Research Article

Human Umbilical Cord Mesenchymal Stem Cells Prevent Bacterial Biofilm Formation

Haoming Yang®, Fang Xu, Xuaner Zheng®, Shumei Yang®, Zhuxiao Ren, and Jie Yang®

Guangdong Women and Children Hospital, Guangzhou, China

Correspondence should be addressed to Jie Yang; jieyang0830@126.com

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Biofilm formation is easily found in patients suffered from ventilator-associated pneumonia (VAP) in neonatal intensive care unit (NICU) and makes the VAP infections not only harder to be treated but easier to relapse. In order to find some novel ways to inhibit biofilm formation, this study describe a previously unrecognized role for the human umbilical cord mesenchymal stem cells (hUCMSCs). In addition to multiple differentiation, hUCMSCs have the ability to prevent the biofilms formation in vitro by secreting antibacterial peptides (LL-37 and hBD-2). This occurred while P. aeruginosa PA27853 and hUCMSCs were cocultured, and the filtrated medium, which was the supernatant containing antibacterial peptides (5.9 ng/ml of LL-37, 1.77 ng/ml of hBD-2), and inhibited the growth of the bacterial biofilm on the surface of tracheal tube (2.5#, for preterm infant). Using microarrays, we were able to demonstrate that the antibacterial peptides from hUCMSC affected biofilm formation by downregulating the gene-encoded polysaccharide biosynthesis protein. In addition, in order to find out the most suitable concentration of hUCMSCs, P. aeruginosa was cocultured with eight-level concentrations of hUCMSCs, and we found that the concentration of LL-37 was positively correlated with the concentration of hUCMSCs. Meanwhile, the concentration of LL-37 became stable while the hUCMSC concentration reaches higher than $5 \times 10^6$ cells/ml. But the concentration of hBD-2 had no significant correlation with hUCMSCs. The collection of these stem cells is not only limited by ethics but also reduces host rejection. This makes it possible to use autologous hUCMSCs to treat neonatal VAP.

1. Introduction

VAP is one of the common nosocomial infections in the NICU, and the incidence rate was about 5.2 per 1000 invasive ventilated patients and results in mortality of 23.5% [1]. In clinical practice, VAP is hard to treat and often causes significant morbidity and mortality. The biofilm on the surface of tracheal tube formed by antibiotic-resistant pathogen caused VAP hard to eradicate [2, 3]. Researches [4–6] report that biofilms are microbial communities encased in extracellular polymeric substances. The main components of biofilm are polysaccharides, proteins, lipids, and nucleic acids. Among them, extracellular polysaccharide substances (EPS) are considered as the skeleton [7]. EPS plays an important role in the formation of biofilm. EPS help bacteria adhere to the solid surface firmly and act as a barrier to protect bacteria [8]. The VAP associated with biofilm is hard to treat and easy to relapse [9]. Thus, we need a novel method to interfere the biofilm formation.

Human host defense peptides, also known as antimicrobial peptides, are secreted in response to infection, or the constant threat of infection. Human antimicrobial peptides, including cathelicidin hCAP-18/LL-37 and human β-defensin 2 (hBD-2), are ubiquitous in nature as components of innate immune defense systems [10]. Subinhibitory concentration of LL-37 (1/4–1/128 of the MIC of 64 μg/ml) was shown to affect biofilm formation by decreasing the attachment of P. aeruginosa cells, stimulating twitching motility, and influencing two major QS systems (Las and Rhl), leading to the downregulation of genes essential for biofilm development [11]. A recent research suggested that both m-RNA and protein expression of cathelicidin hCAP-18/
LL-37 in mesenchymal stem cells increased after bacterial challenge, which make mesenchymal stem cells possess direct antimicrobial activity [12].

Our previous researches show that hUCMSCs also have the ability to synthesize and secrete the LL-37 and HBD-2 in response to the P. aeruginosa [13]. However, researches about using autologus hUCMSCs to inhibit biofilm formation has not been found. Therefore, we designed experiments to find out the optimal concentration of antimicrobial peptides secreted by hUCMSCs and to test the hypothesis that hUCMSCs, by secreting antimicrobial peptides, possessed the ability to interfere the biofilm formation on the tracheal tube. Finally, the differences of P. aeruginosa gene expression were detected by microarrays to reveal the possible mechanism of hUCMSCs inhibiting biofilm.

