

Review Article

An Updated Review on Efficiency of *Penthorum chinense* Pursh in Traditional Uses, Toxicology, and Clinical Trials

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Traditional Chinese medicines (TCM) play an important role in the control and treatment of several animal diseases. *Penthorum chinense* Pursh (PCP) is a famous plant for its use in traditional medication practice and therapeutic effects in numerous pathological conditions. In China, PCP is utilized for both food and medication due to numerous bioactivities. PCP is widely administered in prevention and treatment of traumatic injury, edema, and liver diseases with functions of reducing swelling, support diuresis, blood stasis, and mitigation symptoms of excessive alcohol intake. Recently, PCP highlighted for research trials in various fields including pharmacology, pharmacognosy, cosmeceuticals, nutraceuticals, and pharmaceuticals due to medicinal significance with less toxicity and an effective ethnomedicine in veterinary practice. PCP contains diverse important ingredients such as flavonoids, organic acids, coumarins, lignans, polyphenols, and sterols that are important bioactive constituents of PCP exerting the therapeutic benefits and organ-protecting effects. In veterinary, PCP extract, compound, and phytochemicals/biomolecules significantly reversed the liver and kidney injuries, via antioxidation, oxidative stress, apoptosis, mitochondrial signaling pathways, and related genes. PCP water extract and compounds also proved in animal and humans' clinical trial for their hepatoprotective, antiaging, nephroprotective, anti-inflammatory, antidiabetic, antibacterial, antiapoptotic, immune regulation, and antioxidative stress pathways. This updated review spotlighted the current information on efficiency and application of PCP by compiling and reviewing recent publications on animal research. In addition, this review discussed the toxicology, traditional use, comparative, and clinical application of PCP in veterinary practices to authenticate and find out new perspectives on the research and development of this herbal medicine.

1. Introduction

In recent hundred years, researchers focus in on *Penthorum chinense* Pursh (PCP), and PCP is a perennial herb, Chinese traditional medicine used for its hepatoprotective effects such as liver edema, infectious hepatitis, and liver injuries [1, 2]. It is well established that PCP extract and PCP biomolecules have a range of pharmacological and health-promoting benefits, including

anti-inflammatory, hepatoprotective, and antioxidant properties [3–5]. Hepatoprotective effect of PCP is due to cure liver damage by keeping oxygen-free radicals out and reducing inflammatory response [6–9]. These pharmacological effects result from the PCPE's important bioactive constituents, such as the flavonoids (quercetin, 5-hydroxy-flavanone-7-O-D-glucoside, and kaempferol), lignans, coumarins, steroids, polyphenols, terpenoids, pinocembrin, catechins, and organic acids [10].

Various researches have shown that many extracts from PCP have significant pharmacological activities of antioxidation and anti-inflammation [3–5]. PCP is a traditional medication that has a high concentration of medicinal ingredients such as organic acids, flavonoids, and terpenoids. It is mostly used to treat liver illness and has excellent curative and little harmful effects. According to some studies, *Penthorum chinense* Pursh extracts contain the compounds quercetin, pinocembrin, catechins, 5-hydroxy-flavanone-7-O-D-glucoside, and kaempferol. It is proved with infrared spectroscopy that flavonoids (kaempferol, quercetin, and pinocembrin-7-O-beta-D-glucoside) are dominant biomolecules found in PCPE, which produced liver-protective effects, antioxidation, and anti-inflammation [11, 12].

PCPEs primarily consist of flavonoids. We also confirmed via infrared spectroscopy that PCPE included a high concentration of flavonoids and polyphenols, which may be responsible for the hepatoprotective effect. For instance, it has been noted that many flavonoids and phenolic compounds extracted from PCP have potent antioxidant and anti-inflammatory properties as well as significant liver-protective effects. These compounds include kaempferol, quercetin, and pinocembrin-7-O-beta-D-glucoside; in China, 2000 years ago, TCM is used to treat a variety of diseases in human clinical practices, and recently, TCM draw more attention and popularity due to their successful and safe experimental clinical trial [13].

The domains of life sciences have been modernized by the application of integrated pharmacology and biological networks. The technique of multiple target control is used to predict the primary active ingredient and potential target populations of traditional Chinese medicine and to develop the mechanism by which TCMs exert curative effects. It is based on the construction of drug-target networks and analysis of network characteristics [14, 15]. However, the main barrier to TCM's acceptability globally is the inability to clearly explain its application method due to its complicated components. Because the primary components are the chemical substances that produce the medication effects, a study of the chemical components in TCM will help to clarify the mechanism. Renewed studies gained interest in PCP research and greatly increased potential use of PCP against liver diseases, but varieties of animal models, disease-induced models, cross studies, and dosage use of PCP rigorous demand to reach the existing findings. Furthermore, studies focused on pharmacology and chemical constituents only found the activities and effects of the active ingredients of PCP such as polyphenols and other compounds and their mechanism of actions. This updated review links a bridge between the previous and recent pharmacological studies of PCP and a revised summary of current progress in the aspect of pharmacology, toxicology, and clinical trials, which highlights the evidence for use of PCP and points out the scientific gaps in the future research.

