

## Research Article

# Effect of Selenium on Expression of Apoptosis-Related Genes in Cryomedia of Mice Ovary after Vitrification

Reza Nori-Garavand,<sup>1,2</sup> Maryam Hormozi,<sup>2,3</sup> Leila Narimani,<sup>1,2</sup> Nasim Beigi Boroujeni,<sup>2</sup> Asghar Rajabzadeh,<sup>1,2</sup> Leila Zarei,<sup>1,2</sup> Masoud Beigi Boroujeni,<sup>4</sup> and Mandana Beigi Boroujeni <sup>1,2</sup>

<sup>1</sup>Department of Anatomical Sciences, Lorestan University of Medical Sciences, Khorramabad, Iran

<sup>2</sup>Razi Herbal Medicines Research Center, Lorestan University of Medical Sciences, Khorramabad, Iran

<sup>3</sup>Department of Biochemistry, Lorestan University of Medical Sciences, Khorramabad, Iran

<sup>4</sup>Department of Biochemistry, Branch of Tehran-Shargh, Payame Noor University, Tehran, Iran

Correspondence should be addressed to Mandana Beigi Boroujeni; [mandana.beigi@lums.ac.ir](mailto:mandana.beigi@lums.ac.ir)

Received 11 July 2020; Revised 15 August 2020; Accepted 14 September 2020; Published 23 September 2020

Academic Editor: Monica Fedele

Copyright © 2020 Reza Nori-Garavand et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Introduction.** Freezing of ovarian tissue is used for preservation of fertility. The freezing-thawing process is accompanied by oxidative stress and induction of apoptosis. Apoptosis is a complex process that has been studied in animal models. The present study was aimed at investigating the effect of selenium on suppression of apoptosis during vitrification-thawing process of mice ovary via studying expression of apoptosis-related genes, and also, we aimed to design statistical models for the roles of single genes and gene-gene interactions in suppression of apoptosis. **Methods.** A total of 10 right ovary samples from 10 mice were randomly divided into two groups of selenium treatment (at dose 5 µg/ml sodium selenite, through adding to the media) and control group. Vitrification-thawing process was done according to the existed protocols. Real-time PCR was used for gene expression study. The apoptosis gene profile included *P53*, *Bax*, *Fas*, and *Bcl-2*. General linear model was applied to study single gene associations and gene-gene interactions. **Results.** From the studied genes, *P53* showed a significant downregulation in the selenium group in comparison to the control group ( $\Delta\Delta CT = 1.96$ ;  $P = 0.013$ ; relative expression (RE) = 0.28). *Bcl-2* showed a significant upregulation in the selenium group in comparison to the control group ( $\Delta\Delta CT = -2.49$ ;  $P < 0.001$ ; RE = 3.49). No significant result was found for other genes. According to the multiple models, *Bcl-2* showed a protective single gene association (beta = -0.33;  $P = 0.032$ ), and *Fas* \* *Bcl-2* interaction was significantly positive (beta = 0.19;  $P = 0.036$ ). **Conclusion.** Addition of selenium to cryomedia of vitrification-thawing process could reduce the apoptosis induced by freezing-thawing stress in mice ovary via downregulation of *P53* and upregulation of *Bcl-2* at transcription level. Multivariable statistical models should be performed in future researches to study biological systems.

## 1. Introduction

**1.1. Background.** Some women need to preserve their fertility for their future life. The most important reason for the necessity of fertility preservation is cancer [1]. During chemotherapy and radiotherapy, ovaries may be affected by chemical and physical cytotoxic agents [2]. To solve this problem, freezing of ovarian samples was suggested [3]. Other than ovarian samples, sperms and spermatogonial stem cells are also frozen. There are usually two freezing methods consist

of slow freezing (conventional cryopreservation) and rapid freezing (vitrification or rapid cryopreservation). Freezing-thawing process is associated with oxidative stress and cell damage. Therefore, using additive antioxidants in cellular or tissue preservation media has been suggested [4].

Selenium is a biological trace element plays a key role in human metabolism. Selenium is used for synthesis of selenoproteins in the liver. This element performs its function as a component of antioxidant enzymes [5]. Other than nutritional values, selenium has an important role in human

immune system [6]. Because of the antioxidant effects of selenium, it is widely used in the studies of the animal models of oxidative stress [7].

Apoptosis is a kind of programmed cell death which is usually a physiological phenomenon. However, if the micro-environment of the cells is not suitable, a pathologic apoptosis may occur [8]. Freezing-thawing damage is an example of this unsuitable environmental condition [9]. So far, many animal studies have been designed to model induction of apoptosis. For instance, chemotherapeutic agents are used for induction of apoptosis in animal models [10]. In a rabbit osteoarthritis model, transection of anterior cruciate ligament was applied for induction of apoptosis in chondrocytes [11]. Other than *in vivo* studies, animal cells and tissues are used *in vitro* for apoptosis modeling. Freezing-thawing process is a common method for induction of apoptosis in a practical condition [12, 13]. Nevertheless, the exact mechanism of apoptosis induction and apoptosis suppression by antioxidants is not completely understood.

**1.2. Objectives.** Since there was a gap in evidence on the role of supplementation of nutritional elements to cryomedia of ovarian freezing-thawing process, this study was aimed at evaluating the role of selenium supplementation on reduction of apoptosis after vitrification of mice ovarian tissue through study of apoptosis-related genes including *P53*, *Bax*, *Bcl-2*, and *Fas*. In addition, a statistical modeling was designed to study the roles of single genes and gene-gene interactions in suppression of apoptosis. Our aims were both basic and practical.

## 2. Material and Methods

**2.1. Study Design.** An experimental study was conducted with *in vitro* design. This study consisted of cellular media processing and gene expression assay.

