

Review Article

Analytical Methods for Detection of Gasotransmitter Hydrogen Sulfide Released from Live Cells

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Hydrogen sulfide (H₂S) plays an important role in mammals as a signaling molecule. Recently, abnormal H₂S concentration has been associated with several pathophysiological states, such as diabetes mellitus, hypertension, Alzheimer's disease, and Parkinson's disease. As regulating H₂S concentration can be a very prominent way of developing new drugs, many researchers have paid great attention to H₂S research. To understand the role of H₂S in pathophysiology and develop H₂S-based therapies, it is necessary to measure the exact concentration of H₂S within biological systems. But, H₂S is volatile and can be easily oxidized. Besides, the active sites for several biological effects of H₂S are inside the cell. Therefore, there is a need for the development of new methods for the accurate and reliable detection of H₂S within live cells. This review provides a summary of recent developments in H₂S detection methods for live cell analysis.

1. Introduction

Hydrogen sulfide (H₂S) is a biologically relevant gaseous signaling molecule, i.e., a gasotransmitter, collectively with nitric oxide (NO) and carbon monoxide (CO) [1–3]. The endogenous production and signaling capability of H₂S in mammalian tissues were firstly demonstrated by Abe and Kimura in 1996, showing that H₂S is an endogenous modulator in the central nervous system [4]. Since then, there has been a dramatic shift from the belief that H₂S works entirely as an environmental toxin to the understanding that H₂S plays an important role in organ function and homeostasis [5]. H₂S has been revealed to take part in the regulation of various pathophysiological conditions within mammalian systems, such as vascular tone and blood pressure [6, 7], neurotransmission [8], angiogenesis [9], cardiac function [10], various leukocytic functions [11], penile erectile function [12], and gastrointestinal tract function [13]. In general, when the concentration of H₂S in tissues or cells is high, H₂S is regarded as a toxic substance and its oxidation products—persulfide, sulfite, thiosulfate, and sulfate—may give rise to cytotoxic

effects through inhibiting mitochondrial cytochrome C oxidase and disrupting cell energy production, resulting in tissue inflammation or DNA damage [14]. On the other hand, H₂S at low concentrations can lead to different effects on biological processes including DNA repair and metabolism, cellular division, regulation of cell cycle, modulation of protein kinase, and organization of cytoskeletal framework [15]. But the exact physiological role of H₂S depends on the specific circumstance, its concentration, and the interplays with other signaling molecules—NO and CO.

Recently, there have been a few trials to classify the roles of H₂S as “H₂S-poor” and “H₂S-rich” under pathophysiological conditions. First, there exist a few disease states where local or systemic H₂S deficiency either due to inhibition of H₂S biosynthesis and/or due to increased H₂S consumption such as asthma, diabetic vascular complications, and aging [16]. Especially, H₂S has been known to be associated to the pathogenesis of cardiovascular diseases including hypertension [17], atherosclerosis [18], and myocardial injury [19], and the severity of these diseases is negatively related to plasma H₂S levels [20, 21]. Besides, the mean serum

H₂S level in preeclampsia patients was significantly lower than controls [22]. And Renieris et al. suggested that H₂S could be a potential marker for severity and final outcome of pneumonia by the SARS-CoV-2 coronavirus [23]. They showed that mortality was significantly greater among patients with a decrease of serum H₂S levels (a cut-off point of 150.44 μM) from day 1 to day 7 greater than or equal to 36%. Second, in some diseases like various forms of critical illness and multiple forms of cancer, H₂S biosynthesis is increased due to upregulation of H₂S-synthesizing enzymes [16]. But, there is a lack of understanding of the ideal level of H₂S in physiology and in therapy, as well as the normal concentration range of H₂S in circulation. One of the main obstacles is the insufficiency of an accurate and efficient detection method of H₂S for the screening and/or identification of possible H₂S donors and inhibitors.

H₂S is a colorless and flammable gas with the unique odor of rotten eggs [24]. As H₂S is a highly lipophilic molecule, it can easily penetrate the plasma membrane of all cells without any specific transporter or receptor [25]. Acute exposure to high amounts (more than 500 ppm) of H₂S can give rise to human death [14]. H₂S is a weak acid and easily dissolved in water with a solubility of about 80 mmol/L at 37°C [26]. Generally, it can dissociate into H⁺ and hydrosulfide anion (HS⁻), which may subsequently dissociate to H⁺ and sulfide anion (S₂⁻) in aqueous solution. Because the two acid dissociation constants, pK_{a1} and pK_{a2}, of this reaction are 6.9 and >11, respectively, H₂S is present in the approximate ratio of 20% H₂S and 80% HS⁻ at physiological pH [27, 28]. Nevertheless, it remains unclear whether H₂S, HS⁻, or both are biologically active. In addition to these free H₂S such as H₂S gas, HS⁻, and S₂⁻, H₂S can exist in other bound sulfide pools in the biological systems including the acid-labile, alkaline-labile, and reducible sulfur, which are different from the conditions under which free H₂S is released [29, 30]. For example, acid-labile sulfide which is derived from iron-sulfur centers in mitochondrial enzymes [27] releases H₂S under an acidic condition (pH < 5.0). And H₂S is released from bound sulfane sulfur under reducing conditions including excess reduced glutathione (GSH), L-cysteine (Cys), and dithiothreitol (DTT) [27]. These complicated chemical species make it difficult to accurately measure free H₂S in biological systems. Actually, a lot of reports do not distinguish between the three most important biologic pools of labile sulfur: free H₂S, acid-labile, and DTT-labile sulfide, which may have completely different biological functions [31]. In addition, the volatility of H₂S adds complications to experiments [32]. For example, there are some published reports showing that half of H₂S can be rapidly released from culture medium in tissue culture wells within 5 min and in an even shorter time in a bubbled tissue bath [33, 34]. This may have in part contributed to large variations on the reported level of H₂S in plasma, tissues, and certain experiments [3, 35, 36].

During the past decade, several analytical methods such as methylene blue assay [37], gas chromatography [35], and sulfide-selective electrode [7] have been developed to detect H₂S in biological tissues or fluids. Earlier studies using the methylene blue assay reported that the H₂S level was 26–300 μM in mammalian plasma. However, later studies have

shown that the high level of H₂S may be attributed to the use of a strong acid in the methylene blue method [38, 39], because H₂S can be released from acid-labile sulfur under a condition of a strong acid. Other methods have also been utilized to measure plasma H₂S levels in the rat: sulfide-selective electrode showed approximately 50 μM, and gas chromatography–mass spectrometry showed approximately 80 μM [40]. As gas chromatography is evidently sensitive and specific, we think that it can be helpful for the detection of low physiological H₂S levels. As a result, the large discrepancy among various reports may be attributed to the following reasons: (1) complications to experiments due to the intrinsic properties of H₂S including the instability of sulfide, its high volatility, its great susceptibility to oxidation, and its adherence to various materials (for example, glass); (2) improper experimental conditions such as the wrong release of sulfide out of some rubbers used; (3) there were no distinction between free H₂S, acid-labile, and DTT-labile sulfide; (4) different H₂S levels according to age, tissue, and species; and (5) different measuring methods.

To understand biological roles of free H₂S in health and disease state and develop H₂S-based therapies, it is necessary to detect the exact concentration of free H₂S within biological systems. As the active sites for various biological effects of H₂S are inside the cell, the H₂S level measured either in plasma or in homogenized tissue is not reflective of its cellular site of action [40]. To get more insight in their physiological roles, it is necessary to measure the H₂S level released from cells. However, it is difficult to apply conventional methods, such as methylene blue assay, gas chromatography, and sulfide-selective electrode, to live cells owing to their destructive nature. Therefore, there had been a need for the development of novel methods for the accurate and reliable detection of endogenous free H₂S within live cells.

Recently, a few studies regarding simple, facile, and inexpensive detection methods for the reliable detection of free H₂S in live cells have been reported. In this review, we have focused on recent developments in H₂S-sensing methods for live cells. We summarize the key characteristics of the analytical tools, cell types, experimental conditions for H₂S production, and H₂S concentration. In addition, the advantages and limitations of these methods are presented to provide a guideline for researchers to measure the H₂S levels released from live cells.

2. Spectrophotometric Methods for Live Cell Analysis

Among all the reaction-based spectrophotometric methods, the methylene blue method is the traditional standard. This method was introduced by Fischer in 1883 [41] and has been utilized for H₂S determination in many studies. As H₂S is very volatile and can be easily oxidized, sample preparation using Zn²⁺ is generally required for the stabilization of H₂S. In this method, the acidic condition is generally employed to liberate H₂S from zinc sulfide complex. Subsequently, H₂S reacts with N,N-dimethyl-p-phenylene diamine (N,N-dpd) in the presence of an oxidizing agent (usually FeCl₃), producing methylene blue that strongly

absorbs at 670 nm [42]. The absorbance is proportional to sulfide concentration. This method has been approved by the US Environmental Protection Agency as a standard method for sulfide quantization [42]. But the use of toxic and corrosive agents was the main limiting factor in terms of disturbing the application of this method to live cells.