2. Medical Resource and Chemical Constituents

Most flavonoids have been found such as pinocembrin, quercetin, and kaempferol derivatives and isomers. 2,3'-

Dihydroxy-3-methoxy-6'-methanone-benzophenone-4-O-glucoside and 2,4-dihydroxy-3-methoxy-6'-methanone-benzo-phenone-3'-O-glucoside are two of the four phenylpropanoids [16, 17]. Quercetin, the primary active component of PCP found to be useful for treating alcoholic liver injury, was discovered in the PCP aqueous extract [18].

The structural formula of 38 active compounds, isomers, and derivatives is given in Figure 1 including chebulic acid, gallic acid, ethyl gallate, bergenin, penthorummin C, 2,6-dihydroxyacetophenone-4-O-glucoside, 2,6-dihydroxyacetophenone-O-[4',6'-hexahydroxydiphenyl]-glucoside, quercetin-di-O-glucoside, pinocembrin-7-O-glucoside, pinostrobin, pinocembrin, pinocembrin-7-O-[4',6'-hexahydroxydiphenyl]-glucoside, pinocembrin-7-O-[3'-O-galloyl]-glucoside, isomer of pinocembrin-7-O-[3'-O-galloyl]-glucoside, quercetin-3-O-glucoside, kaempferol-3-O-rutinoside, quercetin-3-O-rhamnoside, kaempferol-3-O-arab infuranoside, kaempferol-3-O-rhamnopyranoside, luteolin, quercetin-di-O-glucoside, rutin, quercetin-3-xyloside, quercetin-3-O-arabinofuranoside, quercetin, apigenin, 2,4-dihydro-3-methoxy-6'-methanone-benzophenone-4-O-glucoside, 2,4-dihydro-3-methoxy-6'-methanone-benzophenone-3'-O-glucoside, penchinone A, penchinone B, catechin, epicatechin, brevifolin carboxylic acid, penthorummin C, and penthorummin B, with molecular formula, i.e., $C_{14}H_{13}O_{11}$, $C_7H_7O_5$, $C_9H_{11}O_5$, $C_{14}H_{17}O_9$, $C_{26}H_{25}O_{17}$, $C_{14}H_{19}O_9$, $C_{21}H_{23}O_9$, $C_{16}H_{15}O_4$, $C_{35}H_{29}O_{17}$, $C_{28}H_{27}O_{13}$, $C_{28}H_{27}O_{13}$, $C_{21}H_{21}O_{12}$, $C_{27}H_{31}O_{15}$, $C_{21}H_{21}O_{11}$, $C_{20}H_{19}O_{10}$, $C_{21}H_{21}O_{10}$, $C_{15}H_{11}O_6$, $C_{27}H_{31}O_{17}$, $C_{27}H_{31}O_{16}$, $C_{20}H_{19}O_{11}$, $C_{20}H_{19}O_{11}$, $C_{15}H_{11}O_7$, $C_{15}H_{11}O_5$, $C_{25}H_{29}O_{11}$, $C_{25}H_{27}O_{11}$, $C_{19}H_{19}O_6$, $C_{19}H_{19}O_6$, $C_{15}H_{15}O_6$, $C_{15}H_{15}O_6$, $C_{13}H_9O_8$, $C_{27}H_{25}O_{17}$, and $C_{12}H_{11}O_8$, respectively [9, 16, 17, 19, 20].

Recently, there are new phenolic compound, flavonoids, and neolignans, namely, (4'E)-2,3'-dihydroxy-3-methoxy-6'-methanone-benzophenone-4-O-β-D-glucopyranoside, (4'E)-2,4-dihydroxy-3-methoxy-6'-methanone-benzophenone-3'-O-β-D-glucopyranoside [21], 20,60-dihydroxydihydrochalcone-40-O-[200-O-galloyl-400,600-hexahydroxydiphenyl]-b-D-glucopyranoside, pinocembrin-7-O-[300-O-galloyl]-b-D-glucose, pinocembrin-7-O-[200-O-galloyl-400,600-hexahydroxydiphenyl]-b-D-glucose [9], (70 Z,8S)-3,8-dihydroxy-4-methoxy-2,40-epoxy-8,50-neolign-70-en-7-one-20-O-b-D-glucopyranose, and (70 Z,8S)-4,8-dihydroxy-3-methoxy-2,40-epoxy-8,50-neolign-70-en-7-one-20-O-b-D-glucopyranose [22], which have protective effect on the liver and are helpful in free radical scavenging activities, antihyperlipidemic activities, and anti-proliferative on hepatic stellate T6 cells (HSC-T6 cells), respectively.

