

Review Article

CX3CL1/CX3CR1 in Alzheimer's Disease: A Target for Neuroprotection

Peiqing Chen,¹ Wenjuan Zhao,¹ Yanjie Guo,² Juan Xu,¹ and Ming Yin¹

¹School of Pharmacy, Shanghai Jiao Tong University, 800 Dongchuan Road, Shanghai 200240, China

²Department of Neurology, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, No. 100 Haining Road, Shanghai 200080, China

Correspondence should be addressed to Wenjuan Zhao; zhaowj@sjtu.edu.cn and Ming Yin; myin@sjtu.edu.cn

Received 18 March 2016; Accepted 5 June 2016

Academic Editor: Tauheed Ishrat

Copyright © 2016 Peiqing Chen et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

CX3C chemokine ligand 1 (CX3CL1) is an intriguing chemokine belonging to the CX3C family. CX3CL1 is secreted by neurons and plays an important role in modulating glial activation in the central nervous system after binding to its sole receptor CX3CR1 which mainly is expressed on microglia. Emerging data highlights the beneficial potential of CX3CL1-CX3CR1 in the pathogenesis of Alzheimer's disease (AD), a common progressive neurodegenerative disease, and in the progression of which neuroinflammation plays a vital role. Even so, the importance of CX3CL1/CX3CR1 in AD is still controversial and needs further clarification. In this review, we make an attempt to present a concise map of CX3CL1-CX3CR1 associated with AD to find biomarkers for early diagnosis or therapeutic interventions.

1. Introduction

Alzheimer's disease (AD), a common progressive neurodegenerative disease, is the most frequent cause of cognitive decline and dementia, which affects more than 46 million people worldwide. The etiology of AD is still unclear now. One of the main pathological characteristics is extracellular deposits of β -amyloid ($A\beta$) peptides in senile plaques. $A\beta$ cascade-inflammatory hypothesis has been elucidated to look forward to seeking treatment for AD [1]. Some scholars believe that $A\beta$ -burdened neurons may play a crucial role in initiating microglial activation and eliciting chronic inflammation which lead to synaptic dysfunction, neurotoxicity, and behavioral deficits in the progression of AD [2–6]. Reactive microglia is also related to driving tau pathology and correlating with the spread of tau pathology [7], which induces neurofibrillary tangles (NFT), another major pathological characteristic of AD. Consistently, depleting microglia dramatically suppressed the propagation of tau in the brain [8].

CX3C chemokine ligand 1 (CX3CL1, also named fractalkin) plays an important role in reducing neuroinflammation and is highly expressed in the main area of pathological changes in AD, such as the hippocampus and cerebral cortex,

and the expression level of CX3CL1 reflects the progression of the disease [9]. CX3CL1 has been demonstrated to play a neuroprotective role in CNS by reducing neurotoxicity and microglial activation [10–12]. Consistent with this is the fact that treatment of aged rats with CX3CL1 attenuates the age-related increase in microglial activation [13]. Moreover, CX3CL1 also has an effect on $A\beta$ clearance and p-tau accumulation in AD [14]. All the above show that CX3CL1 has a major role in the progression of AD. In this review, we summarize the multiple roles of CX3CL1 in neuroinflammation, neurotoxicity, and synaptic plasticity in AD pathogenesis.

2. CX3CL1/CX3CR1 and Microglia

CX3CL1 is a large cytokine protein of 373 amino acids with an extended mucin-like stalk and a chemokine domain on top. It is the only member of CX3C family which belongs to the large family of small secreted chemotactic cytokines. CX3CL1 is expressed with particularly high levels in hippocampal and cortical neurons constitutively but none on microglia [15]. It exists in both secreted and membrane-bound form and its membrane-tethered mucin stalk acts as a cell adhesion

molecule adhering to microglia during an inflammatory reaction [16]. The membrane-bound form can be cleaved in the condition of cathepsin S, ADAM-10, and ADAM-17; then the soluble one can serve as a signaling molecule mediating neural/microglial interactions via its sole receptor CX3CR1 that is mainly expressed on microglia and partly on astrocyte as well as on neurons in the CNS [17–19]. These suggest that CX3CL1/CX3CR1 is an important bridge to connect neuron and microglia.

Microglia, resident mononuclear phagocytes in the CNS, intimately involved in the development of the nervous system, are highly active in their presumed resting state, continually surveying their microenvironment with extremely motile processes and protrusions [20, 21]. It has been demonstrated that A β burdened neurons inducing microglial activation may be an early phenomenon in the procession of AD [22]. However, microglia activation in AD is suggested to be heterogeneous: beneficial or harmful [23]. This may be associated with microglia activation phenotype which includes M1 (iNOS⁺ microglia) and M2 (Arg⁺ microglia); iNOS⁺ microglia induce production of neuroinflammation factors while Arg⁺ microglia have enhanced phagocytic activity. In accordance with this, greater numbers of Arg⁺ microglia containing A β were found when compared to iNOS⁺ microglia in the inflamed hemisphere [24]. Moreover, amounts of evidence indicate that microglia phenotype changes from M2 to M1 in the progression of AD [25].

Neuronal soluble CX3CL1 is likely to alter the microglial state to a more neuroprotective one by acting on CX3CR1 in microglia [26]. This also has been confirmed that disruption of CX3CL1-CX3CR1 leads to dysregulate microglial responses and neuronal damage [12, 18]. Besides, hAPP-CX3CR1^{-/-} mice as well as hTau-CX3CR1^{-/-} mice showed increased expression of inflammatory factors, enhanced tau phosphorylation, and exacerbated plaque-independent neuronal dysfunction and cognitive deficits [27, 28], while researches also demonstrated that both APP-PS1/CX3CR1^{-/-} and CRND8/CX3CR1^{-/-} mice showed reduction in A β deposition with increased number of microglia [29, 30]. Moreover, the suppression of CX3CL1-CX3CR1 alleviated A β -induced neurotoxicity and memory deficiency [31, 32]. Well, CX3CL1/CX3CR1 may play a beneficial role in controlling the progression of AD by inhibiting the inflammation and tau phosphorylation but at a cost of the increased A β deposition. Overexpression of soluble CX3CL1 by adeno-associated viral (AAV) vectors plays an active role in reducing tau pathology and neuron loss, while it has no effect on A β deposition indicating that additional CX3CL1 signaling has no additive effect on A β deposition [26, 33]. Surprisingly, neither enhanced tau phosphorylation nor reduced A β deposition in CX3CL1-deficient APP-PS1 animals was altered by soluble CX3CL1 isoform, which was introduced by bacterial artificial chromosome (BAC) transgene encoding truncated CX3CL1 [34]. Thus making the function of soluble CX3CL1 is full of doubt. A possible explanation is that AAV vectors might make soluble CX3CL1 build the required local gradient and it should suffice, while the only soluble CX3CL1 can be diluted rapidly [35]. This needs to be further clarified.

The expression of CX3CL1 is decreased in cerebral cortex and hippocampus of APP transgenic mice while it is increased in tau-injured neurons [36, 37]. Moreover, the level of plasma soluble CX3CL1 is significantly greater in the patients with mild to moderate AD than in the patients with severe AD, and the level of CX3CL1 is inversely correlated to AD severity [38]. Together, these studies suggest that CX3CL1/CX3CR1 associated with neuroinflammation, neurotoxicity, and synaptic plasticity plays variable roles in different stages of AD pathogenesis. Considering this, we conjecture that mild decreased CX3CL1-CX3CR1 due to intraneuronal A β accumulation in the early stage of AD increases clearance of A β deposition by enhancing the phagocytosis of microglia while resulting in tau hyperphosphorylation and severe downgraded CX3CL1-CX3CR1 signal gives rise to deregulated microglia and abnormally excited neuron which lead to neuron damage and loss in the progression of AD.

