

## Clinical Study

# Evaluation of Granulocyte Colony-Stimulating Factor Effects on Treatment-Resistant Thin Endometrium in Women Undergoing *In Vitro* Fertilization

Michał Kunicki,<sup>1</sup> Krzysztof Łukaszuk,<sup>2,3,4</sup> Izabela Woclawek-Potocka,<sup>5</sup> Joanna Liss,<sup>4</sup> Patrycja Kulwikowska,<sup>4</sup> and Joanna Szczyptańska<sup>4</sup>

<sup>1</sup> INVICTA Fertility and Reproductive Center, 00-019 Warszawa, Poland

<sup>2</sup> Department of Obstetrics and Gynecological Nursing, Faculty of Health Sciences, Medical University of Gdansk, 80-952 Gdańsk, Poland

<sup>3</sup> Department of Obstetrics and Gynecology, Faculty of Medical Sciences, University of Warmia and Masuria, 10-561 Olsztyn, Poland

<sup>4</sup> INVICTA Fertility and Reproductive Center, 80-850 Gdańsk, Poland

<sup>5</sup> Department of Reproductive Immunology and Pathology, Institute of Animal Reproduction and Food Research, Polish Academy of Sciences, 10-747 Olsztyn, Poland

Correspondence should be addressed to Michał Kunicki; [mkunicki@op.pl](mailto:mkunicki@op.pl)

Received 29 November 2013; Accepted 20 December 2013; Published 12 February 2014

Academic Editor: Irma Virant-Klun

Copyright © 2014 Michał Kunicki et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The aim of the study was to assess the granulocyte colony-stimulating factor (G-CSF) effects on unresponsive thin (<7 mm) endometrium in women undergoing *in vitro* fertilization (IVF). We included thirty-seven subjects who had thin unresponsive endometrium on the day of triggering ovulation. These patients also failed to achieve an adequate endometrial thickness in at least one of their previous IVF cycles. In all the subjects at the time of infusion of G-CSF, endometrial thickness was  $6,74 \pm 1,75$  mm, and, after infusion, it increased significantly to  $8,42 \pm 1,73$  mm. When we divided the group into two subgroups according to whether the examined women conceived, we showed that the endometrium expanded significantly from  $6,86 \pm 1,65$  to  $8,80 \pm 1,14$  mm in the first group (who conceived) and from  $6,71 \pm 1,80$  to  $8,33 \pm 1,85$  mm in the second, respectively. There were no significant differences between the two subgroups in respect to the endometrial thickness both before and after G-CSF infusion. The clinical pregnancy rate was 18,9%. We concluded that the infusion of G-CSF leads to the improvement of endometrium thickness after 72 hours.

## 1. Introduction

Many factors could have impact on the *in vitro* fertilization-embryo transfer IVF-ET success. The main independent variables are: the age of women, antimüllerian hormone (AMH) concentrations, number of embryos transferred and their quality [1]. It was also demonstrated that endometrial thickness <7 mm negatively affected pregnancy rate [2, 3]. Moreover, Sharkey showed that immunological mechanisms in the endometrium are very important and crucial in the implantation process [4]. Some investigators demonstrated that the growth factors, hormones, and cytokines, which are produced by decidual cells, are involved in the implantation process [5]. Preliminary studies demonstrated that G-CSF

stimulated neutrophilic granulocyte proliferation and differentiation, acted on macrophages of decidual cells, and finally affected the implantation [6, 7]. What is more, known and reported immune effects of G-CSF are recruitment of dendritic cells, promoting Th-2 cytokine secretion, activating T regulatory cells, and also stimulation of various proangiogenic effects [7, 8]. On the other hand, the receptor for G-CSF is expressed by the trophoblastic cells and by human luteinized granulosa cells [9, 10]. It was also stated that G-CSF prevented repeated miscarriages and implantation failures [11, 12]. In the last two years, Gleicher et al. presented two clinical studies with limited number of participants regarding the usefulness of G-CSF treatment in endometrium expansion in women who had previously cancelled cycles because of

the unresponsive endometrium [13, 14]. Taking into account all these data, the aim of the study was to examine G-CSF effects on unresponsive thin (<7 mm) endometrium in women undergoing IVF.