**2.2. Animals and Study Groups.** A total of 10 balb/c mice aged 6-8 weeks were bought from an animal laboratory of Lorestan University of Medical Sciences, Khorramabad, Lorestan Province, West of Iran. After approving the study protocol by the ethics committee of Lorestan University of Medical Sciences (registration number: IR.LUMS.REC.1397.195), the animals were imported to the study according to ethical guidelines for working with laboratory animal. All the animals were kept at standard condition of light, temperature, moisture, and nutrition. Pseudopregnancy was induced using a vaginal swab. The animals were sacrificed with cervical dislocation. The corpses were kept in supine position and fixed, and the abdominal hair was shaved. In a sterile condition, midline excision was conducted from xiphoid to pubis, and then, peritoneum was excised, and abdominal viscera were removed. The right ovaries were removed by surgical pans and knife for the study. The extra tissues around the ovaries were removed under a stereomicroscope. The ovarian particles were kept in Dulbecco's modified eagle medium (DMEM) culture environment with 10% fetal bovine serum (FBS). The samples were randomly divided into two groups: selenium group and control group.

**2.3. Sample Processing.** Ethylene glycol ficoll sucrose (EGFS) media was used for vitrification process according to a protocol mentioned by Chang et al. [14]. EGFS solution was prepared in a DMEM solvent containing of 40% ethylene glycol, 30% ficoll 70, and 0.5 mole sucrose. For the selenium group, 5  $\mu\text{g}/\text{ml}$  sodium selenite was added to EGFS solution [15]. Then, the EGFS was filtered with a 0.45  $\mu\text{m}$  millipore filter and kept at  $-20^{\circ}\text{C}$ . After preparation of the solutions EGFS and EGFS+Se, the ovarian samples were added to the solutions, and after dehydration, they were put in the cryovials. The dehydration process took about 5 minutes. The cryovials were put on nitrogen vapor for 30 seconds, and then, they were put at temperature  $-196^{\circ}\text{C}$  in liquid nitrogen. For thawing process (rapid method), serial concentrations of sucrose solution were used as 1, 0.5, and 0.25 mole per one liter of DMEM, and the forth step was pure DMEM. The hydration process took about 5 minutes. After hydration process, the ovarian samples were kept in DMEM with 10% FBS, under  $\text{CO}_2$  incubator at temperature  $37^{\circ}\text{C}$  for 24 hours. During this period, the samples were washed for some times. Finally, the ovarian samples were subjected for RNA extraction and gene expression study.

**2.4. Molecular Assay.** Total RNA was extracted using a column-based RNA extraction kit (Yektatajhez Azma, Iran). About 20-30 mg of each ovarian tissue was used. The extracted RNA was preserved in an RNase free elution solution. A NanoDrop spectrophotometer was used to read the concentrations. Then, the extracted RNA was kept at  $-70^{\circ}\text{C}$ . cDNA was synthesized with oligo dT primers using cDNA synthesis kit (Sina Clone, Iran). Primers were designed with help of NCBI database (Table 1). GAPDH was used as internal control. Gene expression was studied via real-time polymerase chain reaction (real-time PCR). SYBR green master mix was used for amplification of the genes. The amplification process was observed and documented in Rotor Gene program. Each gene of each sample was run for 3 times, and then, the best results were chosen for analysis according to the melting curves and reaction efficiencies.

**2.5. Data Analysis.** Amounts of CTs were calculated based on takeoff and amplification cycles at the exponential phase. Reaction efficiencies were obtained from REST2009 program.  $\Delta\text{CT}$  amounts were calculated via CT of target gene minus CT of reference gene.  $\Delta\Delta\text{CT}$  amounts were also calculated in which one  $\Delta\text{CT}$  was for the treated group and the other  $\Delta\text{CT}$  was for the control group. Relative expression (RE) was calculated with Pfaffl's efficiency calibrated method in which each  $\Delta\text{CT}$  was defined as CT of sample minus CT of control for each gene (equation (1)) [16]. RE was also calculated for comparison of apoptosis stimulating genes with *Bcl-2* apoptosis inhibitory gene. For statistical analysis, independent *t*-test was used for comparison of  $-\Delta\text{CT}$ s between the groups, and one sample *t*-test was used for comparison of  $-\Delta\Delta\text{CT}$ s with zero (as the null hypothesis). In order to adjust the associations of the target genes and their interactions with *Bcl-2* gene, a general linear model was used, and beta coefficient with 95% confidence interval (CI) was reported. All the analyses were done before computing RE.

TABLE 1: Primer sequences for amplification of cDNA.

Gene	Primer sequence (5'-3')	Product length (bp)
GAPDH	Forward: CAATGTGTCCGTCGTGGATCT Reverse: GTCCTCAGTGTAGCCCAAGATG	208
P53	Forward: GTTCCTCTTGGGCTTAGGG Reverse: CTTCTGTACGGCGGTCTCTC	255
Bax	Forward: CGAGCTGATCAGAACCATCA Reverse: GAAAAATGCCTTCCCTTC	277
Fas	Forward: GAGAATTGCTGAAGACATGACAATCC Reverse: GTAGTTTTCACTCCAGACATTGTCC	314
Bcl-2	Forward: TAAGCTGTCACAGAGGGGCT Reverse: TGAAGAGTTCCTCCACCACC	344

Heat map was used to cluster the individual sample data. All the mathematical and statistical process was performed in the R 3.6.3 software (R foundation for statistical computing, Austria) [17].

$$RE = \frac{(E_{\text{target}})^{-\Delta CT_{\text{(sample-control)}}}}{(E_{\text{reference}})^{-\Delta CT_{\text{(sample-control)}}}}, \quad (1)$$

where  $E = 2^{\text{Efficiency (\%)}}$ .