Kartha et al. [43] reported enhanced detection of H₂S generated in cell culture using an agar trap method (in situ methylene blue assay). As shown in Figure 1, they cultured the cells in 50 mL cell culture flasks that had a preset layer of zinc-agar on the nonadherent surface inside each flask. The H₂S produced following incubation of the cell culture with H₂S-releasing compounds was trapped as zinc sulfide in the zinc-agar layer. At the end of the incubation, media were carefully eliminated from the flasks without disturbing the agar layer. Then, the flasks were orientated with the agar layer, and H₂S trapped in the agar as zinc sulfide was released and analyzed in situ utilizing a modified methylene blue reaction. This method was used to determine the activity of H₂S-generating enzymes in intact cells, following a 24 h incubation of human endothelial-like immortalized cells Ea.hy 926 or rat vascular smooth muscle cells A10 with 1000, 3000, and 5000 μM Cys. As a result, A10 cells showed about twice the activity of H₂S-generating enzymes, compared to Ea.hy 926 cells. The authors suggested that H₂S-generating enzymes showed different activity depending on the cell type.

The modified methylene blue-based method is inexpensive, nondestructive, adaptable to most lab settings, and more convenient than conventional methylene blue method. But the method still requires several complex steps including medium elimination, H₂S release, and analysis of methylene blue. In addition, this method is also vulnerable to interference with colored substances, lowering its sensitivity.

Fu and Duan [44] reported a sensitive and selective method for H₂S detection based on in situ formation of silver nanoparticles (AgNPs) on the Ag₂S NP surface (Ag₂S@Ag) in a layer-by-layer polyelectrolyte multilayer film using Ag amplification. The UV absorption at 430 nm showed a good linear relationship with the concentration of Na₂S ranging from 10 nM to 5 mM. They measured free H₂S gas generated from live liver cancer cell line HepG2 cells after treatment with Cys and pyridoxal phosphate (PLP). Although this method is sensitive and shows a wide linear range for H₂S, it is necessary to long extra reaction time (2 h) for Ag amplification.

Ahn et al. [45] suggested a simple and cost-effective colorimetric system for selective H₂S detection in live cells utilizing a Ag-embedded Nafion/PVP membrane applied onto a polystyrene microplate cover. The basic principle of H₂S gas detection is the reaction between Ag and sulfide to form brown-colored Ag₂S. A schematic illustration of the colorimetric assay is shown in Figure 2. This assay detected H₂S release in live C6 glioma cells under stimulation of S-adenosyl methionine (SAM), thus confirming the activating effect of SAM and two substrates, Cys and homocysteine (hCys), on the pathway of cystathionine β-synthase- (CBS-) dependent H₂S production [45]. Consequently, SAM-stimulated H₂S release (10.82 ± 1.66 μM) in C6 glioma cells

treated with both Cys and hCys was higher compared with the H₂S production by Cys alone (8.27 ± 0.83 μM, *p* < 0.05) or without Cys and hCys (1.16 ± 0.80 μM, *p* < 0.001) (Figure 2). The results were similar to the Western blot analysis of CBS expression. Using this microplate cover-based colorimetric assay, Kim et al. [46] analyzed the H₂S-releasing properties of seven different H₂S donors, including sodium sulfide (Na₂S), NaHS, diallyl disulfide, diallyl trisulfide, sodium thiosulfate (Na₂S₂O₃), morpholin-4-ium 4-methoxyphenyl-morpholino-phosphinodithioate (GYY4137), and Lawesson's reagent. Besides, Youness et al. [47] utilized this assay to measure the H₂S levels released from MDA-MB-231 and MCF7 after silencing of CBS and cystathionine γ-lyase (CSE).

Although this method was the first report on the measurement of free H₂S release in live cells utilizing the simple colorimetric method, the concentrations of SAM, Cys, and hCys used in this study were not physiological. But, this assay may be more helpful to explore the potential for Cys analogs and prodrugs to promote cytoprotection through the H₂S pathway.

Zeng et al. [48] developed a colorimetric method for detection of H₂S using gold (Au)/AgI dimeric NPs as optical probes. When Au/AgI NPs were reacted with H₂S, AgI was changed to Ag₂S, causing a shift in the plasmonic band of the AuNPs. The color and absorption changes were observed by naked eyes or measured by UV-vis spectroscopy (Figures 3(a) and 3(b)). In addition, the Au/AgI NPs were immobilized in agarose gels as test strips. These agarose gels were placed on the inner surface of the culture plate cover and then used for HepG2 cell culture. As a result, the H₂S concentration was calculated as 167 nmol h⁻¹ · 10⁻⁶ cells after treatment with Cys (2 mM) and PLP (0.5 mM) for 24 h (Figure 3(c)).

3. Fluorescence Detection and Imaging for Live Cell Analysis

Recently, small-molecule fluorescent probes have been attracted attention as an effective tool for detection and imaging of H₂S in biological specimens such as tissues or cells due to their nondestructive property. The H₂S-responsive fluorescent probes are mainly divided into four different categories depending upon their reaction types such as azide-to-amine reduction, nitro-to-amine reduction, copper sulfide precipitation, and nucleophilic addition [49, 50]. Early work in this field utilized the selective H₂S-mediated reduction of azides and sulfonylazides, respectively, to develop first-generation reagents for fluorescence H₂S detection [51]. Since then, Lin et al. [52] reported a family of azide-based fluorescent H₂S indicators which had enhanced sensitivity and cellular trappability. In particular, sulfidefluor-7 acetoxymethyl ester enabled direct and real-time visualization of endogenous H₂S release in live human umbilical vein endothelial cells under stimulation with vascular endothelial growth factor. And Yang et al. [50] developed a red-emitting fluorescent probe for H₂S using the reduction of the azido group. This probe represented a

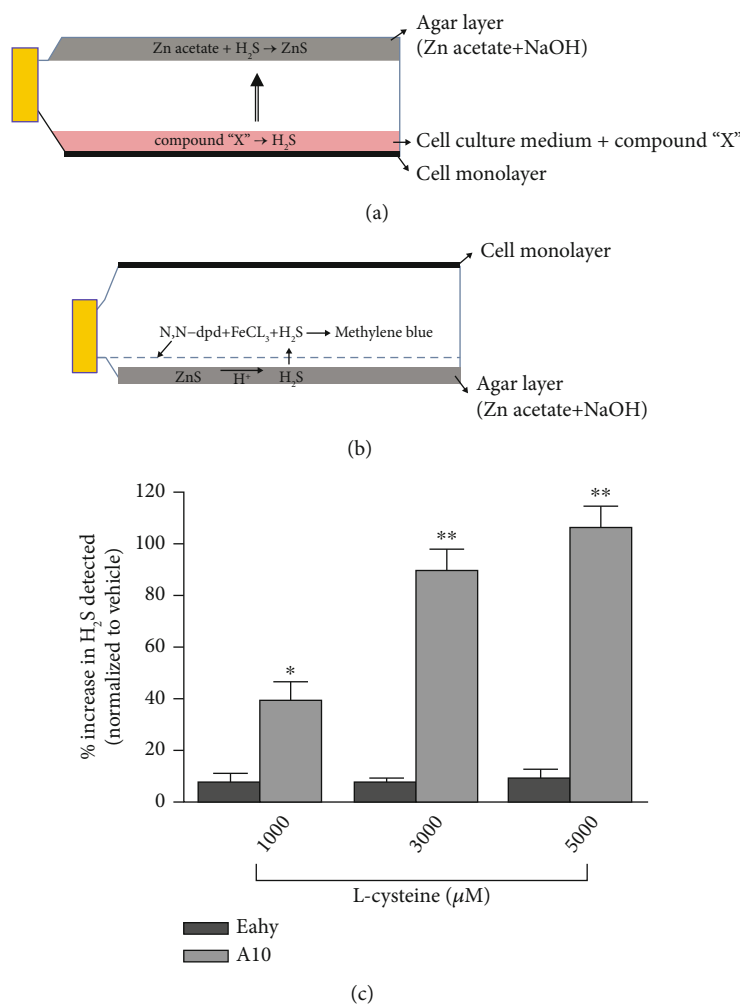


FIGURE 1: Schematic illustration of the experimental setup for H₂S trapping method in (a) cell culture system and (b) in situ methylene blue assay. (c) Comparison of amount of H₂S generated from Ea.hy 926 (checked bars) and A10 (dotted bars) cells. Cells were treated with 1000, 3000, and 5000 μM L-cysteine for 24 h. Reprinted from [43] with permission from publisher.

striking fluorescence enhancement (10-fold) with a large Stokes shift (125 nm), and the detection limit was 5.7 nM. They detected exogenous and endogenous H₂S in live HeLa cells.

Furthermore, Cheng et al. [53] developed a probe for the fluorescence switch-on detection of H₂S, by employing dinitrophenyl ether functionality as both a fluorescence quencher and an H₂S-reaction trigger. It was easily synthesized via nucleophilic substitution of 3,4-dinitrofluorobenzene by the BODIPY fluorophore 1. Its ability to image H₂S in live cells was demonstrated using HeLa cells and NaHS as the H₂S source. For specific and sensitive imaging of H₂S in the cellular lysosome, Wu et al. [54] developed activable fluorescence nanoprobe-based quantum dots. This nanoprobe consisted of p-amino thiophenol-capped AgNPs and thioglycolic-acid-stabilized quantum dots, called QD/AgNP nanocomplexes. The detection limit of this nanoprobe was 15 nM. And they showed high ability to enter into cellular lysosome in live HeLa cells.

