Proper placentation in the first trimester is essential for a healthy pregnancy in humans. A recent proteomics study of human placental tissue has identified that tripeptidyl peptidase 1 (TPP1) production is reduced in the placenta in early-onset preeclampsia compared to uncomplicated pregnancy. However, it remains to be investigated if TPP1 plays a role in regulating trophoblast cell function during early pregnancy. In this study, immunohistochemistry was used to determine the production and localization of TPP1 in human placenta throughout gestation and the first-trimester decidua/implantation sites. TPP1 siRNA (20 nM) was transfected into a human trophoblast cell line (HTR8/SVneo) to knock down TPP1, and functional consequences on cell adhesion, proliferation, migration, and invasion were analyzed via xCELLigence real-time monitoring. The expression of TPP1 downstream targets was examined by qPCR. Our data show that TPP1 localized to the discrete foci in the cytoplasm in syncytiotrophoblast, cytotrophoblast, and decidual cells across all trimesters of pregnancy. In the first-trimester human decidua, TPP1 exhibited similar staining patterns in the cytotrophoblast cells based at the cell columns. However, minimal/no staining was identified in the HLA-G positive extravillous trophoblast cells (EVTs), especially in the EVT that invaded in the decidua. Knockdown of TPP1 in HTR8/SVneo cells by 95% significantly impaired cell adhesion and proliferation without affecting cell migration and invasion. qPCR revealed that the expression of cell proliferation markers P21 and MKI67 and TPP1-related genes MRE11, CLN3, and CLN8 was significantly changed after TPP1 knockdown in HTR8/SVneo cells compared to control. Overall, our data demonstrate that TPP1 alters trophoblast cell line function suggesting that it may be involved in regulating human placentation in the first trimester via controlling trophoblast cell adhesion and proliferation.

1. Introduction

Normal placentation in the first trimester is crucial for a healthy pregnancy. Human placentation initiates after embryo implantation to form a functional placenta. Cytotrophoblast cells in the placental villi form the anchoring chorionic villi, which anchor the placenta to the decidua where they proliferate to form cell columns. Within the columns, the trophoblast differentiates from a non-invasive, proliferative phenotype towards an invasive but less proliferative phenotype [1, 2]. These invasive cells (called extravillous trophoblast [EVT]) migrate and invade into maternal decidua, whereupon they engraft and remodel maternal spiral arteries to create high-flow, low-resistance vessels by the end of the first trimester ensuring blood supply to the placenta [2, 3]. Dysregulation of trophoblast cell adhesion, proliferation, migration, and invasion can lead to dysfunctional placentation and obstetric complications such as preeclampsia [4].

Tripeptidyl peptidase 1 (TPP1) is a lysosomal serine protease widely expressed in higher organisms [5]. Defective TPP1 function results in abnormal accumulation of protein and lipofuscin-like material in cells eventually leading to cell dysfunction [6, 7] and diseases including the neurodegenerative disease neuronal ceroid lipofuscinosis type 2 (CLN2). In a zebrafish model of human CLN2 disease, TPP1 deficiency leads to sustained reduction of cell proliferation in
the central nervous system [8]. In hepatocellular carcinoma, knockdown of TPP1 inhibits cell proliferation, migration, and invasion in vitro [9]. Our recent work has demonstrated that TPP1 is expressed at significantly lower levels in human luminal epithelial cells of infertile endometrium compared to fertile. Functional analysis reveals that knockdown of TPP1 in human endometrial epithelial cells impairs cell adhesion capacity [10]. Since placentation requires highly regulated cell adhesion, proliferation, migration, and invasion, the involvement of TPP1 in mediating these activities support its potential roles in regulating normal placentation.

Although the localization and function of TPP1 in human placenta have not been determined, a recent proteomics analysis of human placental tissue has identified reduced TPP1 expression in the placenta in early-onset pre-eclampsia compared to uncomplicated pregnancy [11]. This study aimed to determine the localization of TPP1 in human placenta and its function on trophoblast cell (HTR8/SVneo) adhesion, proliferation, migration, and invasion.