### 3. Results

After extraction of RNA, the concentration of RNA was obtained at range 69.60-79.40 ng/ $\mu$ l. Using 1  $\mu$ l of cDNA and 1  $\mu$ l of 10 pmol primers resulted in successful amplification with one melting peak at a suitable temperature. From the repetitive reactions, those with better reaction efficiencies were selected. REs were calculated based on  $\Delta CT_{\text{(sample-control)}}$  amounts adjusted with the efficiencies while statistical analyses (and calculation of P values) were done based on  $\Delta CT_{\text{(target-reference)}}$  and  $\Delta\Delta CT$  amounts (Tables 2 and 3). Our sample size had about 79% power to detect at least two amplification cycle differences with standard deviation of one cycle at the point of 5% type one error.

Among the four target genes, *P53* showed a significant downregulation ( $P = 0.013$ ;  $RE = 0.28$ ), and *Bcl-2* showed a significant upregulation ( $P < 0.001$ ;  $RE = 3.49$ ) in the treated group. The results of *Bax* and *Fas* were nonsignificant and inconclusive (Table 2) (Figures 1 and 2). In order to assay overall status of apoptosis with a gene expression approach, expression of target genes per *Bcl-2* was studied. All the target genes showed downregulation in comparison to *Bcl-2* antiapoptotic gene that the most downregulation was for *P53* ( $RE = 0.08$ ) followed by *Bax* ( $RE = 0.25$ ) and *Fas* ( $RE = 0.30$ ) (Table 3) (Figure 3). Clustering of the individual samples showed similar results in the samples, except for sample 5 of the selenium group which was clustered in the cluster of the control group. Clustering of the genes showed that in condition of apoptosis, *Bcl-2* had more expression followed by *P53*, *Fas*, and *Bax*, of course without considering better efficiency of *P53* in our study (Figure 4).

The general linear model was used to reach a per se association of each gene without the confounding effect of other

genes. Accordingly, each unit of increase in  $\Delta CT$  (i.e., down-regulation up to two folds) for *Bcl-2* was independently associated with about 33% reduction in the probability of being in the selenium-treated group ( $\beta = -0.33$ ;  $P = 0.032$ ). No significant association was found for other genes (Table 4, model 1) (Figure 5, model 1). Since *Bcl-2* had a better per se association and also was the only antiapoptotic gene, its interactions with the other target genes were also imported to the model. Accordingly, *Fas* \* *Bcl-2* interaction was positive ( $\beta = 0.19$ ;  $P = 0.036$ ). No significant result was obtained for interactions *Bax* \* *Bcl-2* and *P53* \* *Bcl-2*. Interestingly, among the single genes, the association of *Fas* became significant ( $\beta = -0.81$ ;  $P = 0.045$ ), and *Bcl-2* lost its significance (Table 4, model 2) (Figure 5, model 2). Further addition of other possible interactions to the model resulted in loss of significance or software error due to low number of observations (data not shown).

### 4. Discussion

**4.1. Interpretation.** The aim of this study was to investigate mRNA expression profile of apoptosis-related genes in an animal tissue model of apoptosis induction by vitrification. This study consisted of the profile response to selenium administration as an antioxidant and investigation of gene-gene interactions in the mentioned profile. The practical aim was to reach a model for fertility preservation in conditions of different human pathologies such as cancers in young women.

Among the investigated genes, changes in the expression of *P53* and *Bcl-2* were statistically significant. Other than statistical analysis based on the number of PCR cycles (i.e.,  $\Delta CT$  and  $\Delta\Delta CT$ ), efficiency calibrated method approved the associations of *P53* and *Bcl-2* (95% CIs of their REs had a remarkable distance from  $RE = 1$  as a null hypothesis). In addition, proportions of *P53/Bcl-2*, *Bax/Bcl-2*, and *Fas/Bcl-2* mRNA expression were downregulated in the treated group as a manifestation of the general status of apoptosis at transcription level. It demonstrated that in biological systems, proportions of the levels of biomarkers might be more important than the pure levels to perform their biological functions.

Multivariable and multivariate analyses are a way for a better discovery and interpretation of biological systems. According to the heat map, the individual samples were

TABLE 2: Expressions of the target genes (selenium group vs. control group).