3. CX3CL1/CX3CR1 and Neuroinflammation

Neuroinflammation is classically attributed to A β deposition and plays a vital role in the pathological progress of AD [5, 39]. It is always correlated with increased levels of proinflammatory cytokines including tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6), IL-1 β , interferon gamma (IFN- γ), and chemokine (C-C motif) ligand 2 (CCL2) and C-X-C motif chemokine 10 (CXCL10/IP-10) [40]. CX3CL1, which is identified inhibiting the production of TNF- α , nitric oxide (NO), and superoxide in neuron-glia cell cultures [41], has been implicated as an endogenous neuronal modulator and may limit microgliosis in AD by reducing the inflammatory reaction [37, 42, 43].

TNF- α , a prototypic proinflammatory cytokine, is mainly released by activated microglia, colocalized with A β deposition, and is elevated in the cortex of animal models and human with AD [44–46]. It has been shown that glial TNF- α enhances A β deposition through inhibiting BACE1 expression and A β clearance and promotes neuronal cell cycle events which are toxic for terminally differentiated neurons in the pathogenesis of AD [47, 48]. Besides, Lourenco et al. have proved that A β oligomers lead to synapse loss and memory impairment in a TNFR1 dependent manner [49]. TNF- α activates TNFR1 leading to neuron death while TNFR2 which is expressed primarily by microglia [50] is beneficial to control microglia activity in the progression of AD [51].

Fewer A β plaques and A β -related lesions developed in APP23/TNFR1^{-/-} mice when compared with APP23/TNFR1^{+/+} littermates [52]. However, Barger et al. suggested that TNF- α protects hippocampal neurons against A β toxicity [53]. Both 3xTg-AD lacking TNF-R1+R2 and 3xTg-ADxTNF-R1/R2 knock-out exhibit enhanced A β and tau-related pathological features by the age of 15 months, in stark contrast to age-matched 3xTg-AD counterparts [54]. Loss of opposing TNFR2 leads to a stage-independent increase in Iba-1 positive microglia, and TNFR1 mediated exacerbation of A β and tau pathology in aged 3xTg-AD mice [55]. Thus suggesting the role of CX3CL1/CX3CR1 which inhibits TNF- α secretion [56] may be divaricated dependent on TNFR. But

in view of the fact that TNFR1 is increased by 17–28% and TNFR2 is significantly decreased by 35–43% in AD brains [57], CX3CL1/CX3CR1 inclines to play a beneficial role in the pathogenesis of AD.

The expression of another inflammatory cytokine IL-1 β is also increased in the CX3CR1-deficient APP/PS1 animals [29]. The major role of increased IL-1 β in neuroinflammation and subsequent induction of the microglial autophagy potentially are contributed to AD [58, 59]. CX3CR1 deficiency promotes impairment of cognitive function, synaptic plasticity, and tau hyperphosphorylation via increasing action of IL-1 β and the impairment could be reversed by infusion with IL-1 β receptor antagonist significantly [28, 42]. On the other hand, the upregulated expression of chronic IL-1 β increases plaque-associated microglia and ameliorates amyloid pathology in the APP/PS1 mouse model of AD [60, 61]. The generation of this contradiction is likely to depend on the stage of AD, which may be coordinated with CX3CL1 functions in different period.

In addition, CX3CL1 dose-dependently suppressed the production of nitric oxide (NO) [10]. NO, related to the increased levels of IFN- γ and TNF- α [62], has been involved in neuroinflammation with increased expression of inducible NO synthase (iNOS) at mild and severe stages of AD [63]. Inhibition of iNOS which mediates CNS inflammatory processes reduces the risk of AD [64]. In all, CX3CL1-CX3CR1 inhibits microglia activity via controlling the overproduction of inflammatory mediators. The distinctly decreased expression of CX3CL1 gives rise to dysregulated microglia, leading to neuroinflammation. Drugs that attenuate neuronal degeneration and improve learning and memory ability are accompanied by reduced TNF- α , IL-1 β , TGF- β , and NO levels induced by A β in CSF in mouse models and patients with AD [65–70]. Apart from AD, CX3CL1/CX3CR1 is also involved in other neuroinflammation disorders, including Parkinson's Disease (PD) [71, 72], multiple sclerosis (MS) [73], tauopathies [33], and age-related macular degeneration (ARMD) [74]. These neurodegenerative disorders are all associated with chronic neuroinflammation caused by activated microglia [75], indicating that CX3CL1/CX3CR1 may have the similar mechanisms between AD and other neurodegenerative disorders in regulating neuroinflammation. The complex roles of CX3CL1/CX3CR1 are still being studied.

4. CX3CL1/CX3CR1 Regulates Synaptic Plasticity

Synaptic plasticity plays an important role in learning and memory, and A β -induced synaptic dysfunction is strongly associated with AD [76]. CX3CL1 is upregulated in the rat hippocampus during memory-associated synaptic plasticity [77]. It is considered as a potent neuromodulator of the evoked excitatory synaptic transmission and plays a major role in synaptic plasticity and neuroprotection [78]. Furthermore, the functions of CX3CL1 rely on CX3CR1, as long-lasting-enriched environment failed to affect hippocampal-dependent plasticity in the absence of CX3CR1 [79]. Although the underlying mechanisms have

been underexplored, CX3CL1/CX3CR1 may mediate synaptic plasticity and cognitive function mainly by regulating long-term potentiation (LTP) [80], NO signaling, and production of brain-derived neurotrophic factor (BDNF) [81].

LTP is thought to be related to the storage of declarative memory in the mammalian brain [82]. CX3CL1 clearly interferes with LTP mechanisms and its modulation of neuronal plasticity appears to be mediated through activation of adenosine [80]. Adenosine acts as a neuromodulator with four types of G protein-coupled receptors, termed A1, A2A, A2B, and A3, and exerts important functions in the synaptic plasticity [83]. The downstream pathways branch because of the different types of adenosine receptor. Intracerebroventricular injection of A β _{1–42} inhibited not only NMDA receptor-dependent LTP but also voltage-activated Ca²⁺-dependent LTP induced by strong conditioning stimulation during NMDAR blockade [84], indicating that there is a non-NMDAR-dependent but Ca²⁺-dependent pathway involved in synaptic dysfunction in AD. CX3CL1 increases NMDA-fast excitatory postsynaptic potentials by a mechanism involving the activity of the adenosine receptor type A2 (A2AR) and the release of the NMDAR coagonist D-serine [85]. NMDAR activation affects the threshold for LTP induction which is strongly influenced by the recent history of synaptic activity [86]. An increased density of A2AR on microglia has been detected in human cortex from AD patients [87]. Thus indicating CX3CL1/CX3CR1 may activate A2AR by increasing adenosine and promote the release of D-serine; then D-serine enhances the function of NMDAR and facilitates LTP. Moreover, CX3CL1 causes a reversible depression of excitatory postsynaptic current (EPSC), which is abolished by the A3R antagonist [88], and the inhibition failed to occur in CX3CR1 null mice [80]. Stimulation of A3R induces an intracellular signaling that increases calcium concentrations [89]. The phosphorylation of CAMKII and cyclic adenosine monophosphate response element-binding protein (CREB) is important to hippocampal long-term synaptic plasticity [90]. α CaMKII autophosphorylation is also required for synaptic plasticity induced by a short and precise stimulus, but maybe not for a longer and stronger stimulation [91]. Besides, the reduction of CREB activation also leads to memory impairment [92]. Based on the information given above, we can hypothesize the way CX3CL1 affects LTP; that is, CX3CL1 acts with CX3CR1 on the surface of the microglia and stimulates the release of adenosine; adenosine then activates A2AR and promotes synaptic facilitation by NMDAR-dependent pathway, activates A3R simultaneity, and induces synaptic inhibition by a Ca²⁺-dependent pathway.

