

Review Article

A Candidate Drug for Nonalcoholic Fatty Liver Disease: A Review of Pharmacological Activities of *Polygoni Multiflori Radix*

Mengting Zhou ^{1,2,3}, Naihua Hu,^{1,2,3} Meichen Liu ^{1,2,3}, Ying Deng ^{1,2,3}, Linfeng He ^{1,2,3},
Chaocheng Guo ^{1,2,3}, Xingtao Zhao ^{1,2,3} and Yunxia Li ^{1,2,3}

¹School of Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu 611137, China

²Key Laboratory of Standardization for Chinese Herbal Medicine, Ministry of Education, Chengdu 611137, China

³National Key Laboratory Breeding Base of Systematic Research, Development and Utilization of Chinese Medicine Resources, Chengdu 611137, China

Correspondence should be addressed to Yunxia Li; lyxcdutcm@126.com

Received 18 November 2019; Accepted 6 April 2020; Published 22 April 2020

Academic Editor: Ken-ichi Aihara

Copyright © 2020 Mengting Zhou et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Nonalcoholic fatty liver disease, a type of metabolic syndrome, continues to rise globally. Currently, there is no approved drug for its treatment. Improving lifestyle and exercise can alleviate symptoms, but patients' compliance is poor. More and more studies have shown the potential of *Polygoni Multiflori Radix* (PMR) in the treatment of NAFLD and metabolic syndrome. Therefore, this paper reviews the pharmacological effects of PMR and its main chemical components (tetrahydroxystilbene glucoside, emodin, and resveratrol) on NAFLD. PMR can inhibit the production of fatty acids and promote the decomposition of triglycerides, reduce inflammation, and inhibit the occurrence of liver fibrosis. At the same time, it maintains an oxidation equilibrium status in the body, to achieve the therapeutic purpose of NAFLD and metabolic syndrome. Although more standardized studies and clinical trials are needed to confirm its efficacy, PMR may be a potential drug for the treatment of NAFLD and its complications. However, the occurrence of adverse reactions of PMR has affected its extensive clinical application. Therefore, it is necessary to further study its toxicity mechanism, enhance efficacy and control toxicity, and even reduce toxicity, which will contribute to the safe clinical use of PMR.

1. Introduction

Nonalcoholic fatty liver disease (NAFLD) is one of the most prevalent chronic liver diseases, especially in developed countries, and is considered to be liver manifestations of metabolic syndrome which includes obesity, hypertension, pathoglycemia, and dyslipidemia and leads to atherosclerosis, type 2 diabetes, and so on [1]. NAFLD is characterized by the abnormal accumulation of intracellular triglycerides without excess alcohol intake and is a progressive form of liver disease that includes a large range of diseases from steatosis to steatohepatitis, cirrhosis, and hepatocellular carcinoma eventually [2–4]. Histopathological examination of the occurrence of triglyceride accumulation in more than 5% of hepatocytes was defined as NAFLD [5, 6]. NAFLD is reversible in its early stage and can be intervened through

lifestyle and medical treatment. If not diagnosed and treated, NAFLD may develop into nonalcoholic steatohepatitis (NASH), which may lead to irreversible liver cancer [7]. NAFLD threatens a third of the world's population, across all ages and races [8]. In China, the incidence of NAFLD continues to rise, reaching 15 percent in fast-growing cities [9]. What is worse, in recent years, research studies show that NAFLD is closely related to cognitive performance [10, 11], polycystic ovary syndrome [12], cardiovascular disease, chronic kidney disease, and other extrahepatic diseases [13]. With the change of people's unhealthy lifestyle, the incidence of NAFLD is continuously increasing, which has attracted wide attention worldwide.

At present, the main recommended treatment method is a healthy lifestyle, including strengthening physical exercise and a reasonable diet. However, according to the poor patient

compliance, the treatment effect does not work well [14]. Some scholars have divided potential therapeutic drugs into four categories according to different mechanisms while they have a common goal: improving metabolic problems caused by simple fat accumulation, inhibiting nonalcoholic steatohepatitis, then alleviating liver fibrosis, and finally regulating intestinal flora to reduce intestinal fat absorption, respectively. And some drugs can have multiple effects [15]. Be that as it may, there are no approved drugs on the market for the treatment of NAFLD [16–18]. The drugs used to treat NAFLD mainly inhibit the accumulation of lipids, including insulin sensitizers and lipid-lowering drugs [19, 20]. However, insulin sensitizers have side effects such as edema and hemodilution, while statins may increase the burden on the liver [21–23]. Therefore, it is necessary to find effective therapeutic methods and drugs to control the occurrence and development of NAFLD and solve this problem.

As an important part of the world medical system, traditional Chinese medicine (TCM) has a long history in effectively various diseases. It has the characteristics of multipathway and multitarget and can be used for holistic treatment from different levels because of its remarkable curative effect and small side effects [24–26]. More and more studies show that TCM is effective in treating NAFLD [27, 28]. It is found that the extract of TCM or effective components can address not only NAFLD but also other illnesses of the metabolic syndrome, such as obesity, diabetes, and dyslipidemia. *Polygoni Multiflori Radix* (PMR), as a tonic medicine recorded in “Kaibao Bencao” firstly, has a history of hundreds of years in China. PMR has rich chemical compositions such as stilbenes, quinones, flavonoids, and phospholipids [29]. PMR has a wide range of pharmacological effects such as antiaging, antihyperlipidemia, anticancer, and anti-inflammatory effects, promoting immune regulation as well as nerve protection and healing, and is determined by its various components [30, 31]. Modern studies have shown that PMR has the potential to treat Alzheimer’s disease, hyperlipidemia, Parkinson’s disease, and inflammation. Growing evidence shows that PMR and its compounds are effective in treating NAFLD and the related complications, which is worthy of further study and discussion. Therefore, this review summarizes a series of evidence for the therapeutic action of PMR and its main components in NAFLD.

2. Pharmacological Effects of PMR in NAFLD

The diverse and complex pathogenesis of NAFLD is associated with insulin resistance (IR) which causes the excessive accumulation of free fatty acids. Without timely treatment, it may cause more serious problems such as hepatic inflammation, oxidative stress, mitochondrial dysfunction, endoplasmic reticulum (ER) stress, and apoptosis eventually which are explained by “multiple hit” hypothesis [32]. In addition to the liver injury caused by fat accumulation, the perspective of the relationship between intestinal flora and liver disease has become a research focus recently [33, 34]. Changes in the composition of intestinal microbial communities and their metabolites can also cause liver damage, such

as short-chain fatty acids (SCFA), endogenous ethanol, and bile acids [35]. Therefore, maintaining intestinal flora homeostasis plays an important role in the prevention and treatment of NAFLD. The antisteatosis, antioxidation, anti-inflammation, liver protection, antiobesity, bile acid metabolism adjustment, and intestinal flora regulation effects of PMR will contribute to the treatment of NAFLD (Figure 1). More and more shreds of evidence link NAFLD to metabolic syndrome, so several aspects could be listed to state the pharmacological effects of PMR against NAFLD and metabolic syndrome according to the following seven aspects (Table 1).

2.1. Antisteatosis Activity. The overproduction of total cholesterol (TC) and triglyceride (TG) is considered the sign of hepatic steatosis. In the normal human body, the average content of TC and TG is 3.9 and 19.5 mg/g wet weight in the liver, respectively [6]. At the same time, hepatocytes play a vital role in biosynthesis, biodegradation of low-density lipoprotein (LDL), high-density lipoprotein (HDL), and other related lipoproteins [61, 62]. The control of hepatic steatosis is an important approach to prevent NAFLD and affect its progression to NASH, liver cirrhosis, and hepatocellular carcinoma.

PMR can regulate lipid production and metabolism to alleviate simple fatty hepatocytes. PMR and *Polygoni Multiflori Radix Praeparata* (PMRP) steamed with black beans showed good inhibition of hepatic steatosis. Compared to PMRP, the water extract of PMR displayed a more remarkable effect on regulating the level of TC and TG [36, 38, 40] and the effect of PMR on lipid regulation was more obvious in liver tissues of early NAFLD [37]. Research showed intuitively that PMR and PMRP could inhibit lipase with IC₅₀ values of 38.84 $\mu\text{g/mL}$ and 190.6 $\mu\text{g/mL}$ by a bioactivity-based method, respectively [39]. PMR inhibited the formation of fat and increased the degradation of fat and the oxidation of fatty acids by upregulating the expression of peroxisome proliferator-activated receptor α (PPAR α), carnitine palmitoyltransferase 1 (CPT1), CPT2, uncoupling protein 1 (UCP1), and hormone-sensitive lipase (HSL) and downregulating adipogenic transcription factors and PPAR γ and diacylglycerol O-acyltransferase 2 (DGAT2) mRNA expression in 3T3-L1 preadipocyte cells and high-fat diet models [41, 42].

2.2. Antioxidant Activity. Oxidative stress is an imbalance of oxidation and antioxidation in the body, which produces a large number of oxide intermediates such as reactive oxygen species (ROS) and reactive nitrogen species (RNS). It leads to neutrophil inflammatory infiltration and increased protease secretion [63, 64]. Oxidative stress leads to the progression of NAFLD to NASH, exacerbating the disease [65]. Excessive fat accumulation can lead to an increase of the oxidation of fatty acids in the mitochondrion controlled by PPAR α and the production of excessive ROS [66]. Then, ROS mainly attacks the liver [67] and recruits Kupffer cells which can produce a variety of cytokines like tumor necrosis factor- α (TNF- α) later. As regards hepatic stellate cells, lipid peroxidation can result in proliferation and collagen synthesis caused by oxidative stress [68]. Therefore, treatment for

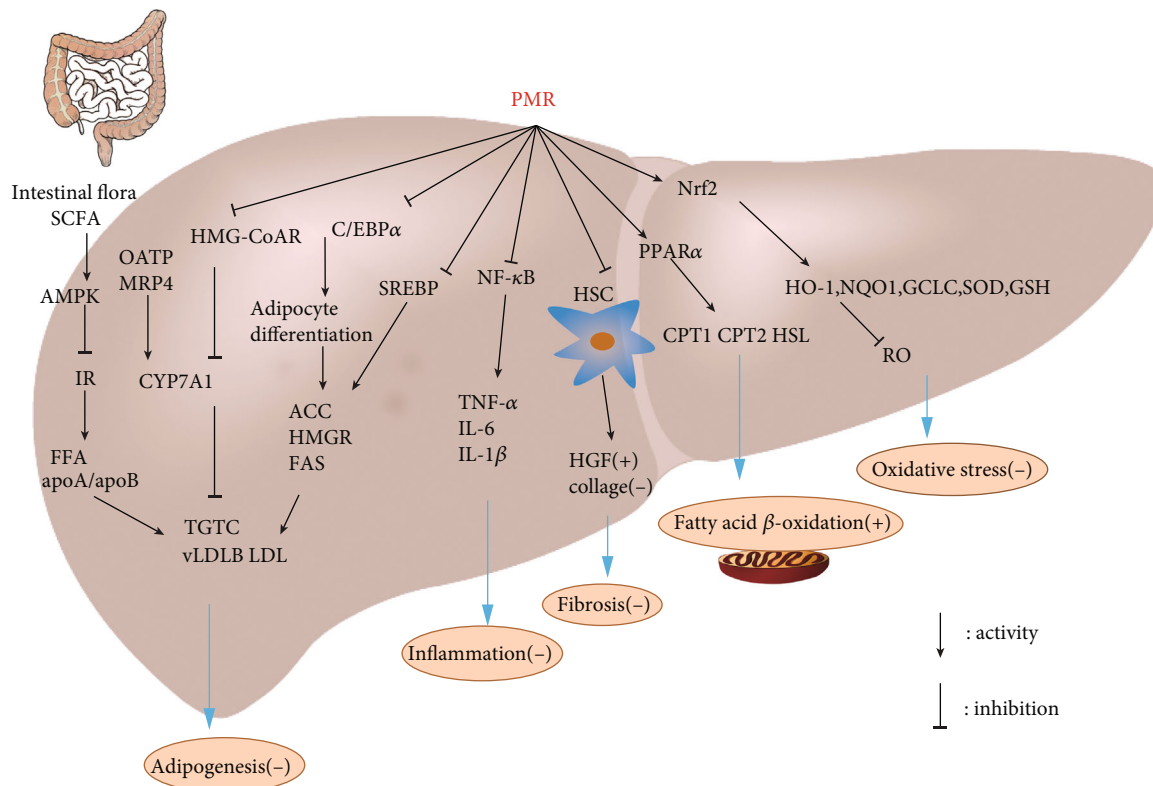


FIGURE 1: Molecular mechanism of PMR in the treatment of NAFLD. PMR exerted pharmacological effects by regulating lipid metabolism, reducing inflammation and fibrosis, improving fatty acid β -oxidation, alleviating oxidative stress, protecting the liver, and adjusting bile acid metabolism. PMR maintained intestinal flora homeostasis via decreasing IR and alleviating inflammation, and PMR reduced the reabsorption of fatty acids to improve NAFLD.