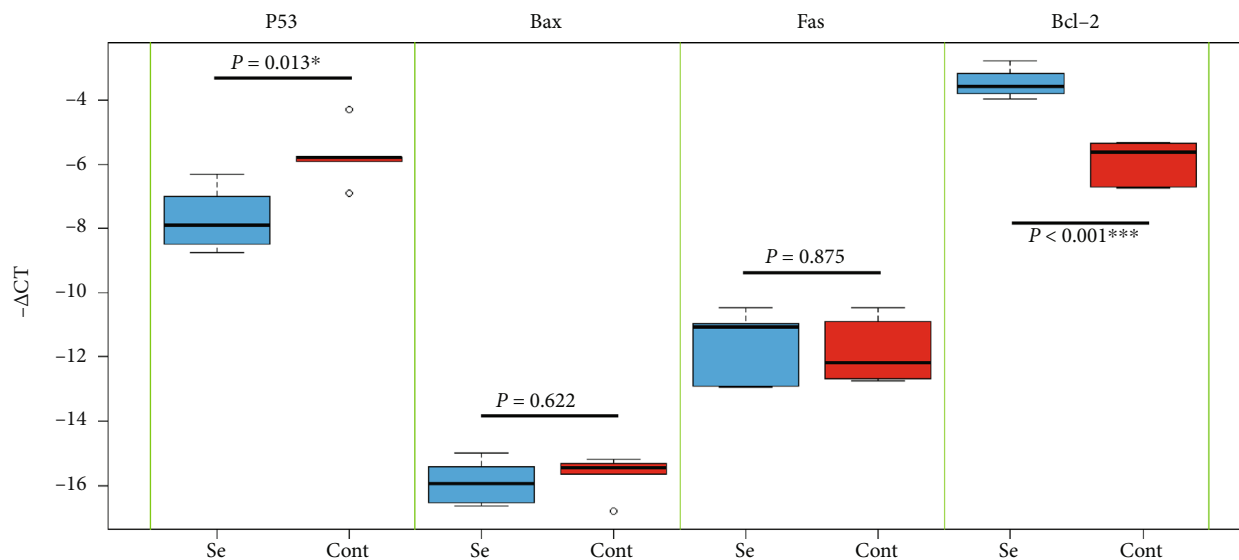
Gene	$\Delta\Delta CT$ (95% CI)	One sample $P$	Two sample $P$	Efficiency	RE (95% CI)
GAPDH	Reference			0.702	
P53	1.96 (0.69, 3.22)	0.013*	0.013*	0.921	0.28 (0.16, 0.47)
Bax	0.22 (-0.66, 1.11)	0.530	0.622	0.728	0.88 (0.78, 1.00)
Fas	-0.11 (-1.56, 1.33)	0.838	0.875	0.707	1.06 (0.68, 1.63)
Bcl-2	-2.49 (-3.09, -1.89)	<0.001***	<0.001***	0.734	3.49 (3.22, 3.78)

\*Significant at 0.05. \*\*\*Significant at 0.001. CIs of RE were computed using RE formula on lower and upper bounds of  $\Delta CT$  (sample-control for each gene separately).

TABLE 3: Expression of the target genes/Bcl-2 (selenium group vs. control group).

Gene	$\Delta\Delta CT$ (95% CI)	One sample $P$	Efficiency	RE (95% CI)
GAPDH	Reference		0.702	
Bcl-2	Reference		0.734	
P53	4.45 (2.78, 6.11)	0.002**	0.921	0.08 (0.05, 0.12)
Bax	2.71 (1.97, 3.44)	<0.001***	0.728	0.25 (0.24, 0.26)
Fas	2.37 (0.61, 4.14)	0.020*	0.707	0.30 (0.21, 0.43)

\*Significant at 0.05. \*\*Significant at 0.01. \*\*\*Significant at 0.001.

FIGURE 1: Gene expression comparison in selenium (Se) and control (Cont) groups (independent  $t$ -test).

clustered in their real groups except one sample of the selenium group that was clustered as the final branch of the control group. The clustering of the genes demonstrated that even in condition of apoptosis *Bcl-2* had further expression and in response to antiapoptotic agents its interval with apoptotic genes is increased. According to the general linear models, only *Bcl-2* had an independent association with the groups of study (in favor of being in the control group, i.e., antiapoptotic effect). After adjusting the model with the interactions of *Bcl-2*, the only significant associations were for *Fas* (in favor of being in the control group, i.e., antiapoptotic effect) and *Fas \* Bcl-2* interaction (positive interaction, i.e., they help each other for upregulation). It showed that

either *Fas* had a secret effect on upregulation of *Bcl-2*, or its secret upregulation was due to a compensatory effect in response to *Bcl-2* upregulation. It was worth noting that the interaction *P53 \* Bcl-2* did not have significant interaction in spite of their significant ratio of expression.

*P53* is a tumor suppressor gene involved in both intrinsic and extrinsic pathways of apoptosis. Therefore, downregulation of *P53* is associated with reduction in apoptosis [18, 19]. *Bcl-2* is an antiapoptotic protein, and therefore, upregulation of *Bcl-2* is associated with suppression of apoptosis [20]. Proportion of *Bax/Bcl-2* is a widely used marker to study apoptosis. Downregulation of this proportion at transcriptome level may indicate downregulation of apoptosis. Das et al. (2007)

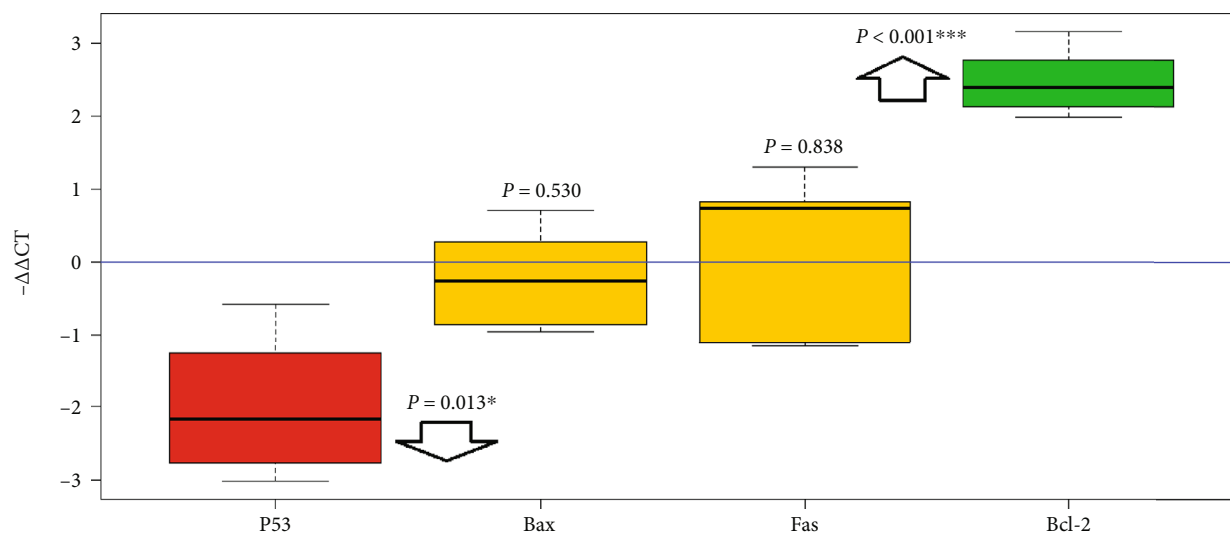


FIGURE 2: Gene expression of the target genes if the treated group calibrated with the control group (one sample *t*-test).