## 2. Materials and Methods

We presented a series of 37 patients who have undergone IVF procedure.

The inclusion criteria were as follows:

- (a) women aged 18–45 years,
- (b) previously cancelled at least one cycle because of thin unresponsive endometrium (<7 mm) during IVF programs,
- (c) inadequate thin endometrium (<7 mm) on the day of hCG injection,
- (d) the lack of contraindications for G-CSF treatment (sickle cell disease, chronic neutropenia, known past or present malignancy, renal insufficiency, upper respiratory infection, pneumonia, and congenital fructose intolerance),
- (e) personal agreement for such still experimental therapy,
- (f) no prenatal genetic screening,
- (g) no Asherman's syndrome, fibroids, and polyps in diagnostic hysteroscopy.

The study protocol was approved by the Institutional Review Board of Varmia and Masuria, Olsztyn, Poland, and written informed consent was given by each participating women.

The primary end point was the endometrial thickness measured in transvaginal sonography. The second point was clinical pregnancy after embryo transfer. Identification of an intrauterine gestational sac by transvaginal ultrasonography together with an increasing serum  $\beta$ -hCG constituted a clinical state of pregnancy. All included women had previously long agonist protocol in all cases.

During the study, all patients received oral contraceptive pill (OCP) starting on days 2–5 of spontaneous menses of the cycle prior to the treatment cycle. The OCP contained 0.03 mg ethinyl estradiol (E2) and 0.15 mg desogestrel Ovulastan (Polfa, Poland). OCPs were taken daily for 21 days. Patients were administered s.c. GnRH agonist 0.1 mg gonapeptyl (Ferring, The Netherlands) daily. The agonist was started 4–5 days before discontinuation of the OCP. When desensitization was achieved as evidenced by plasma E2 levels of <50 pg/mL [15], daily s.c. injection of highly purified menotropin (Menopur, Ferring, The Netherlands) was commenced. When at least two follicles reached a minimum of 17 mm diameter, 5000 U of hCG (Choragon Ferring, The Netherlands) was applied. Transvaginal pick-up was performed 36 h hours after hCG administration under transvaginal sonography.

The infusion of G-CSF was made according to Gleicher et al.'s procedure with full bladder before transfer [14]. Frydman

catheter was introduced to the uterine cavity. We infused under ultrasound guidance 30 mL (300 mg/1 mL) of G-CSF (Neupogen, Filgastrim, Amgen Inc., Thousand Oaks, CA, USA).

Endometrium was reassessed after 72 hours. If it expanded, all transfers were performed by two doctors. When endometrium was below 7 mm after G-CSF, women could choose two options: to have blastocyst transfer, despite inadequate endometrium, or to cancel the cycle. In that case, embryos were frozen. The number of embryos transferred varied from one to three.

## 3. Statistical Analysis

Continuous variables were presented as mean  $\pm$  SD; categorical variables were presented by ratio. Differences between dependent variables (before and after) were checked by paired *t*-test. Differences between independent variables were checked by *t*-test.  $P < 0.05$  was considered statistically significant. The statistical package STATISTICA (data analysis software system), version 10.0 (StatSoft Inc., Tulsa, OK; <http://www.statsoft.com/>), was used for data analysis.

## 4. Results

The baseline characteristics of participants are shown in Table 1. Table 2 presents endometrial thickness in women before and after infusion of G-CSF who had IVF-ET. In all the subjects at the time of infusion of G-CSF, endometrial thickness was  $6,74 \pm 1,75$  mm, and, after infusion, it increased to  $8,42 \pm 1,73$  mm ( $P < 0.001$ ). When we divided the group into two subgroups according to whether they conceived, we showed that the endometrium increased from  $6,86 \pm 1,65$  to  $8,80 \pm 1,14$  mm in the first one and from  $6,71 \pm 1,80$  to  $8,33 \pm 1,85$  mm in the second one ( $P < 0.001$ ). There were no significant differences between the two subgroups in respect to the endometrial thickness both before ( $P = 0.84$ ) and after infusion ( $P = 0.86$ ). The clinical pregnancy rate was 18.9%. Seven women conceived and delivered. Four women had one single sac and one single term birth. We recorded three women with two gestational sacs. One of them had preterm single live birth, whereas the second delivered prematurely single birth—the baby had intrauterine death—and the third had term birth of twins. No triplets were recorded.

