and successful treatment are still of major interest. The commonly performed screening of the prostate-specific antigen (PSA) is controversially discussed, as in many patients the prostate-specific antigen levels are chronically elevated in the absence of cancer. Due to the unsatisfying efficiency of available prostate cancer screening markers and the current treatment outcome of the aggressive hormone refractory prostate cancer, the evaluation of novel molecular markers and targets is considered an issue of high importance. MicroRNAs are relatively stable in body fluids orchestrating simultaneously the expression of many genes. These molecules are currently discussed to bear a greater diagnostic potential than protein-coding genes, being additionally promising therapeutic drugs and/or targets. Herein we review the potential impact of the microRNA *let-7* family on prostate cancer and show how deregulation of several of its target genes could influence the cellular equilibrium in the prostate gland, promoting cancer development as they do in a variety of other human malignant neoplasias.

1. Introduction

Prostate cancer (PC) is a heterogeneous disease ranging from an asymptomatic to a fatal systemic malignancy [1]. According to the World Health Organization (WHO) 1,111,689 men were estimated to be diagnosed with PC in the year 2012 (<http://globocan.iarc.fr/>). Accounting worldwide for 6.6% (307,471) of all cancer death in men in 2012, PC is one of the most common malignant neoplasias and the sixth leading cause of cancer related death in men (<http://globocan.iarc.fr/>).

The development of PC is considered to be a multi-step process initiated by genetic and epigenetic changes [1]. Human PC is commonly accepted to be an androgen dependent malignancy.

An analysis of PC related metastatic pattern in 1,589 patients by Bubendorf et al. revealed that 35% of the analyzed tumors spread to other organs with preference to the bones (90%), lungs (46%), liver (25%), pleura (21%), and adrenals (13%) [2].

The androgen deprivation therapy is actually the most effective palliative standard treatment for primary advanced

PCs with bone metastasis (effective in up to ~90% of patients). However, the great majority of patients relapse subsequently due to the development of castration resistance [3].

Since the introduction of the prostate-specific antigen (PSA) test in the 1990s, the number of diagnosed cases has been rapidly rising being initially associated with a reduced mortality. However, the recent decline in PC related mortality rates is now being discussed to be partially explained by the improved treatment and earlier diagnosis due to a broad standard PSA screening in economically developed countries [4, 5]. As the standard PSA screening in the early diagnosis of human PC remains a very controversial issue, novel, reliable molecular PC markers are needed [6–8].

A promising marker candidate gene is the miRNA *let-7*, which was reported to be down regulated among others in human PC [9–11]. Further, the reconstitution of the *let-7* expression resulted in suppression of PC cell proliferation [10, 12]. In general a single miRNA is able to regulate a huge number of genes. Concerning *let-7* the respective acting ways are actually not entirely deciphered.

Nevertheless, it is to be expected that a deeper understanding of the molecular interactions of *let-7* and associated genes will significantly contribute to the development of novel diagnostic and therapeutic treatment modalities for PC.

Due to the complex regulation mechanisms of *let-7* and its potential role in PC development and relapse the present review highlights *let-7* and its direct and downstream targets in the context of PC.

2. Micro RNA *Let-7* Family

MicroRNAs (miRNAs) are small, non-protein-coding RNAs derived from long, endogenously expressed primary RNA (pri-miRNA) molecules. These pri-miRNAs are processed by the nuclear enzyme Drosha to precursor RNAs (pre-miRNAs), exported by Exportin-5 [13] and matured by the cytoplasmic enzyme Dicer [14]. Finally the guide strand of the mature miRNA is incorporated into the RNA-induced silencing complex (RISC), which blocks the translation of the target mRNA by binding to its 5'-, 3'-prime, or exon regions [15, 16]. The passenger strand is usually degraded [17] (Figure 1).

Mature miRNAs are known to be part of the gene expression regulating machinery at transcriptional [18, 19] and as well posttranscriptional level [13]. It was reported that a single miRNA can orchestrate the expression of several genes and a single gene can be regulated by a set of different miRNAs [20–22]. Several observations suggest that more than 60% of all protein coding genes are regulated by miRNAs [23].

One of the first described members of the large class of non-protein-coding RNAs is *let-7* which was the second miRNA discovered and designated as *lethal-7* (*let-7*) according to the phenotype of a *let-7* deficient *C. elegans* mutant [20]. Soon thereafter, further *let-7* homologs were identified in a variety of species ranging from vertebrates to mollusks [24].

In contrast to “less complex” organisms such as worms, vertebrates show a higher number of *let-7* isoforms coded by different genes [16]. In humans, 13 *let-7* family precursor miRNAs were described (*let-7a-1*, *let-7a-2*, *let-7a-3*, *let-7b*, *let-7c*, *let-7d*, *let-7e*, *let-7f*, *let-7g*, *let-7i*, *miR-98*, and *mir-202*) which code for 10 different mature *let-7* miRNA isoforms [25].

Although the role of *let-7* is still not fully understood, it is evident that the *let-7* family members have a distinct expression pattern in animal development [26]. In the embryonic stage the *let-7* miRNAs were found to be barely detectable, but having an increased expression in differentiated cells [20, 27]. Furthermore, aberrant *let-7* expression was associated with a variety of human diseases as, for example, cardiovascular diseases [28], liver fibrosis [29], lung diseases [30], and cancer [9–12, 26, 31–34]. Interestingly several *let-7* family members were found to be located at fragile sites of human chromosomes potentially contributing to aberrant *let-7* transcript levels [35].

Cancer initiation, progression, and aggressiveness are hypothesized to be driven by cancer stem cells (CSCs)

[36, 37]. Inflammatory microenvironment [38] as well as epithelial-to-mesenchymal transition (EMT), which is tightly linked with CSC biology [39], seems to play a substantial role in cancer etiology as well. Remarkably, a linkage between these factors is the *let-7* miRNA family. As described above *let-7* was shown to be downregulated in prostatic CSCs [36] whereas reconstitution of the *let-7* suppressed the growth of PC cells [10, 12]. Additionally, a direct causal link between cancer and inflammation is given by the association of *let-7*, IL6, and NFκB, which are major players involved in the epigenetic switch from inflammation to cell transformation [31]. The connection between EMT and *let-7* is represented by the *HMGAI* and *HMG2* genes, which are directly regulated by *let-7* and were found to be implicated in EMT [40, 41].

Further, miRNAs of the *let-7* family were reported to directly, negatively regulate *IL6* [24], *NRAS* [42], *c-Myc*, *HMGAI* [43, 44], *HMG2* [45], and *CCND2* [11]. Notably, these *let-7* targets are involved in a wide range of diverse cellular processes interwoven with *let-7* and each other in a fine balanced way (Figure 10).

The *c-Myc* protein regulates the biogenesis of *let-7* by stimulating *Lin28* [46], *Lin28* in turn blocks the maturation of *let-7* [47]. Additionally, *c-Myc* stimulates the expression of *HMGAI* [48], *AR* [12], and *IL6* [49]. *NRAS* is suggested to have an impact on *HMG2* biogenesis [45]. *HMG2* on the other hand influences *HMGAI*, its gene product in turn regulates the expression of *c-Myc* [50] and *HMGBI* [51]. *HMGBI* was found to bind the *AR* promoter [52], *AR* protein was described itself to stimulate *let-7* expression [53] (Figure 10).

Interestingly, the *let-7* family [10, 11] and some of its above mentioned targets were already found to be implicated in PC. As *let-7* is linked with all these protein-coding genes a deeper insight into these connections is of great interest. Thus, these interactions are reviewed more detailed in the following parts.

3. *HMGAI*

The high mobility group proteins (HMG) are chromatin associated nonhistone proteins constituting three superfamilies (HMGA, HMGB, and HMGN) which are classified by their characteristic functional DNA-binding motifs [54]. Expression of these proteins was described to be involved in a variety of biological processes as, for example, transcription, embryogenesis, differentiation, neoplastic transformation, apoptosis, and inflammation [52, 55, 56].

In human neoplasias the HMGA genes are among the most commonly rearranged genes [57]. Deregulation of the *HMGAI* expression was described in human PC [58, 59], lung cancer [60], and breast cancer cells [61]. Its oncogenic property is speculated to be partly mediated through the cytoplasmic relocation of the HIPK2 which is a proapoptotic activator of the tumor suppressor p53 [51] (Figures 2 and 10). *HMGAI* was reported to enhance the proliferation rate and invasion of PC cells [62, 63] potentially through the implication in epithelial-to-mesenchymal transition (EMT) [61]. In line with this, *HMGAI* knock down in the human triple-negative breast cancer cell lines MDA-MB-231 and Hs578T

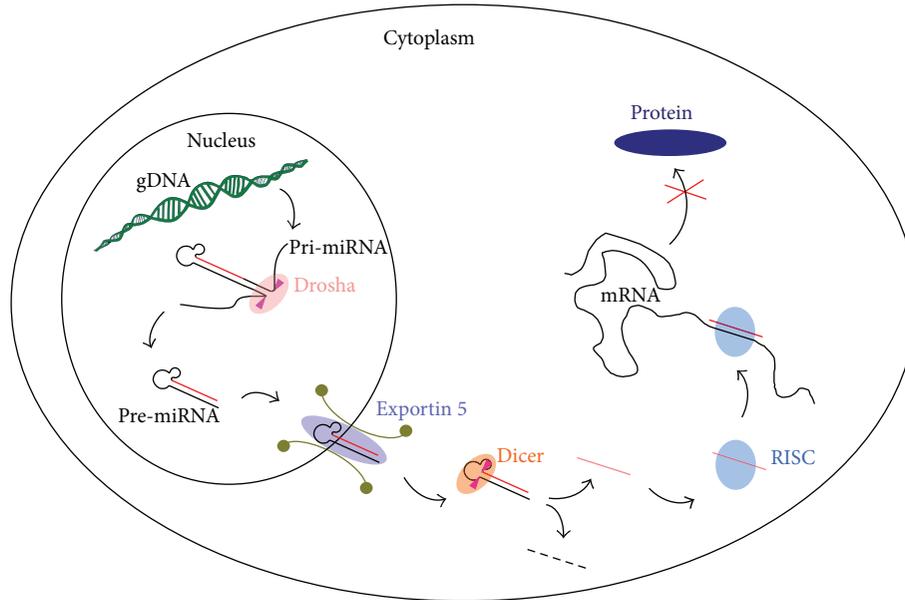


FIGURE 1: Schematic overview of the miRNA maturation and the way of function in eukaryotic cells. The endogenously expressed primary miRNA (pri-miRNA) is processed in the nucleus of a cell by the enzyme Drosha. The emerging precursor miRNA (pre-miRNA) product is exported into the cytoplasm by Exportin-5 and matured by Dicer. Finally the guide strand is incorporated into the RNA-induced silencing complex (RISC), which blocks the translation of the target mRNA.