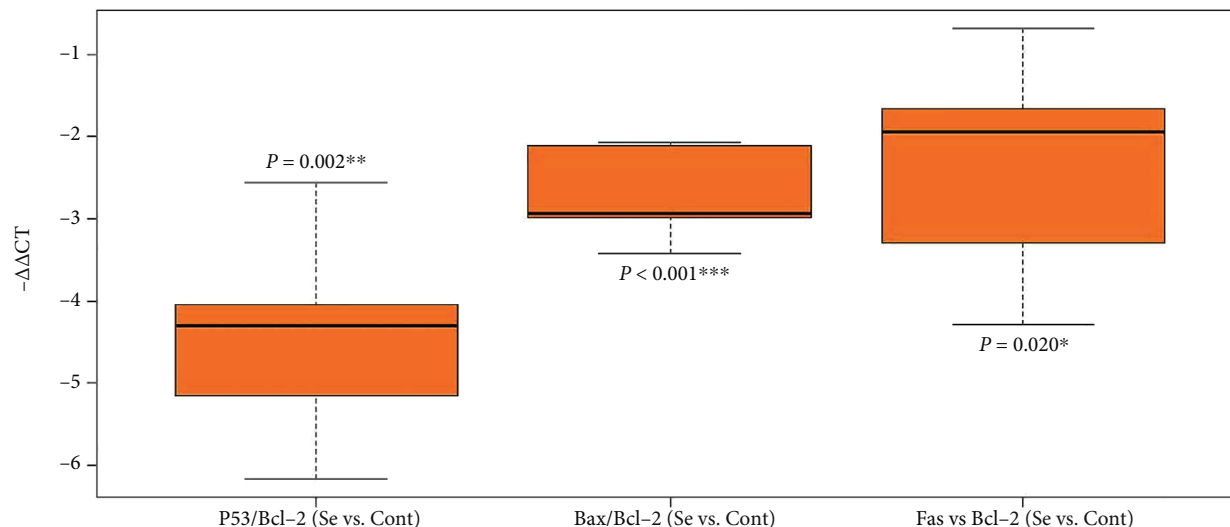


FIGURE 3: Gene expression of the target genes per expression Bcl-2 in the treated group calibrated with the control group (one sample *t*-test).

and Ghiasi et al. (2016) used this ratio via real-time PCR to detect apoptosis [21, 22]. However, in our study, the mRNA ratio *P53/Bcl-2* was more dominant, and the significant interaction was for *Fas \* Bcl-2*.

In general, our study showed an antiapoptotic effect for selenium. The antiapoptotic effect of selenium may be resulted from its antioxidant effects [23]. A controversial hypothesis is that anticancer effect of selenium is through induction of apoptosis [6]. In fact, selenium participates in DNA repair, and it is resulted from helping antioxidant enzymes [24].

**4.2. Previous Literature.** Agrawal et al. (2006) reported that during freezing-thawing process reactive oxygen species (ROS) were increased and induced apoptosis [25]. It has been shown that the effect of ROS on induction of apoptosis is mainly through activation of *P53* [26]. Since in the present study *P53* was downregulated, it seems that protective effect

of selenium may be due to its antioxidant effects. Rimón et al. (2005) believed that apoptosis is the final response of cell to inappropriate conditions, and therefore, apoptosis study is a suitable tool in stressful conditions of freezing protocols [27].

A lot of researches have been performed on freezing media of ovary and sperm samples. Slaweta et al. (1988) had studied the relationship between selenium content of bull semen with sperm motility after freezing-thawing damage. They found that lower selenium content was associated with lower sperm motility [28]. Dorostkar et al. (2012) studied the effect of selenium additive on sperm characteristics in water buffaloes before and after freezing, and they reached a better sperm quality [29]. Rezaeian et al. (2016) studied the effects of selenium (at dose 5 μg/ml) on human sperm parameters after freezing-thawing process. They found a better sperm quality and suggested selenium supplementation for use in laboratories of infertility clinics [15]. Although