2. Materials and Methods

2.1. Human Placental Tissue Collection. Placental tissues were collected from healthy women undergoing pregnancy termination for psychosocial reasons (first and second trimesters, 7–22 weeks; n = 9) and delivery at term (37–42 weeks; n = 4) at Monash Health and The Royal Women’s Hospital (ethics number: #09317B). Written informed consent was obtained from each patient before surgery. Collected placental tissues were quickly washed with Hanks’ Balanced Salt Solution and subjected to 10% formalin fixation for immunohistochemistry.

2.2. Antibodies and Cell Line. Mouse monoclonal antibody against TPP1 (#AB54685) was purchased from Abcam (Cambridge, UK), and mouse monoclonal antibody against HLA-G (#557577) was purchased from BD Pharmingen (San Diego, CA, USA). Mouse IgG isotype control (#X0931) was purchased from Dako (Glostrup, Municipality, Denmark). HTR8/SVneo cells (CRL-3271, ATCC) were cultured as per instructions.

2.3. Immunohistochemistry and Immunocytochemistry. Human placental villous and decidua tissues were fixed in 10% formalin, embedded in paraffin, and sectioned at 4 μm for immunohistochemistry staining. Sections were dehydrated, rehydrated, and subjected to antigen retrieval as previously optimized (10 mM sodium citrate for 5 min) [10]. After antigen retrieval, sections were treated with 3% hydrogen peroxide diluted in methanol to quench endogenous peroxidase. Sections were then rinsed with Tris-buffered saline and blocked with 10% goat serum and 2% human serum. Sections were then rinsed with Tris-buffered saline to quench endogenous peroxidase. After antigen retrieval, sections were treated with 3% hydrogen peroxide and permeabilized with 0.1% Triton X-100 in TBS (v/v). Following TBS wash, cells were treated with 3% hydrogen peroxide and immunolabeled as described for placental sections.

2.4. In Vitro siRNA Transfection. Passaged HTR8/SVneo cells were counted and seeded into wells of 12-well plates with the goal to reach 70–80% confluency the next day. The cells were then washed with Dulbecco’s phosphate-buffered saline (DPBS) and transfected with TPP1 siRNA or scrambled negative control (20 nM) (Dharmacon, Lafayette, CO, USA) using the Lipofectamine RNAmax transfection system as instructed by the manufacturer. After 24 h, the transfection medium was replaced with fresh HTR8/SVneo cell culture medium and the cells were cultured for 48 h before being trypsinized, counted, and analyzed in real-time by xCELLigence assays and qPCR.

2.5. xCELLigence Assays. For HTR8/SVneo cell adhesion and proliferation, the transfected cells were trypsinized after 48 h of culture and seeded at 10,000 cells per well in the xCELLigence 96-well E-plate in fresh culture medium supplemented with 5% fetal bovine serum (FBS), as previously described [12]. The plate was monitored for cell index every 15 min for 8 h (for cell adhesion) and every 1 h following this for 72 h (for cell proliferation). HTR8/SVneo cell migration and invasion were measured using the CIM-plate 16 with 8 μm pores (Roche, Sandhofer Straße, Mannheim, Germany). For migration, 40,000 transfected cells were resuspended in 5% FBS medium and seeded into the upper chambers. 10% FBS medium was added to lower chambers, as we previously described [12]. The cell index was recorded every 15 min for up to 18 h. Cell invasion was recorded using the same conditions as cell migration for up to 60 h with upper transwells being precoated with Matrigel (1:10 dilution in RPMI 1640). All data were calculated using the RTCA software 1.2 as supplied with the instrument and exported for statistical analysis.