All the women were supplied with low-dose aspirin, sildenafil citrate (Viagra), or both in the treatment cycle and in previous cycles.

Three out of seven women who conceived after GCSF were pregnant once before entering the study; one woman was pregnant twice. Eight out of thirty in subgroups who did not conceive were pregnant once, whereas four women twice and one women three times, respectively.

The mean endometrial thickness in previous failures was  $5,75$  mm  $\pm$   $1,0$  mm for all women; when we divided women according to whether they conceived, the mean endometrial thickness was  $6,45 \pm 0,38$  mm for those who conceived and  $5,95 \pm 0,76$  mm for those who did not conceive, respectively.

TABLE 1: Baseline patient characteristics and IVF cycle characteristics in women with thin endometrium.

Characteristic	All women <i>n</i> = 37	Women who conceived <i>n</i> = 7	Women who did not conceive <i>n</i> = 30
Age (years)	34.68 ± 4.13 (35)	32.14 ± 2.79 (33)	35.32 ± 4.20 (36)
Primary infertility diagnosis	24/37 (64.86%)	4/7 (57.14%)	20/30 (66.67%)
Secondary infertility diagnosis	13/37 (35.14%)	3/7 (42.86%)	10/30 (30.33%)
BMI (kg/m <sup>2</sup> )	23.09 ± 2.78 (23.31)	21.89 ± 1.37 (22.15)	23.36 ± 2.97 (23.85)
FSH (mIU/mL)	7.18 ± 1.91 (7.3)	6.87 ± 1.40 (7)	7.31 ± 2.19 (7.60)
AMH (ng/mL)	4.28 ± 3.29 (3.8) 0.1–12.8	6.37 ± 4.17 (4.60) 1–12.8	3.78 ± 2.91 (3.4) 0.1–10.8
Cycles	3.46 ± 2.23 (3.00) 1–11	3.29 ± 1.80 (3.00) 2–7	3.5 ± 2.35 (3.00) 1–11

Continuous variables are shown as mean ± standard deviation.

Categorical variables are shown as ratio.

TABLE 2: Endometrial thickness in women before and after infusion of G-CSF who had IVF-ET.

Characteristic	All women <i>n</i> = 37	Women who conceived <i>n</i> = 7	Women who did not conceive <i>n</i> = 30
Endometrial thickness before G-CSF-infusion	6.74 ± 1.75 <sup>1</sup>	6.86 ± 1.65 <sup>2</sup>	6.71 ± 1.80 <sup>3</sup>
Endometrial thickness after G-CSF infusion	8.42 ± 1.73 <sup>1</sup>	8.80 ± 1.14 <sup>2</sup>	8.33 ± 1.85 <sup>3</sup>
Endometrial thickness (Δ)	1.68 ± 1.05	1.94 ± 0.99 <sup>4</sup>	1.62 ± 1.07 <sup>4</sup>

Data are shown as mean ± standard deviation.

*P* value for two dependent samples (before versus after).

<sup>1</sup>*P* < 0.001.

<sup>2</sup>*P* = 0.0020.

<sup>3</sup>*P* < 0.0001.

*P* value for two independent samples (women who conceived versus women who did not conceive).

<sup>4</sup>*P* = 0.8481.

<sup>5</sup>*P* = 0.2444.

<sup>6</sup>*P* = 0.4650.

## 5. Discussion

It has been demonstrated before that <1% of women have thin endometrium [2, 16]. The thin unresponsive endometrium is still the unresolved clinical problem. There are inconclusive data regarding the diameter of so-called thin endometrium. Some investigators stated that the pregnancy occurs when endometrium reaches more than 7 mm and others that more than 9 mm [2, 3, 15, 16]. However, there are also data in the literature that endometrium 5–8 mm is inadequate [17]. Several methods were proposed, to increase thin endometrium in women undergoing IVF. These therapies included tocopherol, pentoxifylline, low-dose aspirin, sildenafil citrate, and estradiol administration [16, 18, 19].