Brain-derived neurotrophic factor (BDNF), an important growth factor in the CNS, is of great significance for neurons to maintain the survival, growth, differentiation, repair, and regeneration after nerve injury as well as increasing synaptic plasticity. A clinical study involving 535 old participants who underwent annual cognitive assessments and brain autopsy at death showed that higher brain BDNF expression is associated with slower cognitive decline and BDNF may also reduce the deleterious effects of AD pathology on cognitive decline [93]. Studies have shown that A β induces decreased anterograde as well as retrograde transport of BDNF vesicles

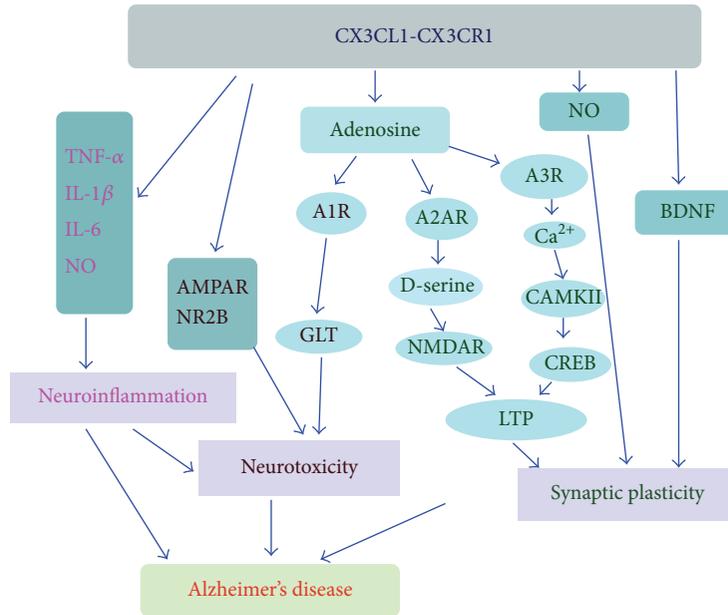


FIGURE 1: The effect of CX3CL1/CX3CR1 in Alzheimer's disease (AD). CX3CL1 binds to CX3CR1 which is its sole receptor and mainly expressed by microglia: (1) regulating introduction of inflammation cytokine (TNF- α , IL-1 β , IL-6, NO, etc.) and reducing neuroinflammation in AD; (2) negatively modulating the function of AMPAR and NR2B, increasing GLT activity through the mechanism dependent on A1R, and then decreasing the neurotoxicity induced by Glu; (3) stimulating the release of adenosine; adenosine then activates A2AR and promotes synaptic facilitation by NMDAR-dependent pathway and simultaneously activates A3R and induces synaptic inhibition by Ca²⁺-dependent pathway. TNF- α : tumor necrosis factor-alpha; IL-1 β : interleukin-1 β ; IL-6: interleukin-6; NO: nitric oxide; A1R: adenosine 1 receptor; A2AR: adenosine A2a receptor; A3R: adenosine 3 receptor; GLT: glutamate transporter; LTP: long-term potentiation; CREB: cyclic adenosine monophosphate response element-binding protein; BDNF: brain-derived neurotrophic factor.

in hippocampal neurons of various AD models [94]. Upregulation of BDNF by activating of ERK/CREB pathway can ameliorate A β -induced neurons loss and dendritic atrophy [95]. Restoration of normal neuronal BDNF expression levels in the cerebral hippocampi and cortices ameliorates the impairment in recognition memory and associative learning in mice of AD [96]. Importantly, BDNF concentrations are associated with CX3CL1 [97]. Chronic injection of CX3CL1 rescues the hippocampal-dependent memory deficits and reverses the decreased hippocampal neurogenesis in genetic BDNF variation mice [98].

In addition, NO is also consistently involved in recognition memory [99]. As mentioned before, CX3CL1/CX3CR1 inhibits the expression of NO in activated microglia cells [10, 100] and may induce synaptic inhibition. NO signaling through neuronal NO synthase (nNOS) prior to the appearance of cognitive symptoms focuses on early developments of AD [63]. NO/soluble GC (sGC)/cGMP-PKG and ERK signaling is important for modulating synaptic transmission and plasticity in the hippocampus and cerebral cortex, which are critical for learning and memory [101]. Recruitment of NO is serving a compensatory role to boost synaptic transmission and plasticity during early AD stage [102]. NO inhibitors ameliorate overexpressed NMDA receptor subunit NR2B which plays a role in memory formation in an inflammatory model of AD [103]. Besides, endothelial NO deficiency also promotes AD pathology [104].

5. CX3CL1/CX3CR1 Reduces Excitotoxicity

It has been verified that CX3CL1 released from hippocampal cells after excitotoxic insult has an essential role in brain protection by reducing against glutamate mediated excitotoxicity [105]. As mentioned before, microglia shape their neuronal environment actively thanks to their ability to trigger neuronal death [106–108]. Apart from regulating neuroinflammation, CX3CL1/CX3CR1 negatively modulates the function of AMPA receptor at active glutamatergic synapses [109]. CX3CL1 reduces the glutamate mediated excitotoxicity by reducing the influx of Ca²⁺ [105]. Calcium channel blockers also exhibit cognitive enhancing abilities and reduce the risk of dementia genuinely [110]. Moreover, the application of ion channel blockers with specific antagonists of the NR2B subunit could reduce neurotoxicity significantly [111]. Besides, CX3CL1 mediated neuroprotection by increasing glutamate transporter-1 (GLT-1) activity on astrocytes is dependent on the presence and the activity of A1 adenosine receptor (A1R), which can be blocked by the specific antagonist DPCPX and absent in A1R^{-/-} astrocytes [112, 113]. Consistently, hippocampal neurons obtained from A1R^{-/-} mice are not protected by CX3CL1 against Glu excitotoxicity [114]. Collectively, these data indicate that CX3CL1/CX3CR1 reduces excitotoxicity by modulating glutamatergic transmission and may play an important role in cognitive functions in AD.

6. Conclusions and Perspectives

There is persistent neuroinflammation throughout the progression of AD associated with neurotoxicity and synaptic dysfunction [115, 116]. The expression of CX3CL1 is significantly decreased in AD and inversely correlated to AD severity. As shown in Figure 1, CX3CL1/CX3CR1 may regulate the activation of microglia by controlling the release of inflammatory cytokines and synaptic plasticity and cognitive functions by modulating receptors in neurons directly or indirectly. The involvement of CX3CL1/CX3CR1 in AD suggests that CX3CL1/CX3CR1 contributes positively to neuron protective as well as detrimental role in the course of the disease. Therefore, targeting CX3CL1 and/or CX3CR1 may provide novel opportunities for treatment of AD. In particular, the development stage of the disease should be considered to better analyze the functions of CX3CL1/CX3CR1 in the progression of AD.