repressed the mesenchymal gene *SNAIL* and stimulated *CDH1* expression [40] (Figures 2 and 10), both of which are involved in EMT [64, 65]. Furthermore, *HMGAI* was reported to drive tumor progression by reprogramming cells to a stem-cell-like state [40]. In accordance, Shah et al. reported in human embryonic stem cells (hESCs) a significant downregulation of the stemness-associated genes *OCT4*, *Sox2*, *Lin28*, and *c-Myc* 96 h after *HMGAI* repression [50]. Interestingly *HMGAI* is not only stimulating *c-Myc* expression [50] it was also reported to be itself induced by *c-Myc* [48] (Figures 2 and 10). It is remarkable that *HMGAI* is implicated in the upregulation of several miRNAs in murine embryonic fibroblasts. Among these miRNAs is the *miR-196a-2*, which in turn is predicted to target its sister gene *HMGAI* [66] (Figures 2 and 10). Furthermore, Hillion and colleagues reported a positive correlation between *HMGAI* and *STAT3* in a subset of primary human acute lymphoblastic leukemia samples [67]. In line with this, *HMGAI* was described to bind the *STAT3* promoter and to upregulate its expression in malignant human hematopoietic cells [67] (Figures 2 and 10). The transcription factor *STAT3* mediates uncontrolled growth, angiogenesis, and survival of cells and has a great potential as target in cancer therapies [68]. Remarkably, Iliopoulos et al. identified *STAT3* binding sites in the promoters of the miRNAs *miR-181b* and *miR-21* [69] (Figures 2 and 10). These tiny regulators in turn were found to block *PTEN* (Figures 2 and 10), stimulating the activity of *NFκB* [69]. The tumor suppressor *PTEN* functions as an antagonist of *PI3K* by dephosphorylating its product *PIP3* [70] (Figure 10).

The *HMGAI* and *HMGAI2* genes were reported to be highly expressed during embryogenesis, reexpressed in several cancer types but to be absent or not detectable in most of the adult healthy tissues [57, 71]. The expression of both

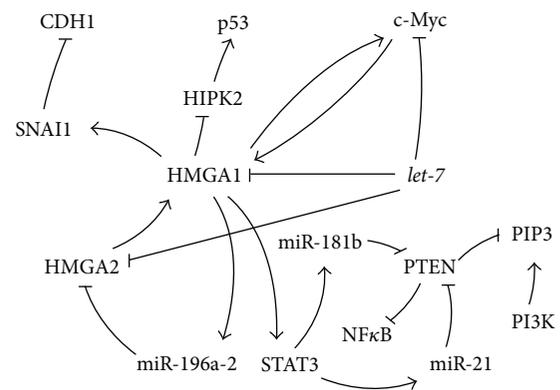


FIGURE 2: Overview of the described interactions between *let-7* and its direct target *HMGAI* with other genes.

HMGAI, *HMGAI2*, and of its regulator *let-7* was shown to be negatively correlating in gastroenteropancreatic neuroendocrine tumors [44] and retinoblastomas [72]. In accordance they were found to be directly, negatively regulated by *let-7* [45, 73, 74] (Figures 2 and 10).

4. *HMGAI2*

Comparable to the described *HMGAI* knock down, the repression of *HMGAI2* in the human PC cell line PC-3 induced an upregulation of *CDH1* indicating an important role in EMT [41]. *SNAIL* and *SNAIL2* are repressors of *CDH1* and were shown to be directly activated by *HMGAI2* [45] (Figures 3 and 10).

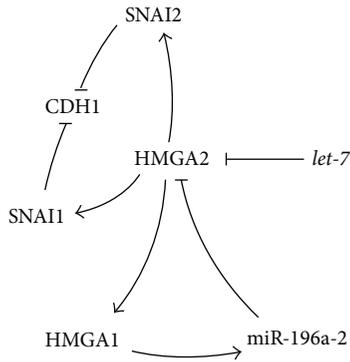


FIGURE 3: Association of the HMGA2/*let-7* axis with the regulation of genes involved in EMT and miRNAs.

Similar to *HMGA1*, an upregulation of *HMGA2* was reported in human lung and breast cancers [75, 76] as well as in a subset of canine PCs [77]. Furthermore, *HMGA2* was recently described to modify gene expression not only as protein but as well as a competing endogenous RNA (ceRNA) by acting as a decoy for mature *let-7* miRNAs [78]. Interestingly, a stimulating *HMGA2* influence on the expression of its sister protein *HMGA1* was found in rat epithelial thyroid cells [79], thus constituting a feedback loop by the stimulation of its suppressor, the *miRNA-196a-2* [66] (Figures 3 and 10). Remarkably *HMGA2* was described to bear seven *let-7*-binding sites in its 3'-untranslated region (3'-UTR) [33]. Aberrations of the chromosomal region 12q14-15 that affect *HMGA2* were frequently found in human cancers [80–82]. Moreover, the disrupted pairing between *let-7* and *HMGA2* by mRNA truncations of the 3'UTR was reported to induce *HMGA2* overexpression leading to tumor formation [33].

5. HMGB1

The high mobility group box 1 (HMGB1) is one of the HMGB superfamily members which was also shown to be implicated in inflammation exercising cytokine like functions [83]. In line with its multiple roles it can be located in the nucleus as well as in the cytoplasm and can even be released passively by necrotic cells or actively secreted in response to inflammatory signals by certain cell types [83, 84].

This proinflammatory cytokine exerts its function by interacting with the toll-like receptors (TLR) 2, and TLR4 and RAGE [85–87] (Figures 4 and 10). Interestingly, the receptor coding gene *TLR4* was found to be a direct *let-7i* target (Figure 4), presenting a mechanism to modify the HMGB1 signaling [88]. The activation of the HMGB1 receptor RAGE results among others in deactivation of MAPK1 and PI3K [89]. PI3K in turn was shown to stimulate NFκB [90]. Furthermore, the TLRs and RAGE were demonstrated to activate NFκB thus, inducing the secretion of angiogenic factors, growth factors, and cytokines [85, 91].

Remarkably, NFκB is able to stimulate RAGE expression by binding to its promoter constituting a positive feedback loop [92] (Figures 4 and 10). Blockade of the RAGE/HMGB1

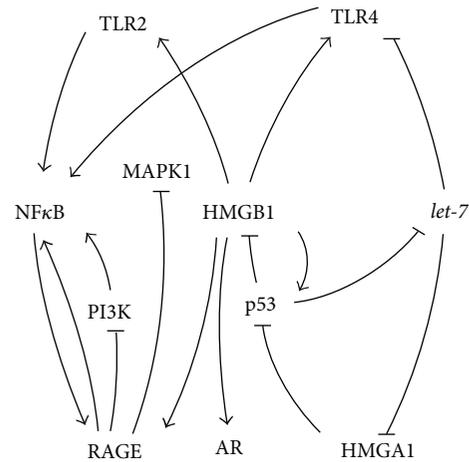


FIGURE 4: Schematic overview over the HMGB1 and *let-7* regulatory pathways affecting each other's activity.

signaling decreased growth and metastasis of implanted and as well of spontaneously developing tumors in susceptible mice [93].

HMGB1 was described to be involved in all proposed hallmarks of cancer and is thus a potential target for therapeutic and diagnostic approaches [94]. Kuniyasu et al. observed the secretion of HMGB1 in primary cultured human prostatic stromal cells after androgen deprivation [95]. *In vitro* suppression of HMGB1 was demonstrated to block the invasion of PC-3 cells which was reversed by culturing the cells in conditioned medium of the above-mentioned stromal cells deprived of androgen [95, 96]. Additionally, HMGB1 was found to stimulate DNA binding of several steroid receptors including the *let-7* downstream target AR (Figure 10) [97]. These facts indicate that *HMGB1* may be a molecular marker for advanced prostate cancer [95, 96].

Although *HMGB1* was not shown to be a direct *let-7* target, its expression is modulated by the direct *let-7* target *HMGA1* [51]. Interestingly HMGB1 was also shown to be involved in the p53 network by facilitating the binding of the tumor suppressor p53 to its cognate DNA [98]. As mentioned before p53 can be inactivated by the HMGB1 sister protein *HMGA1* by translocation of the p53 activator HIPK2 [51] (Figures 4 and 10). The tumor suppressor p53 in turn was found to downregulate the activity of the HMGB1 promoter [99] and to trigger the radiation induced decrease of *let-7a* and *let-7b* expression (Figures 4 and 10) in the human colon cancer cell line HCT116 [100].

6. CCND2

Many tumor cells accumulate mutations resulting in uncontrolled proliferation due to direct or indirect deregulations of the cyclin-dependent kinases (CDKs). Cyclins are known regulating subunits of CDKs being expressed at specific time points during the cell cycle. Consequently cyclin deregulations induce uncontrolled cell proliferation [101].

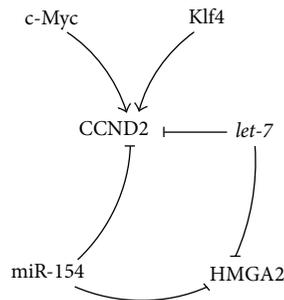


FIGURE 5: *Let-7* and *CCND2* mediated gene regulation.

The cyclin D2 (*CCND2*) is one of the cell cycle regulating factors. This gene, which is highly conserved among mammals, has been associated with human prostate cancer [11], gastric cancer [102], colon cancer [103], and leukemia [104].

Interestingly, *CCND2* was shown to be a direct *let-7* and *miR-154* target like *HMGA2* [11, 41, 45, 105] (Figures 5 and 10). Additionally the *let-7* regulated oncogene *c-Myc* and the stem cell marker *Klf4* were reported to stimulate the *CCND2* transcription [106, 107] (Figures 5 and 10).

Dong et al. described that ectopically overexpressed *let-7a* induced cell cycle arrest at the G1/S phase by suppressing among others the cyclin *CCND2* and additionally inhibited the proliferation of the human prostatic cell lines PC-3 and LnCap [11]. The same group reported that in nude xenograft mice, inoculated with *let-7a* transfected PC-3 cells, the tumor was 80% lighter after 4 weeks of growth compared to controls [11].