The embryos of patients with thin endometrium have to be frozen, which leads to the real clinical dilemma for doctors. On the other side, in some investigations, there was no correlation between the IVF outcome and endometrium thickness [20, 21]. In the pilot study of Gleicher et al., the authors showed preliminary clinical report regarding the role of G-CSF on endometrium expansion in women with unresponsive endometrium [13]. In this case report, the data of four patients infused with G-CSF into the uterus were demonstrated. All these patients finally conceived. Two years later, the same group of authors described 21 patients

with inadequate thin endometrium infused with G-CSF. As a result, 19.1% ongoing clinical pregnancy rate was observed. The findings of Gleicher et al. provided evidence that G-CSF could be promising agent in the treatment of women with thin unresponsive endometrium [14].

In our study, we analyzed 37 women who underwent IVF-ET after G-CSF infusion. The clinical pregnancy rate was very similar to Gleicher study [14]—18.9%. We found that the endometrium significantly increased after infusion of G-CSF when we analyzed all the examined women and when we divided them according to the conception success. The increase of endometrium thickness was greater in group of women who conceived but the difference between groups was not statistically significant. These observations were in accordance with the study presented by Gleicher et al. [14]. In that pilot data, the endometrium increased also in all women but more in the subgroup of women who conceived. In contrast to Gleicher et al.'s study, our population was younger (34,6 versus 40.5 years) and had higher AMH concentrations (4,2 versus 1,5 ng/mL). Another difference was the time interval between G-CSF infusion and the first reassessment of the endometrium; in Gleicher et al. [14] study, it was 48 hours in contrast to 72 hours in our study. However, the question what is the appropriate interval between infusion and second reassessment of endometrium is still open. Secondly, we are

still not sure how many times G-CSF should be applied. In Gleicher et al.'s study [14], three patients (14.3%) reached the minimal thickness after the second infusion of G-CSF. In contrast, in our study, we infused G-CSF only once. We also do not know the extent to which the increase of endometrial thickness is the result of G-CSF function or the synergistic effect of added low-dose aspirin and to the protocol. This supplementation was routinely applied in our study. There are opposing results with aspirin in unselected IVF patients on the endometrium thickness and pregnancy rates [22, 23].

Our study is not without limitations. Firstly, we did not have a control group which received placebo. Thus, the changes of endometrial thickness could be observed only before and after infusion and between subgroups of women who conceived or not. Secondly, the subgroup of women who conceived was very small. Thirdly, we applied aspirin and/or sildenafil citrate which also could have a positive effect on endometrial thickness. For example, aspirin attenuates placental apoptosis, and this could be a possible explanation of how aspirin is beneficial, even in the absence of endometrial or oocyte improvement [24]. We can only speculate that the other factors could have impact on endometrial thickness. For example, we did not measure antiphospholipid antibodies [25]. The presence of them could have influenced endometrial thickness during low-aspirin treatment. But we do not believe that this could have essential impact on our results. We think that taking into account all previous failures the G-CSF effect could play the main role in our study. One should note that women who conceived were younger than women who did not but the difference between groups was not statistically significant.

Therefore, the final assessment on how G-CSF affects the expansion of endometrium thickness remains open until prospectively controlled studies would be performed.

To date, no final conclusions have been also drawn regarding which delivery system is better. Despite the aim of our study, we did not compare the assessment of adverse events and did not record any adverse effect during G-CSF infusion. However, it was demonstrated before that the treatment with G-CSF could lead to bone pain, general fatigue, headaches, insomnia, anorexia, nausea, and/or vomiting [26]. Additionally dyspnea, chest pain, hypoxemia, diaphoresis, anaphylaxis, syncope, and flushing were recorded [27]. There is also a question on how to properly counsel the patients who did not conceive and still have thin unresponsive endometrium despite G-CSF infusion. However, we showed some possibilities to the patients.