In addition, experiment evidences have described the active involvement of CX3CL1/CX3CR1 in many other diseases, such as atherosclerotic, allergic asthma and rhinitis, renal diseases, rheumatoid arthritis (RA), Sjögren's syndrome (SS), systemic lupus erythematosus (SLE), scleroderma, colorectal cancer, and breast cancer [117–123]. For example, a genetically defined less active CX3CL1/CX3CR1 pathway is associated with a reduced risk of atherosclerotic disease in humans and the blockade of the CX3CL1/CX3CR1 pathway ameliorates the severity of atherosclerosis [124, 125]. Moreover, insulin resistance (IR) increases atherosclerotic lesion vulnerability, and this is related to the augment of CX3CL1/CX3CR1 axis [126]. All these indicate that any pharmacological agent that alters CX3CL1 signaling in AD should take into account any other potential effects.

Competing Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This study was supported by the grants from the Natural Science Foundation of China (NSFC) (Grant no. 81471232), the Natural Science Foundation of Shanghai Science and Technology Commission, China (Grant no. 14431901400), and the scientific research fund of Shanghai Jiaotong University (Grant no. 14X130040002).

References

- [1] P. L. McGeer and E. G. McGeer, "The amyloid cascade-inflammatory hypothesis of Alzheimer disease: implications for therapy," *Acta Neuropathologica*, vol. 126, no. 4, pp. 479–497, 2013.
- [2] S. Miyata, Y. Nishimura, and T. Nakashima, "Perineuronal nets protect against amyloid β -protein neurotoxicity in cultured cortical neurons," *Brain Research*, vol. 1150, no. 1, pp. 200–206, 2007.
- [3] V. H. Perry, J. A. R. Nicoll, and C. Holmes, "Microglia in neurodegenerative disease," *Nature Reviews Neurology*, vol. 6, no. 4, pp. 193–201, 2010.
- [4] B. T. Hyman, "Amyloid-dependent and amyloid-independent stages of alzheimer disease," *Archives of Neurology*, vol. 68, no. 8, pp. 1062–1064, 2011.
- [5] C. E. Hanzel, A. Pichet-Binette, L. S. B. Pimentel et al., "Neuronal driven pre-plaque inflammation in a transgenic rat model of Alzheimer's disease," *Neurobiology of Aging*, vol. 35, no. 10, pp. 2249–2262, 2014.
- [6] M. Noda, Y. Doi, J. Liang et al., "Fractalkine attenuates excitoneurotoxicity via microglial clearance of damaged neurons and antioxidant enzyme heme oxygenase-1 expression," *Journal of Biological Chemistry*, vol. 286, no. 3, pp. 2308–2319, 2011.
- [7] N. Maphis, G. Xu, O. N. Kokiko-Cochran et al., "Reactive microglia drive tau pathology and contribute to the spreading of pathological tau in the brain," *Brain: A Journal of Neurology*, vol. 138, part 6, pp. 1738–1755, 2015.
- [8] H. Asai, S. Ikezu, S. Tsunoda et al., "Depletion of microglia and inhibition of exosome synthesis halt tau propagation," *Nature Neuroscience*, vol. 18, no. 11, pp. 1584–1593, 2015.
- [9] S. Strobel, E. Grünblatt, P. Riederer et al., "Changes in the expression of genes related to neuroinflammation over the course of sporadic Alzheimer's disease progression: CX3CL1, TREM2, and PPAR γ ," *Journal of Neural Transmission*, vol. 122, no. 7, pp. 1069–1076, 2015.
- [10] T. Mizuno, J. Kawanokuchi, K. Numata, and A. Suzumura, "Production and neuroprotective functions of fractalkine in the central nervous system," *Brain Research*, vol. 979, no. 1-2, pp. 65–70, 2003.
- [11] M. M. Pabon, A. D. Bachstetter, C. E. Hudson, C. Gemma, and P. C. Bickford, "CX3CL1 reduces neurotoxicity and microglial activation in a rat model of Parkinson's disease," *Journal of Neuroinflammation*, vol. 8, article 9, 2011.
- [12] H. Y. Febinger, H. E. Thomasy, M. N. Pavlova et al., "Time-dependent effects of CX3CR1 in a mouse model of mild traumatic brain injury," *Journal of Neuroinflammation*, vol. 12, no. 1, article 154, 2015.
- [13] A. Lyons, A. M. Lynch, E. J. Downer et al., "Fractalkine-induced activation of the phosphatidylinositol-3 kinase pathway attenuates microglial activation in vivo and in vitro," *Journal of Neurochemistry*, vol. 110, no. 5, pp. 1547–1556, 2009.
- [14] J. Merino, V. Muñetón-Gómez, M. Álvarez, and A. Toledano-Díaz, "Effects of CX3CR1 and fractalkine chemokines in amyloid beta clearance and p-Tau accumulation in Alzheimer's Disease (AD) rodent models: is fractalkine a systemic biomarker for AD?" *Current Alzheimer Research*, vol. 13, no. 4, pp. 403–412, 2016.
- [15] J. K. Harrison, Y. Jiang, S. Chen et al., "Role for neuronally derived fractalkine in mediating interactions between neurons and CX3CR1-expressing microglia," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 18, pp. 10896–10901, 1998.
- [16] P. Hermand, F. Pincet, S. Carvalho et al., "Functional adhesiveness of the CX3CL1 chemokine requires its aggregation: role of the transmembrane domain," *Journal of Biological Chemistry*, vol. 283, no. 44, pp. 30225–30234, 2008.
- [17] G. A. Chapman, K. Moores, D. Harrison, C. A. Campbell, B. R. Stewart, and P. J. Strijbos, "Fractalkine cleavage from neuronal membranes represents an acute event in the inflammatory response to excitotoxic brain damage," *Journal of Neuroscience*, vol. 20, no. 15, article RC87, 5 pages, 2000.