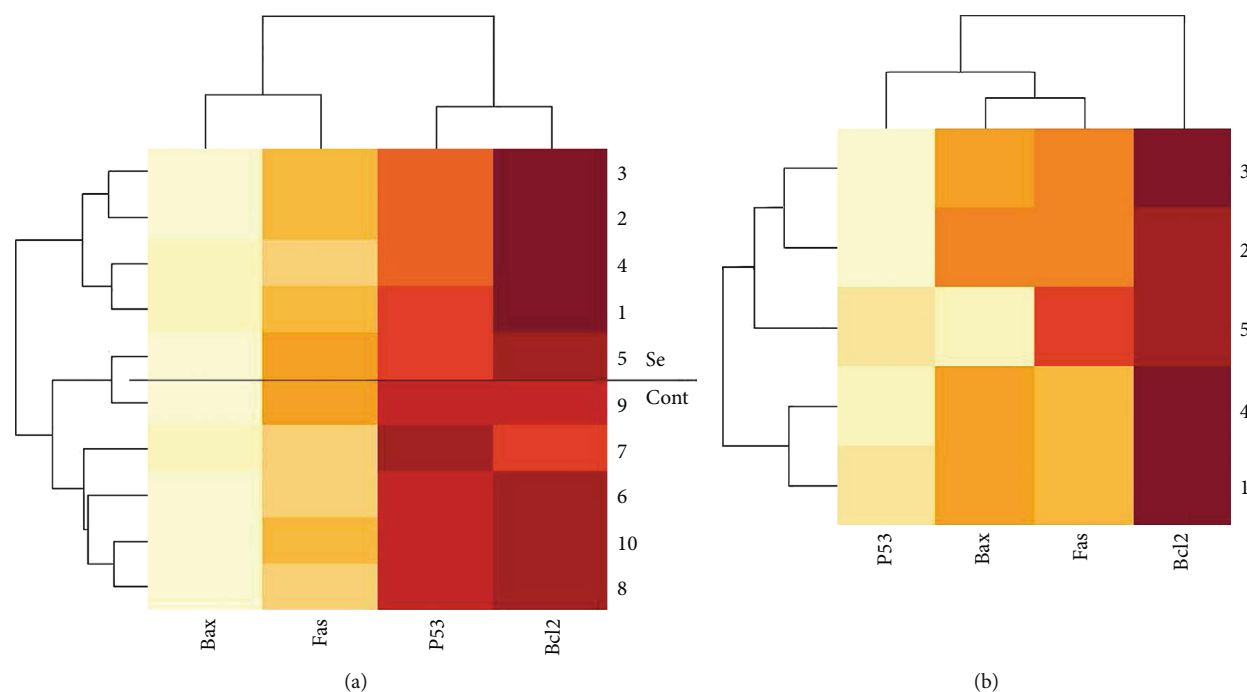


FIGURE 4: Clustering gene expression in individual samples. (a)  $-\Delta CTs$  in selenium (Se) and control (Cont) groups. (b)  $-\Delta\Delta CTs$  in each treated sample.

TABLE 4: General linear model for multiple adjustment of the associations of the genes with being the treated group.

Model/covariate	Beta coefficient (95%, CI)	Wald test <i>P</i>	Residual SE	<i>R</i> -squared
<b>Model 1</b>				
P53	0.01 (-0.28, 0.31)	0.913		
Bax	0.06 (-0.29, 0.42)	0.672		
Fas	0.06 (-0.20, 0.31)	0.598	0.268	0.856
Bcl-2	-0.33 (-0.62, -0.04)	0.032*		
Intercept	0.32 (-6.37, 7.01)			
<b>Model 2</b>				
P53	0.10 (-0.60, 0.79)	0.601		
Bax	0.64 (-0.92, 2.21)	0.218		
Fas	-0.81 (-1.58, -0.05)	0.045*		
Bcl-2	-0.91 (-6.80, 4.96)	0.570	0.111	0.990
Fas*Bcl-2	0.19 (0.03, 0.34)	0.036*		
P53*Bcl-2	-0.01 (-0.14, 0.11)	0.646		
Bax*Bcl-2	-0.10 (-0.45, 0.25)	0.342		
Intercept	1.10 (-23.37, 25.57)			

\*Significant at 0.05 (it is also the interaction sign in the first column). SE: standard error.

there were many studies on the role of selenium in male reproduction system, there was not a study on the role of selenium supplementation in freezing-thawing process of ovary samples via studying apoptosis-related genes. It was the novelty of our study in comparison to the literature. Bozkurt et al. (2012) had found a protective effect for selenium on ischemia reperfusion injury in rat ovary via studying oxidative stress indices [30]. Brito et al. (2013) showed protective effects of selenium in combination with vitamin E on

ovarian samples of monkey via investigating expression of superoxide dismutase 1 gene [31]. In general, protective effect of selenium has been observed in many studies like our study.

**4.3. Limitations.** Lack of study at proteomic level was the most important limitation of our study. Also, frozen oocytes should be studied in future. Although our sample size had an approximately suitable power for univariable analyses like