NAFLD can be initiated by reducing oxidative stress and maintaining an antioxidant balance.

PMR has an antioxidant effect [31, 69] that protects the liver from oxidative stress and may be a potential drug for the treatment of NAFLD. PMR was often used as an antiaging drug. It was reported that the chemical profiles were applied to assess the antioxidant activities by establishing the integrated chemometric fingerprints [70]. Besides, PMR upregulated mRNA expression in the nuclear factor erythroid 2-related factor 2 (Nrf2) signal pathway including heme oxygenase-1 (HO-1), NQO1, and glutamate-cysteine ligase catalytic subunit (GCLc) dose-dependently and influenced the nuclear translocation of Nrf2 as well as reduced the content of ROS in H₂O₂- and acetaminophen- (APAP-) induced cells [43]. The enzyme activities of SOD, GSH, GRD, GSH-Px, and GST were improved by PMR in D-galactose-injected mice and CCl₄-induced mice [44, 46]. PMR and PMRP improved mitochondrial β -oxidation by increasing the activity of CPT1A enzyme in vivo and in vitro [47].

2.3. Anti-inflammatory and Antifibrotic Activity. Inflammation and fibrosis can lead to the progression of simple steatosis to NASH and hepatic fibrosis. Therefore, anti-inflammation and prevention of liver fibrosis are considered a treatment direction to hold back the development of NAFLD. Inflammatory response-related signaling pathways

have been reported to be the main signaling pathways for the development of liver fibrosis. Inflammation plays a major role in liver fibrosis through communication and interaction between inflammatory cells [71], cytokines [72, 73], and related signaling pathways [74].

PMR could regulate inflammatory mediators and inflammatory transcription factors like nuclear factor kappa-B (NF- κ B) for anti-inflammatory purposes. The results proved that the ethanol extract of PMR had an anti-inflammatory effect. The extraction of PMR reduced the expression of TNF- α , GST- α , and interleukin 6 (IL-6) which were regarded as therapeutic targets for hepatic inflammation or fibrosis in high-fat diet (HFD) rats [41, 42]. In CCl₄-induced in vivo and in vitro models, PMR remarkably decreased the content of TNF- α [48]. NF- κ B was an important immune-related transcription factor that regulated many cytokines and adhesion factors. PMR inhibited the NF- κ B transcriptional activity in TNF- α -induced NF- κ B activation compared with the model group evaluated by luciferase reporter gene assays [49]. PMR significantly inhibited the activation of hepatic stellate cells induced by PDGF and facilitated the phagocytic activity of Kupffer cells in a concentration-dependent manner [50]. In CCl₄-induced rats of liver fibrosis, the water extract of PMR improved serum albumin which was an indicator of chronic liver damage and reduced the pathological grade of liver fibrosis as well as the occurrence of ascites [51].

TABLE 1: Pharmacological activities of Polygona Multiflora Radix in NAFLD.

Pharmacological effects	Extraction solvent	Country	Model	Efficient doses	Results	References
	Water	China	1% fat emulsion induced L-02 cells	10, 20, 40, 80, 100 μ g/mL	TG \downarrow , TC \downarrow	[36]
	Water	China	High-fat diet rats	0.405, 0.810, 1.62 g/kg	Liver TG \downarrow , TC \downarrow , LDL-C \downarrow	[37]
	50% ethanol	China	High-fat diet rats	10.5, 3.5, 1.17 g/kg	TC \downarrow , TG \downarrow , LDL-C \downarrow , HDL-C \uparrow	[38]
	Ethanol	China		4-5000 μ g/mL	Inhibit lipase	[39]
Antisteatosis activity	Water	China	CCl ₄ , cortisone, acetate, TAA-induced mice	15 g/kg	The enlargement of liver \downarrow , TG \downarrow	[40]
	70% ethanol	China	High-fat diet rats	2.7, 8.1, 16.2 g/kg	HMGR, FAS, ACC, SREBP1 \downarrow , TC, TG, LDL-C \downarrow	[41]
	70% ethanol	Korea	High-fat diet mice, TCA-treated 3T3-L1 preadipocyte cells	0.05%, 10, 30, 50, 100 μ g/mL	C/EBP α , PPAR γ , FAS, body weight, DGAT2 \downarrow , PPAR α , CPT1, CPT2, UCPI, HSL \uparrow	[42]
Antioxidant activity	50% ethanol	China	APAP, H ₂ O ₂ -treated HepG2	20, 50, 100 μ g/mL	HO-1, NQO1, GCLc mRNA \uparrow , Nrf2 in nuclear fraction \uparrow , Nrf2 in cellular fraction \downarrow , ROS \downarrow , superoxide anion \downarrow , MRP4 \uparrow , survival rate \uparrow , OATP \downarrow	[43]
	75% ethanol	China	D-Galactose-injected mice	1, 0.6, 0.3 mL/kg	SOD \uparrow , GSH-Px \uparrow	[44]
	70% ethanol	China	High-fat diet rats	12, 24 mg/kg	MDA \downarrow , SOD, CAT, GSH-Px, T-POC \uparrow	[45]
	Ethyl acetate	China	CCl ₄ -induced mice	0.5-1.5 g/kg	GSH, GRD, GSH-Px, GST \uparrow , plasma ALT, SDH, MDA \downarrow	[46]
	Water	China	High-fat diet rats; NEFA-induced L-02 cells	70, 140, 280 mg/kg; 3.75, 7.5, 15, 30, 60 μ g/mL	ALT, AST, ROS, TC, TG, lipid droplets \downarrow , mitochondrial β -oxidation, CPT1A \uparrow	[47]
Anti-inflammatory and antifibrotic activity	70% ethanol	China	High-fat diet rats	2.7, 8.1, 16.2 g/kg	TNF- α , GST- α \downarrow	[41]
	70% ethanol	Korea	High-fat diet mice	0.05%	IL-6, TNF- α \downarrow , leptin, ALT, AST \downarrow	[42]
	Water	China	CCl ₄ -induced rat; CCl ₄ -induced BCRC 60201 cells	200, 400 mg/kg; 50-300 μ g/mL	TNF- α \downarrow , fatty degeneration, and necrosis \downarrow	[48]
	70% ethanol	Korea	TNF- α -induced HepG2 cells	0.1, 1, 10 μ M	NF- κ B transcriptional activity \downarrow	[49]
	Methanol	China	DMN-induced mice, hepatic nonparenchymal cells	1-1000 g/mL	HGF, the phagocytic activity of liver Kupffer cells, survival rate \uparrow , proliferation of hepatic stellate cells, hydroxyproline \downarrow	[50]
Water	China	CCl ₄ -induced rats	10 mL/kg	ALB \uparrow , the ratio of ascites, the degree of fibrosis \downarrow	[51]	

TABLE 1: Continued.

Pharmacological effects	Extraction solvent	Country	Model	Efficient doses	Results	References
	70% ethanol	Korea	High-fat diet mice	0.05%	AST↓	[42]
	50% ethanol	China	APAP-induced mouse	120 mg/kg	Plasma AST, ALT↓	[43]
	75% ethanol	China	D-Galactose-injected mice	1, 0.6, 0.3 g/mL/kg	ALT↓, AST↓, MDA↓	[44]
Hepatoprotective activity	Water	China	CCl ₄ -induced rat; CCl ₄ -induced BCRC 60201 cells	200, 400 mg/kg; 50–300 µg/mL	ALT↓, AST↓, MDA↓, glutathione S-transferase and catalase activity↑, serum ALT, AST, MDA↓	[48]
	Methanol	China	Dimethylnitrosamine- induced mice	20, 100 mg/kg	Hydroxyproline↓, hepatocyte growth factor (HGF)↑, survival rate↑	[50]
Hypolipidemic activity	Water	China	High-fat diet rats	0.810, 1.62, 3.24 g/kg	TC, HDL-C↓	[37]
	70% ethanol	China	High-fat diet rats	2.7, 8.1, 16.2 g/kg	Plasma LDL-C, TC, TG, HMGR, FAS, ACC↓	[41]
	70% ethanol	China	High-fat diet rats	12, 24 mg/kg	TC, TG, LDL-C↓, apoA/apoB, HDL- C/TC↑	[45]
	Water	China	Hyperlipidemia patients	10 g/d	TC, TG↓, apoA/apoB↑	[52]
	Water	China	Hyperlipidemia patients	150 mL × 2/d	TC, TG, LDL↓, HDL↑	[53]
	Water	China	Hyperlipidemia patients	3 g/d	TC, TG, LDL↓, HDL↑	[54]
Antiobesity activity	Water	China	High-fat diet rats	PMR 0.4050, 0.8100, 0.1620; PMRP 0.8100, 0.1620, 3.240 g/kg	TC, TG, VLDL, the activity of DGAT↓, HL↑	[55]
	Water	China	High-fat diet rats	5 mL/d	TC, TG↓	[56]
	70% ethanol	Korea	3T3-L1 cells; high-fat diet mice	5, 10 µg/mL; 0.05%	3T3-L1 differentiation, lipid accumulation, TG, C/EBPα, PPARγ, FAS↓; body weight, leptin↓	[42]
Intestinal flora regulatory activity	40% ethanol	China	Rats	2 mL	The activity of FAS, body weight↓	[57]
	Water	China	High-fat diet rats	405, 810 mg/kg	TC, TG, LPS, total SCFA, acetic acid, propionic acid, butyric acid↓	[58]
	Water	China	High-fat diet mice	1.125 mg/g	Firmicutes/Bacteroidetes↑ Firmicutes/Bacteroides↓, Clostridium spp., Bacteroides spp., Bifidobacterium spp.↓	[59]
	80% ethanol	China	High-fat and sugar diet rats	57, 228 mg/kg; 12, 48 mg/kg	Desulfovibrio spp.; Oscillibacter spp.↓, Bacteroides spp., Bifidobacterium spp.↓	[60]

2.4. Hepatoprotective Activity. Patients with NAFLD show elevated levels of ALT and AST, which are important biochemical indicators of liver injury. Without timely treatment and control, NAFLD can progress into cirrhosis.

PMR could alleviate the damage to the liver and might become a hepatoprotective medicine to treat NAFLD. The extract of PMR reduced the contents of AST and ALT in serum [42–44, 48] and the production of malondialdehyde (MDA) compared with CCl_4 -induced liver damage. In addition, the $\text{TNF-}\alpha$ was reduced and histopathology examination showed relieved adipose tissue and necrosis in the PMR treatment group [48]. It not only increased the hepatocyte growth factor (HGF) which played an important role in liver regeneration and attenuated development of liver cirrhosis but also increased hydroxyproline that was an indicator for collagen content. Consequently, the survival rate was enhanced largely in the PMR treatment group [50].

2.5. Hypolipidemic Activity. Hyperlipidemia is a common metabolic syndrome associated with increased TC, TG, and LDL-C, while decreased HDL-C levels [75]. An overload of cholesterol in the liver can lead to fatty liver disease. Therefore, regulating cholesterol balance is an effective means to treat NAFLD.

PMR might control the development of NAFLD by regulating abnormal markers of cholesterol which indicated the severity and progression of NAFLD. Traditional Chinese medicine prescriptions containing PMR have been used for many years to treat NAFLD and hyperlipidemia such as Xuezhining Wan and Shouwu Wan [76]. The extraction of PMR showed a remarkable increase in the activities of 3-hydroxy-3-methylglutaryl-CoA reductase (HMGR) related to TC biosynthesis; meanwhile, fatty acid synthase (FAS) and acetyl-CoA carboxylase (ACC) decreased sufficiently which played an important role in the biosynthesis of TG [41]. However, researchers found that PMRP was more effective in regulating lipids in circulating blood to treat hyperlipidemia [37]. In addition, PMR lowered the plasma LDL-C, TC, and TG levels in high-fat diet rats [41, 55] and hyperlipidemia patients [52–54, 56].

2.6. Antiobesity Activity. Due to people's unhealthy lifestyle, obesity is prevalent all over the world. It is accompanied with many health problems including dyslipidemia, type 2 diabetes, and steatosis [77]. Obesity leads to various metabolic abnormalities, and the proliferation of adipose tissue is closely related to the imbalance of various transcription factors [78].

Based on the antiobesity effect of PMR, it might be developed as a potential weight loss drug to replace the existing weight loss agents with large side effects. In order to reduce the accumulation of fat in the body, lipase inhibitors have been selected as targets to prevent the digestion and absorption of fat for the treatment of obesity [79]. Studies have shown that many ingredients in PMR were screened for potential lipase inhibitors such as stilbenes and anthraquinones, which could be used for curing obesity [39]. 70% ethanol extract of PMR could not only reduce weight but also reduce visceral fat weight including epididymal, retroperito-

neal, perirenal, and mesenteric white adipose tissue in HFD-induced obese mice. PMR reduced the expression of CCAAT/enhancer-binding protein α (C/EBP α) and PPAR γ which played vital roles in controlling the number and size of fat cells. Meanwhile, the expression of FAS also decreased in 3T3-L1 preadipocyte cells cured by PMR [42, 57].