Compared with “turn-on” fluorescent probes, ratiometric fluorescent probes have been proposed to be more accurate for detecting H₂S, independently of variables in

quantitative analysis including variations of excitation intensity, environmental factors, light scattering, and concentration of probe [55]. An et al. [56] reported the quinoline quaternary ammonium salt derivative-based ratiometric fluorescent probe (referred to as QL-N₃). The QL-N₃ probe exhibited two fluorescence emission peaks at 525 and 605 nm with different excitation wavelengths of 385 and 521 nm, and the ratio between fluorescence intensities of two peaks was positively related with the H₂S concentration. This probe could image the changes in exogenous and endogenous H₂S in live HeLa cells.

Nevertheless, there are still several challenging issues in the development of fluorescent probes for H₂S as follows: (i) selectivity over interfering biothiols including GSH, Cys, and hCys; (ii) high sensitivity enough to detect the endogenously produced H₂S; (iii) fast response within a few minutes; (iv) biocompatibility including low toxicity, cell permeability, and intracellular stability; and (v) signaling in the biological optical window. Recently, Singha et al. [57] reported a two-photon fluorescent probe for H₂S which belonged to a Michael acceptor system. They approached the selectivity issue by optimizing the electronic and steric

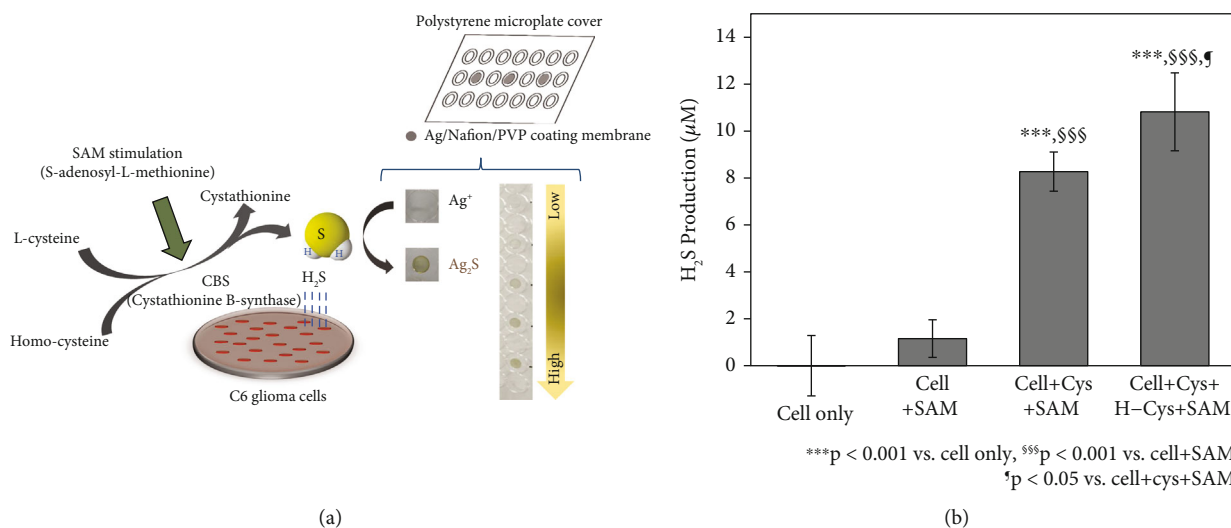


FIGURE 2: (a) Schematic illustration of microplate cover-based colorimetric assay utilizing the Ag/Nafion/PVP membrane. (b) The concentration of endogenous H₂S production in live C6 glioma cells after treatment with s-(5'-adenosyl)-L-methionine (SAM, 2.5 mM), L-cysteine (Cys, 10 mM), and L-homocysteine (hCys, 0.5 mM) treatment in a 5% CO₂ incubator at 37°C. Reprinted from [45] with permission from publisher.

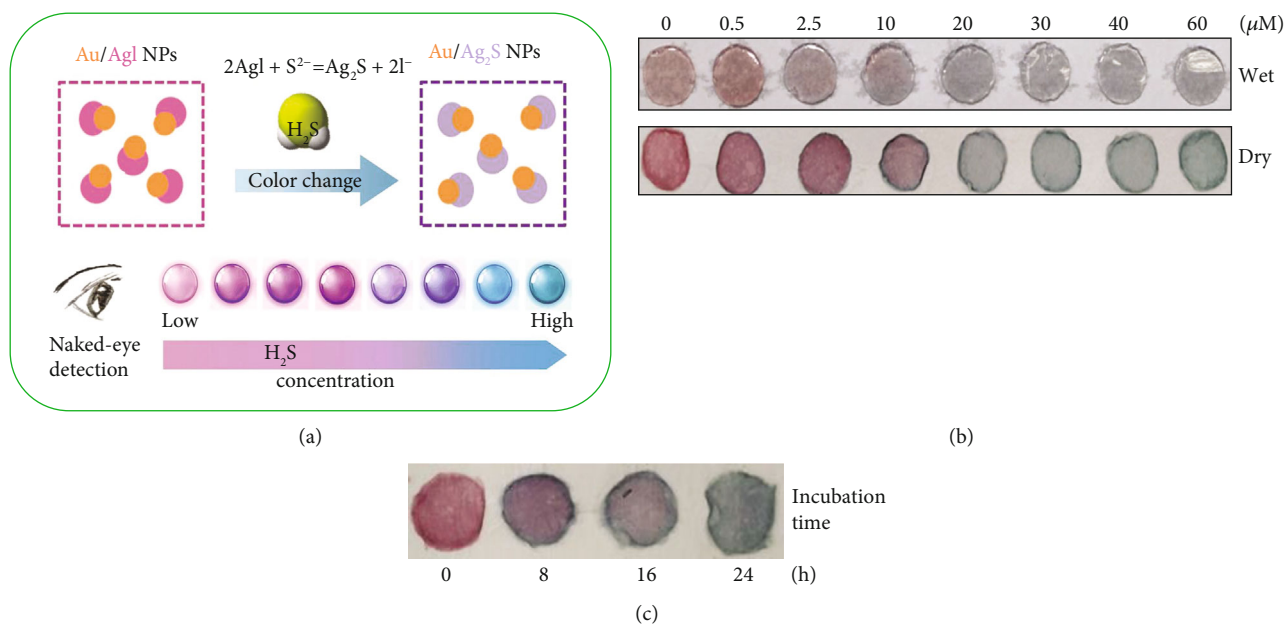


FIGURE 3: (a) The sensing principle of Au/AgI dimeric nanoparticle-based colorimetric assay for detection of H₂S. Photographic images of agarose gels incubated (b) with different concentrations of S²⁻ from 0 to 60 μM and (c) from cell culture for 0, 8, 16, and 24 h. Reprinted from [48] with permission from publisher.

interactions between biothiols and the probe, in addition to gaining very high sensitivity, biocompatibility, and fast response time.

Fluorescence-based detection provides excellent sensitivity, high selectivity, and real-time H₂S monitoring not only within living cells but also within subcellular organelles. Therefore, the progress of H₂S-specific fluorescence probes is regarded one of the fastest-growing areas in the field of H₂S biology [58]. Although fluorescence-based detection has attracted immense attention for detecting H₂S inside living cells, it is essential to use expensive instruments and spe-

cial H₂S probes for live cell fluorescence detection. So, it may be difficult to utilize this method in many labs.

4. Surface-Enhanced Raman Scattering for Live Cell Analysis

Surface-enhanced Raman spectroscopy (SERS) is a promising ultrasensitive spectral analysis technique because of its high selectivity, based on molecular fingerprinting and sensitivity, even at single-molecule detection levels [59–61]. With strong electromagnetic fields and surface chemistry