2. Materials and Methods

2.1. Bacteria, hUCMSCs, Media, and Medical Catheters.

Standard P. aeruginosa (ATCC27853) strains were offered by the Clinical Laboratory Centre of China’s Ministry of Health. The experimental strains were kept at -4°C and grown at 37°C, 5% CO2 on blood agar plate. Before each experiment, the bacterial cells were resuspended in phosphate-buffered saline (PBS) and then adjusted to the concentration of 5 × 10^8 CFU/ml through measuring optical density (OD at λ = 600 nm) of the suspension (according to the equation: OD_{600} = 0.5 corresponds to 5 × 10^8 CFU/ml). For the cultivation of biofilms, P. aeruginosa PA27853 was grown at 37°C, 5% CO2 in complex Luria-Bertani (LB) broth [14]. hUCMSCs were purchased from American Type Culture Collection (ATCC; Manassas, VA). The cells met all the criteria for the classification as MSCs as defined by the International Society of Cellular Therapy. hUCMSCs were cultured at 37°C, 5% CO2 in DMEM/F12 medium (Gibeco, USA), comprising with 10% fetal bovine serum (FBS) on 25 cm^2 cell culture flasks. Sterile tracheal tube (inside diameter 2.5 cm) was cut by sterile scissors into sections with length of 1 cm (according to the scale on the tracheal tube), 5 sections were used for experimental group, 5 sections were used for control group, and 2 for quality control.

2.2. Titration of hUCMSC Concentration. Seven target concentrations of hUCMSCs, diluted from the 5 × 10^6 CFU/ml suspension, were included in the experiments (5 × 10^2 CFU/ml, 5 × 10^3 CFU/ml, 5 × 10^4 CFU/ml, 5 × 10^5 CFU/ml, 5 × 10^6 CFU/ml, 5 × 10^7 CFU/ml, 5 × 10^8 CFU/ml). In coculture experiments, both bacteria cells and hUCMSCs were inoculated in the DMEM/F12 medium in polypropylene 6-well microtiter plates. After 6 hours of incubation at 37°C, the coculture supernatant was collected and filtered twice by 0.22 μm filters. The concentration of LL-37 and HBD-2 in supernatant was determined by the ELISA kit from the Phoenix Pharmaceuticals and Hycult Biotechnology, respectively. All groups were carried out at least in three technical repeats.

2.3. Preparation of Supernatant after Coculture. The bacterial cells and hUCMSCs were cocultured at 50 ml cell culture flasks. One flask as experience group was inoculated with 30 ml of DMEM/F12 medium containing approximately 3 × 10^4 cells/ml of P. aeruginosa PA27853 and 1 × 10^6 cells/ml of hUCMSCs (according to the result of concentration titration). Another flasks, as the control group, were inoculated with 30 ml of DMEM/F12 medium containing 3 × 10^4 cells/ml of P. aeruginosa and 1 × 10^6 cells/ml of normal human lung fibroblast (NHLF). After 6 hours of coculture (37°C, 5% CO2), DMEM/F12 medium from those two flasks was filtered twice by 0.22 μm filters. The supernatant from experimental flask was marked as experience group, and the other was marked as control group.

2.4. Biofilm Experiment. P. aeruginosa biofilm was grown at 37°C, 5% CO2 in two polypropylene 6-well microtiter plates [14], in which 5 holes for experimental group, 5 holes for control group, and 2 holes for quality control. Each hole contains one sterile tracheal tube of 1 cm and 10 μl of complex LB broth containing P. aeruginosa of 0.5 McF. The broth was replaced each 24 hours. After 4 days of incubation, the in vitro model of biofilm on tracheal tube was preliminarily established. Within next 3 days, those tracheal tubes (in vitro model) were incubated with filtered medium as described above, and the filtrated medium was replaced each 24 hours.

After 7 days of incubation, the tracheal tube were rinsed three times with phosphate buffer solution (PBS), and the biofilm was fixed on the surface with formaldehyde. After crystal violet staining, the absorbance was measured in polypropylene 96-well microtiter plates at 595 nm [15].

2.5. Motility Assays. Swimming motility experiment was performed on semisolids plates containing 0.3% (wt/vol) agar. All bacterial motility evaluations have five technical repeats. After poured from the sterilized agar, the semisolids plates were dried in the biological safety cabinet at room temperature for 1 or 2 hours.

2.6. Statistical Method. SPSS (Ver.13) was used in this study as the statistical software. Product-moment correlation coefficient as the statistical method to analyze the correlation between the concentration of hUCMSCs and the level of antibacterial peptides, taking P < 0.05 for the difference was statistically significant.

2.7. DNA Microarray Experiment. The difference of gene expression were determined by microarray chip (Agilent, China). Microarray experiments were performed on those five independent cultures. The P. aeruginosa cells were harvested from the surface of tracheal tube. Only genes that exhibited a change of fivefold or more with a P value of ≤0.05 were considered in this study.