2.6. RNA Isolation and RT-qPCR. The transfected HTR8/SVneo cells were subjected to RNA isolation using TRI Reagent after 48 h of culture. Genomic DNA contamination was removed using the TURBO DNA-free kit (#AM1907). After quantification using NanoDrop, 300 ng RNA was reversed transcribed into cDNA using the SuperScript™ III First-Strand Synthesis System (18080-051, Thermo). qPCR was performed on the Applied Biosystems ViiA7 system using the SYBR Green Master Mix (#4367659, Thermo).
Figure 1: Continued.
and specific primer pairs as summarized in Supplementary Table S1. Gene expression was normalized to 18S and calculated by the comparative cycle threshold method (ΔΔCt).

2.7. Immunoblotting. The organic phase from TRI Reagent was collected for protein extraction as previously optimized [13]. Equivalent amounts of protein were boiled in lysis buffer (5% SDS, 20 mM EDTA, 140 mM NaCl, and 100 mM Tris) at 100 °C for 5 min, prior to being resolved by SDS-PAGE (150 V, 90 min) and transferred to PVDF membranes (350 mA, 70 min). The membranes were blocked with 5% skim milk for 1 h at room temperature and incubated with TPP1 antibody (0.28 μg/mL) at 4 °C overnight. Following washing in TBS, the membranes were probed with HRP-conjugated secondary antibody and detected by chemiluminescence. GAPDH was reprobed, and appropriate protein band intensity was determined by densitometry using the ImageJ.

2.8. Statistics. Experiments were repeated at least four times with numbers indicated in each figure legend. Statistical analysis was performed using PRISM 8.0, two-tailed paired Student’s t-test, or one-way or two-way ANOVA as appropriate with a significance threshold of P < 0.05. Data were presented as mean ± SEM.

3. Results

3.1. Expression of TPP1 in Human Placenta and Decidua. We first sought to determine the relevance of TPP1 on placentation by assessing its expression and immunolocalization in human placenta throughout gestation and the first trimester decidua/implantation sites. In the placental villous, TPP1 exhibited discrete foci of cytoplasmic staining in syncytiotrophoblast, cytotrophoblast, and decidual cells across all trimesters of pregnancy (Figure 1(a) and Supplementary Figure S1). TPP1 immunostaining intensity scores in the decidual cells identified a significant increase (P < 0.05) between second-trimester and term placental villous (Figure 1(a)). TPP1 was also expressed in the decidual cells and glandular epithelial cells in the first trimester decidua/implantation sites (Figure 1(b)). To determine if TPP1 localizes to EVTs, we investigated TPP1 and HLA-G (EVT marker) staining on serial sections of the first trimester decidua basalis. TPP1 exhibited discrete foci of cytoplasmic staining at the base of the cell columns, and the staining intensity was apparently decreased in the HLA-G-positive EVTs that were further away from the cell columns (Figure 1(c)). In the EVTs that invaded into the maternal decidua, no positive staining of TPP1 was identified (Figure 1(c)). An IgG isotype control for TPP1 revealed no positive staining (Figure 1).

3.2. Knockdown of TPP1 in HTR8/SVneo Cells Impaired Cell Adhesion and Proliferation. To determine the function of TPP1 on placentation, HTR8/SVneo cells (first-trimester extravillous trophoblast-derived cell line) were transfected with TPP1 siRNA to knock down the endogenous expression of TPP1 and cells were then monitored for adhesion, proliferation, migration, and invasion in real time via xCELLigence. Before use, we confirmed that HTR8/SVneo cells possess a conserved pattern of TPP1 expression compared to placental tissue (Figure 2(a)). qPCR confirmed that
Figure 2: Examination of the efficiency of TPP1 knockdown in HTR8/SVneo cells after siRNA treatment. (a) Localization of TPP1 in untreated HTR8/SVneo cells (n = 3). (b and c) Cells were transfected with TPP1 siRNA or scrambled control, and after 48 h, qPCR (b) and immunoblotting (c) were used to determine TPP1 expression. For qPCR, expression levels were normalized to 18S (n = 4). TPP1 was reduced by ~95% after siRNA treatment compared to control. For immunoblotting, blots were reprobed with GAPDH for densitometry analysis. (n = 4). **P < 0.01.