- [18] A. E. Cardona, E. P. Pioro, M. E. Sasse et al., "Control of microglial neurotoxicity by the fractalkine receptor," *Nature Neuroscience*, vol. 9, no. 7, pp. 917–924, 2006.
- [19] O. Meucci, A. Fatatis, A. A. Simen, and R. J. Miller, "Expression of CX3CR1 chemokine receptors on neurons and their role in neuronal survival," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 14, pp. 8075–8080, 2000.
- [20] A. Nimmerjahn, F. Kirchhoff, and F. Helmchen, "Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo," *Science*, vol. 308, no. 5726, pp. 1314–1318, 2005.
- [21] K. Helmut, U.-K. Hanisch, M. Noda, and A. Verkhratsky, "Physiology of microglia," *Physiological Reviews*, vol. 91, no. 2, pp. 461–553, 2011.
- [22] E. R. Zimmer, A. Leuzy, A. L. Benedet, J. Breitner, S. Gauthier, and P. Rosa-Neto, "Tracking neuroinflammation in Alzheimer's disease: the role of positron emission tomography imaging," *Journal of Neuroinflammation*, vol. 11, article 120, 2014.
- [23] D. Tejera and M. Heneka, "Microglia in Alzheimer's disease: the good, the bad and the ugly," *Current Alzheimer Research*, vol. 13, no. 4, pp. 370–380, 2016.
- [24] J. D. Cherry, J. A. Olschowka, and M. K. O'Banion, "Arginase 1+ microglia reduce A β plaque deposition during IL-1 β -dependent neuroinflammation," *Journal of Neuroinflammation*, vol. 12, no. 1, article 203, 2015.
- [25] M. M. Varnum and T. Ikezu, "The classification of microglial activation phenotypes on neurodegeneration and regeneration in alzheimer's disease brain," *Archivum Immunologiae et Therapiae Experimentalis*, vol. 60, no. 4, pp. 251–266, 2012.
- [26] K. R. Nash, P. Moran, D. J. Finneran et al., "Fractalkine over expression suppresses α -synuclein-mediated neurodegeneration," *Molecular Therapy*, vol. 23, no. 1, pp. 17–23, 2015.
- [27] S.-H. Cho, B. Sun, Y. Zhou et al., "CX3CR1 protein signaling modulates microglial activation and protects against plaque-independent cognitive deficits in a mouse model of Alzheimer disease," *The Journal of Biological Chemistry*, vol. 286, no. 37, pp. 32713–32722, 2011.
- [28] K. Bhaskar, M. Konerth, O. N. Kokiko-Cochran, A. Cardona, R. M. Ransohoff, and B. T. Lamb, "Regulation of tau pathology by the microglial fractalkine receptor," *Neuron*, vol. 68, no. 1, pp. 19–31, 2010.
- [29] S. Lee, N. H. Varvel, M. E. Konerth et al., "CX3CR1 deficiency alters microglial activation and reduces beta-amyloid deposition in two Alzheimer's disease mouse models," *The American Journal of Pathology*, vol. 177, no. 5, pp. 2549–2562, 2010.
- [30] Z. Liu, C. Condello, A. Schain, R. Harb, and J. Grutzendler, "CX3CR1 in microglia regulates brain amyloid deposition through selective protofibrillar amyloid- β phagocytosis," *Journal of Neuroscience*, vol. 30, no. 50, pp. 17091–17101, 2010.
- [31] J. Dworzak, B. Renvoisé, J. Habchi et al., "Neuronal Cx3cr1 deficiency protects against amyloid β -induced neurotoxicity," *PLoS ONE*, vol. 10, no. 6, Article ID e0127730, 2015.
- [32] J. Wu, B. Bie, H. Yang, J. J. Xu, D. L. Brown, and M. Naguib, "Suppression of central chemokine fractalkine receptor signaling alleviates amyloid-induced memory deficiency," *Neurobiology of Aging*, vol. 34, no. 12, pp. 2843–2852, 2013.
- [33] K. R. Nash, D. C. Lee, J. B. Hunt et al., "Fractalkine overexpression suppresses tau pathology in a mouse model of tauopathy," *Neurobiology of Aging*, vol. 34, no. 6, pp. 1540–1548, 2013.
- [34] S. Lee, G. Xu, T. R. Jay et al., "Opposing effects of membrane-anchored CX3CL1 on amyloid and tau pathologies via the p38 MAPK pathway," *The Journal of Neuroscience*, vol. 34, no. 37, pp. 12538–12546, 2014.
- [35] K.-W. Kim, A. Vallon-Eberhard, E. Zigmond et al., "In vivo structure/function and expression analysis of the CX3C chemokine fractalkine," *Blood*, vol. 118, no. 22, pp. e156–e167, 2011.
- [36] R.-S. Duan, X. Yang, Z.-G. Chen et al., "Decreased fractalkine and increased IP-10 expression in aged brain of APPswe transgenic mice," *Neurochemical Research*, vol. 33, no. 6, pp. 1085–1089, 2008.
- [37] I. Lastres-Becker, N. G. Innamorato, T. Jaworski et al., "Fractalkine activates NRF2/NFE2L2 and heme oxygenase 1 to restrain tauopathy-induced microgliosis," *Brain*, vol. 137, no. 1, pp. 78–91, 2014.
- [38] T.-S. Kim, H.-K. Lim, J. Y. Lee et al., "Changes in the levels of plasma soluble fractalkine in patients with mild cognitive impairment and Alzheimer's disease," *Neuroscience Letters*, vol. 436, no. 2, pp. 196–200, 2008.
- [39] G. Halliday, S. R. Robinson, C. Shepherd, and J. Kril, "Alzheimer's disease and inflammation: a review of cellular and therapeutic mechanisms," *Clinical and Experimental Pharmacology and Physiology*, vol. 27, no. 1-2, pp. 1–8, 2000.
- [40] S. Zaheer, R. Thangavel, Y. Wu, M. M. Khan, D. Kempuraj, and A. Zaheer, "Enhanced expression of glia maturation factor correlates with glial activation in the brain of triple transgenic Alzheimer's disease mice," *Neurochemical Research*, vol. 38, no. 1, pp. 218–225, 2013.
- [41] H. A. Mattison, H. Nie, H. Gao, H. Zhou, J.-S. Hong, and J. Zhang, "Suppressed pro-inflammatory response of microglia in CX3CR1 knockout mice," *Journal of Neuroimmunology*, vol. 257, no. 1-2, pp. 110–115, 2013.
- [42] J. T. Rogers, J. M. Morganti, A. D. Bachstetter et al., "CX3CR1 deficiency leads to impairment of hippocampal cognitive function and synaptic plasticity," *The Journal of Neuroscience*, vol. 31, no. 45, pp. 16241–16250, 2011.
- [43] K. Biber, H. Neumann, K. Inoue, and H. W. G. M. Boddeke, "Neuronal 'On' and 'Off' signals control microglia," *Trends in Neurosciences*, vol. 30, no. 11, pp. 596–602, 2007.
- [44] H. Fillit, W. Ding, L. Buee et al., "Elevated circulating tumor necrosis factor levels in Alzheimer's disease," *Neuroscience Letters*, vol. 129, no. 2, pp. 318–320, 1991.
- [45] E. Tarkowski, K. Blennow, A. Wallin, and A. Tarkowski, "Intracerebral production of tumor necrosis factor- α , a local neuroprotective agent, in Alzheimer disease and vascular dementia," *Journal of Clinical Immunology*, vol. 19, no. 4, pp. 223–230, 1999.
- [46] H.-J. Kang, J.-M. Kim, S.-W. Kim et al., "Associations of cytokine genes with Alzheimer's disease and depression in an elderly Korean population," *Journal of Neurology, Neurosurgery & Psychiatry*, vol. 86, no. 9, pp. 1002–1007, 2015.
- [47] M. Yamamoto, T. Kiyota, M. Horiba et al., "Interferon- γ and tumor necrosis factor- α regulate amyloid- β plaque deposition and β -secretase expression in Swedish mutant APP transgenic mice," *American Journal of Pathology*, vol. 170, no. 2, pp. 680–692, 2007.
- [48] K. Bhaskar, N. Maphis, G. Xu et al., "Microglial derived tumor necrosis factor- α drives Alzheimer's disease-related neuronal cell cycle events," *Neurobiology of Disease*, vol. 62, pp. 273–285, 2014.
- [49] M. V. Lourenco, J. R. Clarke, R. L. Frozza et al., "TNF- α mediates PKR-dependent memory impairment and brain IRS-1