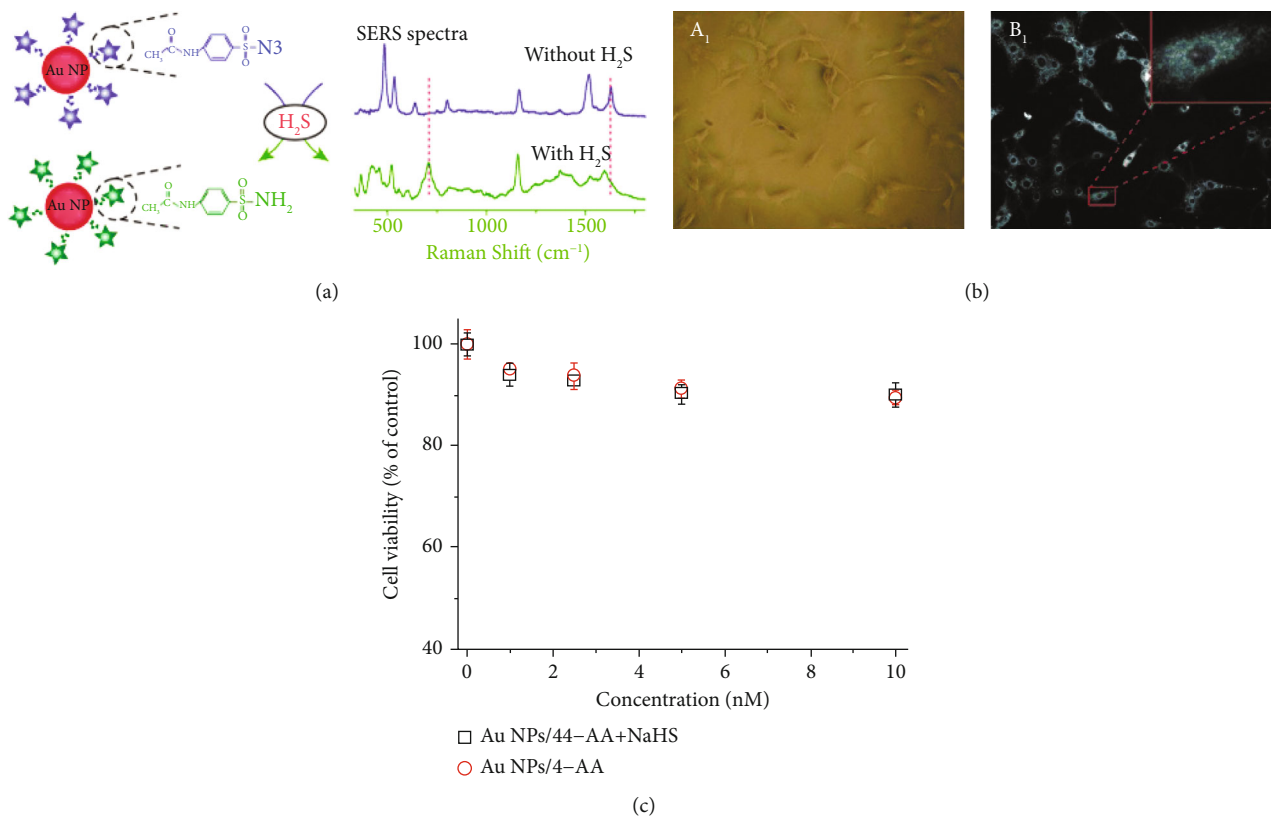


FIGURE 4: (a) Sensing mechanism of SERS nanosensors for detection of endogenous H_2S in living cells. (b) Bright-field microscopy (A1) and dark-field microscopy (B1) images of rat C6 glioma cells after 4 h incubation with AuNPs/4-AA. (c) Cytotoxicity of different concentrations of AuNPs/4-AA in the presence and absence of NaHS after incubation of 48 h. Reprinted from [66] with permission from publisher.

enhancements, SERS can increase the original Raman signal to 10^6 orders or more [62]. Recently, the SERS sensors have been utilized for the analysis of a variety of substances including DNA, protein, metal ions, and pesticides [63–65].

Li et al. [66] reported a novel SERS nanosensor fabricated by functionalizing AuNPs with 4-acetamidobenzenesulfonyl azide (AuNPs/4-AA) for detecting the endogenous H_2S in live cells (Figure 4(a)). The detection was performed with SERS spectrum changes in AuNPs/4-AA coming from the reaction of H_2S with 4-AA on AuNPs (transformation of the azide groups of 4-AA into amino groups). AuNPs/4-AA responded to H_2S within 1 min with a $0.1 \mu\text{M}$ level of sensitivity. Using SERS nanosensor, the H_2S concentration in living glioma cells was found to have approximately 10-fold increase after 2 h stimulation of SAM, confirming that SAM can activate CBS to improve its catalytic ability to produce H_2S . Besides, the viability of glioma cells after the addition of AuNPs/4-AA was higher than 88% at the concentration ranging from 1 to 10 nM, showing the good biocompatibility of AuNPs/4-AA (Figures 4(b) and 4(c)).

And Zhang et al. [67] proposed a smart SERS nanoprobe, Au core-4-mercaptobenzonitrile-Ag shell NP (Au@4-MBN@Ag), for detection of endogenous H_2S in live HepG2 cells. As sulfide in the solution selectively reacted with Ag to transform Ag_2S at room temperature, the SERS intensity of 4-MBN gradually decreased with increasing concentration

of H_2S . It showed a good linearity in the sulfide concentration ranging from 0.05 to $500 \mu\text{M}$, and a detection limit was 0.14 nM .

Though SERS has important advantages over fluorescence-based methods, such as resistance to photobleaching and phototoxicity, and narrow emission peaks for spectral multiplexing, it is still necessitated to develop H_2S -specific SERS probes due to the difficulty in direct sensing of inorganic species. In addition, SERS require expensive instrument—Raman spectrophotometer.

5. Paper-Based Colorimetric Assay for Live Cell Analysis

Paper-based sensors have received great attention in the development of point-of-care (POC) diagnostics owing to the simple fabrication, cost-effectiveness, and user-friendly characteristics. The distinct properties of paper which enable passive liquid transport and compatibility with chemicals or biochemicals are the main reasons on which paper is utilized as a sensing platform [68]. In addition, the white paper is suitable for colorimetric detection because it gives strong contrast with a colored substrate [69]. So, it enables readers to check the results with the naked eye.

Rosolina et al. [70] reported a bismuth-based disposable sensor using a wet, porous, and paper-like substrate coated with alkaline bismuth hydroxide, $\text{Bi}(\text{OH})_3$. The alkaline,

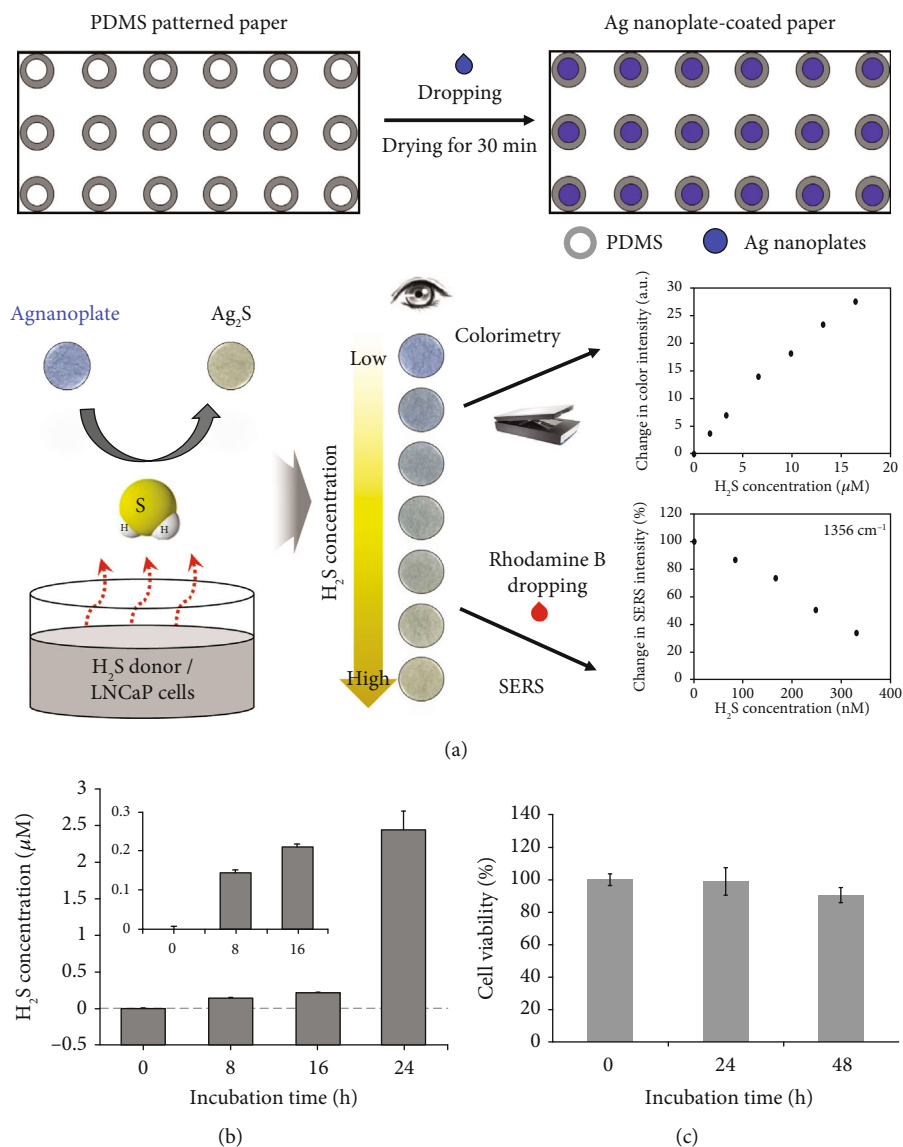


FIGURE 5: (a) Schematics of the preparation of Ag nanoplate-based H₂S-sensing paper and its principle of colorimetric and SERS dual-mode detection of H₂S. (b) The concentration of endogenous H₂S release from live LNCaP cells, varying with incubation times. (c) Evaluation of cellular toxicity of 5 mM L-cysteine (Cys) and 1 mM homocysteine (hCys) cotreatment in LNCaP cells. After incubation for 24 and 48 h, cellular toxicity was measured using water-soluble tetrazolium salt (WST) assay and expressed as a percentage of the control without Cys or hCys. Reprinted from [79] with permission from publisher.

wet coating helped the trapping of acidic H₂S gas and its reaction with Bi(III) species, forming colored Bi₂S₃ (yellow/brown). This sensor responded to ≥ 30 ppb H₂S in a total volume of 1.35 L of gas. However, its alkalinity (pH 11) required special care in handling. And this sensor should be kept inside an inert gas to prevent neutralization by acidic CO₂ in air. Carpenter et al. [71] reported a new probe using the copper(II) complex of 1-(2-pyridylazo)-2-naphthol (Cu-PAN) on the same paper-like substrate. The reaction between H₂S gas and the copper complex led to a striking change in color from purple to yellow/orange. This color change was visible to the naked eye at the concentration as low as 30 ppb H₂S in a 1.35 L of gas which was considered as a typical volume of human breath. They suggested that the colorimetric paper probe is easy to fabricate, cost-effective,

disposable, and a green alternative to the commonly used lead acetate test papers. Recently, Ahn et al. [72] developed a rapid and simple colorimetric paper sensor using an etching-resistant effect on Ag nanoprisms. The detection principle was that Ag NPRs on the paper reacted with H₂S gas to form Ag₂S on their surfaces, which induced etching-resistant Ag NPRs against Cl⁻ ions. As a result, the color of Ag NPR-coated paper varied from yellow to purplish brown, depending on the concentration of H₂S gas after KCl treatment. This H₂S-sensing paper showed good sensitivity with a linear range of 1.03 to 32.9 μM H₂S and a fast response time of 1 min. The authors suggested that it could be utilized as a simple and reliable tool for on-site detection of H₂S gas for quality check of dietary supplements and human breath analysis. However, the H₂S-sensing papers

TABLE 1: Some characteristics and H₂S levels of recently reported methods for the detection of free H₂S in live cells.