3. Clinical and Comparative Activities

3.1. Antioxidation. Generally, ROS production is involved in regular cell metabolism, and it is generally compensated through antioxidant defense system to balance the specific redox stability [23]. Oxidative stress is a biological condition in which free radicals across the antioxidant capabilities, and

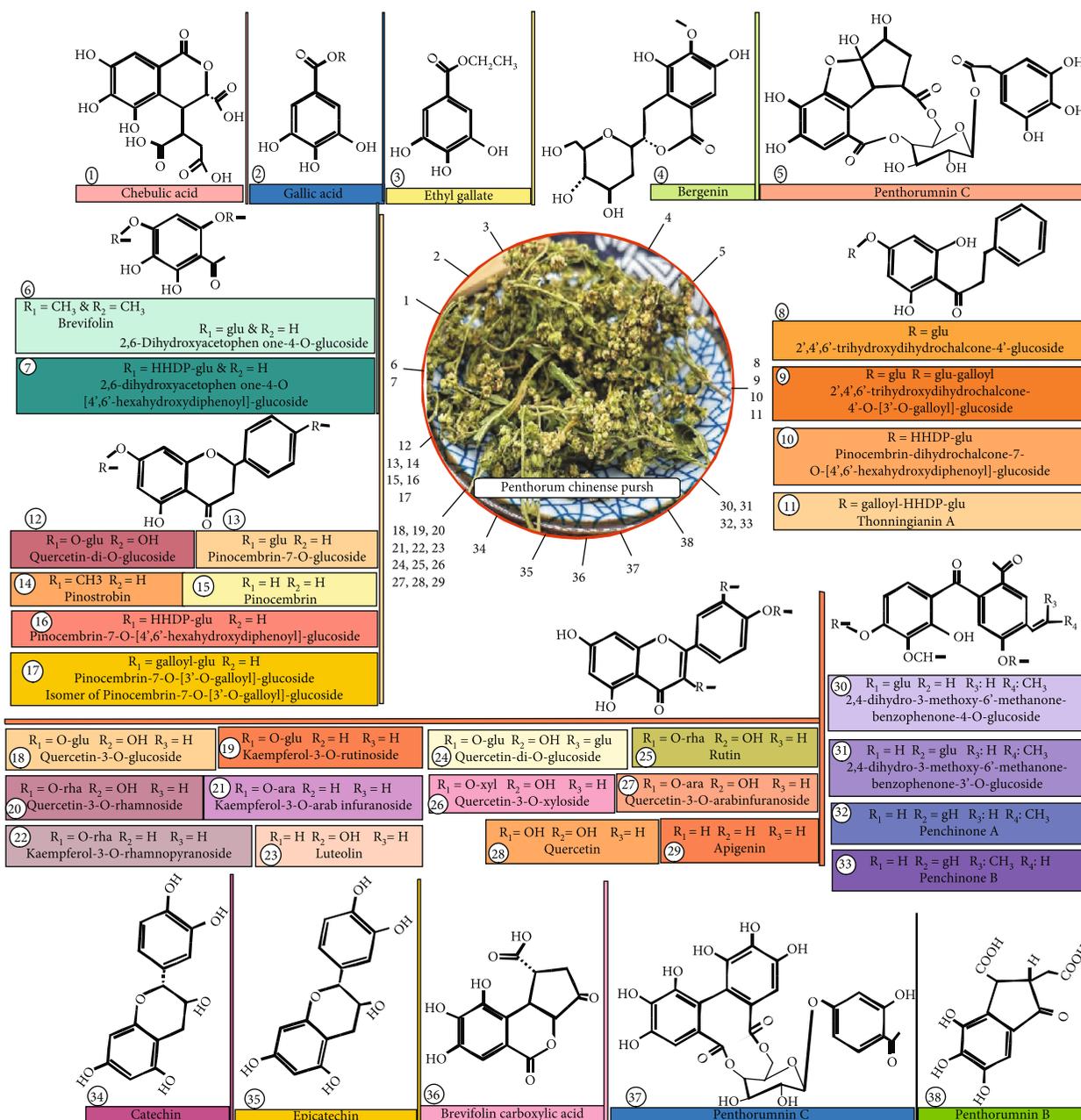


FIGURE 1: Dioramic representation of different compounds, their derivate, and isomers present in PCP.