In summary, we showed that, in women who had thin endometrium in the previous IVF cycles, the infusion of G-CSF increases the endometrial thickness. Additionally, the expanding of endometrial thickness was observed after 72 hours. Because of the limited number of women and no control group, our conclusions are limited. We should also remember that the threshold is different in many other studies; thus, clinical pregnancy was observed even in women with endometrium <4 mm [28]. We think that, despite the obvious limitations, our data are important for doctors and couples seeking fertility assistance. However, further studies are needed in this field.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

## References

- [1] F. J. Broekmans, J. Kwee, D. J. Hendriks, B. W. Mol, and C. B. Lambalk, "A systematic review of tests predicting ovarian reserve and IVF outcome," *Human Reproduction Update*, vol. 12, no. 6, pp. 685–718, 2006.
- [2] A. Al-Ghamdi, S. Coskun, S. Al-Hassan, R. Al-Rejjal, and K. Awartani, "The correlation between endometrial thickness and outcome of in vitro fertilization and embryo transfer (IVF-ET) outcome," *Reproductive Biology and Endocrinology*, vol. 6, article 37, 2008.
- [3] J. D. Isaacs Jr., C. S. Wells, D. B. Williams, R. R. Odem, M. J. Gast, and R. C. Strickler, "Endometrial thickness is a valid monitoring parameter in cycles of ovulation induction with menotropins alone," *Fertility and Sterility*, vol. 65, no. 2, pp. 262–266, 1996.
- [4] A. Sharkey, "Cytokines and implantation," *Reviews of Reproduction*, vol. 3, no. 1, pp. 52–61, 1998.
- [5] A. Psychoyos, "Uterine receptivity for nidation," *Annals of the New York Academy of Sciences*, vol. 476, pp. 36–42, 1986.
- [6] Y. W. Loke, A. King, and T. D. Burrows, "Decidua in human implantation," *Human Reproduction*, vol. 10, supplement 2, pp. 14–21, 1995.
- [7] A. Barash, N. Dekel, S. Fieldust, I. Segal, E. Schechtman, and I. Granot, "Local injury to the endometrium doubles the incidence of successful pregnancies in patients undergoing in vitro fertilization," *Fertility and Sterility*, vol. 79, no. 6, pp. 1317–1322, 2003.
- [8] S. Rutella, F. Zavala, S. Danese, H. Kared, and G. Leone, "Granulocyte colony-stimulating factor: a novel mediator of T cell tolerance," *Journal of Immunology*, vol. 175, no. 11, pp. 7085–7091, 2005.
- [9] A. Salmassi, A. G. Schmutzler, L. Huang, J. Hedderich, W. Jonat, and L. Mettler, "Detection of granulocyte colony-stimulating factor and its receptor in human follicular luteinized granulosa cells," *Fertility and Sterility*, vol. 81, supplement 1, pp. 786–791, 2004.
- [10] H. Uzumaki, T. Okabe, N. Sasaki et al., "Identification and characterization of receptors for granulocyte colony-stimulating factor on human placenta and trophoblastic cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 86, no. 23, pp. 9323–9326, 1989.
- [11] F. Scarpellini and M. Sbracia, "Use of granulocyte colony-stimulating factor for the treatment of unexplained recurrent miscarriage: a randomised controlled trial," *Human Reproduction*, vol. 24, no. 11, pp. 2703–2708, 2009.
- [12] H. Wang, Y. Wen, M. L. Polan, R. Boostanfar, M. Feinman, and B. Behr, "Exogenous granulocyte-macrophage colony-stimulating factor promotes follicular development in the newborn rat in vivo," *Human Reproduction*, vol. 20, no. 10, pp. 2749–2756, 2005.
- [13] N. Gleicher, A. Vidali, and D. H. Barad, "Successful treatment of unresponsive thin endometrium," *Fertility and Sterility*, vol. 95, no. 6, pp. 2123.e13–2123.e17, 2011.
- [14] N. Gleicher, A. Kim, T. Michaeli et al., "A pilot cohort study of granulocyte colony-stimulating factor in the treatment of unresponsive thin endometrium resistant to standard therapies," *Human Reproduction*, vol. 28, no. 1, pp. 172–177, 2013.