3. Results

3.1. Relationship between Antibacterial Peptide Levels and hUCMSC concentration. In this study, bacteria were cocultured with eight levels of hUCMSCs. The level of LL-37
increased significantly with increasing the concentration of hUCMSCs (Figure 1). The incremental effect did not appear while the hUCMSC concentration reach higher than $5 \times 10^6$ cells/ml, and the highest level of LL-37 came from the $5 \times 10^7$ hUCMSCs cells/ml. Since the concentration of LL-37 becomes stable while the hUCMSC concentration reach $5 \times 10^6$ cells/ml, we excluded group 7 ($1 \times 10^6$ cells/ml) and group 8 ($5 \times 10^7$ cells/ml) and studied the correlation between the concentration of hUCMSCs and the level of LL-37. It indicated that the level of LL-37 in supernatant positively correlated with the concentration of hUCMSCs (Table 1).

In addition, we excluded group 8 ($5 \times 10^7$ cells/ml) and tried to build the regression model of LL-37 level versus concentration of hUCMSCs. Figure 2 suggested that the level of LL-37 in supernatant may increase with the concentration of hUCMSCs.

The concentration of hBD-2 showed very low level similarly, but did not increased significantly with increasing the concentration of hUCMSCs (Figure 3). The highest level came from the group of $1 \times 10^6$ hUCMSCs cells/ml. The statistical results indicated that the level of HBD-2 was not associated with the concentration of hUCMSCs (Table 2).

### 3.2 Concentrations of LL-37 and hBD-2 in the Coculture Medium

To test whether the inhibitory effect on biofilm formation was related to LL-37 and HBD-2 secreted by hUCMSCs in the filtrated medium, the concentrations of

<table>
<thead>
<tr>
<th>Groups*</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
</tr>
</thead>
<tbody>
<tr>
<td>hUCMSCS (cells/ml)</td>
<td>$1 \times 10^4$</td>
<td>$5 \times 10^4$</td>
<td>$1 \times 10^5$</td>
<td>$5 \times 10^5$</td>
<td>$1 \times 10^6$</td>
<td>$5 \times 10^6$</td>
</tr>
<tr>
<td>LL-37 (ng/ml)</td>
<td>3.52</td>
<td>4.36</td>
<td>5.24</td>
<td>5.87</td>
<td>6.31</td>
<td>7.96</td>
</tr>
</tbody>
</table>

$r = 0.853$, $P = 0.031$

*Groups 7 and 8 were excluded.*
LL-37 and HBD-2 were measured after the coculture experiment using the ELISA kit. After stimulated by \textit{P. aeruginosa}, hUCMSCs secrete LL-37 and HBD-2 in the DMEM/F12 medium. The difference was statistically significant different (Table 3).

### 3.3. Tracheal Tube Biofilm Analyses.

Since the artificial implants are usually performed as the solid surface in the VAP, tracheal tube was chosen as the solid surface for the bacterial attachment and the biofilms formation. After 3 days of incubation in presence of the supernatant, biofilm attached on the surface of tracheal tubes was stained (Figure 4) and measured the absorbance at 595 nm. Obviously, the biofilm cultured in the presence of LL-37 (5.90 ng/ml) and HBD-2 (1.77 ng/ml) was 56% less than the biofilm formed in the untreated control group (Table 4). The difference between two groups demonstrated statistically significant different (Table 5).

### 3.4. Microarray Analysis.

In order to investigate the mechanism of biofilm inhibition, microarray analysis was performed to examine gene expression profile of \textit{P. aeruginosa}. The analysis demonstrated that greater than twofold change in expression levels of 106 genes downregulated (selected examples are shown in Table 6. All regulated genes are shown in the annex). The inhibiting effect of LL-37 at the concentrations of 16 g/ml (one-quarter MIC concentrations) has already been illuminated [11], and the peptides could influence gene expression of type IV pili and the quorum sensing systems in \textit{P. aeruginosa}. However, the filtrated medium contains LL-37 of 5.9 μg/ml and 1.77 ng/ml and seems to downregulate the gene expression of polysaccharide biosynthesis protein but not to influence type IV pili and the quorum sensing systems.