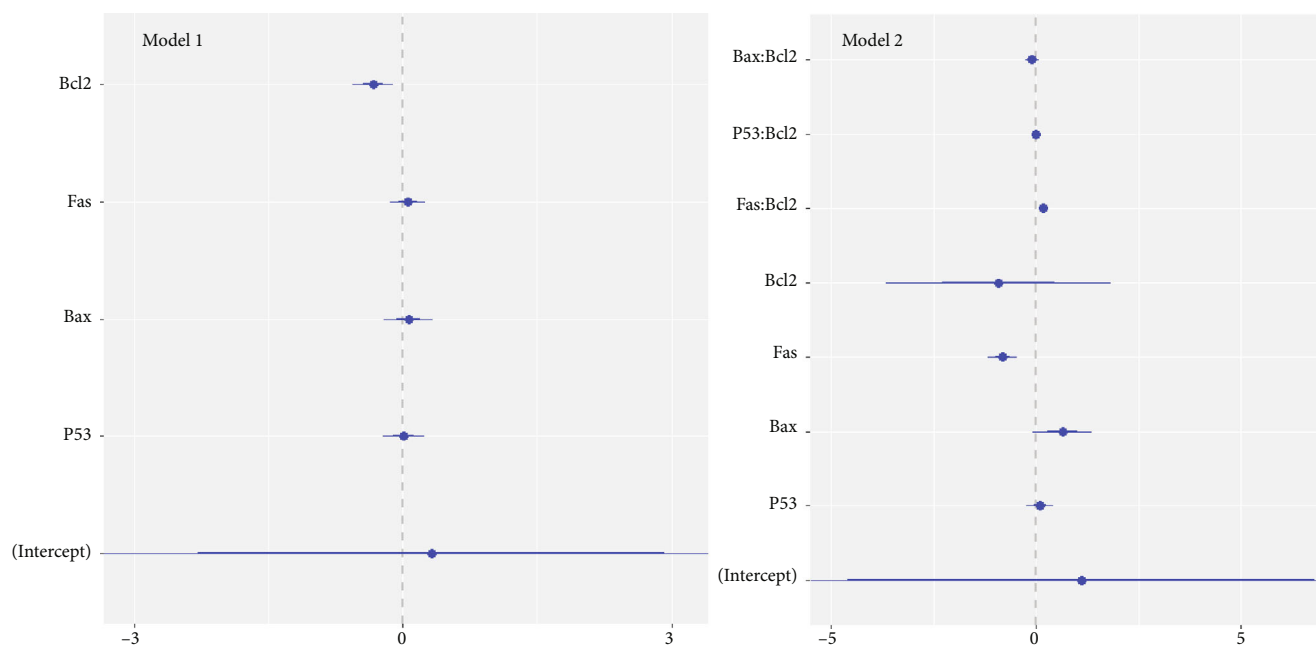


FIGURE 5: Coefficient plot of general linear model for  $+\Delta CTs$  of the genes. Model 1: multiple adjusted model. Model 2: multiple adjusted model with interaction of target genes with Bcl-2.

other animal studies, our multiple models needed further observations to study all the possible interactions with enough power. However, our models were fit (low residual standard error) for individual predictions.

## 5. Conclusion

Addition of selenium at dose  $5 \mu g/ml$  of sodium selenite to cryomeidia of vitrification-thawing process could reduce the apoptosis induced by freezing-thawing stress in mice ovary. This effect was thrown downregulation of P53 and upregulation of Bcl-2 at transcription level. According to the multiple models, Bcl-2 showed a protective single gene association, and Fas \* Bcl-2 interaction was significantly positive. This study can be performed on human ovary samples in the patients who are candidate for wedge sample or whole ovary freezing regarding ethical and legal issues. Multivariable statistical models should be performed in future researches to study biological systems.

## Data Availability

The data (raw amounts of CTs of the real-time PCR) used to support the findings of this study are available from the corresponding author upon request.

## Ethical Approval

The ethics committee of Lorestan University of Medical Sciences (# IR.LUMS.REC.1397.195) approved the protocol of the study. The animals were imported to the study according to ethical guidelines for working with laboratory animal.

## Conflicts of Interest

The authors declare no conflict of interest.

## Authors' Contributions

All the authors had equal contribution and approved the final manuscript.

## Acknowledgments

This study was MSc thesis of R Nori-Garavand under supervision of M Beigi Broujeni. We kindly acknowledge Razi Herbal Medicines Research Center of LUMS for giving the props of study.

## References

- [1] J. Donnez and M.-M. Dolmans, "Fertility preservation in women," *New England Journal of Medicine*, vol. 377, no. 17, pp. 1657–1665, 2017.
- [2] M. Lambertini, L. del Mastro, M. C. Pescio et al., "Cancer and fertility preservation: international recommendations from an expert meeting," *BMC Medicine*, vol. 14, no. 1, p. 1, 2016.
- [3] S. Silber, "Ovarian tissue cryopreservation and transplantation: scientific implications," *Journal of Assisted Reproduction and Genetics*, vol. 33, no. 12, pp. 1595–1603, 2016.
- [4] M. Gholami, S. A. Y. Ahmadi, A. Abaszadeh, and A. Khaki, "Protective effects of melatonin and ghrelin on spermatogenesis: a narrative review of the literature," *International Journal of Reproductive Biomedicine*, vol. 15, no. 5, pp. 265–272, 2017.
- [5] R. F. Burk, "Selenium, an antioxidant nutrient," *Nutrition in Clinical Care*, vol. 5, no. 2, pp. 75–79, 2002.
- [6] J. C. Avery and P. R. Hoffmann, "Selenium, selenoproteins, and immunity," *Nutrients*, vol. 10, no. 9, article 1203, 2018.