7. *c-Myc*

c-Myc is an oncogene frequently activated in human cancers, but is low expressed or absent in quiescent cells [108–110]. In contrast, its overexpression has been connected with PC formation and progression [111, 112]. This gene encodes a transcription factor that has a great impact on the global gene expression pattern and, thus, influences cell-cycle progression, glucose and glutamine metabolism, lipid synthesis, and many other processes, which contribute to tumor progression [109].

Mitogen activated protein kinases (MAPK), glycogen synthase kinase 3 (GSK3), and other CDKs play a key role in the biological function and half-life of *c-Myc* proteins by posttranslational phosphorylation of the Thr58 and Ser62 sites [113] (Figures 6 and 10). Apart from various posttranslational protein modifications and transcriptional regulations of the *c-Myc* gene products, this gene was reported to be directly negatively regulated by members of the *let-7* family [114, 115] (Figures 6 and 10). Additionally, elevated MAPK1 activity, which was associated with advanced, androgen independent human PCs, [116] was demonstrated to influence the *c-Myc* protein, resulting in prolonged function in a human muscle-derived rhabdomyosarcoma cell line [117]. In line with the functions of *c-Myc*, MAPK1 controls diverse cellular processes as growth, differentiation, migration, and apoptosis, its deregulation has often been described to be

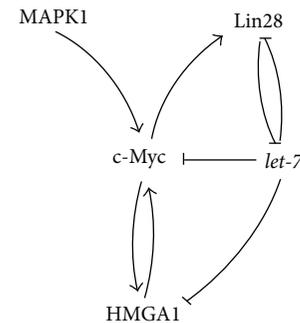


FIGURE 6: Interactions of the oncogene *c-Myc* with *let-7* and MAPK1, Lin28, and HMGA1.

associated with cancer [118]. Furthermore, *c-Myc* was shown to transcriptionally activate *Lin28* [119], which in turn inhibits the biogenesis of its regulator *let-7* constituting a double negative feedback loop [47] (Figures 6 and 10). Interestingly the expression of the direct *let-7* target *HMGA1* is as well induced by *c-Myc* [48], which constitutes a positive feedback loop, stimulating *c-Myc* expression [50] (Figures 6 and 10).

8. IL6

Chronic inflammation of the prostate gained major attention as it is considered to account to the factors contributing to PC [120]. In previous reports a direct causal link between cancer and inflammation has been described with IL6, *let-7*, Lin28, and $\text{NF}\kappa\text{B}$ being the major players involved in the epigenetic switch from inflammation to cell transformation [31].

Originally identified as an inducer of the terminal differentiation of B-cells into antibody-producing cells [121] interleukin-6 (IL6) appears to be a major regulator of prostate cancer progression [122]. Notably, IL6 is not only released by inflammatory cells but also found to be released by hormone insensitive cell lines DU145 and PC-3 but not by the hormone sensitive LNCaP cells [123]. Furthermore, this pleiotropic cytokine stimulates growth and survival of human PC and promotes its progression [123, 124]. In accordance, increased IL6 levels were found in epithelial cells of PC compared to benign tissues [125]. Moreover, Giri et al. reported a ~18 times higher IL6 expression in malignant prostate tissues compared to “normal” prostate specimens [126]. Michalaki and colleagues described significantly higher IL6 serum levels in patients with metastatic prostatic disease [127].

The biological activities of IL6 are mediated by binding to the α -subunit receptor IL6R and the following association with the ubiquitously expressed signal-transducing β -subunit gp130 [128]. Upon engagement of gp130 various Janus tyrosine kinase (JAK) family members (JAK1, JAK2, JAK3, and Tyk2) [129] are activated by ligand induced receptor oligomerization phosphorylating themselves and the intracellular domains of the receptors [130]. Once gp130 is phosphorylated the second protein family, the signal transducer and activator of transcription (STAT), binds to the intracellular domain of the receptor. This leads to the activation of STATs and the subsequent dissociation, allowing

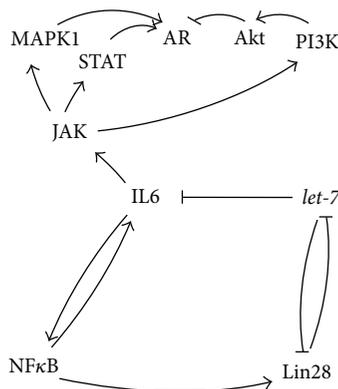


FIGURE 7: IL6 regulated genes.

STAT dimerization and translocation into the nucleus where they act as transcription factors [131]. Additionally IL6 was shown to stimulate the PI3K and MAPK pathways by signaling through activated gp130 [132, 133].

Interestingly, LnCaP cells stimulated with IL6 presented an enhanced AR activity in the absence of a ligand [134, 135]. The IL6 mediated activation of the human AR was indicated to be mediated by STAT3 and MAPK signaling [134, 136], which potentially contribute to recurrence of hormone refractory PCs. Whereas the AR transactivation can be suppressed by the PI3K/AKT pathway. Thus, these three pathways are suggested to coordinately regulate AR activation [136].

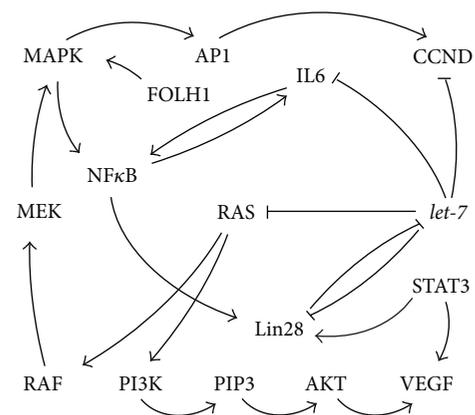
Acquiring resistance to apoptosis appears an important feature for the development of hormone resistant and aggressive human prostate cancer. Furthermore, IL6 was shown to act as a survival factor, blocking apoptosis induced by Bcl-xl, p53, TGF β [137], and cytotoxic agents such as doxorubicin [138] and enzalutamide [139]. Whereas siRNA or STAT3-inhibitor-AG490 mediated suppression of the downstream acting STAT diminished the IL6 induced antiapoptotic function [138, 139].

NF κ B is a regulator of the transcription of *IL6* [140] and *Lin28B* [31, 141] (Figures 7 and 10). Lin28B was demonstrated to block the maturation of *let-7* [46]. Additionally, members of the miRNA *let-7* family directly target *IL6*, which in turn constitutes a positive feedback loop on NF κ B [31, 49] (Figures 7 and 10).

Remarkably, while only a few cells express membrane bound IL6R all cells display gp130 on their surface [132]. This is an interesting feature as IL6 can also bind to a soluble IL6R (sIL6R) variant, which interacts in an IL6R agonistic manner with gp130, thus, enabling the stimulation of cells lacking endogenous IL6R [142].

9. RAS

The founding members of the *RAS* gene superfamily *N-RAS*, *H-RAS*, and *K-RAS* are coding for small GTP-binding proteins [143]. Originally identified as retroviral oncogenes in rat sarcomas, *RAS* were the first human oncogenes discovered, shown to be mutated in around 30% of all human tumors

FIGURE 8: Schematic overview over some of the numerous pathways modified by RAS and *let-7*.

[144, 145]. The very common mutations in the residues G12, G13, and Q61 lock RAS in a constitutively activated state by impairing the intrinsic GTP hydrolysis [145, 146]. RAS proteins are active when they have bound GTP. By hydrolyzing GTP to GDP they become inactive. The intrinsic GTPase activity of the RAS proteins is very low relying on the help of specialized GTP hydrolysis accelerating factors called GTPase activating proteins (GAP) which increase the hydrolysis by more than 100,000 fold [145].

RAS-GTPs are acting as signal transducers across membranes by binding various effector proteins to stimulate signaling pathways [143, 147]. Among these factors are the Raf serine/threonine MAPKK kinases (ARAF, BRAF, and RAF1) which in turn activate MEK-MAPK cascades [148] (Figures 8 and 10). Accordingly, the mammalian MAPK pathways are estimated to be deregulated in one-third of all human cancers [149]. MAPKs activate cytosolic and nuclear factors like JUN and ELK1, which are regulating FOS expression. JUN and FOS are forming the activator protein 1 (AP1) and, thus, influencing the expression of proteins such as CCNDs which are involved in cell-cycle progression [150] (Figures 8 and 10).

Furthermore, RAS-GTPs induce the translocation and subsequent activation of phosphatidylinositol 3-kinase (PI3Ks) by binding to its catalytic subunit [151] (Figures 7 and 10). PI3K signaling is one of the most often deregulated systems in human cancer [152]. Taylor et al. described that the PI3K expression is altered in 42% of the primary and in 100% of the metastatic cases in the analyzed set of human prostatic cancers [153]. PI3Ks belong to one of the main effector molecules of RAS [151]. This enzyme type phosphorylates primarily to the 3'-OH group of the membrane bound phosphatidylinositol-4,5-biphosphate (PIP) to generate the messenger phosphatidylinositol-3,4,5-biphosphate (PIP3) [154]. PIP3 activates itself several pleckstrin homology domain-containing proteins as Akt by directly binding and recruiting it to the plasma membrane [154] (Figures 8 and 10). The activated Akt promotes many processes contributing to a malignant tumor phenotype [155]. Ectopic expression of a constitutively active Akt in the thyroid cell line SW579 was reported to significantly increase VEGF levels [156] (Figures 8 and 10). Neovascularization and angiogenesis are essential

features for the progression of a growing tumor VEGF is one of the most important inducers of angiogenesis [157]. Niu et al. demonstrated a positive correlation between VEGF and a constitutively active STAT3 [157]. In accordance, it was found that STAT3 binds to the *VEGF* promoter [157] (Figures 8 and 10). Additionally STAT3 was reported to bind the promoter of the *let-7* biogenesis regulating gene *Lin28*, resulting in the concomitant upregulation of the *let-7* targets *RAS*, *c-Myc*, and *HMGA2* [158].

In human tissues the activation of RAS and Rac-MAPK pathways was described to be induced by the extracellular signal transducer FolH1 [159] (Figures 8 and 10). FolH1 is expressed in most of the human prostate cancers and is thus a potential target for diagnostic and therapeutic strategies [160]. The elicited phosphorylation of MAPK1 and MAPK14 induces in turn the activation of the transcription factor NFκB (Figures 8 and 10) which controls the expression of various genes including the *let-7* biogenesis-controlling *Lin28* [47] and the cytokine IL6 [31, 161] (Figures 8 and 10). Additionally, NFκB was also shown to enhance the endogenous transcription of the primary miRNAs *let-7a-3* and *let-7b* through NFκB responsive binding sites in the promoter regions [141] (Figures 8 and 10).