Analytical methods for H ₂ S	Cell types	Experimental conditions for H ₂ S production	H ₂ S concentration	Advantages & limitations	Ref.	
Spectrophotometric method	<i>In situ</i> methylene blue assay	Rat A10 cells	24 h incubation with 1, 3, and 5 mM of Cys and 1, 2, and 5 mM NAC	40.67 ± 4.8 μM (with 3 mM Cys) 51.5 ± 7.5 μM (with 2 mM NAC)	(Adv.) cost-effective & adaptable to most lab. settings (Limit.) low sensitivity, several complex steps & interferences with colored substances	[43]
	Ag ₂ S@AgNPs in a layer-by-layer film	HepG2 cells	2 h incubation with Cys and PLP (+Ag amplification for 2 h)	—	(Adv.) high sensitivity (10 nM) (Limit.) additional amplification time (2 h), except for the reaction time (2 h)	[44]
	Ag-embedded Nafion/PVP membrane	Rat C6 glioma cells	SAM treatment and with a combination of Cys (10 mM) and hCys (0.5 mM) for 48 h	10.82 ± 1.66 μM	(Adv.) simple, facile, cost-effective, & adaptable to most lab. settings (Limit.) low sensitivity & treatment with high levels of substrates	[45]
	Au/AgI dimeric NPs	HepG2 cells	24 h incubation with 2 mM of Cys and 0.5 mM PLP	167 nmol h ⁻¹ · 10 ⁻⁶ cells	(Adv.) good sensitivity (500 nM) (Limit.) long response time	[48]
Fluorescence detection and imaging	Reduction of the azido group (λ _{em} = 610 nm)	HeLa cells	Prestimulation with 100 μM of SNP (NO donor) for 60 min	—	(Adv.) high sensitivity (LOD 5.7 nM) (Limit.) only fluorescence cell imaging & long response time	[50]
	Reduction of azide to amine (λ _{em} = 526 nm)	HUVECs	Stimulation with VEGF (40 ng/mL) for 30 min	Intracellular fluorescence ratio ¹ F _f /F _i ≈ 1.27 (vs. 1.07 of control)	(Adv.) good sensitivity (LOD 500 nM) (Limit.) long response time	[52]
	Nucleophilic cleavage of the ether bond (λ _{em} = 570 nm)	HeLa cells	1 h incubation with NaHS (100, 200, and 300 μM)	—	(Adv.) good sensitivity (LOD 500 nM) (Limit.) only fluorescence imaging & long response time	[53]
	H ₂ S-triggered disassembly of QDs/AgNP complexes (λ _{em} = 530 nm)	HeLa cells	Pretreatment with 300 μM NaHS for 30 min	—	(Adv.) high sensitivity (LOD 15 nM) (Limit.) only fluorescence imaging & long response time	[54]
	Reduction of azide to amino group	HeLa cells	60 min incubation with Na ₂ S (100 μM)	The fluorescence intensity (red, 605 nm)/blue, 525 nm) ratio 2.416 (vs. 1.498 of untreated cells)	(Adv.) enhanced detection accuracy (ratiometric analysis) (Limit.) long response time	[56]
Surface-enhanced Raman scattering	Rat C6 glioma cells & human U251 MG glioma cells	SAM stimulation	Ratiometric Raman peak intensity I ₇₀₉ /I ₁₁₆₁ : about 10-fold increase after 2 h stimulation of SAM	(Adv.) good sensitivity (LOD 0.1 μM) & fast response time (1 min) (Limit.) needs specific SERS probe and expensive instrument	[66]	

TABLE 1: Continued.

Analytical methods for H ₂ S	Cell types	Experimental conditions for H ₂ S production	H ₂ S concentration	Advantages & limitations	Ref.	
Paper-based colorimetric assay	LNCap cells	72 h incubation with 5 mM of Cys and 1 mM hCys	17.48 ± 3.80 μM (72 h)	(Adv.) simple, low-cost, practical, & moderate sensitivity (LOD 1.4 μM) (Limit.) treatment with high levels of substrates	[73]	
Dual-mode detection	Colorimetry & near-IR fluorescence	MCF-7 cells	30 min incubation with NaHS (10, 50, and 100 μM)	—	(Adv.) moderate sensitivity (LOD 3.09 μM) & fluorescence “off-on” response (Limit.) only fluorescence imaging	[77]
	Colorimetry & fluorescence	NHOF cells	0, 20, and 60 min incubation with NaHS (10 μM)	—	(Adv.) good sensitivity (LOD 0.17 μM) (Limit.) only fluorescence imaging	[78]
	Colorimetry & SERS	LNCap cells	Cys (5 mM) and hCys (1 mM) treatment for 8, 16, and 24 h	0.144 ± 0.007 μM (8 h) 0.211 ± 0.007 μM (16 h) 2.45 ± 0.26 μM (24 h)	(Adv.) high sensitivity (LOD 15 nM for SERS detection & LOD 520 nM for colorimetry) (Limit.) treatment with substrates	[79]
	Colorimetry & luminescence	HeLa cells	30 min incubation with H ₂ S (200 μM)	—	(Adv.) high sensitivity (LOD 53.9 nM) (Limit.) only luminescence imaging	[80]

¹F_i: initial mean fluorescence intensity; F_f: final mean fluorescence intensity. Cys: L-cysteine; NAC: N-acetylcysteine; VEGF: vascular endothelial growth factor; SNP: sodium nitroprusside; HUVEC: human umbilical vein endothelial cells; SAM: s-adenosyl methionine; hCys: homocysteine. PVP: polyvinylpyrrolidone; QDs: quantum dots; AgNP: silver nanoparticle; PLP: pyridoxal phosphate; A10: vascular smooth muscle cell; NHOF: normal human oral fibroblast.

mentioned above were not confirmed whether they worked properly at cell culture environment or in live cells.

Lee et al. [73] reported a new paper-based colorimetric assay by fabricating a 96-well microplate format for cell culture for sensing H₂S gas in live cancer cells. Microplate-like hydrophobic walls designed using AutoCAD were printed using a Xerox ColorQube 8570 N printer. Wax-printed paper (Whatman grade 1 chromatography paper) consisted of 96 circular reservoirs with a 4 mm detection zone. A PVP membrane containing Ag/Nafion was coated on the H₂S detection zones. Finally, this paper-based colorimetric assay successfully measured the difference in endogenous H₂S level between live prostate cancer LNCaP and PC-3 cells, which showed differential expression of H₂S-producing enzymes. Though this paper-based assay was simple, inexpensive, and feasible, it could not measure the low concentration of H₂S released from LNCaP cells within 24 h.

6. Dual-Mode Detection for Live Cell Analysis

Although colorimetry-based sensing method is a simple and rapid technique for POC diagnostics and high-speed bioana-

lysis, the detection limit or sensitivity may be disappointing [74]. The low sensitivity can somewhat restrict the application of colorimetric methods for quantitative analysis of endogenous H₂S in live cells under shorter time. A dual-mode detection based on colorimetry and other sensing methods such as SERS, fluorescence, or wettability can improve the sensitivity of conventional colorimetric sensors, because they give two different types of output signals covering a wider detection range [75]. Gahlaut et al. [76] introduced a dual-mode H₂S detection combined colorimetric principle and wettability of Ag nanorod arrays on glass substrates. The surface color and water wetting properties of nanorods were found to be highly sensitive toward the H₂S gas environment (5 ppm of gas with an exposure time of only 30 s). Together with high sensitivity and selectivity, the response time was found to be significantly low (within 5 s). The authors suggested that this method could be applied for the future study of H₂S release from biosystems (live cells), as well as art conservation.