![Figure 3: Level of HBD-2 secreted by different concentration of hUCMSCs.](image)

**Table 2: The correlation between the concentration of hUCMSCs and the level of LL-37.**

<table>
<thead>
<tr>
<th>Groups</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
</tr>
</thead>
<tbody>
<tr>
<td>hUCMSCS (cells/ml)</td>
<td>(1 \times 10^4)</td>
<td>(5 \times 10^4)</td>
<td>(1 \times 10^5)</td>
<td>(5 \times 10^5)</td>
<td>(1 \times 10^6)</td>
<td>(5 \times 10^6)</td>
<td>(1 \times 10^7)</td>
<td>(5 \times 10^7)</td>
</tr>
<tr>
<td>HBD-2 (pg/ml)</td>
<td>1.62</td>
<td>1.55</td>
<td>1.6</td>
<td>1.75</td>
<td>1.87</td>
<td>1.83</td>
<td>1.85</td>
<td>1.83</td>
</tr>
</tbody>
</table>

\(r = 0.409, P = 0.314\)

**Table 3: Concentrations of LL-37 and hBD-2 (±s).**

<table>
<thead>
<tr>
<th>Group</th>
<th>Cathelicidin/LL-37 (ng/ml)</th>
<th>(P) value*</th>
<th>HBD-2 (ng/ml)</th>
<th>(P) value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experimental group (hUCMSCs + PA27853)</td>
<td>5.90 ± 0.51</td>
<td>&lt;0.001</td>
<td>1.77 ± 0.25</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Control group (NHLF + PA27853)</td>
<td>1.98 ± 0.34</td>
<td></td>
<td>0.72 ± 0.25</td>
<td></td>
</tr>
</tbody>
</table>

*Statistically significant level \(P \leq 0.05\).
and resulted in the formation of biofilm on various bacteria have been concentrated in the supernatant of the experimental group (hUCMSCs + PA27853). Nevertheless, there was no difference about the motility between two groups (Table 7). The antibacterial peptides secreted by hUCMSCs may not stimulate motility in the presence of those two groups. In the same way, we observed that the motility of bacteria in presence of HBD-2 (17) and their broad-spectrum antimicrobial effects after being stimulated by bacteria, presenting antibiosis effect. The major antibacterial peptides secreted by human epithelial tissues are LL-37 and HBD-2 (17), and their broad-spectrum antimicrobial effects on various bacteria have been confirmed (18, 19). The hUCMSCs used in this study also secreted LL-37 and HBD-2 after stimulation, suggesting a possible bacteriostatic effect. The LL-37 in the supernatant of the experimental group can be found by protein electrophoresis (13). Compared with NHLF, hUCMSCs can secrete more antimicrobial peptides into the filtrated medium. It is suggested that the ability of hUCMSCs to secrete antimicrobial peptides may be used in clinical practice. Beside the bacteriostatic effect, the antibiofilm effect of antimicrobial peptides is also concerned. It has been proved that LL-37 can effectively inhibit the growth of biofilm, but the meaningful concentration (0.5 μg/ml) (12) is significantly higher than that in these studies. Although the concentration of LL-37 measured in these experiments is only 5.9 ng/ml, which is significantly lower than that in previous study, similar inhibition effect can still be obtained. In the filtrated medium, the secretion of hBD-2 can also be detected. So, we may believe that LL-37 and HBD-2 can synergize to produce antibiofilm effect. More researches are needed to explore the mechanism that hBD-2 makes LL-37 produce antibiofilm effect at low concentration.

The formation of biofilm is affected by many factors. In this study, it was found that LL-37 (5.9 ng/ml) combined with HBD-2 mainly affected the expression of polysaccharide biosynthesis protein gene, thus reducing the formation of biofilm. But it did not significantly affect the bacterial attachment, mobility, and quorum sensing systems. Polysaccharide biosynthesis protein is one of the most common polysaccharides in bacteria. It has been proved that Polysaccharides in biofilm play as the role of skeleton and are important to biofilm formation (7). If the ability of bacteria to secrete and synthesize polysaccharides is inhibited, the formation of biofilm is also affected. In addition, if the polysaccharide in biofilm can be reduced, the barrier effect on antibacterial drugs may be reduced.

In microarray analysis, it is found that the expression of fimbrin, flagella, and quorum sensing systems was not significantly affected. However, there are still numerous unknown genes that have been affected and downregulated (not listed in Table 5). The biological effects of these genes are not fully understood. We are curious about whether these genes are related to biofilm formation, so it is worth exploring the role of these genes.