Remarkably, Johnson et al. reported numerous *let-7* binding sites in the 3'-UTR of the RAS genes [42]. In conclusion the expression of the oncogenes *NRAS*, *KRAS*, and *HRAS* was described to be negatively regulated by several members of the *let-7* family [42, 162] (Figures 8 and 10).

10. Androgen Receptor (AR)

The gene of the steroid receptor family member *AR* [163] is located on the human chromosome X and codes for a ligand-dependent transcription factor [164, 165]. Upon ligand binding it translocates into the nucleus and regulates its target genes by binding to the androgen response elements (AREs) [166, 167]. Expressed in nearly all primary human PCs, AR plays a pivotal role in carcinogenesis of the prostate. At the initial diagnosis the majority of PCs depends on androgens and progress after hormone therapy to an androgen-independent disease [3, 168].

Continuous androgen expression is required to drive prostate gland formation during embryogenesis and later to maintain the normal function and glandular anatomy in adults [169]. In general the androgen mediated effects in prostate gland development are driven by the interaction with ARs [169]. The bypass mechanisms of *AR* upregulation include among others the HMGB1 enrichment on the *AR* promoter, which enhances the transcription [52] (Figures 9 and 10), an intracrine androgen production [170, 171] additionally ligand independent *AR* activation by cytokines or growth factors were reported as well [172]. Furthermore, altered specificity or sensitivity as for example by alternative splicing is discussed [173].

However, the activated *AR* stimulates the expression of its targets as, for example, the above mentioned VEGF [174] and PSA [175]. PSA is a pivotal downstream target of *AR*, which is used as biomarker for human PC progression [175]. Interestingly, the frequently observed rising of serum PSA in

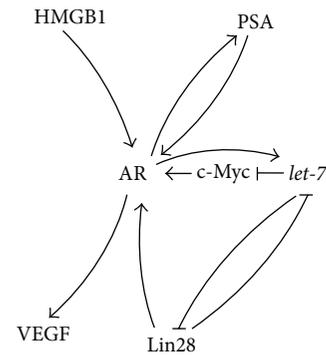


FIGURE 9: Potential AR interaction in prostate cancer development.

castrate-resistant PC patients could in parts be explained by *AR* activity, which is reexpressed/reactivated in advanced PCs [176]. Remarkably, PSA constitutes a positive feedback loop stimulating *AR* expression as was demonstrated *in vitro* [175] (Figures 9 and 10).

Furthermore, Tummala et al. highlighted the impact of the *Lin28/let-7/Myc* axis on PC and demonstrated that *Lin28* activates the *AR* (Figures 9 and 10) and promotes growth of PC [177].

Remarkably *AR* was reported to be regulated in a negative way by the miRNA *let-7c* which suppresses its transcriptional activator *c-Myc* [12] (Figures 9 and 10). Additionally Lyu et al. described an *AR* induced upregulation of *let-7a*, *let-7b*, *let-7c*, and *let-7d* (Figures 9 and 10) in the breast cancer cell lines MDA-MB-231 and MDA-MB-453. At least in the case of *let-7a* this upregulation is indicated to be triggered by *AR* binding to AREs located at the *let-7a* promoter [53] (Figures 9 and 10). Furthermore, it was shown that in these cell lines the expression of the direct *let-7a* targets *c-Myc* and *KRAS* was decreased upon treatment with 5α-dihydrotestosterone and increased after an additional suppression of the miRNA *let-7a* [53].

The spatiotemporal expression of genes and functions depend highly on the cellular and developmental context. Thus, the impact of a single gene can be completely different between diverse tissues and at different time points in development. Nevertheless elucidation of the above described interactions in PC bears great potential due to the ubiquitous existence of the cellular regulatory elements and the potential interactions in each somatic cell of an organism. This idea is supported by the already found implication of each of the described genes in various human cancers. Furthermore several of the reviewed genes are already used as targets for diagnostic, prognostic, and therapeutic approaches. Thus, the master regulator family *let-7* is as well a promising target in cancer of the prostate gland.

For a better overview all described interactions between the master regulator family *let-7* and its major targets are summarized in Figure 10.

11. Conclusion

Although the knowledge of the genetic and epigenetic alterations in prostate cancer has significantly increased in the last

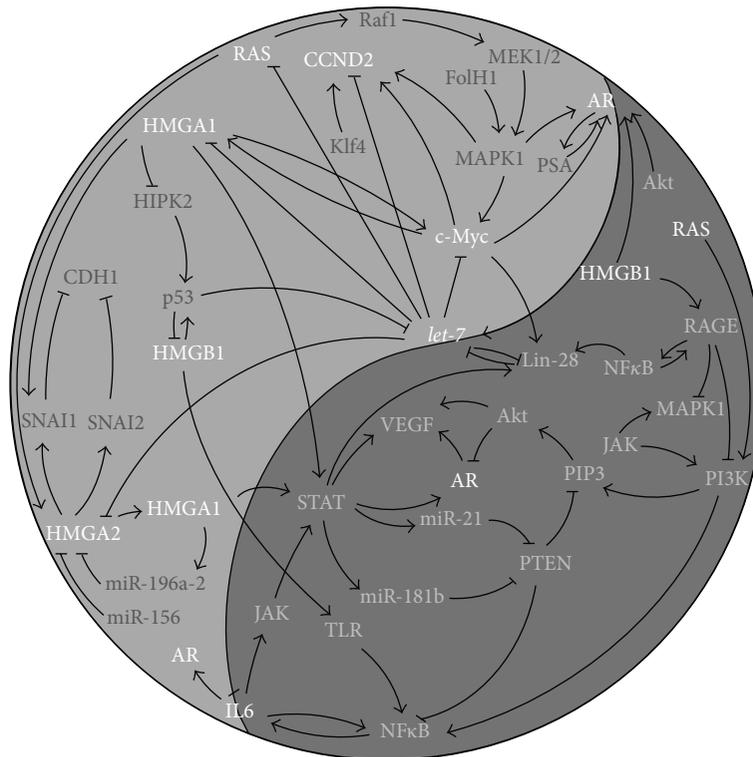


FIGURE 10: This figure represents the described interactions between *let-7* and the reviewed *let-7* associated targets (in white letters) with other genes which are as well commonly deregulated in human cancers (in gray letters). The indicated interactions are on transcriptional, posttranscriptional or posttranslational level.

decades, its diagnosis and therapy still remains a major challenge. The actually described genetic alterations in prostate cancer give more questions than answers. As we could highlight, the genes reviewed in the present paper are not acting in solitude but are closely interwoven with each other (Figure 10). Remarkably, the miRNA *let-7* family members are major players in the regulation of gene expression and appear to contribute greatly to the maintenance of the Ying and Yang in “normal” prostatic cells. However, their impact can be modified greatly by other factors. For that reason the complex intra- and intercellular genetic interactions of *let-7* family members and associated genes must be further investigated and will likely have an impact on diagnostic, prognostic, and treatment modalities in future.

Conflict of Interests

The authors have no conflict of interests.

References

- [1] L. Kopper and J. Tímár, “Genomics of prostate cancer: is there anything to “translate?” *Pathology & Oncology Research*, vol. 11, no. 4, pp. 197–203, 2005.
- [2] L. Bubendorf, A. Schöpfer, U. Wagner et al., “Metastatic patterns of prostate cancer: an autopsy study of 1,589 patients,” *Human Pathology*, vol. 31, no. 5, pp. 578–583, 2000.
- [3] R. T. Divrik, L. Türkeri, A. F. Şahin et al., “Prediction of response to androgen deprivation therapy and castration resistance in primary metastatic prostate cancer,” *Urologia Internationalis*, vol. 88, no. 1, pp. 25–33, 2012.
- [4] A. Jemal, F. Bray, M. M. Center, J. Ferlay, E. Ward, and D. Forman, “Global cancer statistics,” *CA Cancer Journal for Clinicians*, vol. 61, no. 2, pp. 69–90, 2011.
- [5] R. T. Greenlee, T. Murray, S. Bolden, and P. A. Wingo, “Cancer statistics, 2000,” *CA: A Cancer Journal for Clinicians*, vol. 50, no. 1, pp. 7–33, 2000.
- [6] G. L. Andriole, E. D. Crawford, R. L. Grubb III et al., “Mortality results from a randomized prostate-cancer screening trial,” *New England Journal of Medicine*, vol. 360, no. 13, pp. 1310–1319, 2009.
- [7] F. H. Schröder, J. Hugosson, M. J. Roobol et al., “Screening and prostate-cancer mortality in a randomized european study,” *The New England Journal of Medicine*, vol. 360, no. 13, pp. 1320–1328, 2009.
- [8] E. Basch, T. K. Oliver, A. Vickers et al., “Screening for prostate cancer with prostate-specific antigen testing: american society of clinical oncology provisional clinical opinion,” *Journal of Clinical Oncology*, vol. 30, no. 24, pp. 3020–3025, 2012.
- [9] H. Ramberg, A. Alshbib, V. Berge, A. Svindland, and K. A. Taskén, “Regulation of PBX3 expression by androgen and Let-7d in prostate cancer,” *Molecular Cancer*, vol. 10, article 50, 2011.
- [10] N. Nadiminty, R. Tummla, W. Lou et al., “MicroRNA let-7c is downregulated in prostate cancer and suppresses prostate cancer growth,” *PLoS ONE*, vol. 7, no. 3, Article ID e32832, 2012.
- [11] Q. Dong, P. Meng, T. Wang et al., “MicroRNA let-7a inhibits proliferation of human prostate cancer cells *in vitro* and *in vivo*