2.7. Intestinal Flora Regulatory Activity. Intestinal flora is closely related to the development of NAFLD [80]. The accumulating evidence suggests that changes in intestinal flora can promote the deterioration of NAFLD by influencing processes of inflammation, bile acids, and IR, and vice versa [81, 82]. And intestinal flora promotes the development of NAFLD through the enterohepatic axis [83]. SCFA are metabolites produced by intestinal flora rather than the host [84] mainly including acetic acid, propionic acid, and butyric acid, which can mediate the inflammatory response through various channels and directly or indirectly affect NAFLD.

PMR can regulate NAFLD by maintaining intestinal flora homeostasis to change bile acid metabolism and fatty acid absorption. The extraction of PMR could decrease the content of TC and TG in the liver tissue of NAFLD mice fed with a high-fat diet; at the same time, it reduced the total SCFA in the intestinal canal of the model group. However, there were gender differences in the change of different SCFA [58]. PMR could regulate blood glucose and alleviate IR by managing the diversity of intestinal flora such as changing the imbalance of Firmicutes/Bacteroides which was directly proportional to the level of blood sugar [59, 85] and the relative abundance of Proteobacteria and so on [60].

3. Pharmacological Effects of Active Constituents of PMR in NAFLD

There are 133 chemical constituents isolated from PMR, including stilbene glycosides, anthraquinones, flavonoids, phospholipids, and phenylpropanoids [86]. Stilbene glycosides and anthraquinones are the main components of PMR. Studies have shown that tetrahydroxystilbene glucoside, emodin, and resveratrol can effectively improve NAFLD (Figure 2). Therefore, this paper reviews the therapeutic effects of the three components in NAFLD (Table 2).

3.1. Tetrahydroxystilbene Glucoside. Tetrahydroxystilbene glucoside, named 2,3,5,4'-tetrahydroxystilbene-2-O- β -D-glucoside (TSG), is the main component extracted from PMR. It is regarded as a quality control indicator of PMR and is required to contain no less than 1% in Chinese Pharmacopoeia. There was growing evidence that TSG had a wide range of pharmacological effects such as anti-inflammation, antioxidation, and antiapoptosis [124, 125].

TSG attenuated the inflammatory response by downregulating the levels of IL-6 and $\text{TNF-}\alpha$ in HFD-induced apoE^{-/-} mice. In vivo experiment showed that TSG significantly reduced the release of inflammatory factors IL-6, $\text{TNF-}\alpha$, and C-reactive protein in high-fat and high-cholesterol diet rats [89, 90]. Besides, TSG decreased the expression of p-Smad3 that increased NF- κ B inhibitor I κ B α degradation and then promoted the activation of the NF- κ B signaling

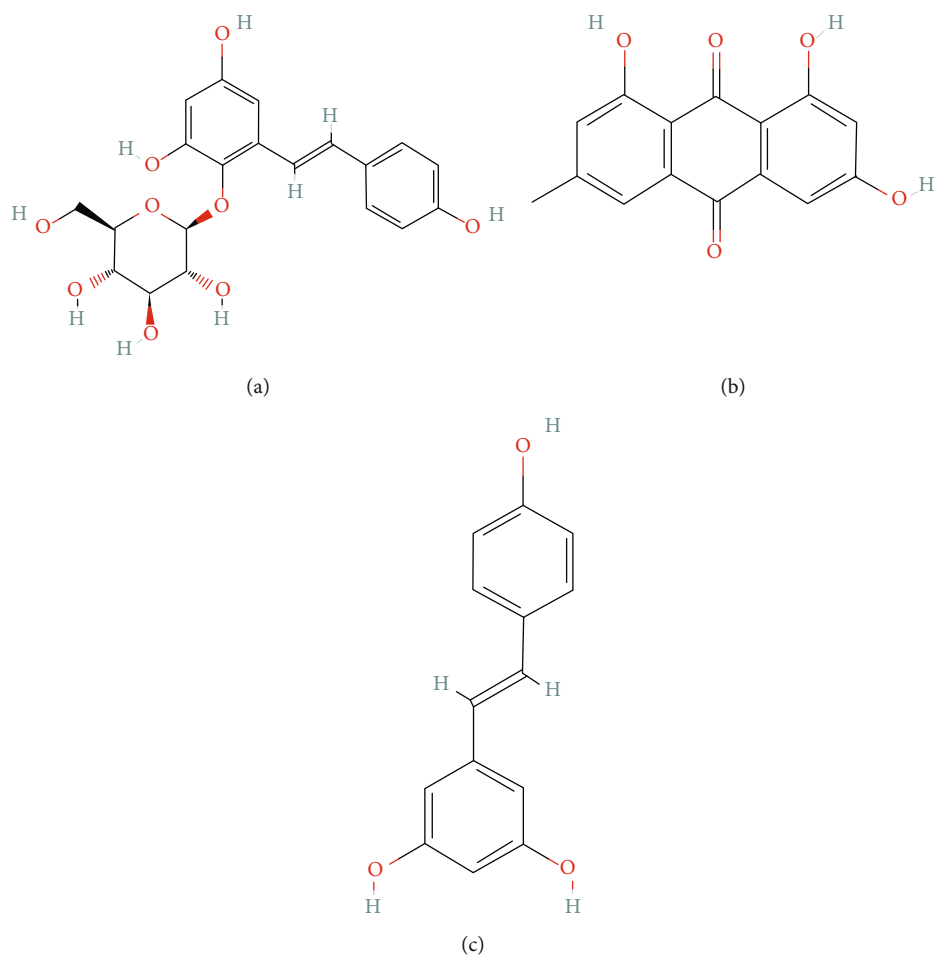


FIGURE 2: Chemical structures of three constituents from PMR. (a) tetrahydroxystilbene glucoside, (b) emodin, and (c) resveratrol.

pathway. 3-Hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors could increase LDL-C uptake and metabolism by increasing the number of LDL receptors on the surface of hepatocytes [126]. TSG could reduce the LDL level by increasing the expression of LDL receptors and TC and TG in hyperlipidemic rats and increase the HDL [45, 89–93]. Hence, TSG may be used as HMG-CoA reductase inhibitors to decrease the level of LDL. In fat emulsion-incubated L-02 cells, TSG effectively reduced the accumulation of triglycerides by inhibiting the expression of related proteins that synthesized triglycerides [87, 88]. Reverse cholesterol transport (RCT) was involved in cholesterol metabolism by transporting cholesterol to the liver; TSG mediated the RCT signaling pathway by upregulating the expression of ATP-binding cassette transporter A1 (ABCA1), ABCG1, and scavenger receptor class B type I (SR-BI) which regulated cholesterol efflux from the macrophage [127] and the expression of cholesterol 7α -hydroxylase (CYP7A1) that was a rate-limiting enzyme of bile acid synthesis [128]. Therefore, the lipid profiles decreased owing to the increased level of excretion [92]. In a study after HFD rats were orally administrated with TSG, the activity of SOD, CAT, GSH-Px, and T-AOC was increased remarkably indicating that TSG had an antioxidant effect to cure hyperlipidemia [90]. Particularly, TSG downregu-

lated the expression of α -SMA associated with the activation of hepatic stellate cells, and $\text{TNF}\beta$ correlated to the fibrosis-related genes [45, 90]. Studies have shown that TSG could also regulate the homeostasis of intestinal flora to rectify lipid metabolism by increasing Akkermansia genera and the ratio of Firmicutes/Bacteroidetes, while the abundance of *Helicobacter pylori* decreased [59].

Taken together, TSG might develop as an underlying agent against NAFLD through mediating liver lipid metabolism, alleviating inflammation, regulating oxidation and fibrosis, and other ways (Figure 3).

3.2. Emodin. Emodin (1,3,8-trihydroxy-6-methylantraquinone) is a hydroxyanthraquinone derivative in PMR and has a wide range of physiological activities. The experimental results demonstrated that it has anti-inflammatory, antioxidant, hepatoprotective, and anticancer activities [129–131]. There was growing evidence that emodin had a significant effect on the treatment of NAFLD.

Emodin alleviated the lipid accumulation and ameliorated hepatic steatosis in vivo and in vitro [96, 97]. It reduced the expression of sterol regulatory element-binding protein 1 (SREBP1) [95] which was an important lipogenic transcription factor associated with triglyceride accumulation [132] and the phosphorylated mTOR (p-mTOR) that positively

TABLE 2: Pharmacological activities of tetrahydroxystilbene glucoside, emodin, and resveratrol in NAFLD.

Pharmacological effects	Country	Type	Doses	Model	Results	References
Tetrahydroxystilbene glucoside						
Antisteatosis activity	China	In vitro	150 μ M/L	Fat emulsion-induced L-02 cells	TG, SREBP1c, ACACA, FASN, FATP4, L-FABP \downarrow , PPAR α \uparrow	[87]
	China	In vitro	50-300 μ M	Fat emulsion-induced L-02 cells	TC, TG, HMG-CoA reductase \downarrow , CYP7A \uparrow	[88]
	China	In vivo	0.035, 0.07 mg/g	HFD-induced mice	IL-6, TNF- α , VCAM-1, MCP-1, TG, ox-LDL \downarrow	[59]
Anti-inflammatory activity	China	In vivo	30, 60, 120 mg/kg	HFD/HCD-induced rats	IL-6, TNF- α , CRP \downarrow	[89]
	China	In vivo	50, 100 mg/kg	HFD-induced mice	CD68, TNF- α , IL-6, ICAM \downarrow	[90]
	China	In vivo	12, 24 mg/kg	HFD-induced rats	TC, TG, LDL-C, apoB, MDA \downarrow	[45]
	China	In vivo	120, 60, 30 mg/kg	HFD/HCD-induced rats	TC, TG, LDL \downarrow , HDL \uparrow	[89]
Hypolipidemic activity	China	In vivo	50, 100 mg/kg	HFD-induced mice	TC, TG, LDL-C \downarrow , HDL-C \uparrow ; ALT, AST \downarrow , SREBP1c, ACC α , FAS \downarrow , PPAR α , CPT1A, ACO, ABCG5, CYP7A1 \uparrow	[90]
	China	In vivo	90, 180 mg/kg	Hyperlipidemic rats	TC, LDL-C, AI \downarrow , LDLR \uparrow	[91]
	China	In vivo	50, 100 mg/kg	HFD-induced apoE $^{-/-}$ mice	TC, TG, LDL, LABCAL, ABCG1, HDL, SR-BI, ABCG5, CYP7A1 \uparrow	[92]
	China	In vivo	30, 60, 120 mg/kg	HFD-induced rats	TC, TG, LDL-C, MDA, TC/HDL-C \downarrow	[93]
	China	In vivo	12, 24 mg/kg	HFD-induced rats	SOD, CAT, GSH-Px, T-AOC \uparrow	[45]
Antioxidant activity	China	In vivo	50, 100 mg/kg	HFD-induced mice	ROS, NOX-2, NOX-4, CYP2E1, MDA \downarrow , SOD, GSH, CAT \uparrow	[90]
Antifibrotic activity	China	In vivo	50, 100 mg/kg	HFD-induced mice	α -SMA and TGF- β \downarrow	[90]
Intestinal flora regulatory activity	China	In vivo	0.035, 0.07 mg/g	HFD-induced mice	Bacteroidetes, Proteobacteria, Tenericutes, Helicobacter pylori \downarrow , Firmicutes, Akkermansia \uparrow	[59]
Emodin						
	China	In vitro	50-300 μ M	Fat emulsion-induced L-02 cells	HMG-CoA reductase, DGAT1 \downarrow , CYP7A \uparrow	[88]
	China	In vivo and in vitro	20, 40, 80 μ M; 40, 80, 160 mg/kg	FFA-induced HepG2 cells; HFD-induced rats	Intracellular lipids, TC, TG, SREBP1, SCD1, FAS, CD36, p-mTOR, P-p70S6K \downarrow , CPT1, PPAR α , P-AMPK, P-ACC \downarrow	[94]
Antisteatosis activity	China	In vivo	40, 80, 160 mg/kg	Fructose-induced rats	SREBP1c, body weight, liver index, serum and hepatic TG, ACC1, FAS, SCD1, GRP78 \downarrow , CPT1, SREBP1c \downarrow	[95]
	Italy	In vivo	40 mg/kg	HFD/HF-induced rats	TG, ALT, glucose, insulin, HOMA-IR \downarrow	[96]
	China	In vivo	40 mg/kg	HCD-induced rats	Body weight, liver index, serum ALT, blood lipids, hepatic triglyceride \downarrow , PPAR γ \uparrow	[97]
	Italy	In vivo	40 mg/kg	HFD/HF-induced rats	Pro SSG/Tot GSH, PTEN phosphorylation/glutathionylation	[96]
Antioxidant activity	China	In vivo	10 mg/kg	HFD/HF-induced rats	SMass, CRE, apoptotic foam cell, MDA, OxLDL \downarrow , SOD \uparrow	[98]

TABLE 2: Continued.