- [15] A. Weissman, A. Barash, H. Shapiro, and R. F. Casper, "Ovarian hyperstimulation following the sole administration of agonistic analogues of gonadotrophin releasing hormone," *Human Reproduction*, vol. 13, no. 12, pp. 3421–3424, 1998.
- [16] L. N. Weckstein, A. Jacobson, D. Galen, K. Hampton, and J. Hammel, "Low-dose aspirin for oocyte donation recipients with a thin endometrium: prospective, randomized study," *Fertility and Sterility*, vol. 68, no. 5, pp. 927–930, 1997.
- [17] Y. Shufaro, A. Simon, N. Laufer, and M. Fatum, "Thin unresponsive endometrium—a possible complication of surgical curettage compromising ART outcome," *Journal of Assisted Reproduction and Genetics*, vol. 25, no. 8, pp. 421–425, 2008.
- [18] N. Lédée-Bataille, F. Olivennes, J. L. Lefaix, G. Chaouat, R. Frydman, and S. Delanian, "Combined treatment by pentoxifylline and tocopherol for recipient women with a thin endometrium enrolled in an oocyte donation programme," *Human Reproduction*, vol. 17, no. 5, pp. 1249–1253, 2002.
- [19] G. Sher and J. D. Fisch, "Effect of vaginal sildenafil on the outcome of in vitro fertilization (IVF) after multiple IVF failures attributed to poor endometrial development," *Fertility and Sterility*, vol. 78, no. 5, pp. 1073–1076, 2002.
- [20] C. de Geyter, M. Schmitter, M. de Geyter, E. Nieschlag, W. Holzgreve, and H. P. G. Schneider, "Prospective evaluation of the ultrasound appearance of the endometrium in a cohort of 1,186 infertile women," *Fertility and Sterility*, vol. 73, no. 1, pp. 106–113, 2000.
- [21] J. B. A. Oliveira, R. L. R. Baruffi, A. L. Mauri, C. G. Petersen, M. S. Campos, and J. G. Franco Jr., "Endometrial ultrasonography as a predictor of pregnancy in an in-vitro fertilization programme," *Human Reproduction*, vol. 8, no. 8, pp. 1312–1315, 1993.
- [22] M. Rubinstein, A. Marazzi, and E. P. de Fried, "Low-dose aspirin treatment improves ovarian responsiveness, uterine and ovarian blood flow velocity, implantation, and pregnancy rates in patients undergoing in vitro fertilization: a prospective, randomized, double-blind placebo-controlled assay," *Fertility and Sterility*, vol. 71, no. 5, pp. 825–829, 1999.
- [23] B. Urman, R. Mercan, C. Alatas, B. Balaban, A. Isiklar, and A. Nuhoglu, "Low-dose aspirin does not increase implantation rates in patients undergoing intracytoplasmic sperm injection: a prospective randomized study," *Journal of Assisted Reproduction and Genetics*, vol. 17, no. 10, pp. 586–590, 2000.
- [24] P. Bose, S. Black, M. Kadyrov et al., "Heparin and aspirin attenuate placental apoptosis in vitro: implications for early pregnancy failure," *The American Journal of Obstetrics and Gynecology*, vol. 192, no. 1, pp. 23–30, 2005.
- [25] G. Sher, C. Zouves, M. Feinman et al., "A rational basis for the use of combined heparin/aspirin and IVIG immunotherapy in the treatment of recurrent IVF failure associated with antiphospholipid antibodies," *The American Journal of Reproductive Immunology*, vol. 39, no. 6, pp. 391–394, 1998.
- [26] H. Khoury, D. Adkins, R. Brown et al., "Adverse side-effects associated with G-CSF in patients with chronic myeloid leukemia undergoing allogeneic peripheral blood stem cell transplantation," *Bone Marrow Transplantation*, vol. 25, no. 11, pp. 1197–1201, 2000.
- [27] A. D'Souza, I. Jaiyesimi, L. Trainor, and P. Venuturumili, "Granulocyte colony-stimulating factor administration: adverse events," *Transfusion Medicine Reviews*, vol. 22, no. 4, pp. 280–290, 2008.
- [28] P. Sundström, "Establishment of a successful pregnancy following in-vitro fertilization with an endometrial thickness of no more than 4 mm," *Human Reproduction*, vol. 13, no. 6, pp. 1550–1552, 1998.



# Hindawi

Submit your manuscripts at  
<http://www.hindawi.com>