- by targeting E2F2 and CCND2," *PLoS ONE*, vol. 5, no. 4, Article ID e10147, 2010.
- [12] N. Nadiminty, R. Tummala, W. Lou et al., "MicroRNA let-7c suppresses androgen receptor expression and activity via regulation of myc expression in prostate cancer cells," *Journal of Biological Chemistry*, vol. 287, no. 2, pp. 1527–1537, 2012.
- [13] D. P. Bartel, "MicroRNAs: genomics, biogenesis, mechanism, and function," *Cell*, vol. 116, no. 2, pp. 281–297, 2004.
- [14] Y. Lee, C. Ahn, J. Han et al., "The nuclear RNase III Drosha initiates microRNA processing," *Nature*, vol. 425, no. 6956, pp. 415–419, 2003.
- [15] G. Meister, "Argonaute proteins: functional insights and emerging roles," *Nature Reviews Genetics*, vol. 14, no. 7, pp. 447–459, 2013.
- [16] V. Mondol and A. E. Pasquinelli, "Let's make it happen: the role of let-7 microRNA in development," *Current Topics in Developmental Biology*, vol. 99, pp. 1–30, 2012.
- [17] W. Filipowicz, S. N. Bhattacharyya, and N. Sonenberg, "Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight?" *Nature Reviews Genetics*, vol. 9, no. 2, pp. 102–114, 2008.
- [18] H.-W. Hwang, E. A. Wentzel, and J. T. Mendell, "A hexanucleotide element directs microRNA nuclear import," *Science*, vol. 315, no. 5808, pp. 97–100, 2007.
- [19] R. F. Place, L.-C. Li, D. Pookot, E. J. Noonan, and R. Dahiya, "MicroRNA-373 induces expression of genes with complementary promoter sequences," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 5, pp. 1608–1613, 2008.
- [20] B. J. Reinhart, F. J. Slack, M. Basson et al., "The 21-nucleotide let-7 RNA regulates developmental timing in *Caenorhabditis elegans*," *Nature*, vol. 403, no. 6772, pp. 901–906, 2000.
- [21] P.-S. Chen, J.-L. Su, S.-T. Cha et al., "miR-107 promotes tumor progression by targeting the let-7 microRNA in mice and humans," *The Journal of Clinical Investigation*, vol. 121, no. 9, pp. 3442–3455, 2011.
- [22] J. Winter, S. Jung, S. Keller, R. I. Gregory, and S. Diederichs, "Many roads to maturity: microRNA biogenesis pathways and their regulation," *Nature Cell Biology*, vol. 11, no. 3, pp. 228–234, 2009.
- [23] B. N. Davis and A. Hata, "Regulation of MicroRNA Biogenesis: a miRiad of mechanisms," *Cell Communication and Signaling*, vol. 7, article 18, 2009.
- [24] A. E. Pasquinelli, B. J. Reinhart, F. Slack et al., "Conservation of the sequence and temporal expression of let-7 heterochronic regulatory RNA," *Nature*, vol. 408, no. 6808, pp. 86–89, 2000.
- [25] S. Roush and F. J. Slack, "The let-7 family of microRNAs," *Trends in Cell Biology*, vol. 18, no. 10, pp. 505–516, 2008.
- [26] B. Boyerinas, S.-M. Park, A. Hau, A. E. Murmann, and M. E. Peter, "The role of let-7 in cell differentiation and cancer," *Endocrine-Related Cancer*, vol. 17, no. 1, pp. F19–F36, 2010.
- [27] J. M. Thomson, M. Newman, J. S. Parker, E. M. Morin-Kensicki, T. Wright, and S. M. Hammond, "Extensive post-transcriptional regulation of microRNAs and its implications for cancer," *Genes and Development*, vol. 20, no. 16, pp. 2202–2207, 2006.
- [28] M. H. Bao, X. Feng, Y. W. Zhang, X. Y. Lou, Y. Cheng, and H. H. Zhou, "Let-7 in cardiovascular diseases, heart development and cardiovascular differentiation from stem cells," *International Journal of Molecular Sciences*, vol. 14, no. 11, pp. 23086–23102, 2013.
- [29] W. Hou, Q. Tian, N. M. Steuerwald, L. W. Schrum, and H. L. Bonkovsky, "The let-7 microRNA enhances heme oxygenase-1 by suppressing Bach1 and attenuates oxidant injury in human hepatocytes," *Biochimica et Biophysica Acta: Gene Regulatory Mechanisms*, vol. 1819, no. 11–12, pp. 1113–1122, 2012.
- [30] S. Polikepahad, J. M. Knight, A. O. Naghavi et al., "Proinflammatory role for let-7 microRNAs in experimental asthma," *Journal of Biological Chemistry*, vol. 285, no. 39, pp. 30139–30149, 2010.
- [31] D. Iliopoulos, H. A. Hirsch, and K. Struhl, "An epigenetic switch involving NF- κ B, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation," *Cell*, vol. 139, no. 4, pp. 693–706, 2009.
- [32] C. D. Johnson, A. Esquela-Kerscher, G. Stefani et al., "The let-7 microRNA represses cell proliferation pathways in human cells," *Cancer Research*, vol. 67, no. 16, pp. 7713–7722, 2007.
- [33] C. Mayr, M. T. Hemann, and D. P. Bartel, "Disrupting the pairing between let-7 and Hmga2 enhances oncogenic transformation," *Science*, vol. 315, no. 5818, pp. 1576–1579, 2007.
- [34] F. Meng, R. Henson, H. Wehbe-Janek, H. Smith, Y. Ueno, and T. Patel, "The microRNA let-7a modulates interleukin-6-dependent STAT-3 survival signaling in malignant human cholangiocytes," *The Journal of Biological Chemistry*, vol. 282, no. 11, pp. 8256–8264, 2007.
- [35] G. A. Calin, C. Sevignani, C. D. Dumitru et al., "Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 9, pp. 2999–3004, 2004.
- [36] C. Liu, K. Kelnar, A. V. Vlassov, D. Brown, J. Wang, and D. G. Tang, "Distinct microRNA expression profiles in prostate cancer stem/progenitor cells and tumor-suppressive functions of let-7," *Cancer Research*, vol. 72, no. 13, pp. 3393–3405, 2012.
- [37] T. Reya, S. J. Morrison, M. F. Clarke, and I. L. Weissman, "Stem cells, cancer, and cancer stem cells," *Nature*, vol. 414, no. 6859, pp. 105–111, 2001.
- [38] L. M. Coussens and Z. Werb, "Inflammation and cancer," *Nature*, vol. 420, no. 6917, pp. 860–867, 2002.
- [39] D. Kong, S. Banerjee, A. Ahmad et al., "Epithelial to mesenchymal transition is mechanistically linked with stem cell signatures in prostate cancer cells," *PLoS ONE*, vol. 5, no. 8, Article ID e12445, 2010.
- [40] S. N. Shah, L. Cope, W. Poh et al., "HMGA1: a master regulator of tumor progression in triple-negative breast cancer cells," *PLoS ONE*, vol. 8, no. 5, Article ID e63419, 2013.
- [41] C. Zhu, J. Li, G. Cheng et al., "MiR-154 inhibits EMT by targeting HMGA2 in prostate cancer cells," *Molecular and Cellular Biochemistry*, vol. 379, no. 1–2, pp. 69–75, 2013.
- [42] S. M. Johnson, H. Grosshans, J. Shingara et al., "RAS is regulated by the let-7 microRNA family," *Cell*, vol. 120, no. 5, pp. 635–647, 2005.
- [43] K. Patel, A. Kollory, A. Takashima, S. Sarkar, D. V. Faller, and S. K. Ghosh, "MicroRNA let-7 downregulates STAT3 phosphorylation in pancreatic cancer cells by increasing SOCS3 expression," *Cancer Letters*, vol. 347, no. 1, pp. 54–64, 2014.
- [44] M. M. Rahman, Z. R. Qian, E. L. Wang et al., "Frequent overexpression of HMGA1 and 2 in gastroenteropancreatic neuroendocrine tumours and its relationship to let-7 downregulation," *British Journal of Cancer*, vol. 100, no. 3, pp. 501–510, 2009.