Pharmacological effects	Country	Type	Doses	Model	Results	References
	Italy	In vivo	40 mg/kg	HFD/HF-induced rats	TNF- α ↓	[96]
	China	In vivo	40 mg/kg	MCD-induced mice	ALT, AST, IL-1 β , IL-6↓	[99]
Anti-inflammatory activity	USA.	In vivo and in vitro	40 mg/kg; 25 μ M	LPS-induced hyperlipidemic mice and macrophages	Liver weight, total liver infiltrating cells, liver infiltrating cells, leukocyte number, ALT, AST, ORO positive area, cholesterol↓; TNF- α , IL-6, IL-1 β , iNOS, P-Erk/t-Erk↓	[100]
Antifibrotic activity	China	In vivo	20, 40, 80 mg/kg	HFD-induced rats	TNF- α , IL-1, TLR4, MyD88, TRAF-6↓	[101]
	China	In vitro	3, 10, 30 μ M	SB203580 and TGF- β -neutralizing antibody-treated HSC-T6 cells	α -SMA, fibronectin, type I collagen, TGF- β 1, TGF- β R I, TGF- β R II↓	[102]
	China	In vivo	10, 20, 40 mg/kg	CCl ₄ -induced rats	Collagen, TGF- β 1↓	[103]
	China	In vivo	20 mg/kg	CCl ₄ -induced mice	TGF- β 1, IL-1 β , TNF- α , GRN, MCP-1, CCL7↓	[104]
Resveratrol	China	In vivo	15 mg/kg	HFD-induced rats	TC, TG, LDL-C↓, HDL-C↑	[105]
Antisteatosis activity	China	In vivo and in vitro	100 mg/kg; 40 μ M	HFD-induced rats; PA-induced HepG2 cells	Body weight, liver index, TC, TG, LDL-C↓	[106]
	China	In vivo	100 mg/kg	HFD-induced mice	TC, HDL-C, glucose, insulin, HOMA-IR↓	[107]
	Poland	In vitro	10, 20 μ M	HG-induced HepG2 cells	Lipid accumulation↓	[108]
	China	In vivo and in vitro	400 mg/kg; 10, 20, 40 μ M	HFD-induced mice; PA-induced HepG2 cells	SIRT1↑, ATF6, Fsp27 β /CIDEA, CREBH, PLIN1↓	[109]
	Poland	In vitro	10, 20 mol/L	OA- and PA-induced HepG2 cells	Lipid accumulation↓	[110]
	Serbia	In vivo	20 mg/kg	Cholesterol and cholic acid-induced rats	HDL↑, LDL, TG↓	[111]
	China	In vivo and in vitro	15 mg/kg; 20 μ M	HFD-induced mice; FFA-induced HepG2 cells	TG, body weight, lipid accumulation, ROS↓	[112]
	China	In vivo	15 mg/kg	HFD-induced rats	ALT, AST, TBIL, DBIL, IBIL↓	[105]
	China	In vivo and in vitro	100 mg/kg; 40 μ M	HFD-induced rats; PA-induced HepG2 cells	ALT, AST↓	[106]
	China	In vivo	100 mg/kg	HFD-induced mice	ALT, AST↓	[107]
Hepatoprotective activity	Cyprus	In vivo	50 mg	Patients	SGPT, g-GT, IR↓	[113]
Antioxidant activity	China	In vivo and in vitro	100 mg/kg; 40 μ M	HFD-induced rats; PA-induced HepG2 cells	SREBP1c, FAS, mROS↓, PPAR α , AMPK↑	[106]
	China	In vivo	100 mg/kg	HFD-induced mice	MDA, T-SOD, GPx, CD36↓	[107]
	Poland	In vitro	10, 20 mol/L	OA- and PA-induced HepG2 cells	Apoptotic cells, oxidative stress intensity↓, mitochondrial membrane potential↑	[110]
	Egypt	In vivo	20 mg/kg	HFD-induced rats	Proteolytic cleavage of SREBP1 and SREBP2, CPT1, UCP2↓	[114]

TABLE 2: Continued.

Pharmacological effects	Country	Type	Doses	Model	Results	References
	Iran	In vivo and in vitro	0.4%; 20 μ M	HFD-induced mice; HD-induced HepG2 cells	Nrf2, HO-1, NQO1, SOD \uparrow , TG, FAS, FBS, SREBP1c \downarrow	[115]
	China	In vivo	100 mg/kg	HFD-induced mice	TNF- α , TLR4 \downarrow	[107]
	China	In vivo and vitro	30 mg/kg; 50, 100 μ M	HFD-induced mice; NEFA-induced primary hepatocytes of mice	IL-1 β , IL-6, TNF- α , I κ B α , NF- κ B, p65 \downarrow , AMPK α , SIRT1 \uparrow	[116]
Anti-inflammatory activity	Iran	In vivo	500 mg	NAFLD patient	ALT, hs-CRP, IL-6, NF- κ B, cytokeratin-18 M30 \downarrow	[117]
	Brazil	In vivo	30 mg/kg	HFD-induced mice	TC, TG, transaminases, insulin, TNF- α , IL-6, NF- κ B, ACC, PPAR γ , SREBP1 \downarrow	[118]
	China	In vivo	50 mg/kg	HFD-induced ULK1-deficient mice	IL-6, TNF- α , p65 \downarrow , I κ B α \uparrow	[119]
	China	In vivo	50 mg/kg	HFD-induced ULK1-deficient mice	Lipid droplets, the inflammatory infiltrate, ALT, AST, insulin, glucose, SREBP1c, MDA, 8-isoprostane \downarrow , adiponectin, GPx \uparrow	[119]
Antifibrotic activity	Japan	In vivo	2, 20 mg/kg	HFD/LPS-induced mice	CD14, ALT, TNF- α , IL-6, p-STAT3 \downarrow	[120]
	Iran	In vivo	10 mg/kg	CCl $_4$ -induced rats	ALT, AST, ALP, hydroxyproline, LOX, TOS, MDA \downarrow , TAC, -SH \uparrow	[121]
Inducing autophagy activity	China	In vitro and in vivo	20, 40, 80 μ M; 0.4%	PA-induced HepG2 cells; HFD-induced mice	cAMP, SIRT1, pPRKA, P-AMPK, SIRT1 \uparrow	[122]
Regulating FXR activity	Iran	In vitro	25 mg/kg	HFD-induced rats	SIRT1, LXR, FXR \uparrow , AST, ALT, ALP \downarrow	[123]

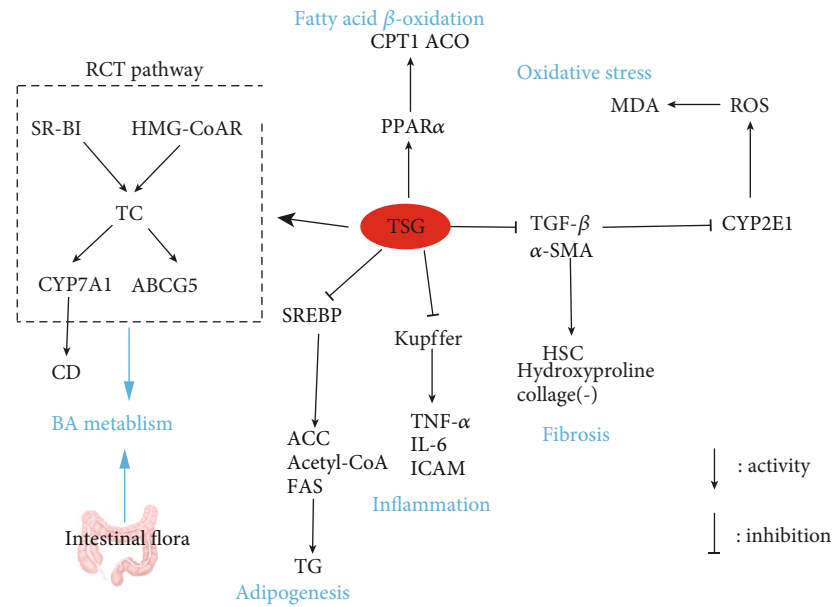


FIGURE 3: Schematic illustration of mechanism of TSG on improving NAFLD. TSG could not only improve bile acid metabolism abnormalities caused by NAFLD through the PCT signaling pathway and intestinal flora but also inhibit fat production, inflammation, and oxidative stress pathways, while promoting the β -oxidation of fatty acids.

regulated the activity of SREBP1, while the expression of AMP-activated protein kinase (AMPK) which was an indirect upstream kinase of SREBP1 was increased. Hence, emodin effectively regulated lipid metabolism via the CaMKK-AMPK-mTOR-p70S6K signaling pathway [94]. Furthermore, emodin inhibited the expression of HMG-CoA reductase and DGAT1 associated with the synthesis of TC and TG [88]. In addition, emodin showed a powerful effect on lowering blood lipids by inhibiting the activity of SMase, the content of CRE, and the quantity of apoptotic foam cell and promoting antioxidant ability at the same time [96, 98]. Emodin also alleviated inflammation by reducing leukocyte infiltration as well as the expression of inflammatory factors. Further study showed that extracellular regulated protein kinases 1/2 (Erk1/2), p38, toll-like receptor 4 (TLR4), and NF- κ B signaling pathways were inhibited dramatically [96, 100, 101], so we could conclude that emodin could make a contribution to steatohepatitis in a way. However, it was reported that emodin could aggravate liver damage and inflammation in MCD diet-induced NAFLD in mice along with the increased serum ALT and AST levels and the expression of inflammatory factors IL-1 β and IL-6 [99]. Two completely opposite results might be due to the different modeling methods and model animals, which needed further exploration. In addition, emodin improved liver fibrosis via decreasing transforming growth factor- β 1 (TGF- β 1) to inhibit the activation of hepatic stellate cells and the infiltration of Gr1^{hi} monocytes [102–104].

In conclusion, the emodin could develop into a potential agent to prevent the progression of NAFLD to NASH owing to a variety of pharmacological activities (Figure 4).

3.3. Resveratrol. Resveratrol is a polyphenol named trans-3,5,4'-trihydroxy-trans-stilbene (RES). Resveratrol had both *cis* and *trans* optical isomers, and studies had shown that

the latter was more stable and active [133]. A large number of studies had shown that it had a powerful effect on the prevention and treatment of NAFLD.

Most of the available experimental data came from two models including in vivo experiments of mice or rats with high-fat diet as well as in vitro tests with primary hepatocytes or HepG2 cells. RES could improve the symptom of NAFLD by protecting the liver, adjusting lipid metabolism, alleviating inflammation and fibrosis, regulating the oxidation equilibrium status, and enhancing autophagy [119, 120, 122] as well as controlling the farnesoid X receptor (FXR) [123]. The increase of serum TC, TG, LDL-C, ALT, and AST content and the reduction of HDL-C were as serum markers of NAFLD, and RES could return them to normality effectively due to the hepatoprotective and lipid metabolic activity [105–107, 111, 113]. In addition, in FFA-, PA-, OA-, or HG-induced HepG2 cell models, RES reduced lipid droplet accumulation indirectly [108, 110, 112]. Sirtuin 1 (SIRT1) was an important regulator associated with glucose and fat acid metabolism in the liver. A study showed that RES could remarkably activate the expression of SIRT1. At the same time, a series of proteins related to lipid droplets were down-regulated such as activating transcription factor 6 (ATF6), cAMP response element-binding protein H (CREBH), and perilipin 1 (PLIN1) [109, 116]. Liver inflammation was accompanied with the increase of inflammatory cytokines. RES reduced the expression and secretion of proinflammatory cytokines (IL-1 β , IL-6, TNF- α , and TLR4), and further studies also suggested that RES suppressed NF- κ B which was a transcription factor combined with its inhibitor I κ B α and bound to DNA and then promoted the expression of cytokines when it was activated by an external stimulus [134] via activating the phosphorylation of AMPK α and the expression of SIRT1 [116–119]. In addition, RES reduced collagen fiber bundles, hydroxyproline, and lysyl oxidase

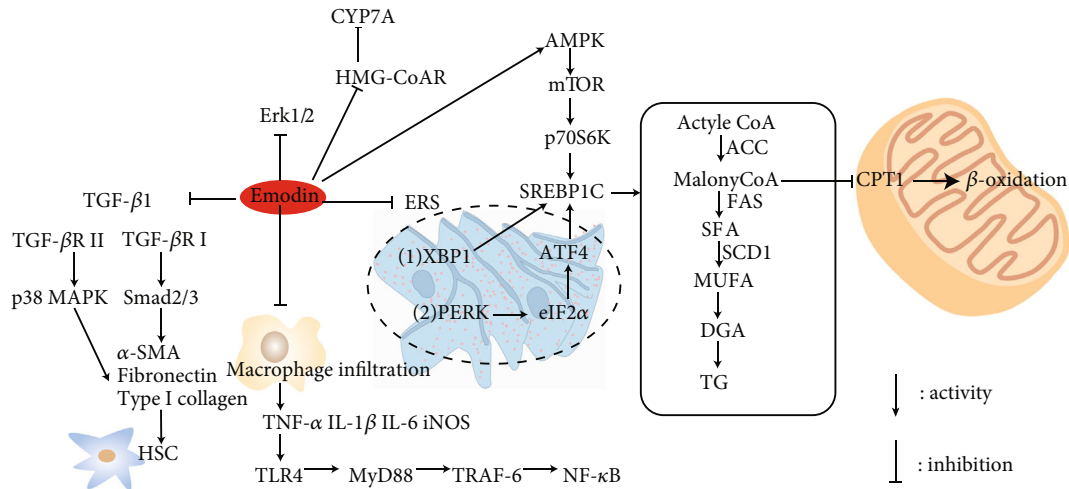


FIGURE 4: Schematic illustration of mechanism of emodin on improving NAFLD. Emodin mainly reduced fat production and increased β -oxidation of fatty acids by inhibiting the oxidative stress of the endoplasmic reticulum and alleviated the inflammatory response by inhibiting the Erk1/2, p38, and NF- κ B signaling pathway. Emodin suppressed the activation of hepatic stellate cells via inhibiting the expression of TGF- β 1.