oxidative stress is identified as major element in several diseases such as chronic liver diseases, such as hepatitis and alcoholic and nonalcoholic fatty liver diseases [24]. The liver is a vital organ with metabolic activities and normal physiological functions. The functions of the liver are correlated with gastrointestinal tract (GIT), and the disorder between this balance may danger to drug toxicity and introduction of xenobiotics within the organism [25, 26]. Therefore, the antioxidant treatment is very effective for the controlling the oxidative stress conditions and correct or control the equilibrium between the antioxidant and oxidants in the encouragement of the liver pathogenesis and protection of hepatocytes from excessive exposure to oxidative stress.

PCP-rich source of natural antioxidants is recently the focus of pharmaceutical, cosmaceutical, nutraceutical, and food industries [27]. Polyphenols demonstrated potent hepatoprotective benefits against oxidative injury by directly scavenging ROS and lowering liver enzymes as well as indirectly increasing antioxidant levels [12]. It was proved that activation of Nrf2 signaling pathway by PCPE produced hepatoprotective effect in CCl4-induced oxidative stress model [28].

Flavonoids upregulated the activities of superoxide dismutase and catalase while downregulating the level of malondialdehyde [29]. PCP and polysaccharide fraction PCPP-1a possessed strong hydroxyl radical scavenging activity, Fe²⁺ chelating, and DPPH radical scavenging [30, 31].

Pinocembrin decreases the oxidation by increasing scavenging of the free radical and antisuperoxide formation when taken in 30 μM in rat [32] while increasing the cellular antioxidant defense and metal ion chelation [33]. Quercetin with different concentrations (10, 50, and 100 μM) in rat resulted in decreases of H_2O_2 stress, ROS production, and ER stress [34, 35]. Kaempferol downregulated the production of ROS, mitochondrial membrane potential, and intracellular and intracellular ROS production [36, 37], due to scavenging ROS and activating the Nrf2-antioxidant signaling pathway. Polyphenols from PCP demonstrated a potent protective effect against high glucose- (HG-) associated vascular inflammation [3–5].

Thonningianin A (TA), a substance derived from PCP, effectively decreased the quantity of ROS in human umbilical vein endothelial cells that are stimulated by H_2O_2 HUVECs. Additionally, following the administration of TA, the expression of pro- and cleaved-IL-1 in the aortic artery of ApoE-KO mice was also reduced at the transcriptional and posttranscriptional levels, which may be related to the decrease in oxidative stress-related Nod-like receptor protein 3 (NLRP3) in the aortic arteries of ApoE-KO mice [5]. The structure of ThA is a combination of gallic acid esters of glucose and dihydrochalcone. The antioxidant properties of ThA have been previously reported [32]. Synthetic antioxidant (tannic acid) and ThA found similar activities; however, ThA is more effective than gallic acid, vitamins E and C in LPO-induced, and the deoxyribose assay. The properties of ThA are important for the free radical-mediated disease inhibition, and further studies are needed to explore the in vitro and in vivo experiments. Figure 2 represents the oxidative stress production and protective effect of PCP.

3.2. Anti-Inflammatory. Inflammation process is well-known process to promote the pathological and physiological pathways by activating the various other systems such as the immune system, vascular system, and other cells within the damaged tissues [38]. The acute and chronic inflammation is caused by the several factors including some microorganism infection, chemical, surgical, and physical irritation. The classic type of inflammations is heat, edema, pain swelling, and redness [39]. Chronic inflammation or prolonged inflammation may also affects many other organs systems such as the heart, lungs, brain, and skin [40]; however, the chronic inflammation also connected with various pathogenesis and tissues damages can cause the serious cellular injury and variety of disease conditions such as Alzheimer's disease, diabetes, and carcinogenesis [41].

Penthorum chinense Pursh shows anti-inflammatory capabilities due to the presence of several polyphenol compounds that inhabits inflammation by activating the nuclear erythroid 2-related factor 2 in the liver of mice while downregulation of heme oxygenase⁻¹ (Nrf2/HO⁻¹) signaling pathway in the hepatocytes of human beings [42]. Other compound present in PCP decreases the amount of proprotein convertase subtilisin/kexin type 9 (PCSK9) and activates the low-density lipoprotein receptor (LDLR) in the hepatics cells [43]. It protects cell by anti-inflammatory mechanism like increasing the expression of anti-inflammatory cytokines interleukin-10