- inhibition induced by Alzheimer's β -amyloid oligomers in mice and monkeys," *Cell Metabolism*, vol. 18, no. 6, pp. 831–843, 2013.
- [50] M. K. McCoy and M. G. Tansey, "TNF signaling inhibition in the CNS: implications for normal brain function and neurodegenerative disease," *Journal of Neuroinflammation*, vol. 5, article 45, 2008.
- [51] X. Cheng, Y. Shen, and R. Li, "Targeting TNF: a therapeutic strategy for Alzheimer's disease," *Drug Discovery Today*, vol. 19, no. 11, pp. 1822–1827, 2014.
- [52] P. He, Z. Zhong, K. Lindholm et al., "Deletion of tumor necrosis factor death receptor inhibits amyloid β generation and prevents learning and memory deficits in Alzheimer's mice," *The Journal of Cell Biology*, vol. 178, no. 5, pp. 829–841, 2007.
- [53] S. W. Barger, D. Hörster, K. Furukawa, Y. Goodman, J. Kriegelstein, and M. P. Mattson, "Tumor necrosis factors alpha and beta protect neurons against amyloid beta-peptide toxicity: evidence for involvement of a kappa B-binding factor and attenuation of peroxide and Ca²⁺ accumulation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 92, no. 20, pp. 9328–9332, 1995.
- [54] S. L. Montgomery, M. A. Mastrangelo, D. Habib et al., "Ablation of TNF-RI/RII expression in Alzheimer's disease mice leads to an unexpected enhancement of pathology: Implications for chronic pan-TNF- α suppressive therapeutic strategies in the brain," *American Journal of Pathology*, vol. 179, no. 4, pp. 2053–2070, 2011.
- [55] S. L. Montgomery, W. C. Narrow, M. A. Mastrangelo, J. A. Olschowka, M. K. O'Banion, and W. J. Bowers, "Chronic neuron- and age-selective down-regulation of TNF receptor expression in triple-transgenic alzheimer disease mice leads to significant modulation of amyloid- and Tau-related pathologies," *American Journal of Pathology*, vol. 182, no. 6, pp. 2285–2297, 2013.
- [56] V. Zujovic, J. Benavides, X. Vigé, C. Carter, and V. Taupin, "Fractalkine modulates TNF- α secretion and neurotoxicity induced by microglial activation," *Glia*, vol. 29, no. 4, pp. 305–315, 2000.
- [57] X. Cheng, L. Yang, P. He, R. Li, and Y. Shen, "Differential activation of tumor necrosis factor receptors distinguishes between brains from Alzheimer's disease and non-demented patients," *Journal of Alzheimer's Disease*, vol. 19, no. 2, pp. 621–630, 2010.
- [58] P. Wang, X. Yu, P. P. Guan et al., "Magnesium ion influx reduces neuroinflammation in A β precursor protein/Presenilin 1 transgenic mice by suppressing the expression of interleukin-1 β ," *Cellular and Molecular Immunology*, 2015.
- [59] A. François, F. Terro, T. Janet, A. R. Bilan, M. Paccalin, and G. Page, "Involvement of interleukin-1 β in the autophagic process of microglia: relevance to Alzheimer's disease," *Journal of Neuroinflammation*, vol. 10, article 151, 2013.
- [60] F. Rivera-Escalera, S. B. Matousek, S. Ghosh, J. A. Olschowka, and M. K. O'Banion, "Interleukin-1 β mediated amyloid plaque clearance is independent of CCR2 signaling in the APP/PS1 mouse model of Alzheimer's disease," *Neurobiology of Disease*, vol. 69, pp. 124–133, 2014.
- [61] S. B. Matousek, S. Ghosh, S. S. Shaftef, S. Kyrkanides, J. A. Olschowka, and M. K. O'Banion, "Chronic IL-1 β -mediated neuroinflammation mitigates amyloid pathology in a mouse model of alzheimer's disease without inducing overt neurodegeneration," *Journal of Neuroimmune Pharmacology*, vol. 7, no. 1, pp. 156–164, 2012.
- [62] M. Belkhelda, H. Rafa, O. Medjebber et al., "IFN- γ and TNF- α are involved during Alzheimer disease progression and correlate with nitric oxide production: a study in Algerian patients," *Journal of Interferon and Cytokine Research*, vol. 34, no. 11, pp. 839–847, 2014.
- [63] R. Balez and L. Ooi, "Getting to NO Alzheimer's disease: neuroprotection versus neurotoxicity mediated by nitric oxide," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 3806157, 8 pages, 2016.
- [64] P. Jiang, C. Li, Z. Xiang, and B. Jiao, "Tanshinone IIA reduces the risk of Alzheimer's disease by inhibiting iNOS, MMP-2 and NF- κ Bp65 transcription and translation in the temporal lobes of rat models of Alzheimer's disease," *Molecular Medicine Reports*, vol. 10, no. 2, pp. 689–694, 2014.
- [65] S. Shi, D. Liang, Y. Chen et al., "Gx-50 reduces β -amyloid-induced TNF- α , IL-1 β , NO, and PGE2 expression and inhibits NF-kappaB signaling in a mouse model of Alzheimer's disease," *European Journal of Immunology*, vol. 46, no. 3, pp. 665–676, 2016.
- [66] W. Li, J.-W. Zhang, F. Lu et al., "Effects of telmisartan on the level of A β 1–42, interleukin-1 β , tumor necrosis factor α and cognition in hypertensive patients with Alzheimer's disease," *Zhonghua Yi Xue Za Zhi*, vol. 92, no. 39, pp. 2743–2746, 2012.
- [67] Y. Y. Zhang, Y. C. Fan, M. Wang et al., "Atorvastatin attenuates the production of IL-1 β , IL-6, and TNF- α in the hippocampus of an amyloid β 1–42-induced rat model of Alzheimer's disease," *Journal of Clinical Interventions in Aging*, vol. 8, pp. 103–110, 2013.
- [68] X. Cheng, Q. Wang, N. Li, and H. Zhao, "Effects of resveratrol on hippocampal astrocytes and expression of TNF- α in Alzheimer's disease model rats," *Wei Sheng Yan Jiu*, vol. 44, no. 4, pp. 610–614, 2015.
- [69] A. K. Sachdeva and K. Chopra, "Lycopene abrogates A β (1–42)-mediated neuroinflammatory cascade in an experimental model of Alzheimer's disease," *Journal of Nutritional Biochemistry*, vol. 26, no. 7, pp. 736–744, 2015.
- [70] A.-G. Xuan, X.-B. Pan, P. Wei et al., "Valproic acid alleviates memory deficits and attenuates amyloid- β deposition in transgenic mouse model of Alzheimer's disease," *Molecular Neurobiology*, vol. 51, no. 1, pp. 300–312, 2014.
- [71] A. D. Thome, D. G. Standaert, and A. S. Harms, "Fractalkine signaling regulates the inflammatory response in an α -synuclein model of Parkinson disease," *PLoS ONE*, vol. 10, no. 10, Article ID e0140566, 2015.
- [72] J. M. Morganti, K. R. Nash, B. A. Grimmig et al., "The soluble isoform of CX3CL1 is necessary for neuroprotection in a mouse model of Parkinson's disease," *The Journal of Neuroscience*, vol. 32, no. 42, pp. 14592–14601, 2012.
- [73] L. Stojković, T. Djurić, A. Stanković et al., "The association of V249I and T280M fractalkine receptor haplotypes with disease course of multiple sclerosis," *Journal of Neuroimmunology*, vol. 245, no. 1–2, pp. 87–92, 2012.
- [74] M. K. Falk, A. Singh, C. Faber, M. H. Nissen, T. Hviid, and T. L. Sørensen, "CX3CL1/CX3CR1 and CCL2/CCR2 chemokine/chemokine receptor complex in patients with AMD," *PLoS ONE*, vol. 9, no. 12, Article ID e112473, 2014.
- [75] T. C. Frank-Cannon, L. T. Alto, F. E. McAlpine, and M. G. Tansey, "Does neuroinflammation fan the flame in neurodegenerative diseases?" *Molecular Neurodegeneration*, vol. 4, article 47, 2009.
- [76] Z.-C. Wang, J. Zhao, and S. Li, "Dysregulation of synaptic and extrasynaptic N-methyl-D-aspartate receptors induced by