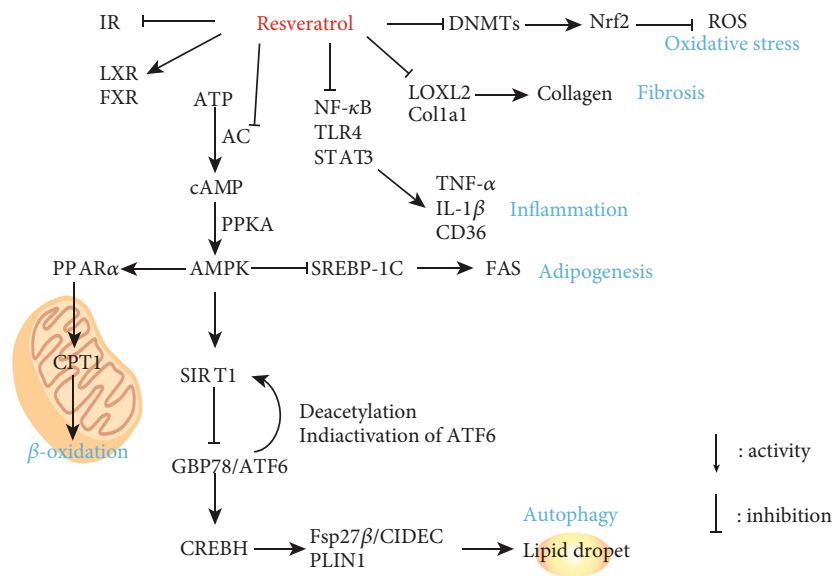


FIGURE 5: Schematic illustration of mechanism of RES on improving NAFLD. RES promoted autophagy to reduce the accumulation of lipid droplets and inhibited β -oxidation of fatty acids by activating the AMPK signaling pathway. RES alleviated the symptom caused by inflammation, liver fibrosis, and oxidative stress and improved the expression of LXR and FXR genes.

(LOX) to alleviate liver fibrosis [121]. A lot of evidence showed RES improved redox balance by activating PPAR α related to fatty acid oxidation and inhibiting SREBP1c associated to lipogenesis [106, 110, 114]. Meanwhile, the content and activity of T-SOD and GPx were improved by the treatment with RES, while the content of MDA decreased [107]. The Nrf2-Keap1 pathway participated in the prevention of metabolic disorders in NAFLD, and RES could activate Nrf2 signaling to inhibit lipogenesis [115]. However, RES presented low bioavailability due to poor solubility. Many researchers were devoted to exploiting new dosage forms, for example, the PLGA nanoparticles loaded with RES, in order to improve the effect [135, 136].

To sum up, RES had varieties of biological activities, which have been proved to play a potential role in treating NAFLD (Figure 5).

4. Knowledge of Toxicity

With the widespread application of PMR and its preparations, adverse reactions related to the hepatotoxicity of PMR have been reported in the early 20th century [137–139]. Therefore, the toxicity of PMR attracted wide attention. The adverse reactions of PMR were jaundice, yellowing urine, cholestasis, liver injury, etc. It was reported that the toxicity of the ethanol extract of PMR was higher than that

of the water extract. Therefore, it was not recommended to make wine with PMR for nourishing. 70% ethanol extract had the highest toxicity [140]. Studies showed that the occurrence of adverse reactions was related to time and dose, and a long-term large dose was more likely to cause hepatotoxicity [139]. The other researchers thought that PMR-induced hepatic injury was an idiosyncratic drug-induced liver injury, so they built the lipopolysaccharide- (LPS-) induced model of hepatotoxicity [141–143] to explore the toxic substance basis and mechanism of PMR.

Many claims have been made to clarify its toxic components. Some studies concluded that adverse reactions were mainly due to its anthraquinone components [31, 144, 145]. Emodin and its derivatives were the most likely hepatotoxic components [144] and had a time-dependent intracellular accumulation [146], while TSG and physcion may mitigate the effects of emodin [147]. Nevertheless, some studies held that the hepatotoxicity of PMR depended not only on the composition of emodin but also on the content of TSG [147]. Therefore, there are many uncertainties about the toxic components of PMR, and more toxicological studies are needed.

The clinical application of PMR pays attention to compatibility, and reasonable compatibility can reduce toxicity. PMR can be used with other TCM to increase the curative effect and reduce toxicity. At the same time, its toxicity may be attributed to high doses and prolonged use. Clinical use of PMR should attach great importance to the examination of liver function. PMRP toxicity is more suitable for safe clinical use with lower toxicity [147]. In the meantime, it is important to improve the public's correct understanding of PMR.

5. Discussion

PMR, based on the theory of TCM, PMR, and PMRP, has different effects. PMR can moisten intestines and help defecate, remove toxicity, and eliminate carbuncles, while PMRP which is steamed with black soya beans has a large effect including nourishing the liver and kidney, strengthening bones and muscles, and blackening the beard and hair. Modern studies have shown that PMR and PMRP had therapeutic potential for aging, hair loss, hyperlipidemia, inflammation, and cancer [148]. Numerous experimental data indicated the potential of PMR in the treatment of NAFLD. In the experiments, scholars found that both PMR and PMRP could effectively reduce TC, TG, and LDL-C and increase HDL-C content to regulate lipid metabolism. The effective compounds, TSG, emodin, and resveratrol, might have synergic effects in the body to regulate lipid metabolism in NAFLD. However, studies have found that PMR had a better effect on lowering lipids [38, 39, 76]. The possible reason is that after processing, the content and proportion of active ingredients are changed. For example, conjugated anthraquinone compounds are hydrolyzed at high temperature to increase the content of free anthraquinone [149]. At the same time, the content of TSG decreased significantly [29]. Treatment of the NAFLD process is complicated by PMR because of its rich pharmacological effects and complex ingredients. It can go through the different ways in the different mecha-

nisms to realize regulation by the multicomponent and multiple targets.

To sum up, the therapeutic mechanism of PMR is mainly controlled by the following pathways: (1) reducing lipid formation by downregulating SREBP, ACC, and FAS; (2) suppressing the release of inflammatory cytokines through the NF- κ B signaling pathway; (3) resulting in antifibrosis by inhibiting the activation of hepatic stellate cells; (4) augmenting fatty acid β -oxidation via upregulating the PPAR α ; (5) reducing oxidative stress and improving antioxidant levels through Nrf2; (6) reducing IR and improving bile acid metabolism by regulating intestinal flora and increasing the expression of CYP7A1; and (7) decreasing ALT and AST levels to protect the liver. These different pathways work together to improve NAFLD by regulating lipid metabolism, reducing inflammation and fibrosis, improving antioxidant levels, and protecting the liver.

This review also summarizes the research progress of the three main components of PMR in the treatment of NAFLD. TSG, emodin, and RES whose pharmacological activities are consistent with those of PMR all show antisteatosis, anti-inflammatory, antifibrotic, and antioxidative stress activities and increase β -oxidation of fatty acids in mitochondria. Meanwhile, TSG and emodin can regulate bile acid metabolism by increasing the expression of CYP7A1, while RES can affect bile acid metabolism by regulating LXR and FXR genes which can adjust CYP7A1 indirectly [150]. Therefore, these three components may contribute to the activity of PMR in regulating bile acid metabolism. Of these three components, the current literature has found that only RES has been shown to reduce lipid droplet accumulation by upregulating SIRT1 to activate the autophagy pathway. However, studies on PMR have not mentioned the reduction of lipid droplets through autophagy. The possible reason is that the content of RES in PMR is low, and the administration of the PMR extract does not reach the concentration to render autophagy, while the administration of the RES monomer has obvious effects.

6. Conclusion

This review describes in detail the therapeutic effects of PMR and its chemical components on NAFLD. Its antisteatosis, antioxidation, anti-inflammation, antifibrosis, liver protection, lipid reduction, antiobesity, intestinal flora regulation, and bile acid adjustment effects might contribute to its therapeutic effects. Although stilbene glycosides and anthraquinones are the main components, the relationship between the two is still unclear; whether they act synergically or inhibit each other and the sequence of action need further study. At present, adverse reactions of PMR are frequent, but its therapeutic effect is undeniable. Therefore, it is necessary not only to understand the basis and mechanism of its efficacy in the treatment of NAFLD but also to further study its toxicity mechanism so as to contribute to the safety and wide use of PMR in clinical practice.

Conflicts of Interest

The authors report no conflicts of interest.

Acknowledgments

The work was supported by the National Natural Science Foundation of China (NSFC) (Grant Nos. 81373943 and 81573583) and the Sichuan Provincial Science and Technology Department of Youth Science and Technology Innovation Research Team Program (2017TD0001).

References

- [1] A. M. Escudé, G. Pera, I. Arteaga et al., "Relationship between hypothyroidism and non-alcoholic fatty liver disease in the Spanish population," *Medicina Clínica (English Edition)*, vol. 154, no. 1, pp. 1–6, 2020.
- [2] M. M. de Freitas Carvalho, N. N. Lage, A. H. de Souza Paulino et al., "Effects of açai on oxidative stress, ER stress, and inflammation-related parameters in mice with high fat diet-fed induced NAFLD," *Scientific Reports*, vol. 9, no. 1, p. 8107, 2019.
- [3] B. A. Neuschwander-Tetri and S. H. Caldwell, "Nonalcoholic steatohepatitis: summary of an AASLD Single Topic Conference," *Hepatology*, vol. 37, no. 5, pp. 1202–1219, 2003.
- [4] S. McPherson, T. Hardy, E. Henderson, A. D. Burt, C. P. Day, and Q. M. Anstee, "Evidence of NAFLD progression from steatosis to fibrosing-steatohepatitis using paired biopsies: implications for prognosis and clinical management," *Journal of Hepatology*, vol. 62, no. 5, pp. 1148–1155, 2015.
- [5] S. Tandra, M. M. Yeh, E. M. Brunt et al., "Presence and significance of microvesicular steatosis in nonalcoholic fatty liver disease," *Journal of Hepatology*, vol. 55, no. 3, pp. 654–659, 2011.
- [6] L. S. Szczepaniak, P. Nurenberg, D. Leonard et al., "Magnetic resonance spectroscopy to measure hepatic triglyceride content: prevalence of hepatic steatosis in the general population," *American Journal of Physiology. Endocrinology and Metabolism*, vol. 288, no. 2, pp. E462–E468, 2005.
- [7] F. Tzifi, A. Fretzayas, G. Chrousos, and C. Kanaka-Gantenbein, "Non-alcoholic fatty liver infiltration in children: an underdiagnosed evolving disease," *Hormones*, vol. 18, no. 3, pp. 255–265, 2019.
- [8] K. Tziomalos, V. G. Athyros, P. Paschos, and A. Karagiannis, "Nonalcoholic fatty liver disease and statins," *Metabolism, Clinical and Experimental*, vol. 64, no. 10, pp. 1215–1223, 2015.
- [9] J. Guang, "Relevant issues concerning clinical trials of traditional Chinese medicine treatment of nonalcoholic fatty liver disease," *Journal of Clinical Hepatology*, vol. 30, no. 4, pp. 299–302, 2014.
- [10] G. Weinstein, K. Davis-Plourde, J. J. Himali, S. Zelber-Sagi, A. S. Beiser, and S. Seshadri, "Non-alcoholic fatty liver disease, liver fibrosis score and cognitive function in middle-aged adults: the Framingham study," *Liver International*, vol. 39, no. 9, pp. 1713–1721, 2019.
- [11] A. Pinçon, O. De Montgolfier, N. Akkoyunlu et al., "Non-alcoholic fatty liver disease, and the underlying altered fatty acid metabolism, reveals brain hypoperfusion and contributes to the cognitive decline in APP/PS1 mice," *Metabolites*, vol. 9, no. 5, p. 104, 2019.
- [12] N. Salva-Pastor, N. C. Chávez-Tapia, M. Uribe, and N. Nuño-Lámbarri, "Understanding the association of polycystic ovary syndrome and non-alcoholic fatty liver disease," *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 194, article 105445, 2019.
- [13] J. A. Velarde-Ruiz Velasco, E. S. García-Jiménez, K. R. García-Zermeño et al., "Extrahepatic complications of non-alcoholic fatty liver disease: its impact beyond the liver," *Revista de Gastroenterología de México*, vol. 84, no. 4, pp. 472–481, 2019.
- [14] S. Zelber-Sagi, R. Lotan, A. Shlomai et al., "Predictors for incidence and remission of NAFLD in the general population during a seven-year prospective follow-up," *Journal of Hepatology*, vol. 56, no. 5, pp. 1145–1151, 2012.
- [15] L. Drew, "Drug development: sprint finish," *Nature*, vol. 551, no. 7681, pp. S86–S89, 2017.
- [16] A. F. G. Cicero, A. Colletti, and S. Bellentani, "Nutraceutical approach to non-alcoholic fatty liver disease (NAFLD): the available clinical evidence," *Nutrients*, vol. 10, no. 9, p. 1153, 2018.
- [17] A. C. Sheka, O. Adeyi, J. Thompson, B. Hameed, P. A. Crawford, and S. Ikramuddin, "Nonalcoholic steatohepatitis: a review," *JAMA*, vol. 323, no. 12, pp. 1175–1183, 2020.
- [18] H. Cui and X. Zhang, "Occurrence and clinical management of nonalcoholic fatty liver disease in obesity patients: a literature review," *Journal of Pediatric Endocrinology & Metabolism*, 2020.
- [19] E. Bugianesi, E. Gentilcore, R. Manini et al., "A randomized controlled trial of metformin versus vitamin E or prescriptive diet in nonalcoholic fatty liver disease," *The American Journal of Gastroenterology*, vol. 100, no. 5, pp. 1082–1090, 2005.
- [20] R. J. Ford, M. D. Fullerton, S. L. Pinkosky et al., "Metformin and salicylate synergistically activate liver AMPK, inhibit lipogenesis and improve insulin sensitivity," *The Biochemical Journal*, vol. 468, no. 1, pp. 125–132, 2015.
- [21] Y. Lin-yuan, G. Li-hong, and T. Yun-qiu, "Nonalcoholic fatty liver disease effect of emodin based on AMPK signaling pathway," *Chinese Journal of Experimental Traditional Medical Formulae*, vol. 26, pp. 203–209, 2020.
- [22] A. B. Walker, E. K. Naderali, P. D. Chattington, R. E. Buckingham, and G. Williams, "Differential vasoactive effects of the insulin sensitizers rosiglitazone (BRL 49653) and troglitazone on human small arteries in vitro," *Diabetes*, vol. 47, no. 5, pp. 810–814, 1998.
- [23] D. Pastori, L. Polimeni, F. Baratta, A. Pani, M. Del Ben, and F. Angelico, "The efficacy and safety of statins for the treatment of non-alcoholic fatty liver disease," *Digestive and Liver Disease*, vol. 47, no. 1, pp. 4–11, 2015.
- [24] C. M. Chong, H. Su, J. J. Lu, and Y. Wang, "The effects of bioactive components from the rhizome of *Salvia miltiorrhiza* (Danshen) on the characteristics of Alzheimer's disease," *Chinese Medicine*, vol. 14, no. 1, p. 19, 2019.
- [25] T. H. Huang, C. F. Lin, A. Alalaiwe, S. C. Yang, and J. Y. Fang, "Apoptotic or antiproliferative activity of natural products against keratinocytes for the treatment of psoriasis," *International Journal of Molecular Sciences*, vol. 20, no. 10, 2019.
- [26] Y. Wang, C. Liu, H. Wang, Y. Jiang, P. Wang, and H. Shang, "Systematic Review of Basic Research on Alzheimer's Disease with Shen Zhi Ling Oral Liquid," *Evidence-based Complementary and Alternative Medicine*, vol. 2019, Article ID 8216714, 10 pages, 2019.
- [27] X.-L. Hu, Y.-J. Niu, M. Chen et al., "Preventive effects of total flavonoid C-glycosides from *Abrus mollis* on nonalcoholic