Zhong et al. [77] reported a colorimetric and near-infrared fluorescent probe (L) with a donor-π-acceptor structure derived from 4-diethylaminosalicylaldehyde and

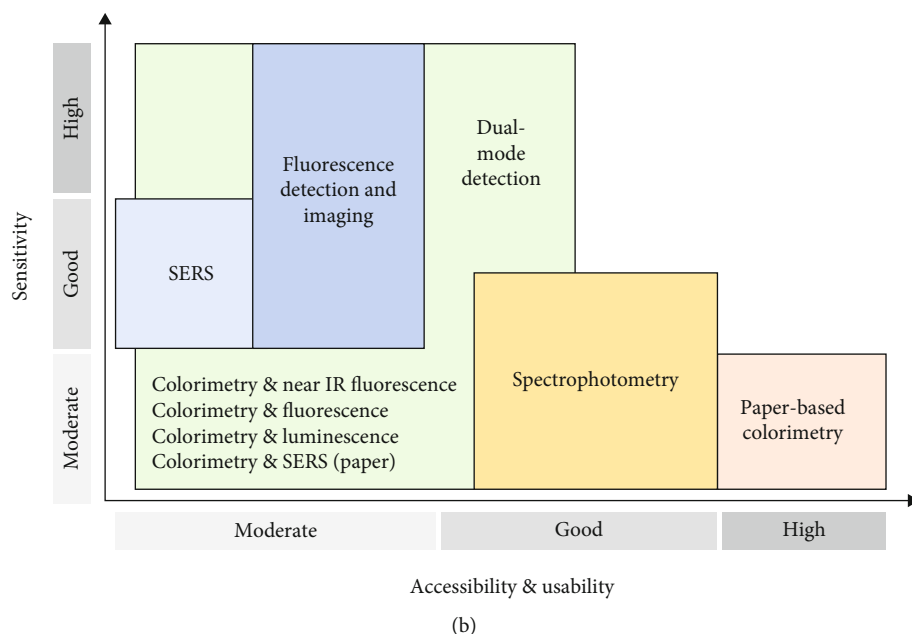
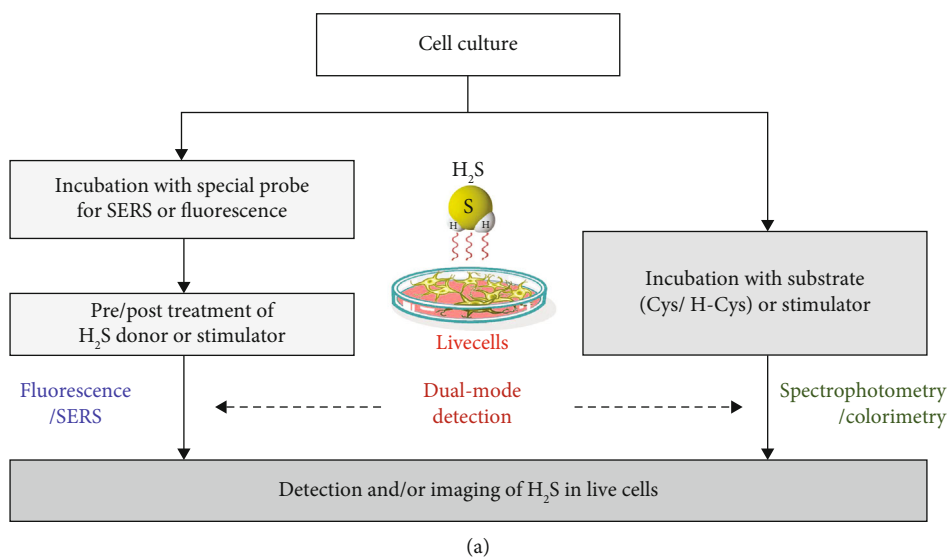


FIGURE 6: (a) Schematic diagram of the H_2S detection process in live cells. (b) Comparison plot of sensitivity versus accessibility & usability of various analytical methods for detection of H_2S in live cells.

2-(3-cyano-4,5,5-trimethylfuran-2(5H)-ylidene)malononitrile. A distinct color change of L solution from colorless to bluish-purple took place after treatment with H_2S . Using this probe, they detected H_2S vapor, H_2S in water and wine samples, and H_2S imaging in live MCF-7 cells. Besides, Paul et al. [78] reported a colorimetric and fluorescence turn-off probe 10-(4-azido phenyl)-5,5-difluoro-5h-dipyrrolo[1,2-c:1',2'-f] [1-3] diazaborinin-4-ium-5-uide, 1, for selective detection of H_2S . The detection limit of this probe was $0.17 \mu M$ for H_2S . They successfully detected exogenous H_2S in live normal human oral fibroblast (NHOF) cells.

In addition, Ahn et al. [79] suggested a colorimetric and SERS-sensing system using Ag nanoplates on the paper. This dual-mode system could be helpful for detection of low concentrations of H_2S in live cells because SERS could greatly

improve the detection limit (Figure 5(a)). As a result, this simple paper assay could measure H_2S with wider ranging from nano- to micromolar levels. And it was able to measure endogenous H_2S in live LNCaP cells even at 8 h of incubation after cotreatment with Cys (5 mM) and hCys (1 mM) (Figure 5(b)). Besides, the viability of LNCaP cells was greater than 90% for 48 h, indicating good cell proliferation of live cells (Figure 5(c)).

Liu et al. [80] reported a ruthenium (Ru) (II) complex-based probe for colorimetric and luminescent detection and imaging of H_2S in live cells and organisms. This Ru(II) complex was yellow color and nonluminescent in aqueous solution. But, when it reacted with H_2S , the color of the solution changed from yellow to pink for colorimetric analysis and the emission intensity was about 65-fold increased for

luminescent analysis. As this probe had low cytotoxicity and good permeability to cell membrane, it could be utilized for luminescence imaging of H₂S in live HeLa cells.

Until now, we described the recent developments in H₂S detection methods for live cell analysis. Table 1 summarized some characteristics, H₂S levels, advantages, and limitations of recently reported methods for the detection of free H₂S from live cells. And Figures 6(a) and 6(b) show a schematic diagram of the H₂S detection process in live cells without causing any destruction to the cells and a comparison plot of sensitivity versus accessibility of various analytical methods, respectively.

7. Conclusions

In this review, we have summarized the methods for the detection of H₂S in live cells without causing any destruction to the cells. To measure H₂S level in conventional biology, an indirect method that analyzes the expression of H₂S synthase, such as CBS and CSE, in cell lysate or tissue homogenate using Western blot analysis is widely used. But the total analysis time for the Western blot from cell seeding is approximately 7 days including all the necessary processes such as treatment of substrates and cell lysis. To eliminate or reduce the complicated and labor-intensive analytical approach, several methods that are simple, efficient, and reliable for detection of H₂S in live cells have been developed.

Microplate cover-based and paper-based colorimetric assays utilizing Ag/Nafion/PVP membrane can quantitatively analyze the endogenous H₂S levels in live cancer cells, without expensive instruments and special H₂S probes. Although these colorimetric assays are simple, easy to use, and cost-effective, they still have the limitation of low sensitivity. Recent developments in fluorescent probes for reactive sulfur species can further facilitate analysis based on fluorescence bioimaging technology. But, there are still challenging issues including high sensitivity and selectivity in the presence of many interfering biomolecules, water solubility, and low cytotoxicity. And SERS detection using H₂S-responsive SERS probe shows great promise for the real-time monitoring of H₂S produced in live cells, though it requires a Raman spectrophotometer. The dual-mode detection based on colorimetry and other detection methods such as SERS or fluorescence can improve the sensing performance such as high sensitivity and wide detection range, as well as easy to use. And further work for developing the highly sensitive, specific, biocompatible, and reproducible detection method is required to measure free H₂S in live cells in the absence of additional substrates or stimulator.

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this paper.

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References

- [1] C. Szabo, "Hydrogen sulphide and its therapeutic potential," *Nature Reviews Drug Discovery*, vol. 6, no. 11, pp. 917–935, 2007.
- [2] Q. Xiao, J. Ying, L. Xiang, and C. Zhang, "The biologic effect of hydrogen sulfide and its function in various diseases," *Medicine*, vol. 97, no. 44, article e13065, 2018.
- [3] X. Cao, L. Ding, Z. Xie et al., "A review of hydrogen sulfide synthesis, metabolism, and measurement: is modulation of hydrogen sulfide a novel therapeutic for Cancer?," *Antioxidants & Redox Signaling*, vol. 31, no. 1, pp. 1–38, 2019.
- [4] K. Abe and H. Kimura, "The possible role of hydrogen sulfide as an endogenous neuromodulator," *Journal of Neuroscience*, vol. 16, no. 3, pp. 1066–1071, 1996.
- [5] Z. Li, D. J. Polhemus, and D. J. Lefer, "Evolution of hydrogen sulfide therapeutics to treat cardiovascular disease," *Circulation Research*, vol. 123, no. 5, pp. 590–600, 2018.
- [6] W. Zhao, J. Zhang, Y. Lu, and R. Wang, "The vasorelaxant effect of H₂S as a novel endogenous gaseous K(ATP) channel opener," *EMBO Journal*, vol. 20, no. 21, pp. 6008–6016, 2001.
- [7] G. Yang, L. Wu, B. Jiang et al., "H₂S as a physiologic vasorelaxant: hypertension in mice with deletion of Cystathionine-Lyase," *Science*, vol. 322, no. 5901, pp. 587–590, 2008.
- [8] X. Zhang and J. S. Bian, "Hydrogen sulfide: a neuromodulator and neuroprotectant in the central nervous system," *ACS Chemical Neuroscience*, vol. 5, no. 10, pp. 876–883, 2014.
- [9] C. Szabo, "Hydrogen sulfide, an enhancer of vascular nitric oxide signaling: mechanisms and implications," *American Journal of Physiology Cell Physiology*, vol. 312, no. 1, pp. C3–C15, 2017.
- [10] D. K. Polhemus and D. J. Lefer, "Emergence of hydrogen sulfide as an endogenous gaseous signaling molecule in cardiovascular disease," *Circulation Research*, vol. 114, no. 4, pp. 730–737, 2014.
- [11] J. L. Wallace, G. Caliendo, V. Santagada, and G. Cirino, "Markedly reduced toxicity of a hydrogen sulphide-releasing derivative of naproxen (ATB-346)," *British Journal of Pharmacology*, vol. 159, no. 6, pp. 1236–1246, 2010.
- [12] di Villa Bianca Rd, G. Cirino, and R. Sorrentino, "Hydrogen sulfide and urogenital tract," *Handbook of Experimental Pharmacology*, vol. 230, pp. 111–136, 2015.
- [13] N. Voloshchuk, I. Taran, O. Pashynska, A. Melnyk, and S. Magdebura, "The role of hydrogen sulfide in gastrointestinal tract functioning (review)," *Current Issues in Pharmacy and Medical Sciences*, vol. 33, no. 1, pp. 45–50, 2020.
- [14] R. J. Reiffenstein, W. C. Hulbert, and S. H. Roth, "Toxicology of hydrogen sulfide," *Annual Reviews of Pharmacology and Toxicology*, vol. 32, no. 1, pp. 109–134, 1992.
- [15] C. Szabo, "A timeline of hydrogen sulfide (H₂S) research: from environmental toxin to biological mediator," *Biochemical Pharmacology*, vol. 149, pp. 5–19, 2018.
- [16] C. Szabo and A. Papapetropoulos, "International union of basic and clinical pharmacology. CII: pharmacological modulation of H₂S levels: H₂S donors and H₂S biosynthesis