and TGF- β in fish intestine cells [44]. Similarly, PCP shows anti-inflammatory effect by downregulating mitogen-activated protein kinase (MAPK) and nuclear factor κB (NF- κB) signaling pathways [45]. PCP and its related compounds proved for anti-inflammatory effects could reverse inflammation of kidney, liver, and nervous tissues by interacting with discussed signaling pathways. Recent studies have also shown that PCP downregulated the overexpression of inflammatory cytokines and mediators in LPS-induced inflammatory response in animals and significantly inhibited the expression level of NO, TNF- α , and IL-1 β , and it shows the significant anti-inflammatory effect together inhibiting the MyD88/TLR4/NF- κB signaling pathways [46, 47]. Tao et al. studied the hepatoprotective effect of the PCPE in CCl₄-induced liver injury in dogs and found that the NF- κB and MAPK signaling pathways in dogs are associated to the antioxidant and anti-inflammatory effects of PCPE. Inflammation that occurred in liver tissues induced by CCl₄ has been controlled through the administrations of PCPE in dogs, and inflammatory and proinflammatory factors (IL-1 β , IL-6, and TNF) are the important mediators, involved in the response of inflammation, and anti-inflammatory factor such as IL-10 turns as an antagonistic inflammatory mediator pathways controlled in acute liver injury in dogs [45].

3.3. Antitoxic. In the view of the toxic effects of synthetic drugs, collected data resulted that existing treatment options have been limited the therapeutic success in animals and humans. Considerably over the last decade, the therapeutic use of herbal/plant medicines has been increased over the world. Recent severe complications and new disease with no treatment has promoted the belief that natural medicines are safe and less toxic. PCP is a nontoxic herbal medicine, and well-established reports indicated that PCP singly or with combined therapy is proved to be safest herbal medicine with no or negligible toxicity or side effects whether administered in dose/or time-dependent way [1]. It is reported that PCP induced cytotoxic effect possibly due to accumulation of reactive oxygen species (ROS) [2], while the extract of bioactive parts of PCP like stem, leaves, and flowers is used to protect cells from oxidants toxic damages as induced with H_2O_2 [48] [12]. The flavonoid contents from leaf of PCP protected the liver cells from lipotoxicity injury [1]. The toxic compound produced by several bacterial is also regularized by extract fraction of PCP like PGF and other components, i.e., pinocembrin-7-O-[4'',6''-hexahydroxydiphenoyl]- β -D-glucose, thonningianin A, and pinocembrin-7-O-[3''-O-galloyl-4',6'-(s)-HHDP]- β -D-glucose (PGHG), that reduce the activity of methicillin resistance *S. aureus* (MRSA) [49]. The toxicity produced by aflatoxin β 1 is one of the major toxicities in broiler chicken; this toxicity can be prevented by the use of PCPC as a natural and safe agent with supplemented in the diet [50]. PCP and its compounds could be included in the human and veterinary diet due to the best detoxification properties.

Pinocembrin, the primary flavonoid isolated from PCP, can be used as neuroprotective agent in cerebral ischemic injury along with the pharmacological effects almost in various systems. This flavonoid attracted recent interest due to the