- [7] H. Ahmadvand, E. Babaenezhad, H. Nayeri, and Z. Zarei Nezhad, "Selenium effects on antioxidant and inflammatory indices in renal ischemia-reperfusion injury in rats," *Journal of Renal Injury Prevention*, vol. 8, no. 2, pp. 71–77, 2019.
- [8] D. R. Green, *Apoptosis: Physiology and Pathology*, Cambridge University Press, 2011.
- [9] N. Zribi, N. F. Chakroun, F. Ben Abdallah et al., "Effect of freezing–thawing process and quercetin on human sperm survival and DNA integrity," *Cryobiology*, vol. 65, no. 3, pp. 326–331, 2012.
- [10] K. J. Yagle, J. F. Eary, J. F. Tait et al., "Evaluation of 18F-annexin V as a PET imaging agent in an animal model of apoptosis," *Journal of Nuclear Medicine*, vol. 46, no. 4, pp. 658–666, 2005.
- [11] A. Kourtis, P. G. Adamopoulos, A. Papalois, D. C. Iliopoulos, G. C. Babis, and A. Scorilas, "Quantitative analysis and study of the mRNA expression levels of apoptotic genes BCL2, BAX and BCL2L12 in the articular cartilage of an animal model of osteoarthritis," *Annals of Translational Medicine*, vol. 6, no. 12, p. 243, 2018.
- [12] E. Labrune, P. Jaeger, C. Santamaria et al., "Cellular and molecular impact of vitrification versus slow freezing on ovarian tissue," *Tissue Engineering Part C: Methods*, vol. 26, no. 5, pp. 276–285, 2020.
- [13] M. B. Boroujeni, F. Peidayesh, A. Pirnia, N. B. Boroujeni, S. A. Y. Ahmadi, and M. Gholami, "Effect of selenium on freezing–thawing damage of mice spermatogonial stem cell: a model to preserve fertility in childhood cancers," *Stem Cell Investigation*, vol. 6, p. 36, 2019.
- [14] H. J. Chang, J. H. Moon, J. R. Lee, B. C. Jee, C. S. Suh, and S. H. Kim, "Optimal condition of vitrification method for cryopreservation of human ovarian cortical tissues," *Journal of Obstetrics and Gynaecology Research*, vol. 37, no. 8, pp. 1092–1101, 2011.
- [15] Z. Rezaeian, H. Yazdekhashti, S. Nasri, Z. Rajabi, P. Fallahi, and F. Amidi, "Effect of selenium on human sperm parameters after freezing and thawing procedures," *Asian Pacific Journal of Reproduction*, vol. 5, no. 6, pp. 462–466, 2016.
- [16] J. S. Yuan, A. Reed, F. Chen, and C. N. Stewart Jr., "Statistical analysis of real-time PCR data," *BMC Bioinformatics*, vol. 7, no. 1, 2006.
- [17] R Core Team, *R: a language and environment for statistical computing*, R Foundation for Statistical Computing, Vienna, Austria, 2020.
- [18] S. Krautwald, E. Ziegler, L. Rölver et al., "Effective blockage of both the extrinsic and intrinsic pathways of apoptosis in mice by TAT-crmA," *Journal of Biological Chemistry*, vol. 285, no. 26, pp. 19997–20005, 2010.
- [19] J. D. Amaral, J. M. Xavier, C. J. Steer, and C. M. Rodrigues, "The role of p53 in apoptosis," *Discovery Medicine*, vol. 9, no. 45, pp. 145–152, 2010.
- [20] K. Fuenzalida, R. Quintanilla, P. Ramos et al., "Peroxisome proliferator-activated receptor  $\gamma$  up-regulates the Bcl-2 anti-apoptotic protein in neurons and induces mitochondrial stabilization and protection against oxidative stress and apoptosis," *Journal of Biological Chemistry*, vol. 282, no. 51, pp. 37006–37015, 2007.
- [21] A. Das, N. L. Banik, and S. K. Ray, "Differentiation decreased telomerase activity in rat glioblastoma C6 cells and increased sensitivity to IFN- $\gamma$  and taxol for apoptosis," *Neurochemical Research*, vol. 32, no. 12, pp. 2167–2183, 2007.
- [22] P. Ghiasi, S. Hosseinkhani, H. Ansari, N. Aghdami, and H. Baharvand, "Comparison of BAX and Bcl-2 expression during human embryonic stem cell differentiation into cardiomyocytes and doxorubicin-induced apoptosis," *Biomacromolecular Journal*, vol. 2, no. 2, pp. 162–168, 2016.
- [23] A. Valadbeygi, T. Naji, A. Pirnia, and M. Gholami, "Supplementation freeze-thawed media with selenium protect adipose-derived mesenchymal stem cells from freeze-thawed induced injury," *Cryobiology*, vol. 73, no. 2, pp. 135–139, 2016.
- [24] S. Bera, V. D. Rosa, W. Rachidi, and A. M. Diamond, "Does a role for selenium in DNA damage repair explain apparent controversies in its use in chemoprevention?," *Mutagenesis*, vol. 28, no. 2, pp. 127–134, 2013.
- [25] A. Agarwal, S. Gupta, and S. Sikka, "The role of free radicals and antioxidants in reproduction," *Current Opinion in Obstetrics and Gynecology*, vol. 18, no. 3, pp. 325–332, 2006.
- [26] B. Y. Sheikh, M. M. R. Sarker, M. N. A. Kamarudin, and G. Mohan, "Antiproliferative and apoptosis inducing effects of citral via p53 and ROS-induced mitochondrial-mediated apoptosis in human colorectal HCT116 and HT29 cell lines," *Biomedicine & Pharmacotherapy*, vol. 96, pp. 834–846, 2017.
- [27] E. Rimón, T. Cohen, A. Dantes et al., "Apoptosis in cryopreserved human ovarian tissue obtained from cancer patients: a tool for evaluating cryopreservation utility," *International Journal of Oncology*, vol. 27, no. 2, pp. 345–353, 2005.
- [28] R. Slaweta, W. Wařowicz, and T. Laskowska, "Selenium content, glutathione peroxidase activity, and lipid peroxide level in fresh bull semen and its relationship to motility of spermatozoa after freezing–thawing," *Journal of Veterinary Medicine Series A*, vol. 35, no. 1–10, pp. 455–460, 1988.
- [29] K. Dorostkar, S. M. Alavi-Shoushtari, and A. Mokarizadeh, "Effects of in vitro selenium addition to the semen extender on the spermatozoa characteristics before and after freezing in water buffaloes (*Bubalus bubalis*)," in *Veterinary research forum*, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran, 2012.
- [30] S. Bozkurt, D. C. Arıkan, E. B. Kurutas et al., "Selenium has a protective effect on ischemia/reperfusion injury in a rat ovary model: biochemical and histopathologic evaluation," *Journal of Pediatric Surgery*, vol. 47, no. 9, pp. 1735–1741, 2012.
- [31] D. C. Brito, A. B. Brito, S. R. R. A. Scalerio et al., "Vitamin E-analog Trolox prevents endoplasmic reticulum stress in frozen–thawed ovarian tissue of capuchin monkey (*Sapajus apella*)," *Cell and Tissue Research*, vol. 355, no. 2, pp. 471–480, 2014.