- fatty liver disease through activating the PPAR α signaling pathway," *Planta Medica*, vol. 85, no. 8, pp. 678–688, 2019.
- [28] O. Ahmad, B. Wang, K. Ma et al., "Lipid modulating anti-oxidant stress activity of gastrodin on nonalcoholic fatty liver disease larval zebrafish model," *International Journal of Molecular Sciences*, vol. 20, no. 8, p. 1984, 2019.
- [29] Y. Liu, Q. Wang, J. Yang et al., "Polygonum multiflorum Thunb.: a review on chemical analysis, processing mechanism, quality evaluation, and hepatotoxicity," *Frontiers in Pharmacology*, vol. 9, p. 364, 2018.
- [30] G. Bounda and Y. Feng, "Review of clinical studies of Polygonum multiflorum Thunb. and its isolated bioactive compounds," *Pharmacognosy Research*, vol. 7, no. 3, pp. 225–236, 2015.
- [31] L. Lin, B. Ni, H. Lin et al., "Traditional usages, botany, phytochemistry, pharmacology and toxicology of Polygonum multiflorum Thunb.: a review," *Journal of Ethnopharmacology*, vol. 159, pp. 158–183, 2015.
- [32] E. Buzzetti, M. Pinzani, and E. A. Tsochatzis, "The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD)," *Metabolism*, vol. 65, no. 8, pp. 1038–1048, 2016.
- [33] I. Pierantonelli and G. Svegliati-Baroni, "Nonalcoholic fatty liver disease: basic pathogenetic mechanisms in the progression from NAFLD to NASH," *Transplantation*, vol. 103, no. 1, pp. e1–e13, 2019.
- [34] M. J. Ma and J. Wu, "Association between intestinal flora imbalance and nonalcoholic fatty liver disease," *Zhonghua Gan Zang Bing Za Zhi*, vol. 25, no. 10, pp. 789–793, 2017.
- [35] D. Zhou and J. G. Fan, "Microbial metabolites in non-alcoholic fatty liver disease," *World Journal of Gastroenterology*, vol. 25, no. 17, pp. 2019–2028, 2019.
- [36] M. Wang, R. Zhao, W. Wang, X. Mao, and J. Yu, "Lipid regulation effects of Polygoni Multiflori Radix, its processed products and its major substances on steatosis human liver cell line L02," *Journal of Ethnopharmacology*, vol. 139, no. 1, pp. 287–293, 2012.
- [37] N. Li, Z. Chen, X. Mao, J. Yu, and R. Zhao, "Effects of lipid regulation using raw and processed radix polygoni multiflori in rats fed a high-fat diet," *Evidence-based Complementary and Alternative Medicine*, vol. 2012, Article ID 329171, 10 pages, 2012.
- [38] J. Chen, H. Zhao, Y. Yang, B. Liu, J. Ni, and W. Wang, "Lipid-lowering and antioxidant activities of Jiang-Zhi-Ning in traditional Chinese medicine," *Journal of Ethnopharmacology*, vol. 134, no. 3, pp. 919–930, 2011.
- [39] Y. X. Chang, A. H. Ge, Y. Jiang, J. Teye Azietaku, J. Li, and X. M. Gao, "A bioactivity-based method for screening, identification of lipase inhibitors, and clarifying the effects of processing time on lipase inhibitory activity of Polygonum multiflorum," *Evidence-based Complementary and Alternative Medicine*, vol. 2016, Article ID 5965067, 10 pages, 2016.
- [40] C. Liu, Q. Zhang, and J. Lin, "Effect of the root of Polygonum multiflorum Thunb. and its processed products on fat accumulation in the liver of mice," *Zhongguo Zhong Yao Za Zhi*, vol. 17, no. 10, pp. 639–646, 1992, 639.
- [41] Z. Xian, Y. Liu, W. Xu, F. Duan, Z. Guo, and H. Xiao, "The anti-hyperlipidemia effects of raw Polygonum multiflorum extract in vivo," *Biological & Pharmaceutical Bulletin*, vol. 40, no. 11, pp. 1839–1845, 2017.
- [42] R. Y. Choi, H. I. Lee, J. R. Ham, S. T. Yee, K. Y. Kang, and M. K. Lee, "Heshouwu (Polygonum multiflorum Thunb.) ethanol extract suppresses pre-adipocytes differentiation in 3T3-L1 cells and adiposity in obese mice," *Biomedicine & Pharmacotherapy*, vol. 106, pp. 355–362, 2018.
- [43] E. Y. Lin, A. Chagnaadorj, S. J. Huang, C. C. Wang, Y. H. Chiang, and C. W. Cheng, "Hepatoprotective Activity of the Ethanolic Extract of Polygonum multiflorum Thunb. against Oxidative Stress-Induced Liver Injury," *Evidence-based Complementary and Alternative Medicine*, vol. 2018, Article ID 4130307, 9 pages, 2018.
- [44] J. Yang, Y. He, J. Zou, L. Xu, F. Fan, and Z. Ge, "Effect of Polygonum multiflorum Thunb on liver fatty acid content in aging mice induced by D-galactose," *Lipids in Health and Disease*, vol. 18, no. 1, p. 128, 2019.
- [45] X. Congkun, W. Rui, and Y. Zhifang, "Study on effect of Polygonum multiflorum extract on lipid metabolism and its anti-oxidation in SD rats with hyperlipemia," *China Pharmaceuticals*, vol. 18, no. 24, pp. 19–20, 2009.
- [46] P. Y. Chiu, D. H. Mak, M. K. Poon, and K. M. Ko, "In vivo antioxidant action of a lignan-enriched extract of Schisandra fruit and an anthraquinone-containing extract of Polygonum root in comparison with schisandrin B and emodin," *Planta Medica*, vol. 68, no. 11, pp. 951–956, 2002.
- [47] L. Lin, Z. Hao, S. Zhang et al., "Study on the protection of water extracts of Polygoni Multiflori Radix and Polygoni Multiflori Radix Praeparata against NAFLD and its mechanism," *Journal of Ethnopharmacology*, vol. 252, article 112577, 2020.
- [48] B.-H. Lee, Y.-Y. Huang, P.-D. Duh, and S.-C. Wu, "Hepato-protection of emodin and Polygonum multiflorum against CCl₄-induced liver injury," *Pharmaceutical Biology*, vol. 50, no. 3, pp. 351–359, 2012.
- [49] Y. N. Sun, W. Li, S. B. Song, X. T. Yan, S. Y. Yang, and Y. H. Kim, "Nuclear factor kappa B activation and peroxisome proliferator-activated receptor transactivational effects of chemical components of the roots of Polygonum multiflorum," *Pharmacognosy Magazine*, vol. 12, no. 45, pp. 31–35, 2016.
- [50] C. H. Huang, L. Y. Horng, C. F. Chen, and R. T. Wu, "Chinese herb Radix Polygoni Multiflori as a therapeutic drug for liver cirrhosis in mice," *Journal of Ethnopharmacology*, vol. 114, no. 2, pp. 199–206, 2007.
- [51] W. Jin, *Studies of Zhiheshouwu on CCl₄-induced liver fibrosis in rats*, Master. Dalian Medical University, 2004.
- [52] J. H. Yin, X. Y. Zhou, and X. Q. Zhu, "Pharmacological and clinical studies on the processed products of radix Polygoni multiflori," *Zhongguo Zhong Yao Za Zhi*, vol. 17, no. 12, pp. 722–724, 1992, 62–3.
- [53] L. X., "Clinical observation on Shouwu Jiangzhi decoction for treatment of hyperlipemia," *Zhejiang Journal of Traditional Chinese Medicine*, vol. 10, pp. 588–589, 2006.
- [54] W. Z, Y. Y, and L. Y, "Observation of therapeutic effects of Shouwu granules for patients with hyperlipidemia and hypercoagulable state," *Chinese Traditional Patent Medicine*, vol. 12, pp. 28–30, 2000.
- [55] P. Lin, Y. R. He, J. M. Lu et al., "In vivo lipid regulation mechanism of polygoni multiflori radix in high-fat diet fed rats," *Evidence-based Complementary and Alternative Medicine*, vol. 2014, Article ID 642058, 8 pages, 2014.
- [56] W. X, X. G, and S. X, "The effect of different kinds of Chinese herb medicine such as Padix Polygoni Multiflori on blood biochemical indexes in rats of fatty liver," *Journal of Anhui University of Chinese Medicine*, vol. 5, pp. 39–40, 2006.