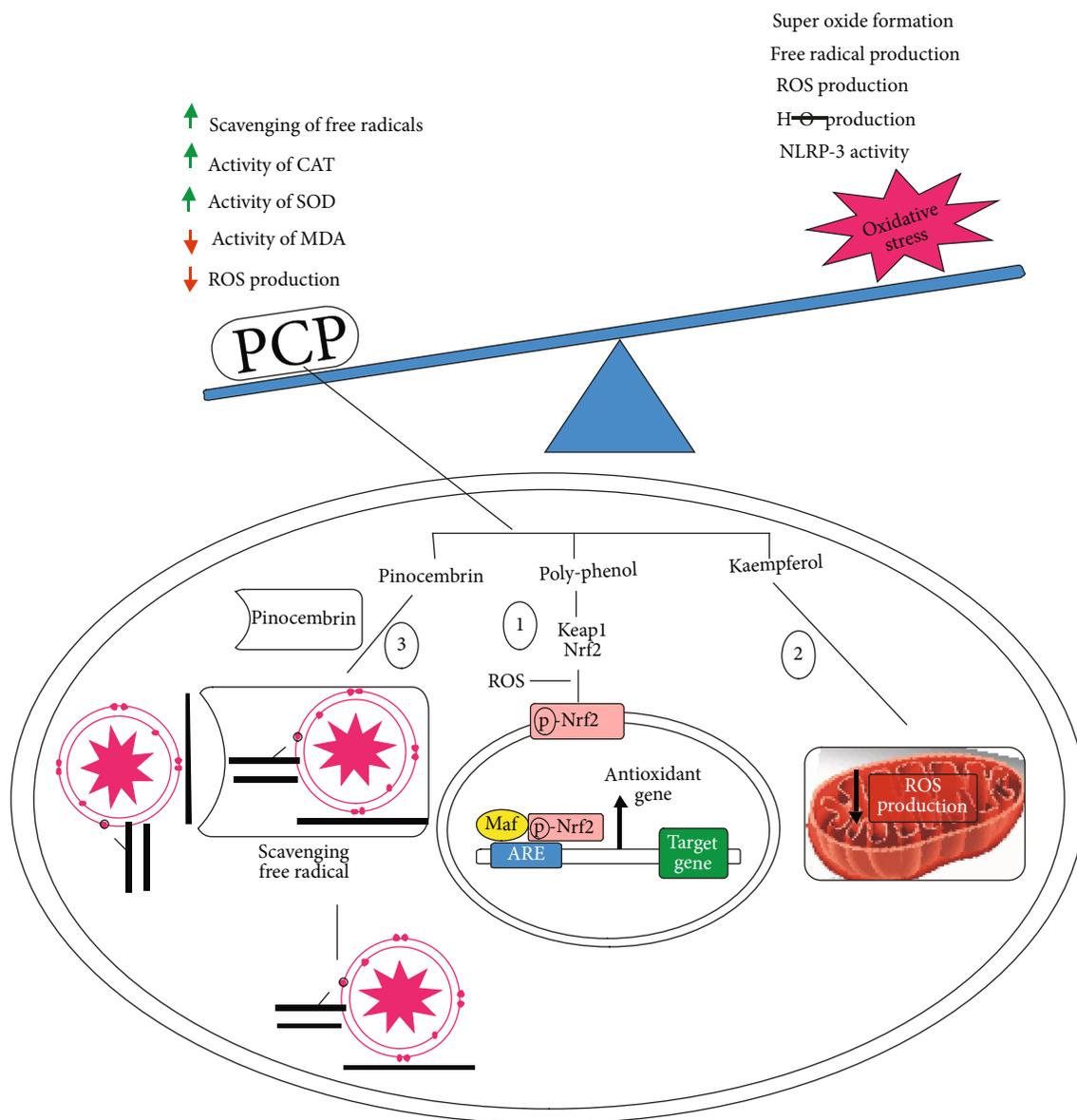


FIGURE 2: Diagram representation of oxidative stress production and protective effects of PCP. (1) Polyphenol reduces the ROS production by Nrf2 pathway. (2) Kaempferol downregulates ROS production in mitochondria. (3) Pinocembrin reduces the oxidative stress by scavenging free radicals

antitoxic effects [6, 51]. Quercetin is one of the polyphenolic compounds isolated from PCP and widely studied for anti-inflammatory and antioxidant compound. Quercetin prevents alcohol-induced hepatotoxicity [52], has scavenging properties [53], decreased the cytotoxicity [54], inhibits neurotoxicity [55], and possesses several medicinal benefits. Herbal antioxidants are major supporting agents in the battle of diseases and infections. There is a wide published data regarding PCP, their extract, or compound, there is no published information on their toxicity, and importantly, most of published data is available on their beneficial, scavenging, and biological effect when administered traditionally in a mixture or single form in animals. It is crucial to evaluate the advantageous application of these herbs when used in a combination or single according to a precisely defined recipe.

3.4. *Antimicrobial.* Flavonoids present in PCP showed inhibitory effects on many strains of bacteria including *Escherichia coli*, *Pseudomonas aeruginosa*, and *Cryptococcus neoformans* by destroying the microbial membrane, inhibiting the invasion of bacteria into the host cell, increasing the likelihood of bacterial apoptosis, and ceasing the bacterial fatty acid synthesis [56].

Flavonoids present in PCP are proved for antiviral capability [44, 57, 58], so PCP could be included in medicinal preparation of acute liver disease (acute viral hepatitis, chronic active viral hepatitis, and hepatitis B virus) [6, 28, 59] interfering with viral replication [1, 60]. Similarly, gallic acid also protects the liver from viral infections [61]; similarly, flavonoids also affected the indices of TNF- α , IL-6, and IL-1 β in infection [56].