As a novel antibiofilm pathway, the application of the hUCMSCs may have same obvious advantages especially in NICU. The Centers for Disease Control and Prevention (CDC; Atlanta, Ga., USA) defines VAP as "a nosocomial infection diagnosed in patients undergoing mechanical ventilation for at least 48 h" (20). The formation of biofilm on the tracheal tube were easily to be found in preterm neonates

### Table 4: Absorbance of biofilm experiment.

<table>
<thead>
<tr>
<th>Group/hole</th>
<th>Absorbance</th>
<th>Mean (x ± s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experimental group (hUCMSCs + PA27853)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>0.152</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>0.170</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>0.210</td>
<td>0.214 ± 0.054</td>
</tr>
<tr>
<td>4</td>
<td>0.266</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>0.270</td>
<td></td>
</tr>
<tr>
<td>Control group (NHLF + PA27853)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>0.517</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>0.418</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>0.525</td>
<td>0.482 ± 0.043</td>
</tr>
<tr>
<td>4</td>
<td>0.486</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>0.463</td>
<td></td>
</tr>
</tbody>
</table>

*33% glacial acetic acid was set as the reagent blank group. The situation that absorbance value are ≥ twice as reagent blank group is considered as biofilms formation [3].

### Table 5: Absorbance statistic by Student’s t-test.

<table>
<thead>
<tr>
<th>Group</th>
<th>Cases (n)</th>
<th>Mean (x ± s)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experimental group</td>
<td>5</td>
<td>0.214 ± 0.054</td>
<td>0.000*</td>
</tr>
<tr>
<td>Control group</td>
<td>5</td>
<td>0.482 ± 0.043</td>
<td></td>
</tr>
</tbody>
</table>

*Statistically significant level P = 0.05.

### 3.5. The Motility of Bacteria Was Not under the Influence

Low concentration of LL-37 (even at 4 μg/ml) could stimulate bacterial surface motility. In the same way, we observed the motility of bacteria in presence of those two groups filtrated medium. Nevertheless, there was no difference about the motility between two groups (Table 7). The antibacterial peptides secreted by hUCMSCs may not stimulate motility of *P. aeruginosa*.

### 4. Discussion

Previous research (16) showed that the growth of colonies was inhibited in the cocultured in vitro model of bone marrow mesenchymal stem cells with Gram-negative bacteria. The possible mechanism is that the stem cells secrete more antimicrobial peptides after being stimulated by bacteria, thus presenting antibiosis effect. The major antibacterial peptides secreted by human epithelial tissues are LL-37 and HBD-2 (17), and their broad-spectrum antimicrobial effects on various bacteria have been confirmed (18, 19). The hUCMSCs used in this study also secreted LL-37 and HBD-2 after stimulation, suggesting a possible bacteriostatic effect. The LL-37 in the filtrated medium of the experimental group can be found by protein electrophoresis (13). Compared with NHLF, hUCMSCs can secrete more antimicrobial peptides into the filtrated medium. It is suggested that the ability of hUCMSCs to secrete antimicrobial peptides may be used in clinical practice.

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hUCMSCs could secrete the highest concentration of hBD-2 at a lower concentration.

It is worth noting that higher concentration of hUCMSCs would be used to obtain higher concentration of LL-37; therefore, it may increase the cost of application. As the experimental group needed a large amount of supernatant, the concentration of LL-37 was chosen in the coculture experiment under the condition of not affecting the experimental results. At the same time, we considered that hUCMSCs with higher concentration than $1 \times 10^6$ cells/ml should also show the ability to inhibit the biofilm formation.

Table 6: Selected *P. aeruginosa* genes that were dysregulated.

<table>
<thead>
<tr>
<th>Gene name</th>
<th>Fold increase</th>
<th>Protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>CP000744.1_cds_PSPA7_0598_597</td>
<td>-9.61</td>
<td>Allophanate hydrolase</td>
</tr>
<tr>
<td>CP000744.1_cds_PSPA7_2187_2163</td>
<td>-8.16</td>
<td>Probable hydrolase</td>
</tr>
<tr>
<td>CP000438.1_cds_PA14_23390_1869</td>
<td>-20.02</td>
<td>Polysaccharide biosynthesis protein</td>
</tr>
<tr>
<td>CP000744.1_cds_PSPA7_2314_2288</td>
<td>-11.10</td>
<td>Enoyl-CoA hydratase</td>
</tr>
<tr>
<td>CP000744.1_cds_PSPA7_5599_5524</td>
<td>-8.84</td>
<td>Termination factor rho</td>
</tr>
</tbody>
</table>

Data Availability

Data will be available from the corresponding author if required.

Ethical Approval

The name of the ethics committee that approved the study is Guangdong Women and Children Hospital Ethics committee.

Consent

Written informed consent was obtained from all participants. The authors consent to publish this article. All authors read and approved the final manuscript.

Conflicts of Interest

The authors report no conflict of interest.
Acknowledgments

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References