- [57] L. L. W. X. and T. W., "Inhibition to fatty acid synthase with extract of tuber fleeceflower root," *Chinese Journal of Biochemistry and Molecular Biology*, vol. 3, pp. 297–304, 2003.
- [58] Y. Wang, P. Lin, L. Jianmei, M. Zhang, and L. Li, "Effect of Radix *Polygoni Multiflori* and TSG on short-chain fatty acids in intestinal tract of NAFLD rats," *Modern Chinese Medicine*, vol. 19, no. 9, pp. 1254–1261, 2017.
- [59] F. Li, T. Zhang, Y. He et al., "Inflammation inhibition and gut microbiota regulation by TSG to combat atherosclerosis in ApoE^{-/-} mice," *Journal of Ethnopharmacology*, vol. 247, article 112232, 2020.
- [60] B. Q., *Pharmacological and metagenomic evidences for the alleviation of insulin resistance by Polygoni Multiflori Radix Preparta*, Master. Yunnan University of Traditional Chinese Medicine, 2018.
- [61] Z. Chen, H. Qin, S. Qiu, G. Chen, and Y. Chen, "Correlation of triglyceride to high-density lipoprotein cholesterol ratio with nonalcoholic fatty liver disease among the non-obese Chinese population with normal blood lipid levels: a retrospective cohort research," *Lipids in Health and Disease*, vol. 18, no. 1, p. 162, 2019.
- [62] L. Wu and K. G. Parhofer, "Diabetic dyslipidemia," *Metabolism*, vol. 63, no. 12, pp. 1469–1479, 2014.
- [63] Y. Z. Fang, S. Yang, and G. Wu, "Free radicals, antioxidants, and nutrition," *Nutrition*, vol. 18, no. 10, pp. 872–879, 2002.
- [64] C. Peng, X. Wang, J. Chen et al., "Biology of ageing and role of dietary antioxidants," *BioMed Research International*, vol. 2014, Article ID 831841, 13 pages, 2014.
- [65] Y. Sumida, E. Niki, Y. Naito, and T. Yoshikawa, "Involvement of free radicals and oxidative stress in NAFLD/NASH," *Free Radical Research*, vol. 47, no. 11, pp. 869–880, 2013.
- [66] D. H. Ipsen, J. Lykkesfeldt, and P. Tveden-Nyborg, "Molecular mechanisms of hepatic lipid accumulation in non-alcoholic fatty liver disease," *Cellular and Molecular Life Sciences*, vol. 75, no. 18, pp. 3313–3327, 2018.
- [67] V. Sánchez-Valle, N. C. Chávez-Tapia, M. Uribe, and N. Méndez-Sánchez, "Role of oxidative stress and molecular changes in liver fibrosis: a review," *Current Medicinal Chemistry*, vol. 19, no. 28, pp. 4850–4860, 2012.
- [68] S. Li, H. Y. Tan, N. Wang et al., "The role of oxidative stress and antioxidants in liver diseases," *International Journal of Molecular Sciences*, vol. 16, no. 11, pp. 26087–26124, 2015.
- [69] H. N. Kim, Y. R. Kim, J. Y. Jang et al., "Neuroprotective effects of Polygonum multiflorum extract against glutamate-induced oxidative toxicity in HT22 hippocampal cells," *Journal of Ethnopharmacology*, vol. 150, no. 1, pp. 108–115, 2013.
- [70] H. F. Chen, Y. H. Chen, C. H. Liu et al., "Integrated chemometric fingerprints of antioxidant activities and HPLC-DAD-CL for assessing the quality of the processed roots of Polygonum multiflorum Thunb. (Heshouwu)," *Chinese Medicine*, vol. 11, no. 1, p. 18, 2016.
- [71] T. Khoury, A. Mari, W. Nseir, A. Kadah, W. Sbeit, and M. Mahamid, "Neutrophil-to-lymphocyte ratio is independently associated with inflammatory activity and fibrosis grade in nonalcoholic fatty liver disease," *European Journal of Gastroenterology & Hepatology*, vol. 31, no. 9, pp. 1110–1115, 2019.
- [72] Y. Kamari, A. Shaish, E. Vax et al., "Lack of interleukin-1 α or interleukin-1 β inhibits transformation of steatosis to steatohepatitis and liver fibrosis in hypercholesterolemic mice," *Journal of Hepatology*, vol. 55, no. 5, pp. 1086–1094, 2011.
- [73] A. Berlanga, E. Guiu-Jurado, J. A. Porras, and T. Auguet, "Molecular pathways in non-alcoholic fatty liver disease," *Clinical and Experimental Gastroenterology*, vol. 7, pp. 221–239, 2014.
- [74] H. J. Zhangdi, S. B. Su, F. Wang et al., "Crosstalk network among multiple inflammatory mediators in liver fibrosis," *World Journal of Gastroenterology*, vol. 25, no. 33, pp. 4835–4849, 2019.
- [75] E. Dembowski and M. H. Davidson, "A review of lipid management in primary and secondary prevention," *Journal of Cardiopulmonary Rehabilitation and Prevention*, vol. 29, no. 1, pp. 2–12, 2009.
- [76] Commission of Chinese Pharmacopoeia, *Pharmacopoeia of the People's Republic of China*, China Medicopharmaceutical Science & Technology Publishing House, Beijing, China, 2015.
- [77] D. P. Schuster, "Obesity and the development of type 2 diabetes: the effects of fatty tissue inflammation," *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 3, pp. 253–262, 2010.
- [78] B. R. Sharma, H. J. Kim, M. S. Kim, C. M. Park, and D. Y. Rhyu, "Caulerpa okamurae extract inhibits adipogenesis in 3T3-L1 adipocytes and prevents high-fat diet-induced obesity in C57BL/6 mice," *Nutrition Research*, vol. 47, pp. 44–52, 2017.
- [79] M. Hanefeld and G. Sachse, "The effects of orlistat on body weight and glycaemic control in overweight patients with type 2 diabetes: a randomized, placebo-controlled trial," *Diabetes, Obesity & Metabolism*, vol. 4, no. 6, pp. 415–423, 2002.
- [80] J. Ma, Q. Zhou, and H. Li, "Gut microbiota and nonalcoholic fatty liver disease: insights on mechanisms and therapy," *Nutrients*, vol. 9, no. 10, p. 1124, 2017.
- [81] X. He, G. Ji, W. Jia, and H. Li, "Gut microbiota and nonalcoholic fatty liver disease: insights on mechanism and application of metabolomics," *International Journal of Molecular Sciences*, vol. 17, no. 3, p. 300, 2016.
- [82] J. Shen, M. S. Obin, and L. Zhao, "The gut microbiota, obesity and insulin resistance," *Molecular Aspects of Medicine*, vol. 34, no. 1, pp. 39–58, 2013.
- [83] D. Compare, P. Coccoli, A. Rocco et al., "Gut–liver axis: The impact of gut microbiota on non alcoholic fatty liver disease," *Nutrition, Metabolism, and Cardiovascular Diseases*, vol. 22, no. 6, pp. 471–476, 2012.
- [84] A. L. McOrist, G. C. Abell, C. Cooke, and K. Nyland, "Bacterial population dynamics and faecal short-chain fatty acid (SCFA) concentrations in healthy humans," *The British Journal of Nutrition*, vol. 100, no. 1, pp. 138–146, 2008.
- [85] K. Nishitsuji, J. Xiao, R. Nagatomo et al., "Analysis of the gut microbiome and plasma short-chain fatty acid profiles in a spontaneous mouse model of metabolic syndrome," *Scientific Reports*, vol. 7, no. 1, p. 15876, 2017.
- [86] W. H. Y. J. and Z. L., "Chemical compositions and pharmacological activities of Polygoni Multiflori Radix," *Chinese Journal of Experimental Traditional Medical Formulae*, vol. 25, no. 13, pp. 192–205, 2019.
- [87] P. Lin, J. M. Lu, Y. F. Wang, W. Gu, R. H. Zhao, and J. Yu, "Prevention mechanism of 2,3,5,4'-tetrahydroxy-stilbene-2-O- β -D-glucoside on lipid accumulation in steatosis hepatic L-02 cell," *Pharmacognosy Magazine*, vol. 13, no. 50, pp. 245–253, 2017.
- [88] W. Wang, Y. He, P. Lin et al., "In vitro effects of active components of Polygonum multiflorum Radix on enzymes

- involved in the lipid metabolism,” *Journal of Ethnopharmacology*, vol. 153, no. 3, pp. 763–770, 2014.
- [89] W. Zhang, C. H. Wang, F. Li, and W. Z. Zhu, “2,3,4,5-Tetrahydroxystilbene-2-O-beta-D-glucoside suppresses matrix metalloproteinase expression and inflammation in atherosclerotic rats,” *Clinical and Experimental Pharmacology & Physiology*, vol. 35, no. 3, pp. 310–316, 2008.
- [90] J. Xu, Y. Peng, Y. Zeng, Y. Q. Hua, and X.-l. Xu, “2, 3, 4, 5-Tetrahydroxystilbene-2-O-β-d glycoside attenuates age- and diet-associated non-alcoholic steatohepatitis and atherosclerosis in LDL receptor knockout mice and its possible mechanisms,” *International Journal of Molecular Sciences*, vol. 20, no. 7, p. 1617, 2019.
- [91] X. Gao, Y. J. Hu, and L. C. Fu, “Blood lipid-regulation of stilbene glycoside from polygonum multiflorum,” *Zhongguo Zhong Yao Za Zhi*, vol. 32, no. 4, pp. 323–326, 2007.
- [92] X. Chen, K. Tang, Y. Peng, and X. Xu, “2,3,4,5-tetrahydroxystilbene-2-O-β-d-glycoside attenuates atherosclerosis in apolipoprotein E-deficient mice: role of reverse cholesterol transport,” *Canadian Journal of Physiology and Pharmacology*, vol. 96, no. 1, pp. 8–17, 2018.
- [93] W. C. Z. L. and Y. Z., “Blood lipid regulation of ethyl acetate extracting fraction and stilbene glycoside from tuber of Polygonum multiflorum,” *Chinese Traditional and Herbal Drugs*, vol. 1, pp. 78–83, 2008.
- [94] S. Wang, X. Li, H. Guo et al., “Emodin alleviates hepatic steatosis by inhibiting sterol regulatory element binding protein 1 activity by way of the calcium/calmodulin-dependent kinase kinase-AMP-activated protein kinase-mechanistic target of rapamycin-p70 ribosomal S6 kinase signaling pathway,” *Hepatology Research*, vol. 47, no. 7, pp. 683–701, 2017.
- [95] X. Li, Z. Xu, S. Wang et al., “Emodin ameliorates hepatic steatosis through endoplasmic reticulum-stress sterol regulatory element-binding protein 1c pathway in liquid fructose-feeding rats,” *Hepatology Research*, vol. 46, no. 3, pp. E105–E117, 2016.
- [96] A. Alisi, A. Pastore, S. Ceccarelli et al., “Emodin prevents intrahepatic fat accumulation, inflammation and redox status imbalance during diet-induced hepatosteatosis in rats,” *International Journal of Molecular Sciences*, vol. 13, no. 2, pp. 2276–2289, 2012.
- [97] H. Dong, F. E. Lu, Z. Q. Gao, L. J. Xu, K. F. Wang, and X. Zou, “Effects of emodin on treating murine nonalcoholic fatty liver induced by high caloric laboratory chaw,” *World Journal of Gastroenterology*, vol. 11, no. 9, pp. 1339–1344, 2005.
- [98] Z.-q. Hei, H.-q. Huang, H.-m. Tan et al., “Emodin inhibits dietary induced atherosclerosis by antioxidation and regulation of the sphingomyelin pathway in rabbits,” *Chinese Medical Journal*, vol. 119, no. 10, pp. 868–870, 2006.
- [99] Q. Liu, F. Yu, N. Yu, M. Shi, and F. Wang, “Emodin worsens methionine-choline-deficient diet-induced non-alcoholic fatty liver disease in mice,” *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*, vol. 31, no. 5, pp. 620–624, 2015.
- [100] X. Jia, S. Iwanowycz, J. Wang et al., “Emodin attenuates systemic and liver inflammation in hyperlipidemic mice administered with lipopolysaccharides,” *Experimental Biology and Medicine (Maywood, N.J.)*, vol. 239, no. 8, pp. 1025–1035, 2014.
- [101] T. Liu, Q. Xu, and Y. Zhao, “Effect of emodin on inhibiting hepatic TLR4 signaling expression in NAFLD rats,” vol. 27, no. 2, pp. 201–205, 2016.
- [102] X. Wang, C. Niu, X. Zhang, and M. Dong, “Emodin suppresses activation of hepatic stellate cells through p38 mitogen-activated protein kinase and Smad signaling pathways in vitro,” *Phytotherapy Research*, vol. 32, no. 12, pp. 2436–2446, 2018.
- [103] F. Liu, J. Zhang, J. Qian, G. Wu, and Z. Ma, “Emodin alleviates CCl4-induced liver fibrosis by suppressing epithelial-mesenchymal transition and transforming growth factor-β1 in rats,” *Molecular Medicine Reports*, vol. 18, no. 3, pp. 3262–3270, 2018.
- [104] X. A. Zhao, G. Chen, Y. Liu et al., “Emodin Alleviates Liver Fibrosis of Mice by Reducing Infiltration of Gr1hi Monocytes,” *Evidence-based Complementary and Alternative Medicine*, vol. 2018, Article ID 5738101, 11 pages, 2018.
- [105] X. X. Chen, Y. Y. Xu, R. Wu et al., “Resveratrol reduces gluco-lipid metabolic dysfunction and learning and memory impairment in a NAFLD rat model: involvement in regulating the imbalance of nesfatin-1 abundance and copine 6 expression,” *Frontiers in Endocrinology*, vol. 10, p. 434, 2019.
- [106] Y. Huang, H. Lang, K. Chen et al., “Resveratrol protects against nonalcoholic fatty liver disease by improving lipid metabolism and redox homeostasis via the PPARα pathway,” *Applied Physiology, Nutrition, and Metabolism*, vol. 45, no. 3, pp. 227–239, 2020.
- [107] K. Cheng, Z. Song, H. Zhang et al., “The therapeutic effects of resveratrol on hepatic steatosis in high-fat diet-induced obese mice by improving oxidative stress, inflammation and lipid-related gene transcriptional expression,” *Medical Molecular Morphology*, vol. 52, no. 4, pp. 187–197, 2019.
- [108] M. Izdebska, M. Herbet, M. Gawrońska-Grzywacz et al., “Resveratrol limits lipogenesis and enhance mitochondrial activity in HepG2 cells,” *Journal of Pharmacy & Pharmaceutical Sciences*, vol. 21, no. 1, pp. 504–515, 2018.
- [109] R. Zhou, L. Yi, X. Ye et al., “Resveratrol ameliorates lipid droplet accumulation in liver through a SIRT1/ATF6-dependent mechanism,” *Cellular Physiology and Biochemistry*, vol. 51, no. 5, pp. 2397–2420, 2018.
- [110] M. Izdebska, I. Piątkowska-Chmiel, A. Korolczuk et al., “The beneficial effects of resveratrol on steatosis and mitochondrial oxidative stress in HepG2 cells,” *Canadian Journal of Physiology and Pharmacology*, vol. 95, no. 12, pp. 1442–1453, 2017.
- [111] A. Rašković, V. Čučuč, L. Torović et al., “Resveratrol supplementation improves metabolic control in rats with induced hyperlipidemia and type 2 diabetes,” *Saudi Pharmaceutical Journal*, vol. 27, no. 7, pp. 1036–1043, 2019.
- [112] L. Gong, S. Guo, and Z. Zou, “Resveratrol ameliorates metabolic disorders and insulin resistance in high-fat diet-fed mice,” *Life Sciences*, vol. 242, article 117212, 2020.
- [113] M. Theodotou, K. Fokianos, D. Moniatis et al., “Effect of resveratrol on non-alcoholic fatty liver disease,” *Experimental and Therapeutic Medicine*, vol. 18, no. 1, pp. 559–565, 2019.
- [114] E. F. Khaleel, G. A. Abdel-Aleem, and D. G. Mostafa, “Resveratrol improves high-fat diet induced fatty liver and insulin resistance by concomitantly inhibiting proteolytic cleavage of sterol regulatory element-binding proteins, free fatty acid oxidation, and intestinal triglyceride absorption,” *Canadian Journal of Physiology and Pharmacology*, vol. 96, no. 2, pp. 145–157, 2018.
- [115] H. Hosseini, M. Teimouri, M. Shabani et al., “Resveratrol alleviates non-alcoholic fatty liver disease through epigenetic modification of the Nrf2 signaling pathway,” *The*