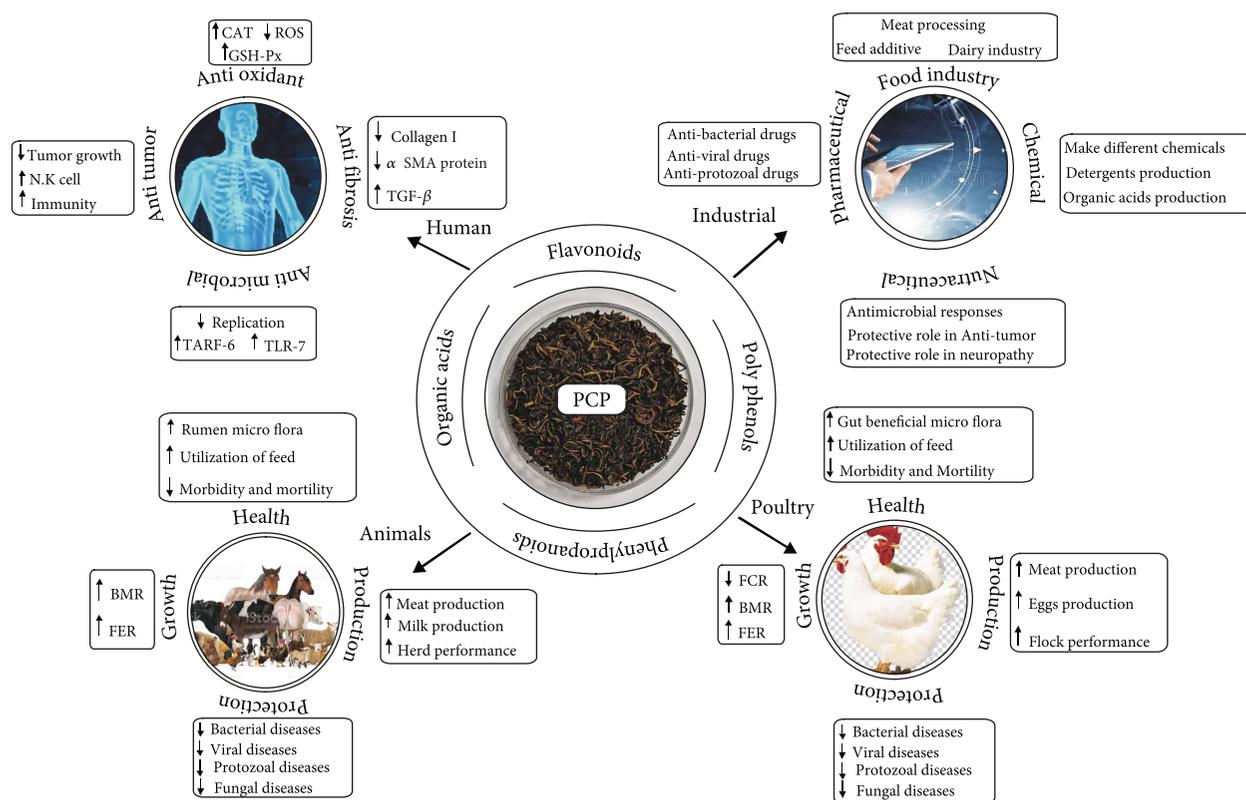


FIGURE 3: Schematic diagram illustrating the therapeutic effects and underlying mechanism of action, which showed the beneficial effect on poultry and animal health and production, highlighting the industrial application of PCP.

The flavonoid reported for antiviral effects against influenza A virus by an interaction with various mechanisms increases the expression of PKC α (protein kinase C alpha), VIPR1 (vasoactive intestinal polypeptide receptor 1), retinoic acid-inducible gene (RIG)-1, TRAF6 (TNF receptor associated factor 6), and Toll-like receptor (TLR)-7 [28] and by regulating the eTLR7, RIG-1, and AQP5 signaling pathways [56]. Furthermore, biomolecules of PCP can be investigated for their interactions with various signaling pathways and in various species/or strains of microbes.

3.5. Health Benefits in Animals. PCP (Penthoraceae) traditionally has been used in China for the treatment of liver-related problems (infectious hepatitis, cholecystitis, jaundice, infectious hepatitis, edema, and antidrunk hangover). PCP showed hepatoprotective effects in broiler and other animals by decreasing liver injuries via various mechanisms. In ethanol-induced liver injury mice model, aqueous extract of PCP shows protective effect via decreasing CYP2E1-mediated oxidative stress and boosting oxidant defense mechanisms through activation of the Nrf2/HO-1 pathway. PCP also reported for correction of heat-related problems, diuresis, circulation activation, and protection of the spleen and liver [62]. PCP is involved in the reversion of jaundice and viral hepatitis and further corrected the serum indices of various biochemicals (insulin, triglycerides, TC, LDL-C, and HbA1c) and oral glucose tolerance test [63]. The liver injuries by carbon tetrachloride (CCl₄) can be improved by treatment of PCP. Malondialdehyde levels are reduced, glu-

tathione (GSH) is restored, superoxide dismutase (SOD) and catalase (CAT) activities are improved, hepatic cytochrome P450 2E1 (CYP2E1) is prevented from degrading, and nuclear factor erythroid 2-related factor 2 (Nrf2) and its target proteins are improved in CCl₄ treated mice [28]. In many studies, PCP has been shown to lessen inflammation, liver fibrosis, viral infection, the balance of important liver enzymes, the activation of hepatic stellate cells, and hepatic virus DNA replication. Animal testing, however, reveals neither toxicity nor negative effects [60].