- inhibitors," *Pharmacological Reviews*, vol. 69, no. 4, pp. 497–564, 2017.
- [17] N. L. Sun, Y. Xi, S. N. Yang, Z. Ma, and C. S. Tang, "Plasma hydrogen sulfide and homocysteine levels in hypertensive patients with different blood pressure levels and complications," *Zhonghua Xin Xue Guan Bing Za Zhi*, vol. 35, no. 12, pp. 1145–1148, 2007.
- [18] H. Laggner, M. K. Muellner, S. Schreier et al., "Hydrogen sulphide: a novel physiological inhibitor of LDL atherogenic modification by HOCl," *Free Radical Research*, vol. 41, no. 7, pp. 741–747, 2007.
- [19] C. K. Nicholson and J. W. Calvert, "Hydrogen sulfide and ischemia-reperfusion injury," *Pharmacological Research*, vol. 62, no. 4, pp. 289–297, 2010.
- [20] A. M. Koning, W. C. Meijers, I. Minović et al., "The fate of sulfate in chronic heart failure," *American Journal of Physiology Heart and Circulatory Physiology*, vol. 312, no. 3, pp. H415–H421, 2017.
- [21] L. Yu, W. Li, B. M. Park, G. J. Lee, and S. H. Kim, "Hypoxia augments NaHS-induced ANP secretion via KATP channel, HIF-1 α and PPAR- γ pathway," *Peptides*, vol. 121, article 170123, 2019.
- [22] S. Chaudhuri, U. Biswas, and A. Kumar, "Association of serum hydrogen sulphide with hypertension and proteinuria in pre-eclampsia," *Asian Journal of Medical Sciences*, vol. 10, no. 6, pp. 33–38, 2019.
- [23] G. Renieris, K. Katrini, C. Damoulari et al., "Serum hydrogen sulfide and outcome association in pneumonia by the SARS-COV-2 coronavirus," *Shock*, vol. 54, no. 5, pp. 633–637, 2020.
- [24] Y. D. Wen, H. Wang, and Y. Z. Zhu, "The drug developments of hydrogen sulfide on cardiovascular disease," *Oxidative Medicine and Cellular Longevity*, vol. 2018, Article ID 4010395, 21 pages, 2018.
- [25] J. C. Mathai, A. Missner, P. Kugler et al., "No facilitator required for membrane transport of hydrogen sulfide," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 39, pp. 16633–16638, 2009.
- [26] O. Kabil and R. Banerjee, "Redox Biochemistry of Hydrogen Sulfide," *Journal of Biological Chemistry*, vol. 285, no. 29, pp. 21903–21907, 2010.
- [27] K. R. Olson, "Is hydrogen sulfide a circulating "gasotransmitter" in vertebrate blood?," *Biochimica et Biophysica Acta - Bioenergetics*, vol. 1787, no. 7, pp. 856–863, 2009.
- [28] H. Kimura, "Hydrogen sulfide and polysulfides as biological mediators," *Molecules*, vol. 19, no. 10, pp. 16146–16157, 2014.
- [29] P. Nagy, Z. Pálinkás, A. Nagy, B. Budai, I. Tóth, and A. Vasas, "Chemical aspects of hydrogen sulfide measurements in physiological samples," *Biochimica et Biophysica Acta - General Subjects*, vol. 1840, no. 2, pp. 876–891, 2014.
- [30] B. Tan, S. Jin, J. Sun et al., "New method for quantification of gasotransmitter hydrogen sulfide in biological matrices by LC-MS/MS," *Scientific Reports*, vol. 7, no. 1, article 46278, 2017.
- [31] A. Tangerman, "Measurement and biological significance of the volatile sulfur compounds hydrogen sulfide, methanethiol and dimethyl sulfide in various biological matrices," *Journal of Chromatography B Analytical Technologies in the Biomedical and Life Sciences*, vol. 877, no. 28, pp. 3366–3377, 2009.
- [32] Y. Zheng, X. Ji, K. Ji, and B. Wang, "Hydrogen sulfide prodrugs—a review," *Acta Pharmaceutica Sinica B*, vol. 5, no. 5, pp. 367–377, 2015.
- [33] E. R. DeLeon, G. F. Stoy, and K. R. Olson, "Passive loss of hydrogen sulfide in biological experiments," *Analytical Biochemistry*, vol. 421, no. 1, pp. 203–207, 2012.
- [34] G. S. Oh, H. O. Pae, B. S. Lee et al., "Hydrogen sulfide inhibits nitric oxide production and nuclear factor- κ B via heme oxygenase-1 expression in RAW264.7 macrophages stimulated with lipopolysaccharide," *Free Radical Biology and Medicine*, vol. 41, no. 1, pp. 106–119, 2006.
- [35] J. Furne, A. Saeed, and M. D. Levitt, "Whole tissue hydrogen sulfide concentrations are orders of magnitude lower than presently accepted values," *American Journal of Physiology, Regulatory, Integrative and Comparative Physiology*, vol. 295, no. 5, pp. R1479–R1485, 2008.
- [36] X. Shen, C. B. Pattillo, S. Pardue, S. C. Bir, R. Wang, and C. G. Kevil, "Measurement of plasma hydrogen sulfide in vivo and in vitro," *Free Radical Biology and Medicine*, vol. 50, no. 9, pp. 1021–1031, 2011.
- [37] J. L. Wallace, L. Vong, W. McKnight, M. Dickey, and G. R. Martin, "Endogenous and exogenous hydrogen sulfide promotes resolution of colitis in rats," *Gastroenterology*, vol. 137, no. 2, pp. 569–578.e1, 2009.
- [38] N. L. Whitfield, E. L. Kreimier, F. C. Verdial, N. Skovgaard, and K. R. Olson, "Reappraisal of H₂S/sulfide concentration in vertebrate blood and its potential significance in ischemic preconditioning and vascular signaling," *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, vol. 294, no. 6, pp. R1930–R1937, 2008.
- [39] M. Ishigami, K. Hiraki, K. Umemura, Y. Ogasawara, K. Ishii, and H. Kimura, "A source of hydrogen sulfide and a mechanism of its release in the brain," *Antioxidants & Redox Signaling*, vol. 11, no. 2, pp. 205–214, 2009.
- [40] L. Li, P. Rose, and P. K. Moore, "Hydrogen sulfide and cell signaling," *Annual Review of Pharmacology and Toxicology*, vol. 51, no. 1, pp. 169–187, 2011.
- [41] E. Fischer, "Bildung von methylenblau als reaction auf schwefelwasserstoff," *European Journal of Inorganic Chemistry*, vol. 16, no. 2, pp. 2234–2236, 1883.
- [42] Z. G. Li, "Quantification of hydrogen sulfide concentration using methylene blue and 5,5'-dithiobis(2-nitrobenzoic acid) methods in plants," in *Hydrogen sulfide in redox biology, Part A, Methods Enzymol*, E. Cadenas and L. Packer, Eds., pp. 101–110, Elsevier, Amsterdam, Netherland, 2015.
- [43] R. V. Kartha, J. Zhou, L. B. Hovde, B. W. Y. Cheung, and H. Schröder, "Enhanced detection of hydrogen sulfide generated in cell culture using an agar trap method," *Analytical Biochemistry*, vol. 423, no. 1, pp. 102–108, 2012.
- [44] H. Fu and X. Duan, "Highly sensitive and colorimetric detection of hydrogen sulphide by in situ formation of Ag₂S@Ag nanoparticles in polyelectrolyte multilayer film," *RSC Advances*, vol. 5, pp. 3508–3511, 2014.
- [45] Y. J. Ahn, Y. J. Lee, J. Lee, D. Lee, H. K. Park, and G. J. Lee, "Colorimetric detection of endogenous hydrogen sulfide production in living cells," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 177, pp. 118–124, 2017.
- [46] T. J. Kim, Y. J. Lee, Y. J. Ahn, and G. J. Lee, "Characterization of H₂S releasing properties of various H₂S donors utilizing microplate cover-based colorimetric assay," *Analytical Biochemistry*, vol. 574, no. 1, pp. 57–65, 2019.
- [47] R. A. Youness, A. Z. Gad, K. Sanber et al., "Targeting hydrogen sulphide signaling in breast cancer," *Journal of Advanced Research*, vol. 27, pp. 177–190, 2021.