Figure 3: Real-time examination of the effects of TPP1 knockdown on cell adhesion (a) and proliferation (b) of HTR8/SVneo cells via xCELLigence (expressed as cell index). (a) TPP1 knockdown in HTR8/SVneo cells significantly decreased their adhesive capacity from 2 h to 4 h after seeding compared to control. (b) TPP1 knockdown in HTR8/SVneo cells significantly decreased cell proliferation from 36 h to 60 h compared to control. Data were presented as mean ± SEM (n = 4). *P < 0.05.
TPP1 siRNA treatment in HTR8/SVneo cells resulted in ~95% decreased TPP1 expression compared to control (P < 0.001; Figure 2(b)). The significant knockdown of TPP1 was also confirmed at the protein level by immuno-blotting (P < 0.01; Figure 2(c)). Cell adhesion was significantly impaired after TPP1 knockdown from 2 h to 4 h of culture, after which the cell index plateaued (P < 0.05; Figure 3(a)). Similarly, cell proliferation was significantly reduced from 36 h to 60 h of culture, after TPP1 knockdown compared to control (P < 0.05; Figure 3(b)). HTR8/SVneo cell migration and invasion after TPP1 knockdown were determined via xCELLigence assays, as previously described [12]. No significant changes in HTR8/SVneo cell migration and invasion between the TPP1 siRNA treated group and control at 18 h and 42 h of culture were identified, respectively (Figure 4).

3.3. The Effect of TPP1 Knockdown on the Expression of Genes Related to Cell Adhesion and Proliferation and TPP1 Functional Partners. The expression of two adhesion-related genes B-cell lymphoma 2 (BCL2) and P53, as we previously identified in Ishikawa cells after TPP1 knockdown [10], was determined by qPCR. No significant changes were identified between groups (Figure 5(a)). With relevance to cell proliferation, among three genes determined, TPP1 knockdown significantly increased P21 expression (P < 0.05 ) and decreased marker of proliferation Ki-67 (MKI67) expression (P < 0.01), compared to control, respectively (Figure 5(b)). No significant difference was identified for the expression of telomerase reverse transcriptase (TERT) between groups (Figure 5(b)). The expression of TPP1 functional partners/targets was also examined in the HTR8/SVneo cells following TPP1 knockdown by qPCR. TPP1 knockdown significantly decreased the expression of MRE11 (P < 0.001) and ceroïd-lipofuscinosis, neuronal (CLN) 3 (P < 0.01) while the expression of CLN8 was significantly increased (P < 0.05) compared to control (Figure 5(c)).

4. Discussion

This study determined the localization of TPP1 in the human placenta throughout gestation and the first-trimester decidua/implantation sites. TPP1 showed discrete foci of cytoplasmic staining in the syncytiotrophoblast, cytotrophoblast, and decidual cells; however, minimal/no staining was identified in the HLA-G-positive EVTs, especially in the EVTs that invaded into the decidua. Functional analysis revealed a role for TPP1 in regulating trophoblast cell adhesion and proliferation, but not migration and invasion.