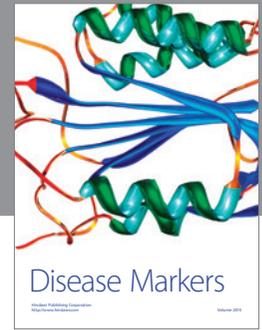
- [45] S. Watanabe, Y. Ueda, S.-I. Akaboshi, Y. Hino, Y. Sekita, and M. Nakao, "HMGA2 maintains oncogenic RAS-induced epithelial-mesenchymal transition in human pancreatic cancer cells," *The American Journal of Pathology*, vol. 174, no. 3, pp. 854–868, 2009.
- [46] E. Piskounova, C. Polyarchou, J. E. Thornton et al., "Lin28A and Lin28B inhibit let-7 MicroRNA biogenesis by distinct mechanisms," *Cell*, vol. 147, no. 5, pp. 1066–1079, 2011.
- [47] A. Rybak, H. Fuchs, L. Smirnova et al., "A feedback loop comprising lin-28 and let-7 controls pre-let-7 maturation during neural stem-cell commitment," *Nature Cell Biology*, vol. 10, no. 8, pp. 987–993, 2008.
- [48] L. J. Wood, M. Mukherjee, C. E. Dolde et al., "HMG-I/Y, a new c-Myc target gene and potential oncogene," *Molecular and Cellular Biology*, vol. 20, no. 15, pp. 5490–5502, 2000.
- [49] S. Y. Sung, C. H. Liao, H. P. Wu et al., "Loss of let-7 microRNA upregulates IL-6 in bone marrow-derived mesenchymal stem cells triggering a reactive stromal response to prostate cancer," *PLoS ONE*, vol. 8, no. 8, Article ID e71637, 2013.
- [50] S. N. Shah, C. Kerr, L. Cope et al., "HMGA1 reprograms somatic cells into pluripotent stem cells by inducing stem cell transcriptional networks," *PLoS ONE*, vol. 7, no. 11, Article ID e48533, 2012.
- [51] G. M. Pierantoni, C. Rinaldo, M. Mottolese et al., "High-mobility group A1 inhibits p53 by cytoplasmic relocalization of its proapoptotic activator HIPK2," *The Journal of Clinical Investigation*, vol. 117, no. 3, pp. 693–702, 2007.
- [52] T. Ueda and M. Yoshida, "HMGB proteins and transcriptional regulation," *Biochimica et Biophysica Acta—Gene Regulatory Mechanisms*, vol. 1799, no. 1-2, pp. 114–118, 2010.
- [53] S. Lyu, Q. Yu, G. Ying et al., "Androgen receptor decreases CMYC and KRAS expression by upregulating let-7a expression in ER-, PR-, AR+ breast cancer," *International Journal of Oncology*, vol. 44, no. 1, pp. 229–237, 2013.
- [54] M. Bustin, "Revised nomenclature for high mobility group (HMG) chromosomal proteins," *Trends in Biochemical Sciences*, vol. 26, no. 3, pp. 152–153, 2001.
- [55] K. R. Diener, N. Al-Dasooqi, E. L. Lousberg, and J. D. Hayball, "The multifunctional alarmin HMGB1 with roles in the pathophysiology of sepsis and cancer," *Immunology and Cell Biology*, vol. 91, pp. 443–450, 2013.
- [56] R. Reeves, "Nuclear functions of the HMG proteins," *Biochimica et Biophysica Acta—Gene Regulatory Mechanisms*, vol. 1799, no. 1-2, pp. 3–14, 2010.
- [57] M. Fedele and A. Fusco, "HMGA and cancer," *Biochimica et Biophysica Acta: Gene Regulatory Mechanisms*, vol. 1799, no. 1-2, pp. 48–54, 2010.
- [58] Y. Tamimi, H. G. van der Poel, M.-M. Denyn et al., "Increased expression of high mobility group protein I(Y) in high grade prostatic cancer determined by in situ hybridization," *Cancer Research*, vol. 53, no. 22, pp. 5512–5516, 1993.
- [59] J.-J. Wei, X. Wu, Y. Peng et al., "Regulation of HMGA1 expression by MicroRNA-296 affects prostate cancer growth and invasion," *Clinical Cancer Research*, vol. 17, no. 6, pp. 1297–1305, 2011.
- [60] J. Hillion, L. J. Wood, M. Mukherjee et al., "Upregulation of MMP-2 by HMGA1 promotes transformation in undifferentiated, large-cell lung cancer," *Molecular Cancer Research*, vol. 7, no. 11, pp. 1803–1812, 2009.
- [61] R. Reeves, D. D. Edberg, and Y. Li, "Architectural transcription factor HMGI(Y) promotes tumor progression and mesenchymal transition of human epithelial cells," *Molecular and Cellular Biology*, vol. 21, no. 2, pp. 575–594, 2001.
- [62] N. Takaha, A. L. Hawkins, C. A. Griffin, W. B. Isaacs, and D. S. Coffey, "High mobility group protein I(Y): a candidate architectural protein for chromosomal rearrangements in prostate cancer cells," *Cancer Research*, vol. 62, no. 3, pp. 647–651, 2002.
- [63] N. Takaha, L. M. S. Resar, D. Vindivich, and D. S. Coffey, "High mobility group protein HMGI(Y) enhances tumor cell growth, invasion, and matrix metalloproteinase-2 expression in prostate cancer cells," *The Prostate*, vol. 60, no. 2, pp. 160–167, 2004.
- [64] A. Cano, M. A. Pérez-Moreno, I. Rodrigo et al., "The transcription factor Snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression," *Nature Cell Biology*, vol. 2, no. 2, pp. 76–83, 2000.
- [65] J. Pérez-Losada, M. Sánchez-Martin, A. Rodríguez-García et al., "Zinc-finger transcription factor slug contributes to the function of the stem cell factor c-kit signaling pathway," *Blood*, vol. 100, no. 4, pp. 1274–1286, 2002.
- [66] I. De Martino, R. Visone, M. Fedele et al., "Regulation of microRNA expression by HMGA1 proteins," *Oncogene*, vol. 28, no. 11, pp. 1432–1442, 2009.
- [67] J. Hillion, S. Dhara, T. F. Sumter et al., "The high-mobility group A1a/signal transducer and activator of transcription-3 axis: an achilles heel for hematopoietic malignancies?" *Cancer Research*, vol. 68, no. 24, pp. 10121–10127, 2008.
- [68] O. A. Timofeeva, N. I. Tarasova, X. Zhang et al., "STAT3 suppresses transcription of proapoptotic genes in cancer cells with the involvement of its N-terminal domain," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 4, pp. 1267–1272, 2013.
- [69] D. Iliopoulos, S. A. Jaeger, H. A. Hirsch, M. L. Bulyk, and K. Struhl, "STAT3 activation of miR-21 and miR-181b-1 via PTEN and CYLD are part of the epigenetic switch linking inflammation to cancer," *Molecular Cell*, vol. 39, no. 4, pp. 493–506, 2010.
- [70] J. Ma, H. Sawai, N. Ochi et al., "PTEN regulate angiogenesis through PI3K/Akt/VEGF signaling pathway in human pancreatic cancer cells," *Molecular and Cellular Biochemistry*, vol. 331, no. 1-2, pp. 161–171, 2009.
- [71] R. Sgarra, S. Zammitti, A. Lo Sardo et al., "HMGA molecular network: from transcriptional regulation to chromatin remodeling," *Biochimica et Biophysica Acta—Gene Regulatory Mechanisms*, vol. 1799, no. 1-2, pp. 37–47, 2010.
- [72] G. Mu, H. Liu, F. Zhou et al., "Correlation of overexpression of HMGA1 and HMGA2 with poor tumor differentiation, invasion, and proliferation associated with let-7 down-regulation in retinoblastomas," *Human Pathology*, vol. 41, no. 4, pp. 493–502, 2010.
- [73] D. Palmieri, D. D'Angelo, T. Valentino et al., "Downregulation of HMGA-targeting microRNAs has a critical role in human pituitary tumorigenesis," *Oncogene*, vol. 31, no. 34, pp. 3857–3865, 2012.
- [74] M. Schubert, M. Spahn, S. Kneitz et al., "Distinct microRNA expression profile in prostate cancer patients with early clinical failure and the impact of let-7 as prognostic marker in high-risk prostate cancer," *PLoS ONE*, vol. 8, no. 6, Article ID e65064, 2013.
- [75] B. Meyer, S. Loeschke, A. Schultze et al., "HMGA2 overexpression in non-small cell lung cancer," *Molecular Carcinogenesis*, vol. 46, no. 7, pp. 503–511, 2007.
- [76] P. Rogalla, K. Drechsler, B. Kazmierczak, V. Rippe, U. Bonk, and J. Bullerdiek, "Expression of HMGI-C, a member of the high mobility group protein family, in a subset of breast cancers:

- relationship to histologic grade," *Molecular Carcinogenesis*, vol. 19, no. 3, pp. 153–156, 1997.
- [77] S. Winkler, H. M. Escobar, B. Meyer et al., "HMGA2 expression in a canine model of prostate cancer," *Cancer Genetics and Cytogenetics*, vol. 177, no. 2, pp. 98–102, 2007.
- [78] M. S. Kumar, E. Armenteros-Monterroso, P. East et al., "HMGA2 functions as a competing endogenous RNA to promote lung cancer progression," *Nature*, vol. 505, no. 7482, pp. 212–217, 2014.
- [79] M. T. Berlingieri, G. Manfioletti, M. Santoro et al., "Inhibition of HMGI-C protein synthesis suppresses retrovirally induced neoplastic transformation of rat thyroid cells," *Molecular and Cellular Biology*, vol. 15, no. 3, pp. 1545–1553, 1995.
- [80] J.-M. Berner, L. A. Meza-Zepeda, P. F. J. Kools et al., "HMGIC, the gene for an architectural transcription factor, is amplified and rearranged in a subset of human sarcomas," *Oncogene*, vol. 14, no. 24, pp. 2935–2941, 1997.
- [81] F. di Cello, J. Hillion, A. Hristov et al., "HMGA2 participates in transformation in human lung cancer," *Molecular Cancer Research*, vol. 6, no. 5, pp. 743–750, 2008.
- [82] M. Fedele, S. Battista, G. Manfioletti, C. M. Croce, V. Giancotti, and A. Fusco, "Role of the high mobility group A proteins in human lipomas," *Carcinogenesis*, vol. 22, no. 10, pp. 1583–1591, 2001.
- [83] S. Müller, P. Scaffidi, B. Degryse et al., "The double life of HMGB1 chromatin protein: architectural factor and extracellular signal," *The EMBO Journal*, vol. 20, no. 16, pp. 4337–4340, 2001.
- [84] E. Pikarsky, R. M. Porat, I. Stein et al., "NF- κ B functions as a tumour promoter in inflammation-associated cancer," *Nature*, vol. 431, no. 7007, pp. 461–466, 2004.
- [85] J. R. van Beijnum, W. A. Buurman, and A. W. Griffioen, "Convergence and amplification of toll-like receptor (TLR) and receptor for advanced glycation end products (RAGE) signaling pathways via high mobility group B1 (HMGB1)," *Angiogenesis*, vol. 11, no. 1, pp. 91–99, 2008.
- [86] J. S. Park, D. Svetkauskaite, Q. He et al., "Involvement of toll-like receptors 2 and 4 in cellular activation by high mobility group box 1 protein," *The Journal of Biological Chemistry*, vol. 279, no. 9, pp. 7370–7377, 2004.
- [87] O. Hori, J. Brett, T. Slattery et al., "The receptor for advanced glycation end products (RAGE) is a cellular binding site for amphotericin. Mediation of neurite outgrowth and co-expression of RAGE and amphotericin in the developing nervous system," *The Journal of Biological Chemistry*, vol. 270, no. 43, pp. 25752–25761, 1995.
- [88] X. M. Chen, P. L. Splinter, S. P. O'Hara, and N. F. LaRusso, "A cellular micro-RNA, let-7i, regulates Toll-like receptor 4 expression and contributes to cholangiocyte immune responses against *Cryptosporidium parvum* infection," *The Journal of Biological Chemistry*, vol. 282, no. 39, pp. 28929–28938, 2007.
- [89] G. Li, J. Xu, and Z. Li, "Receptor for advanced glycation end products inhibits proliferation in osteoblast through suppression of Wnt, PI3K and ERK signaling," *Biochemical and Biophysical Research Communications*, vol. 423, no. 4, pp. 684–689, 2012.
- [90] J. E. Hutti, A. D. Pfefferle, S. C. Russell, M. Sircar, C. M. Perou, and A. S. Baldwin, "Oncogenic PI3K mutations lead to NF- κ B-dependent cytokine expression following growth factor deprivation," *Cancer Research*, vol. 72, no. 13, pp. 3260–3269, 2012.
- [91] M. Yu, H. Wang, A. Ding et al., "HMGB1 signals through toll-like receptor (TLR) 4 and TLR2," *Shock*, vol. 26, no. 2, pp. 174–179, 2006.
- [92] J. Li and A. M. Schmidt, "Characterization and functional analysis of the promoter of RAGE, the receptor for advanced glycation end products," *The Journal of Biological Chemistry*, vol. 272, no. 26, pp. 16498–16506, 1997.
- [93] A. Taguchi, D. C. Blood, G. Del Toro et al., "Blockade of RAGE-amphotericin signalling suppresses tumour growth and metastases," *Nature*, vol. 405, no. 6784, pp. 354–360, 2000.
- [94] D. Tang, R. Kang, H. J. Zeh III, and M. T. Lotze, "High-mobility group box 1 and cancer," *Biochimica et Biophysica Acta—Gene Regulatory Mechanisms*, vol. 1799, no. 1–2, pp. 131–140, 2010.
- [95] H. Kuniyasu, Y. Chihara, H. Kondo, H. Ohmori, and R. Ukai, "Amphotericin induction in prostatic stromal cells by androgen deprivation is associated with metastatic prostate cancer," *Oncology Reports*, vol. 10, no. 6, pp. 1863–1868, 2003.
- [96] M. Gnanasekar, R. Kalyanasundaram, G. Zheng, A. Chen, M. C. Bosland, and A. Kajdacsy-Balla, "HMGB1: a promising therapeutic target for prostate cancer," *Prostate Cancer*, vol. 2013, Article ID 157103, 8 pages, 2013.
- [97] V. Boonyaratanakornkit, V. Melvin, P. Prendergast et al., "High-mobility group chromatin proteins 1 and 2 functionally interact with steroid hormone receptors to enhance their DNA binding in vitro and transcriptional activity in mammalian cells," *Molecular and Cellular Biology*, vol. 18, no. 8, pp. 4471–4487, 1998.
- [98] J. P. Rowell, K. L. Simpson, K. Stott, M. Watson, and J. O. Thomas, "HMGB1-facilitated p53 DNA binding occurs via HMG-Box/p53 transactivation domain interaction, regulated by the acidic tail," *Structure*, vol. 20, no. 12, pp. 2014–2024, 2012.
- [99] H. Uramoto, H. Izumi, G. Nagatani et al., "Physical interaction of tumour suppressor p53/p73 with CCAAT-binding transcription factor 2 (CTF2) and differential regulation of human high-mobility group 1 (HMGI) gene expression," *The Biochemical Journal*, vol. 371, no. 2, pp. 301–310, 2003.
- [100] A. D. Saleh, J. E. Savage, L. Cao et al., "Cellular stress induced alterations in microRNA let-7a and let-7b expression are dependent on p53," *PLoS ONE*, vol. 6, no. 10, Article ID e24429, 2011.
- [101] M. Malumbres and M. Barbacid, "Cell cycle, CDKs and cancer: a changing paradigm," *Nature Reviews Cancer*, vol. 9, no. 3, pp. 153–166, 2009.
- [102] Y. Takano, Y. Kato, P. J. Van Diest, M. Masuda, H. Mitomi, and I. Okayasu, "Cyclin D2 overexpression and lack of p27 correlate positively and cyclin E inversely with a poor prognosis in gastric cancer cases," *The American Journal of Pathology*, vol. 156, no. 2, pp. 585–594, 2000.
- [103] A. Mermelshtein, A. Gerson, S. Walfisch et al., "Expression of D-type cyclins in colon cancer and in cell lines from colon carcinomas," *British Journal of Cancer*, vol. 93, no. 3, pp. 338–345, 2005.
- [104] T. Igawa, Y. Sato, K. Takata et al., "Cyclin D2 is overexpressed in proliferation centers of chronic lymphocytic leukemia/small lymphocytic lymphoma," *Cancer Science*, vol. 102, no. 11, pp. 2103–2107, 2011.
- [105] C. Zhu, P. Shao, M. Bao et al., "miR-154 inhibits prostate cancer cell proliferation by targeting CCND2," *Urologic Oncology: Seminars and Original Investigations*, vol. 32, no. 1, pp. 31.e9–31.e16, 2014.
- [106] J. Klaewsongkram, Y. Yang, S. Golech, J. Katz, K. H. Kaestner, and N.-P. Weng, "Krüppel-like factor 4 regulates B cell number and activation-induced B cell proliferation," *Journal of Immunology*, vol. 179, no. 7, pp. 4679–4684, 2007.