- International Journal of Biochemistry & Cell Biology*, vol. 119, article 105667, 2020.
- [116] Y. Tian, J. Ma, W. Wang et al., "Resveratrol supplement inhibited the NF- κ B inflammation pathway through activating AMPK α -SIRT1 pathway in mice with fatty liver," *Molecular and Cellular Biochemistry*, vol. 422, no. 1-2, pp. 75–84, 2016.
- [117] F. Faghihzadeh, P. Adibi, R. Rafiei, and A. Hekmatdoost, "Resveratrol supplementation improves inflammatory biomarkers in patients with nonalcoholic fatty liver disease," *Nutrition Research*, vol. 34, no. 10, pp. 837–843, 2014.
- [118] J. M. Andrade, A. F. Paraíso, M. V. de Oliveira et al., "Resveratrol attenuates hepatic steatosis in high-fat fed mice by decreasing lipogenesis and inflammation," *Nutrition*, vol. 30, no. 7-8, pp. 915–919, 2014.
- [119] L. Li, J. Hai, Z. Li et al., "Resveratrol modulates autophagy and NF- κ B activity in a murine model for treating non-alcoholic fatty liver disease," *Food and Chemical Toxicology*, vol. 63, pp. 166–173, 2014.
- [120] T. Kessoku, K. Imajo, Y. Honda et al., "Resveratrol ameliorates fibrosis and inflammation in a mouse model of nonalcoholic steatohepatitis," *Scientific Reports*, vol. 6, no. 1, article 22251, 2016.
- [121] R. Mohseni, Z. Arab Sadeghabadi, M. T. Goodarzi, and J. Karimi, "Co-administration of resveratrol and beta-aminopropionitrile attenuates liver fibrosis development via targeting lysyl oxidase in CCl₄-induced liver fibrosis in rats," *Immunopharmacology and Immunotoxicology*, vol. 41, no. 6, pp. 644–651, 2019.
- [122] Y. Zhang, M. L. Chen, Y. Zhou et al., "Resveratrol improves hepatic steatosis by inducing autophagy through the cAMP signaling pathway," *Molecular Nutrition & Food Research*, vol. 59, no. 8, pp. 1443–1457, 2015.
- [123] A. Hajighasem, P. Farzanegi, Z. Mazaheri, M. Naghizadeh, and G. Salehi, "Effects of resveratrol, exercises and their combination on Farnesoid X receptor, liver X receptor and sirtuin 1 gene expression and apoptosis in the liver of elderly rats with nonalcoholic fatty liver," *PeerJ*, vol. 6, article e5522, 2018.
- [124] C. Jiao, F. Gao, L. Ou et al., "Tetrahydroxy stilbene glycoside (TSG) antagonizes A β -induced hippocampal neuron injury by suppressing mitochondrial dysfunction via Nrf2-dependent HO-1 pathway," *Biomedicine & Pharmacotherapy*, vol. 96, pp. 222–228, 2017.
- [125] Z. Jiang, W. Wang, and C. Guo, "Tetrahydroxy stilbene glucoside ameliorates H₂O₂-induced human brain microvascular endothelial cell dysfunction in vitro by inhibiting oxidative stress and inflammatory responses," *Molecular Medicine Reports*, vol. 16, no. 4, pp. 5219–5224, 2017.
- [126] Y. Carpentier, J. Ducobu, and J. Sternon, "Atorvastatin (Lipitor)," *Revue Médicale de Bruxelles*, vol. 20, no. 5, pp. 427–433, 1999.
- [127] D. A. Chistiakov, Y. V. Bobryshev, and A. N. Orekhov, "Macrophage-mediated cholesterol handling in atherosclerosis," *Journal of Cellular and Molecular Medicine*, vol. 20, no. 1, pp. 17–28, 2016.
- [128] W. M. Pandak, C. Schwarz, P. B. Hylemon et al., "Effects of CYP7A1 overexpression on cholesterol and bile acid homeostasis," *American Journal of Physiology. Gastrointestinal and Liver Physiology*, vol. 281, no. 4, pp. G878–G889, 2001.
- [129] D. Shrimali, M. K. Shanmugam, A. P. Kumar et al., "Targeted abrogation of diverse signal transduction cascades by emodin for the treatment of inflammatory disorders and cancer," *Cancer Letters*, vol. 341, no. 2, pp. 139–149, 2013.
- [130] W. T. Wei, S. Z. Lin, D. L. Liu, and Z. H. Wang, "The distinct mechanisms of the antitumor activity of emodin in different types of cancer (review)," *Oncology Reports*, vol. 30, no. 6, pp. 2555–2562, 2013.
- [131] X. Dong, J. Fu, X. Yin et al., "Emodin: a review of its pharmacology, toxicity and pharmacokinetics," *Phytotherapy Research*, vol. 30, no. 8, pp. 1207–1218, 2016.
- [132] M. Kohjima, M. Enjoji, N. Higuchi et al., "Re-evaluation of fatty acid metabolism-related gene expression in nonalcoholic fatty liver disease," *International Journal of Molecular Medicine*, vol. 20, no. 3, pp. 351–358, 2007.
- [133] B. C. Trela and A. L. Waterhouse, "Resveratrol: isomeric molar absorptivities and stability," *Journal of Agricultural and Food Chemistry*, vol. 44, no. 5, pp. 1253–1257, 1996.
- [134] J. A. DiDonato, F. Mercurio, and M. Karin, "NF- κ B and the link between inflammation and cancer," *Immunological Reviews*, vol. 246, no. 1, pp. 379–400, 2012.
- [135] S. Wan, L. Zhang, Y. Quan, and K. Wei, "Correction to 'Resveratrol-loaded PLGA nanoparticles: enhanced stability, solubility and bioactivity of resveratrol for non-alcoholic fatty liver disease therapy'," *Royal Society Open Science*, vol. 6, no. 1, article 182173, 2019.
- [136] S. Wan, L. Zhang, Y. Quan, and K. Wei, "Resveratrol-loaded PLGA nanoparticles: enhanced stability, solubility and bioactivity of resveratrol for non-alcoholic fatty liver disease therapy," *Royal Society Open Science*, vol. 5, no. 11, p. 181457, 2018.
- [137] G. Mazzanti, L. Battinelli, C. Daniele et al., "New case of acute hepatitis following the consumption of Shou Wu Pian, a Chinese herbal product derived from *Polygonum multiflorum*," *Annals of Internal Medicine*, vol. 140, no. 7, 2004.
- [138] B. Panis, D. R. Wong, P. M. Hooymans, P. A. De Smet, and P. P. Rosias, "Recurrent toxic hepatitis in a Caucasian girl related to the use of Shou-Wu-Pian, a Chinese herbal preparation," *Journal of Pediatric Gastroenterology and Nutrition*, vol. 41, no. 2, pp. 256–258, 2005.
- [139] X. Lei, J. Chen, J. Ren et al., "Liver damage associated with *Polygonum multiflorum* Thunb.: a systematic review of case reports and case series," *Evidence-based Complementary and Alternative Medicine*, vol. 2015, Article ID 459749, 9 pages, 2015.
- [140] J. B. Yang, W. F. Li, Y. Liu et al., "Acute toxicity screening of different extractions, components and constituents of *Polygonum multiflorum* Thunb. on zebrafish (*Danio rerio*) embryos in vivo," *Biomedicine & Pharmacotherapy*, vol. 99, pp. 205–213, 2018.
- [141] C. Y. Li, C. Tu, D. Gao et al., "Metabolomic study on idiosyncratic liver injury induced by different extracts of *Polygonum multiflorum* in rats integrated with pattern recognition and enriched pathways analysis," *Frontiers in Pharmacology*, vol. 7, p. 483, 2016.
- [142] C. Y. Li, X. F. Li, C. Tu et al., "The idiosyncratic hepatotoxicity of *Polygonum multiflorum* based on endotoxin model," *Yao Xue Xue Bao*, vol. 50, no. 1, pp. 28–33, 2015.
- [143] X. Q, Z. W, and R. J, "Investigation of *Polygonum multiflorum* radix processing approaches using a lipopolysaccharide model of hepatotoxicity," in *Chinese Archives of Traditional Chinese Medicine*, vol. 38, no. 2, pp. 162–282,

China Association of Chinese Medicine; Liaoning University of Traditional Chinese Medicine, 2020.

- [144] J. Ma, L. Zheng, Y. S. He, and H. J. Li, "Hepatotoxic assessment of Polygoni Multiflori Radix extract and toxicokinetic study of stilbene glucoside and anthraquinones in rats," *Journal of Ethnopharmacology*, vol. 162, pp. 61–68, 2015.
- [145] H. Li, X. Wang, Y. Liu et al., "Hepatoprotection and hepatotoxicity of Heshouwu, a Chinese medicinal herb: context of the paradoxical effect," *Food and Chemical Toxicology*, vol. 108, Part B, pp. 407–418, 2017.
- [146] C. L. Li, J. Ma, L. Zheng, H. J. Li, and P. Li, "Determination of emodin in L-02 cells and cell culture media with liquid chromatography-mass spectrometry: application to a cellular toxicokinetic study," *Journal of Pharmaceutical and Biomedical Analysis*, vol. 71, pp. 71–78, 2012.
- [147] J. Yu, J. Xie, X. J. Mao et al., "Hepatotoxicity of major constituents and extractions of Radix Polygoni Multiflori and Radix Polygoni Multiflori Praeparata," *Journal of Ethnopharmacology*, vol. 137, no. 3, pp. 1291–1299, 2011.
- [148] T. T. Ho, H. N. Murthy, D. Dalawai, M. A. Bhat, K. Y. Paek, and S. Y. Park, "Attributes of Polygonum multiflorum to transfigure red biotechnology," *Applied Microbiology and Biotechnology*, vol. 103, no. 8, pp. 3317–3326, 2019.
- [149] J. Huang, J. P. Zhang, J. Q. Bai et al., "Chemical profiles and metabolite study of raw and processed Polygoni Multiflori Radix in rats by UPLC-LTQ-Orbitrap MSⁿ spectrometry," *Chinese Journal of Natural Medicines*, vol. 16, no. 5, pp. 375–400, 2018.
- [150] Y. Duan, F. Zhang, W. Yuan et al., "Hepatic cholesterol accumulation ascribed to the activation of ileum Fxr-Fgf15 pathway inhibiting hepatic Cyp7a1 in high-fat diet-induced obesity rats," *Life Sciences*, vol. 232, article 116638, 2019.