- amyloid- β ," *Neuroscience Bulletin*, vol. 29, no. 6, pp. 752–760, 2013.
- [77] G. K. Sheridan, A. Wdowicz, M. Pickering et al., "CX3CL1 is up-regulated in the rat hippocampus during memory-associated synaptic plasticity," *Frontiers in Cellular Neuroscience*, vol. 8, article 233, 2014.
- [78] C. Bertollini, D. Ragozzino, C. Gross, C. Limatola, and F. Eusebi, "Fractalkine/CX3CL1 depresses central synaptic transmission in mouse hippocampal slices," *Neuropharmacology*, vol. 51, no. 4, pp. 816–821, 2006.
- [79] L. Maggi, M. Scianni, I. Branchi, I. D'Andrea, C. Lauro, and C. Limatola, "CX₃CR1 deficiency alters hippocampal-dependent plasticity phenomena blunting the effects of enriched environment," *Frontiers in Cellular Neuroscience*, vol. 5, no. 22, 2011.
- [80] L. Maggi, F. Trettel, M. Scianni et al., "LTP impairment by fractalkine/CX3CL1 in mouse hippocampus is mediated through the activity of adenosine receptor type 3 (A3R)," *Journal of Neuroimmunology*, vol. 215, no. 1-2, pp. 36–42, 2009.
- [81] C. N. Parkhurst, G. Yang, I. Ninan et al., "Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor," *Cell*, vol. 155, no. 7, pp. 1596–1609, 2013.
- [82] L. R. Squire and S. Zola-Morgan, "The medial temporal lobe memory system," *Science*, vol. 253, no. 5026, pp. 1380–1386, 1991.
- [83] B. B. Fredholm, M. P. Abbracchio, G. Burnstock et al., "Nomenclature and classification of purinoceptors," *Pharmacological Reviews*, vol. 46, no. 2, pp. 143–156, 1994.
- [84] I. Klyubin, T. Ondrejcek, J. Hayes et al., "Neurotransmitter receptor and time dependence of the synaptic plasticity disrupting actions of Alzheimer's disease A β in vivo," *Philosophical Transactions of the Royal Society B: Biological Sciences*, vol. 369, no. 1633, 2014.
- [85] M. Scianni, L. Antonilli, G. Chece et al., "Fractalkine (CX₃CL1) enhances hippocampal N-methyl-d-aspartate receptor (NMDAR) function via d-serine and adenosine receptor type A2 (A_{2A}R) activity," *Journal of Neuroinflammation*, vol. 10, article 108, 2013.
- [86] Y.-Y. Huang, A. Colino, D. K. Selig, and R. C. Malenka, "The influence of prior synaptic activity on the induction of long-term potentiation," *Science*, vol. 255, no. 5045, pp. 730–733, 1992.
- [87] J. L. Albasanz, S. Perez, M. Barrachina, I. Ferrer, and M. Martin, "Up-regulation of adenosine receptors in the frontal cortex in Alzheimer's disease," *Brain Pathology*, vol. 18, no. 2, pp. 211–219, 2008.
- [88] S. Piccinin, S. Di Angelantonio, A. Piccioni et al., "CX3CL1-induced modulation at CA1 synapses reveals multiple mechanisms of EPSC modulation involving adenosine receptor subtypes," *Journal of Neuroimmunology*, vol. 224, no. 1-2, pp. 85–92, 2010.
- [89] M. E. Olah and G. L. Stiles, "Adenosine receptor subtypes: characterization and therapeutic regulation," *Annual Review of Pharmacology and Toxicology*, vol. 35, pp. 581–606, 1995.
- [90] F. Qiao, X.-P. Gao, L. Yuan, H.-Y. Cai, and J.-S. Qi, "Apolipoprotein E4 impairs in vivo hippocampal long-term synaptic plasticity by reducing the phosphorylation of CaMKII α and CREB," *Journal of Alzheimer's Disease*, vol. 41, no. 4, pp. 1165–1176, 2014.
- [91] A. Villers, K. P. Giese, and L. Ris, "Long-term potentiation can be induced in the CA1 region of hippocampus in the absence of α CaMKII T286-autophosphorylation," *Learning and Memory*, vol. 21, no. 11, pp. 616–626, 2014.
- [92] A. F. Teich, R. E. Nicholls, D. Puzzo et al., "Synaptic therapy in Alzheimer's disease: a CREB-centric approach," *Neurotherapeutics*, vol. 12, no. 1, pp. 29–41, 2015.
- [93] A. S. Buchman, L. Yu, P. A. Boyle, J. A. Schneider, P. L. De Jager, and D. A. Bennett, "Higher brain BDNF gene expression is associated with slower cognitive decline in older adults," *Neurology*, vol. 86, no. 8, pp. 735–741, 2016.
- [94] B. Seifert, R. Eckenstaler, R. Rönicke et al., "Amyloid-beta induced changes in vesicular transport of BDNF in hippocampal neurons," *Neural Plasticity*, vol. 2016, Article ID 4145708, 15 pages, 2016.
- [95] C. Meng, Z. He, and D. Xing, "Low-level laser therapy rescues dendrite atrophy via upregulating BDNF expression: implications for Alzheimer's disease," *The Journal of Neuroscience*, vol. 33, no. 33, pp. 13505–13517, 2013.
- [96] K. Fukumoto, H. Mizoguchi, H. Takeuchi et al., "Fingolimod increases brain-derived neurotrophic factor levels and ameliorates amyloid β -induced memory impairment," *Behavioural Brain Research*, vol. 268, pp. 88–93, 2014.
- [97] M. Pedraz, A. I. Martín-Velasco, N. García-Marchena et al., "Plasma concentrations of BDNF and IGF-1 in abstinent cocaine users with high prevalence of substance use disorders: relationship to psychiatric comorbidity," *PLoS ONE*, vol. 10, no. 3, Article ID e0118610, 2015.
- [98] D.-D. Wang, T. Tian, Q. Dong et al., "Transcriptome profiling analysis of the mechanisms underlying the BDNF Val66Met polymorphism induced dysfunctions of the central nervous system," *Hippocampus*, vol. 24, no. 1, pp. 65–78, 2014.
- [99] N. Pitsikas, "The role of nitric oxide in the object recognition memory," *Behavioural Brain Research*, vol. 285, pp. 200–207, 2015.
- [100] X.-J. Zhu, Y.-F. Song, Q.-Y. Zhang, Y.-P. Cao, W.-Q. Xu, and D.-H. Su, "Effects of fractalkine on the expression of inflammatory substances in LPS-activated microglia cells," *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*, vol. 27, no. 12, pp. 1298–1300, 2011.
- [101] Q. Zhihui, "Modulating nitric oxide signaling in the CNS for Alzheimer's disease therapy," *Future Medicinal Chemistry*, vol. 5, no. 12, pp. 1451–1468, 2013.
- [102] S. Chakroborty, J. Kim, C. Schneider, A. R. West, and G. E. Stutzmann, "Nitric oxide signaling is recruited as a compensatory mechanism for sustaining synaptic plasticity in Alzheimer's disease mice," *Journal of Neuroscience*, vol. 35, no. 17, pp. 6893–6902, 2015.
- [103] A. Maher, N. S.-E. El-Sayed, H.-G. Breitingner, and M. Z. Gad, "Overexpression of NMDAR2B in an inflammatory model of Alzheimer's disease: modulation by NOS inhibitors," *Brain Research Bulletin*, vol. 109, pp. 109–116, 2014.
- [104] S. A. Austin, A. V. Santhanam, D. J. Hinton, D.-S. Choi, and Z. S. Katusic, "Endothelial nitric oxide deficiency promotes Alzheimer's disease pathology," *Journal of Neurochemistry*, vol. 127, no. 5, pp. 691–700, 2013.
- [105] C. Limatola, C. Lauro, M. Catalano et al., "Chemokine CX3CL1 protects rat hippocampal neurons against glutamate-mediated excitotoxicity," *Journal of Neuroimmunology*, vol. 166, no. 1-2, pp. 19–28, 2005.
- [106] J. L. Marín-Teva, I. Dusart, C. Colin, A. Gervais, N. Van Rooijen, and M. Mallat, "Microglia promote the death of developing Purkinje cells," *Neuron*, vol. 41, no. 4, pp. 535–547, 2004.
- [107] C. M. Tyler and L. M. Boulanger, "Complement-mediated microglial clearance of developing retinal ganglion cell axons," *Neuron*, vol. 74, no. 4, pp. 597–599, 2012.
- [108] S. Wakselman, C. Béchade, A. Roumier, D. Bernard, A. Triller, and A. Bessis, "Developmental neuronal death in hippocampus