- [107] C. Bouchard, O. Dittrich, A. Kiermaier et al., "Regulation of cyclin D2 gene expression by the Myc/Max/Mad network: Myc-dependent TRRAP recruitment and histone acetylation at the cyclin D2 promoter," *Genes & Development*, vol. 15, no. 16, pp. 2042–2047, 2001.
- [108] C. E. Nesbit, J. M. Tersak, and E. V. Prochownik, "MYC oncogenes and human neoplastic disease," *Oncogene*, vol. 18, no. 19, pp. 3004–3016, 1999.
- [109] C. V. Dang, "MYC, metabolism, cell growth, and tumorigenesis," *Cold Spring Harbor Perspectives in Medicine*, vol. 3, no. 8, 2013.
- [110] B. Nagy, A. Szendroi, and I. Romics, "Overexpression of CD24, c-myc and phospholipase 2A in prostate cancer tissue samples obtained by needle biopsy," *Pathology and Oncology Research*, vol. 15, no. 2, pp. 279–283, 2009.
- [111] R. Buttyan, I. S. Sawczuk, M. C. Benson, J. D. Siegal, and C. Olsson, "Enhanced expression of the c-myc protooncogene in high-grade human prostate cancers," *The Prostate*, vol. 11, no. 4, pp. 327–337, 1987.
- [112] J. Gil, P. Kerai, M. Leonart et al., "Immortalization of primary human prostate epithelial cells by c-Myc," *Cancer Research*, vol. 65, no. 6, pp. 2179–2185, 2005.
- [113] S. R. Hann, "Role of post-translational modifications in regulating c-Myc proteolysis, transcriptional activity and biological function," *Seminars in Cancer Biology*, vol. 16, no. 4, pp. 288–302, 2006.
- [114] H. K. Hyeon, Y. Kuwano, S. Srikantan, K. L. Eun, J. L. Martindale, and M. Gorospe, "HuR recruits let-7/RISC to repress c-Myc expression," *Genes and Development*, vol. 23, no. 15, pp. 1743–1748, 2009.
- [115] V. B. Sampson, N. H. Rong, J. Han et al., "MicroRNA let-7a down-regulates MYC and reverts MYC-induced growth in Burkitt lymphoma cells," *Cancer Research*, vol. 67, no. 20, pp. 9762–9770, 2007.
- [116] D. Gioeli, J. W. Mandell, G. R. Petroni, H. F. Frierson Jr., and M. J. Weber, "Activation of mitogen-activated protein kinase associated with prostate cancer progression," *Cancer research*, vol. 59, no. 2, pp. 279–284, 1999.
- [117] F. Marampon, C. Ciccarelli, and B. M. Zani, "Down-regulation of c-Myc following MEK/ERK inhibition halts the expression of malignant phenotype in rhabdomyosarcoma and in non muscle-derived human tumors," *Molecular Cancer*, vol. 5, article 31, 2006.
- [118] C. Bradham and D. R. McClay, "p38 MAPK in development and cancer," *Cell Cycle*, vol. 5, no. 8, pp. 824–828, 2006.
- [119] T.-C. Chang, L. R. Zeitels, H.-W. Hwang et al., "Lin-28B transactivation is necessary for Myc-mediated let-7 repression and proliferation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 9, pp. 3384–3389, 2009.
- [120] K. S. Sfanos and A. M. de Marzo, "Prostate cancer and inflammation: the evidence," *Histopathology*, vol. 60, no. 1, pp. 199–215, 2012.
- [121] T. Hirano, K. Yasukawa, H. Harada et al., "Complementary DNA for a novel human interleukin (BSF-2) that induces B lymphocytes to produce immunoglobulin," *Nature*, vol. 324, no. 6092, pp. 73–76, 1986.
- [122] D. P. Nguyen, J. Li, and A. K. Tewari, "Inflammation and prostate cancer: the role of interleukin 6 (IL-6)," *BJU International*, vol. 113, no. 6, pp. 986–992, 2014.
- [123] M. Okamoto, C. Lee, and R. Oyasu, "Interleukin-6 as a paracrine and autocrine growth factor in human prostatic carcinoma cells in vitro," *Cancer Research*, vol. 57, no. 1, pp. 141–146, 1997.
- [124] B. Wegiel, A. Bjartell, Z. Culig, and J. L. Persson, "Interleukin-6 activates PI3K/Akt pathway and regulates cyclin A1 to promote prostate cancer cell survival," *International Journal of Cancer*, vol. 122, no. 7, pp. 1521–1529, 2008.
- [125] P. Sivashanmugam, L. Tang, and Y. Daaka, "Interleukin 6 mediates the lysophosphatidic acid-regulated cross-talk between stromal and epithelial prostate cancer cells," *Journal of Biological Chemistry*, vol. 279, no. 20, pp. 21154–21159, 2004.
- [126] D. Giri, M. Ozen, and M. Ittmann, "Interleukin-6 is an autocrine growth factor in human prostate cancer," *The American Journal of Pathology*, vol. 159, no. 6, pp. 2159–2165, 2001.
- [127] V. Michalaki, K. Syrigos, P. Charles, and J. Waxman, "Serum levels of IL-6 and TNF- α correlate with clinicopathological features and patient survival in patients with prostate cancer," *British Journal of Cancer*, vol. 90, no. 12, pp. 2312–2316, 2004.
- [128] R. J. Simpson, A. Hammacher, D. K. Smith, J. M. Matthews, and L. D. Ward, "Interleukin-6: structure-function relationships," *Protein Science*, vol. 6, no. 5, pp. 929–955, 1997.
- [129] K. Yamaoka, P. Saharinen, M. Pesu, V. E. T. Holt III, O. Silvennoinen, and J. J. O'Shea, "The Janus kinases (Jaks)," *Genome Biology*, vol. 5, no. 12, article 253, 2004.
- [130] X. Wang, P. Lupardus, S. L. LaPorte, and K. C. Garcia, "Structural biology of shared cytokine receptors," *Annual Review of Immunology*, vol. 27, pp. 29–60, 2009.
- [131] J. N. Ihle, "The stat family in cytokine signaling," *Current Opinion in Cell Biology*, vol. 13, no. 2, pp. 211–217, 2001.
- [132] J. Scheller, A. Chalaris, D. Schmidt-Arras, and S. Rose-John, "The pro- and anti-inflammatory properties of the cytokine interleukin-6," *Biochimica et Biophysica Acta: Molecular Cell Research*, vol. 1813, no. 5, pp. 878–888, 2011.
- [133] A. Fahmi, N. Smart, A. Punj, R. Jabr, M. Marber, and R. Heads, "P42/p44-MAPK and PI3K are sufficient for IL-6 family cytokines/gp130 to signal to hypertrophy and survival in cardiomyocytes in the absence of JAK/STAT activation," *Cellular Signalling*, vol. 25, no. 4, pp. 898–909, 2013.
- [134] T. Ueda, N. Bruchovsky, and M. D. Sadar, "Activation of the androgen receptor N-terminal domain by interleukin-6 via MAPK and STAT3 signal transduction pathways," *The Journal of Biological Chemistry*, vol. 277, no. 9, pp. 7076–7085, 2002.
- [135] K. Malinowska, H. Neuwirt, I. T. Cavarretta et al., "Interleukin-6 stimulation of growth of prostate cancer *in vitro* and *in vivo* through activation of the androgen receptor," *Endocrine-Related Cancer*, vol. 16, no. 1, pp. 155–169, 2009.
- [136] L. Yang, L. Wang, H.-K. Lin et al., "Interleukin-6 differentially regulates androgen receptor transactivation via PI3K-Akt, STAT3, and MAPK, three distinct signal pathways in prostate cancer cells," *Biochemical and Biophysical Research Communications*, vol. 305, no. 3, pp. 462–469, 2003.
- [137] Y.-S. Pu, T.-C. Hour, S.-E. Chuang, A.-L. Cheng, M.-K. Lai, and M.-L. Kuo, "Interleukin-6 is responsible for drug resistance and anti-apoptotic effects in prostatic cancer cells," *Prostate*, vol. 60, no. 2, pp. 120–129, 2004.
- [138] Y. Liu, P.-K. Li, C. Li, and J. Lin, "Inhibition of STAT3 signaling blocks the anti-apoptotic activity of IL-6 in human liver cancer cells," *The Journal of Biological Chemistry*, vol. 285, no. 35, pp. 27429–27439, 2010.

