Supplementary materials

yeast culture media and solutions	composition	supplier	notes
YPD	deionized water		autoclaving at 121°C for 20'
	10g/L yeast extract	sigma	
	20g/L peptone	sigma	
	20g/L agar (only for semisolid medium)	sigma	
	2% D-glucose		freshly added after autoclaving
	100U/ml penicillin and 100 µg/ml streptomicin	gibco	
selective medium YNB	deionized water		autoclaving at 121°C for 20'
	6.8 g/L yeast nitrogen base	sigma	
	20g/L agar (only for semisolid medium)	sigma	
	2% D-glucose		freshly added after autoclaving
	1X yeast synthetic drop out without tryptophan		
	100U/ml penicillin and100 µg/ml streptomicin	gibco	
induction medium	deionized water		autoclaving at 121°C for 20'
	6.8 g/L yeast nitrogen base	sigma	
	0.05% D-glucose		freshly added after autoclaving
	2% D-galactose		
	1X yeast synthetic drop out without tryptophan		
	100U/ml penicillin and100 µg/ml streptomicin	gibco	
20% D-glucose stock solution	deionized water		0.22 µm filtration
	200g/L D-glucose	sigma	
20% D-galactose stock solution	deionized water		0.22 µm filtration
	200g/L D-galactose	sigma	
10X yeast synthetic drop out without tryptophan stock solution	deionized water		0.22 µm filtration
	19.2g/L yeast synthetic drop out without tryptophan	sigma	

Figure S1: List and recipes of media and solutions used for yeast cultures.

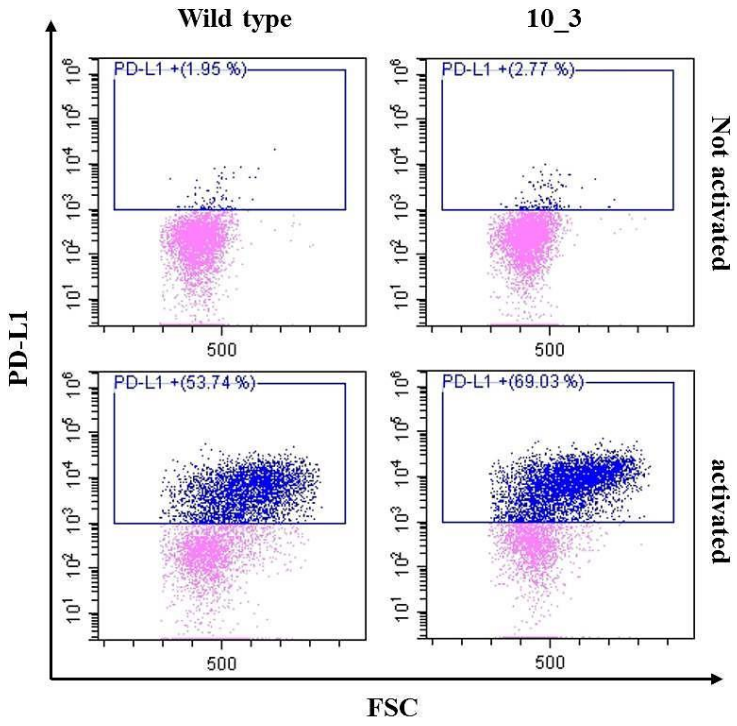

Figure S2: Validation of binding specificity of mAb 10_3 to activated hPBMCs. The panels show the percentages of not activated (upper panels) and activated (lower panels) lymphocytes (CD2-positive cells) which bound to a saturating concentration of the wild type and affinity matured mAb 10_3 (blue cells). The affinity maturation did not change the specificity for PD-L1, as demonstrated by the ability of binding only to activated cells.