- requires the microglial CD11b integrin and DAP12 immunoreceptor," *The Journal of Neuroscience*, vol. 28, no. 32, pp. 8138–8143, 2008.
- [109] D. Ragozzino, S. Di Angelantonio, F. Trettel et al., "Chemokine fractalkine/CX3CL1 negatively modulates active glutamatergic synapses in rat hippocampal neurons," *Journal of Neuroscience*, vol. 26, no. 41, pp. 10488–10498, 2006.
- [110] P. Saravanaraman, R. K. Chinnadurai, and R. Boopathy, "Why calcium channel blockers could be an elite choice in the treatment of Alzheimer's disease: A comprehensive review of evidences," *Reviews in the Neurosciences*, vol. 25, no. 2, pp. 231–246, 2014.
- [111] D. Zádori, G. Veres, L. Szalárdy, P. Klivényi, J. Toldi, and L. Vécsei, "Glutamatergic dysfunctioning in Alzheimer's disease and related therapeutic targets," *Journal of Alzheimer's Disease*, vol. 42, pp. S177–S187, 2014.
- [112] M. Catalano, C. Lauro, R. Cipriani et al., "CX3CL1 protects neurons against excitotoxicity enhancing GLT-1 activity on astrocytes," *Journal of Neuroimmunology*, vol. 263, no. 1-2, pp. 75–82, 2013.
- [113] S. Giunta, V. Andriolo, and A. Castorina, "Dual blockade of the A1 and A_{2A} adenosine receptor prevents amyloid beta toxicity in neuroblastoma cells exposed to aluminum chloride," *International Journal of Biochemistry and Cell Biology*, vol. 54, pp. 122–136, 2014.
- [114] C. Lauro, R. Cipriani, M. Catalano et al., "Adenosine A1 receptors and microglial cells mediate CX3CL1-induced protection of hippocampal neurons against glu-induced death," *Neuropsychopharmacology*, vol. 35, no. 7, pp. 1550–1559, 2010.
- [115] Z. Fan, A. A. Okello, D. J. Brooks, and P. Edison, "Longitudinal influence of microglial activation and amyloid on neuronal function in Alzheimer's disease," *Brain*, vol. 138, no. 12, pp. 3685–3698, 2015.
- [116] J. S. Rao, M. Kellom, H.-W. Kim, S. I. Rapoport, and E. A. Reese, "Neuroinflammation and synaptic loss," *Neurochemical Research*, vol. 37, no. 5, pp. 903–910, 2012.
- [117] R. Shah, G. J. Matthews, R. Y. Shah et al., "Serum fractalkine (CX3CL1) and cardiovascular outcomes and diabetes: findings from the chronic renal insufficiency cohort (CRIC) study," *American Journal of Kidney Diseases*, vol. 66, no. 2, pp. 266–273, 2015.
- [118] X. Zhang, X. Feng, W. Cai et al., "Chemokine CX3CL1 and its receptor CX3CR1 are associated with human atherosclerotic lesion vulnerability," *Thrombosis Research*, vol. 135, no. 6, pp. 1147–1153, 2015.
- [119] E. Ferretti, V. Pistoia, and A. Corcione, "Role of fractalkine/CX3CL1 and its receptor in the pathogenesis of inflammatory and malignant diseases with emphasis on B cell malignancies," *Mediators of Inflammation*, vol. 2014, Article ID 480941, 10 pages, 2014.
- [120] B. Jones, A. E. Koch, and S. Ahmed, "Pathological role of fractalkine/CX3CL1 in rheumatic diseases: a unique chemokine with multiple functions," *Frontiers in Immunology*, vol. 2, article 82, 2012.
- [121] L. W. Huo, Y. L. Ye, G. W. Wang, and Y. G. Ye, "Fractalkine (CX3CL1): a biomarker reflecting symptomatic severity in patients with knee osteoarthritis," *Journal of Investigative Medicine*, vol. 63, no. 4, pp. 626–631, 2015.
- [122] E. Astorri, R. Scrivo, M. Bombardieri et al., "CX3CL1 and CX3CR1 expression in tertiary lymphoid structures in salivary gland infiltrates: fractalkine contribution to lymphoid neogenesis in Sjögren's syndrome," *Rheumatology*, vol. 53, no. 4, pp. 611–620, 2014.
- [123] U. Flierl, J. Bauersachs, and A. Schafer, "Modulation of platelet and monocyte function by the chemokine fractalkine (CX3CL1) in cardiovascular disease," *European Journal of Clinical Investigation*, vol. 45, no. 6, pp. 624–633, 2015.
- [124] S. Apostolakis and D. Spandidos, "Chemokines and atherosclerosis: focus on the CX3CL1/CX3CR1 pathway," *Acta Pharmacologica Sinica*, vol. 34, no. 10, pp. 1251–1256, 2013.
- [125] H. Zhang, C. Guo, D. Wu et al., "Hydrogen sulfide inhibits the development of atherosclerosis with suppressing CX3CR1 and CX3CL1 expression," *PLoS ONE*, vol. 7, no. 7, Article ID e41147, 2012.
- [126] S. Martinez-Hervas, A. Vinue, L. Nunez et al., "Insulin resistance aggravates atherosclerosis by reducing vascular smooth muscle cell survival and increasing CX3CL1/CX3CR1 axis," *Cardiovascular Research*, vol. 103, no. 2, pp. 324–336, 2014.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