- [139] C. Liu, Y. Zhu, W. Lou, Y. Cui, C. P. Evans, and A. C. Gao, "Inhibition of constitutively active Stat3 reverses enzalutamide resistance in LNCaP derivative prostate cancer cells," *The Prostate*, vol. 74, no. 2, pp. 201–209, 2014.
- [140] B. Paule, S. Terry, L. Kheuang, P. Soyeux, F. Vacherot, and A. de Taille, "The NF- κ B/IL-6 pathway in metastatic androgen-independent prostate cancer: new therapeutic approaches?" *World Journal of Urology*, vol. 25, no. 5, pp. 477–489, 2007.
- [141] D. J. Wang, A. Legesse-Miller, E. L. Johnson, and H. A. Collier, "Regulation of the let-7a-3 promoter by NF- κ B," *PLoS ONE*, vol. 7, no. 2, Article ID e31240, 2012.
- [142] S. Rose-John, G. H. Waetzig, J. Cheller, J. Grötzinger, and D. Seegert, "The IL-6/sIL-6R complex as a novel target for therapeutic approaches," *Expert Opinion on Therapeutic Targets*, vol. 11, no. 5, pp. 613–624, 2007.
- [143] K. Rajalingam, R. Schreck, U. R. Rapp, and Š. Albert, "Ras oncogenes and their downstream targets," *Biochimica et Biophysica Acta: Molecular Cell Research*, vol. 1773, no. 8, pp. 1177–1195, 2007.
- [144] A. Fernández-Medarde and E. Santos, "Ras in cancer and developmental diseases," *Genes and Cancer*, vol. 2, no. 3, pp. 344–358, 2011.
- [145] P. Gideon, J. John, M. Frech et al., "Mutational and kinetic analyses of the GTPase-activating protein (GAP)-p21 interaction: the C-terminal domain of GAP is not sufficient for full activity," *Molecular and Cellular Biology*, vol. 12, no. 5, pp. 2050–2056, 1992.
- [146] K. Scheffzek, M. R. Ahmadian, W. Kabsch et al., "The Ras-RasGAP complex: structural basis for GTPase activation and its loss in oncogenic ras mutants," *Science*, vol. 277, no. 5324, pp. 333–338, 1997.
- [147] S. Schubert, K. Shannon, and G. Bollag, "Hyperactive Ras in developmental disorders and cancer," *Nature Reviews Cancer*, vol. 7, no. 4, pp. 295–308, 2007.
- [148] N. Gerits, S. Kostenko, A. Shiryaev, M. Johannessen, and U. Moens, "Relations between the mitogen-activated protein kinase and the cAMP-dependent protein kinase pathways: comradeship and hostility," *Cellular Signalling*, vol. 20, no. 9, pp. 1592–1607, 2008.
- [149] A. S. Dhillon, S. Hagan, O. Rath, and W. Kolch, "MAP kinase signalling pathways in cancer," *Oncogene*, vol. 26, no. 22, pp. 3279–3290, 2007.
- [150] G. A. Repasky, E. J. Chenette, and C. J. Der, "Renewing the conspiracy theory debate: does Raf function alone to mediate Ras oncogenesis?" *Trends in Cell Biology*, vol. 14, no. 11, pp. 639–647, 2004.
- [151] E. Castellano and J. Downward, "Ras interaction with PI3K: more than just another effector pathway," *Genes & Cancer*, vol. 2, no. 3, pp. 261–274, 2011.
- [152] J. A. Engelman, J. Luo, and L. C. Cantley, "The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism," *Nature Reviews Genetics*, vol. 7, no. 8, pp. 606–619, 2006.
- [153] B. S. Taylor, N. Schultz, H. Hieronymus et al., "Integrative genomic profiling of human prostate cancer," *Cancer Cell*, vol. 18, no. 1, pp. 11–22, 2010.
- [154] J. A. Engelman, "The role of phosphoinositide 3-kinase pathway inhibitors in the treatment of lung cancer," *Clinical Cancer Research*, vol. 13, no. 15, part 2, pp. S4637–S4640, 2007.
- [155] M. A. Davies, "Regulation, role, and targeting of Akt in cancer," *Journal of Clinical Oncology*, vol. 29, no. 35, pp. 4715–4717, 2011.
- [156] V. Poulaki, C. S. Mitsiades, C. McMullan et al., "Regulation of vascular endothelial growth factor expression by insulin-like growth factor I in thyroid carcinomas," *The Journal of Clinical Endocrinology & Metabolism*, vol. 88, no. 11, pp. 5392–5398, 2003.
- [157] G. Niu, K. L. Wright, M. Huang et al., "Constitutive Stat3 activity up-regulates VEGF expression and tumor angiogenesis," *Oncogene*, vol. 21, no. 13, pp. 2000–2008, 2002.
- [158] L. Guo, C. Chen, M. Shi et al., "Stat3-coordinated Lin-28-let-7-HMGA2 and miR-200-ZEB1 circuits initiate and maintain oncostatin M-driven epithelial-mesenchymal transition," *Oncogene*, vol. 32, no. 45, pp. 5272–5282, 2013.
- [159] M. Colombatti, S. Grasso, A. Porzia et al., "The prostate specific membrane antigen regulates the expression of IL-6 and CCL5 in prostate tumour cells by activating the MAPK pathways," *PLoS ONE*, vol. 4, no. 2, Article ID e4608, 2009.
- [160] K. Bouchelouche, P. L. Choyke, and J. Capala, "Prostate specific membrane antigen- a target for imaging and therapy with radionuclides," *Discovery Medicine*, vol. 9, no. 44, pp. 55–61, 2010.
- [161] T. Naka, N. Nishimoto, and T. Kishimoto, "The paradigm of IL-6: from basic science to medicine," *Arthritis Research & Therapy*, vol. 4, supplement 3, pp. S233–S242, 2002.
- [162] F. Yu, H. Yao, P. Zhu et al., "let-7 regulates self renewal and tumorigenicity of breast cancer cells," *Cell*, vol. 131, no. 6, pp. 1109–1123, 2007.
- [163] W. Wahli and E. Martinez, "Superfamily of steroid nuclear receptors: positive and negative regulators of gene expression," *The FASEB Journal*, vol. 5, no. 9, pp. 2243–2249, 1991.
- [164] C. J. Brown, S. J. Goss, D. B. Lubahn et al., "Androgen receptor locus on the human X chromosome: regional localization to Xq11-12 and description of a DNA polymorphism," *The American Journal of Human Genetics*, vol. 44, no. 2, pp. 264–269, 1989.
- [165] E. P. Gelmann, "Molecular biology of the androgen receptor," *Journal of Clinical Oncology*, vol. 20, no. 13, pp. 3001–3015, 2002.
- [166] P. J. Roche, S. A. Hoare, and M. G. Parker, "A consensus DNA-binding site for the androgen receptor," *Molecular Endocrinology*, vol. 6, no. 12, pp. 2229–2235, 1992.
- [167] F. Claessens, G. Verrijdt, E. Schoenmakers et al., "Selective DNA binding by the androgen receptor as a mechanism for hormone-specific gene regulation," *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 76, no. 1–5, pp. 23–30, 2001.
- [168] P. E. Lonergan and D. J. Tindall, "Androgen receptor signaling in prostate cancer development and progression," *Journal of Carcinogenesis*, vol. 10, article 20, 2011.
- [169] A. I. So, A. Hurtado-Coll, and M. E. Gleave, "Androgens and prostate cancer," *World Journal of Urology*, vol. 21, no. 5, pp. 325–337, 2003.
- [170] M. A. Titus, B. Zeithaml, B. Kantor et al., "Dominant-negative androgen receptor inhibition of intracrine androgen-dependent growth of castration-recurrent prostate cancer," *PLoS ONE*, vol. 7, no. 1, Article ID e30192, 2012.
- [171] J. A. Locke, E. S. Guns, A. A. Lubik et al., "Androgen Levels increase by intratumoral de novo steroidogenesis during progression of castration-resistant prostate cancer," *Cancer Research*, vol. 68, no. 15, pp. 6407–6415, 2008.
- [172] M.-L. Zhu and N. Kyprianou, "Androgen receptor and growth factor signaling cross-talk in prostate cancer cells," *Endocrine-Related Cancer*, vol. 15, no. 4, pp. 841–849, 2008.

- [173] Z. Guo and Y. Qiu, "A new trick of an old molecule: androgen receptor splice variants taking the stage?!" *International Journal of Biological Sciences*, vol. 7, no. 6, pp. 815–822, 2011.
- [174] K. Eisermann, C. J. Broderick, A. Bazarov, M. M. Moazam, and G. C. Fraizer, "Androgen up-regulates vascular endothelial growth factor expression in prostate cancer cells via an Sp1 binding site," *Molecular Cancer*, vol. 12, no. 1, article 7, 2013.
- [175] P. Saxena, M. Trerotola, T. Wang et al., "PSA regulates androgen receptor expression in prostate cancer cells," *Prostate*, vol. 72, no. 7, pp. 769–776, 2012.
- [176] G. Attard, C. S. Cooper, and J. S. de Bono, "Steroid hormone receptors in prostate cancer: a hard habit to break?" *Cancer Cell*, vol. 16, no. 6, pp. 458–462, 2009.
- [177] R. Tummala, N. Nadiminty, W. Lou et al., "Lin28 promotes growth of prostate cancer cells and activates the androgen receptor," *The American Journal of Pathology*, vol. 183, no. 1, pp. 288–295, 2013.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