- [48] J. Zeng, M. Li, A. Liu et al., "Colorimetric Sulfide Sensing: Au/AgI dimeric nanoparticles for highly selective and sensitive colorimetric detection of hydrogen sulfide (Adv. Funct. Mater. 26/2018)," *Advanced Functional Materials*, vol. 28, no. 26, article 1870176, 2018.
- [49] F. Yu, X. Han, and L. Chen, "Fluorescent probes for hydrogen sulfide detection and bioimaging," *Chemical Communications*, vol. 50, no. 82, pp. 12234–12249, 2014.
- [50] L. Yang, Y. Su, Z. Sha, Y. Geng, F. Qi, and X. Song, "A red-emitting fluorescent probe for hydrogen sulfide in living cells with a large Stokes shift," *Organic & Biomolecular Chemistry*, vol. 16, no. 7, pp. 1150–1156, 2018.
- [51] H. Peng, Y. Cheng, C. Dai et al., "A fluorescent probe for fast and quantitative detection of hydrogen sulfide in blood," *Angewandte Chemie (International Ed. in English)*, vol. 50, no. 41, pp. 9672–9675, 2011.
- [52] V. S. Lin, A. R. Lippert, and C. J. Chang, "Cell-trappable fluorescent probes for endogenous hydrogen sulfide signaling and imaging H₂O₂-dependent H₂S production," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 18, pp. 7131–7135, 2013.
- [53] J. Cheng, B. Shao, S. Zhang, Y. Hu, and X. Li, "Selective and sensitive detection of hydrogen sulfide in live cells," *RSC Advances*, vol. 5, no. 80, pp. 65203–65207, 2015.
- [54] Y. Wu, Q. Wang, T. Wu et al., "Detection and imaging of hydrogen sulfide in lysosomes of living cells with activatable fluorescent quantum dots," *ACS Applied Materials & Interfaces*, vol. 10, no. 50, pp. 43472–43481, 2018.
- [55] L. Zhang, W. Meng, L. Lu et al., "Selective detection of endogenous H₂S in living cells and the mouse hippocampus using a ratiometric fluorescent probe," *Scientific Reports*, vol. 4, p. 5870, 2015.
- [56] B. An, H. Zhang, J. Peng, W. Zhu, N. Wei, and Y. Zhang, "A highly sensitive ratiometric fluorescent probe for imaging endogenous hydrogen sulfide in cells," *New Journal of Chemistry*, vol. 44, no. 46, pp. 20253–20258, 2020.
- [57] S. Singha, D. Kim, H. Moon et al., "Toward a selective, sensitive, fast-responsive, and biocompatible two-photon probe for hydrogen sulfide in live cells," *Analytical Chemistry*, vol. 87, no. 2, pp. 1188–1195, 2015.
- [58] H. Ibrahim, A. Serag, and M. A. Farag, "Emerging analytical tools for the detection of the third gasotransmitter H₂S, a comprehensive review," *Journal of Advanced Research*, vol. 27, pp. 137–153, 2021.
- [59] X. Gao, H. Zhang, X. Fan et al., "Toward the highly sensitive SERS detection of bio-molecules: the formation of a 3D self-assembled structure with a uniform GO mesh between Ag nanoparticles and Au nanoparticles," *Optics Express*, vol. 27, no. 18, pp. 25091–25106, 2019.
- [60] M. Lee, K. Oh, H. K. Choi et al., "Subnanomolar sensitivity of filter paper-based SERS sensor for pesticide detection by hydrophobicity change of paper surface," *ACS Sensors*, vol. 3, no. 1, pp. 151–159, 2018.
- [61] A. I. Pérez-Jiménez, D. Lyu, Z. Lu, G. Liu, and B. Ren, "Surface-enhanced Raman spectroscopy: benefits, trade-offs and future developments," *Chemical Science*, vol. 11, no. 18, pp. 4563–4577, 2020.
- [62] Y. Guo, Y. Tao, X. Ma et al., "A dual colorimetric and SERS detection of Hg²⁺ based on the stimulus of intrinsic oxidase-like catalytic activity of Ag-CoFe₂O₄/reduced graphene oxide nanocomposites," *Chemical Engineering Journal*, vol. 350, pp. 120–130, 2018.
- [63] J. A. Huang, M. Z. Mousavi, Y. Zhao et al., "SERS discrimination of single DNA bases in single oligonucleotides by electroplasmic trapping," *Nature Communications*, vol. 10, no. 1, p. 5321, 2019.
- [64] L. M. Almeahmadi, S. M. Curley, N. A. Tokranova, S. A. Tenenbaum, and I. K. Lednev, "Surface enhanced Raman spectroscopy for single molecule protein detection," *Scientific Reports*, vol. 9, no. 1, p. 12356, 2019.
- [65] G. Bodelón and I. Pastoriza-Santos, "Recent progress in surface-enhanced Raman scattering for the detection of chemical contaminants in water," *Frontiers in Chemistry*, vol. 8, p. 478, 2020.
- [66] D. W. Li, L. L. Qu, K. Hu, Y. T. Long, and H. Tian, "Monitoring of endogenous hydrogen sulfide in living cells using surface-enhanced Raman scattering," *Angewandte Chemie (International Ed. in English)*, vol. 54, no. 43, pp. 12758–12761, 2015.
- [67] W. S. Zhang, Y. N. Wang, and Z. R. Xu, "High sensitivity and non-background SERS detection of endogenous hydrogen sulfide in living cells using core-shell nanoparticles," *Analytica Chimica Acta*, vol. 1094, pp. 106–112, 2020.
- [68] A. T. Singh, D. Lantigua, A. Meka, S. Taing, M. Pandher, and G. Camci-Unal, "Paper-based sensors: Emerging themes and applications," *Sensors*, vol. 18, no. 9, article 2838, 2018.
- [69] A. W. Martinez, S. T. Phillips, G. M. Whitesides, and E. Carrilho, "Diagnostics for the developing world: microfluidic paper-based analytical devices," *Analytical Chemistry*, vol. 82, no. 1, pp. 3–10, 2010.
- [70] S. M. Rosolina, T. S. Carpenter, and Z. L. Xue, "Bismuth-based, disposable sensor for the detection of hydrogen sulfide gas," *Analytical Chemistry*, vol. 88, no. 3, pp. 1553–1558, 2016.
- [71] T. S. Carpenter, S. M. Rosolina, and Z. L. Xue, "Quantitative, colorimetric paper probe for hydrogen sulfide gas," *Sensors and Actuators B: Chemical*, vol. 253, pp. 846–851, 2017.
- [72] Y. J. Ahn, S. H. Han, and G. J. Lee, "Rapid and simple colorimetric detection of hydrogen sulfide using an etching-resistant effect on silver nanoprisms," *Microchimica Acta*, vol. 188, no. 4, p. 129, 2021.
- [73] J. Lee, Y. J. Lee, Y. J. Ahn, S. Choi, and G. J. Lee, "A simple and facile paper-based colorimetric assay for detection of free hydrogen sulfide in prostate cancer cells," *Sensors and Actuators B: Chemical*, vol. 256, pp. 828–834, 2018.
- [74] S. Zong, Z. Wang, H. Chen et al., "Colorimetry and SERS dual-mode detection of telomerase activity: combining rapid screening with high sensitivity," *Nanoscale*, vol. 6, no. 3, pp. 1808–1816, 2014.
- [75] S. Sharma, A. Jaiswal, and K. N. Uttam, "Colorimetric and surface enhanced Raman scattering (SERS) detection of metal ions in aqueous medium using sensitive, robust and novel pectin functionalized silver nanoparticles," *Analytical Letters*, vol. 53, no. 15, pp. 2355–2378, 2020.
- [76] S. K. Gahlaut, K. Yadav, C. Sharan, and J. P. Singh, "Quick and selective dual mode detection of H₂S gas by mobile app employing silver nanorods array," *Analytical Chemistry*, vol. 89, no. 24, pp. 13582–13588, 2017.
- [77] K. Zhong, L. Chen, Y. Pan et al., "A colorimetric and near-infrared fluorescent probe for detection of hydrogen sulfide and its real multiple applications," *Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy*, vol. 221, article 117135, 2019.

- [78] N. Paul, R. Sarkar, R. Sarkar, A. Barui, and S. Sarkar, "Detection of hydrogen sulfide using BODIPY based colorimetric and fluorescent on-off chemosensor," *Journal of Chemical Sciences*, vol. 132, no. 1, p. 21, 2020.
- [79] Y. J. Ahn, Y. G. Gil, Y. J. Lee, H. Jang, and G. J. Lee, "A dual-mode colorimetric and SERS detection of hydrogen sulfide in live prostate cancer cells using a silver nanoplate-coated paper assay," *Microchemical Journal*, vol. 155, p. 104724, 2020.
- [80] C. Liu, J. Liu, W. Zhang et al., "A ruthenium(II) complex-based probe for colorimetric and luminescent detection and imaging of hydrogen sulfide in living cells and organisms," *Analytica Chimica Acta*, vol. 1145, pp. 114–123, 2021.