After blastocyst implantation, the cytotrophoblast cells proliferate to form cell columns based at the anchoring villi which then further differentiate to a less proliferative but more invasive phenotype in order to invade into the maternal decidua [1, 3]. Dysregulation of this process may lead to obstetric complications such as preeclampsia due to inadequate placentation [1, 14]. Our immunohistochemistry staining data confirmed the expression of TPP1 in these proliferative cells in the cell columns, suggesting a direct role of TPP1 in regulating trophoblast cell proliferation. In support of this hypothesis, knockdown of TPP1 in the HTR8/SVneo cells significantly impaired cell proliferation. We also identified an inverse relationship between TPP1 and HLA-G staining in the first trimester human decidua. While the low levels of TPP1 were still identified in the HLA-G-positive EVTs residing at the placental cell columns, no TPP1 staining was revealed in the invaded HLA-G positive EVTs in the decidua where cells are no longer proliferative [2]. Consistently, knockdown of TPP1 in the HTR8/SVneo cells did not compromise cell invasion, suggesting that TPP1 may not be involved in regulating trophoblast cell invasion.
MRE11 has been recently identified as a TPP1 downstream target [15]. In addition to enhancing DNA repair and improving cell survival [15], MRE11 is also required for cell proliferation [16]. In human breast cancer cells, MRE11 regulates cell proliferation via signal transducer and activator of transcription 3 (STAT3) [17]. As a downstream target of TPP1, MRE11 expression was significantly decreased after TPP1 knockdown in the HTR8/SVneo cells. Although we remain uncertain if MRE11 is involved in regulating HTR8/SVneo cell proliferation, the effects of TPP1 knockdown on HTR8/SVneo cell proliferation were confirmed by two other proliferation markers MKI67 and P21. P21 is a cell-cycle inhibitory protein that leads to cell-cycle arrest in the S phase by targeting cyclin-CDK complexes [18]. The expressional changes of P21 in relation to cell proliferation have been confirmed in the HTR8/SVneo cells, and in this study, we identified similarly that cells with defects in proliferation had higher P21 expression levels, compared to control [18].

We have previously shown that in human endometrial epithelial cells, knockdown of TPP1 impairs cell adhesion by targeting BCL2 and P53 [10]. Although TPP1 knockdown similarly affected cell adhesion in the HTR8/SVneo cells, no significant changes were recorded on the expression of BCL2 and P53, suggesting cell type-specific regulation of targeted genes to control cell adhesion. We have also reported that TPP1 knockdown in primary human endometrial stromal cells does not impact decidualization or expression of decidualization markers [10]. In this study, TPP1 was detected at appreciable levels in the decidua and knockdown of TPP1 in HTR8/SVneo cells did not impact cell invasion, we cannot exclude a potential indirect regulation of EVT invasion via decidual cells.

5. Conclusions

Our study identified that TPP1 knockdown in HTR8/SVneo cells impaired cell proliferation, as evidenced by both xCEL-Ligence analysis and effects on the expression of proliferation markers MKI67 and P21. It remains to be determined if TPP1 dysregulation in the first-trimester placenta of women causes obstetric complications.

Abbreviations

EVT: Extravillous trophoblast
TPP1: Tripeptidyl peptidase 1
CLN2: Neuronal ceroid lipofuscinosis type 2
TBS: Tris-buffered saline
DPBS: Dulbecco’s phosphate-buffered saline
FBS: Fetal bovine serum
BCL2: B-cell lymphoma 2
MKI67: Marker of proliferation Ki-67
TERT: Telomerase reverse transcriptase
CLN: Ceroid-lipofuscinosis, neuronal
STAT3: Signal transducer and activator of transcription 3.

Data Availability
All data are provided in the submitted manuscript.

Conflicts of Interest
The authors declare that they have no conflicts of interest.

Acknowledgments
We are grateful to Ms Emily-Jane Bromley, Ms Judi Hocking, Dr Paddy Moore, Dr Ian Roberts, and Dr Jeanette Henderson for orchestrating the tissue collection and the women who donated tissue. We thank Dr Ellen Menkhorst for assistance with the xCELLigence analysis. This work was supported by a project grant (GNT1098332) and a senior research fellowship (#550905; E.D.) from the National Health and Medical Research Council of Australia.

Supplementary Materials
Supplementary 1. Supplementary Figure S1. Immunolocalization of TPP1 in the decidual cells throughout gestation. The immunolocalization of HLA-G was also determined in the first trimester decidua using serial sections to distinguish EVT.

Supplementary 2. Supplemental Table 1. Primers used throughout this study.

References