In dogs, acute liver injury due to CCl₄ and oxidative stress due to which free radicals are produced that generate free radicals can directly cause lipid peroxidation in the cell membrane and cause cell membrane destruction. Hepatocytes can experience oxidative stress, degeneration, hepatocellular injury, and necrosis by the production of ALI. The ALI has shown improvement in vacuolar inflammatory lesions in liver tissues when treated with PCPE, restored glutathione peroxidase, enhanced activity of superoxide dismutase, and significantly lowers the serum levels of nitric oxide and malondialdehyde. By using PCPE, inflammatory factors were downregulated and anti-inflammatory factors were upregulated. In dogs with ALI, PCPE therapy decreased the levels of MEKK4, MKK3, p38MAPK, MSK1, and NF-B and increased the levels of I κ B mRNA [45]. Similarly, PCP blocked the expression of cytochrome P450 2E1 and production of intracellular reactive oxygen species and decreases liver fat accumulation and oxidative damage. It upregulates the nucleus factor E2-related factor 2 (Nrf2)

TABLE 1: A summary of studies with PCP and biological effects on animals.

| Study model/ animal type | Dose and extract type | Results | Treatment and overall effect | Reference |
|--|--|--|---|----------------------|
| Chicken | 1, 2, and 3 g PCPE/kg feed | <p>↑ growth</p> <p>↑ immunoglobulin level</p> <p>↓ oxidative stress</p> <p>↓ Bax, Bak, caspase-9, caspase-3, and p53</p> <p>↓ pathological lesions in liver</p> | Treatment of liver injury and oxidative stress | Nabi et al. [25, 26] |
| Chicken | 5, 10, and 15 mL PCPC/kg feed | <p>↓ Bax, Bak, cytochrome c, caspase-9, and caspase-3)</p> <p>↑ NRF2 and HMOX1</p> <p>↓ apoptosis in the kidneys</p> <p>↓ vacuolar inflammatory lesions in liver tissues</p> | Mitochondrial pathways in the kidneys | Tao et al. [45] |
| Dog | 0.5, 1, and 1.5 mL PCP/dog herb mL/dog | <p>↓ IL-1β, IL-6, TNF-α, MEKK4, MKK3, p38MAPK, MSK1, and NF-κB</p> <p>↑ IL-10, IκB mRNA</p> <p>↑ activity of superoxide dismutase</p> | Hepatoprotective effect in dogs | Tao et al. [45] |
| HSCs LX-2 and HSC-T6 cells in rat | 100 μ g/mL PCE | <p>↓ collagen I</p> <p>↓ α-SMA protein levels</p> <p>↓ PI3K-Akt pathway</p> <p>↑ TGF-β-Smad pathway</p> | Liver fibrosis | Zhou et al. [15] |
| Female Kunming mice 20 \pm 2 g and 6 to 8 week | 100 and 300 μ g/mL purified polysaccharide fraction of leaves of PCP | <p>↑ immunity, i.e., NK cells and lymphocytes</p> <p>↓ tumor growth</p> | Anticancer | Chen et al. [69] |
| Mice | PCE 166.6 \pm 20.1 U/mL | <p>↑ CAT and GSH-Px</p> | Antioxidant | Yin et al. [16] |
| HUVEC cells/Mice | PCE 10-30 μ g/mL | <p>↑ scavenging ROS</p> <p>↑ Nrf2-antioxidant signaling pathway,</p> <p>↓ oxidative-stress,</p> <p>↓ pro- and cleaved-IL-1β</p> | Treatment of cardiovascular diseases, Autophagy induction | Sun et al. [3-5] |
| Liver cells (L-02 cells cultured) | — | <p>↑ expression of TNF-α and IL-6 at mRNA and protein levels</p> <p>↓ mRNA and protein expression of Nrf2 and HO⁻¹</p> | Protect liver injury | Ding et al. [70] |
| Rat | PCE 545 mg/kg/da | <p>↑ GSH-Px, SOD, and CAT</p> <p>↓ levels of MDA</p> <p>↑ scavenging ROS</p> | Diabetic treatment | Hu et al. [71] |
| Normal rat's liver cell (BRL-3A) | 6.25-100 μ g/mL stem extract | <p>↓ liver enzymes directly</p> <p>↑ antioxidant levels</p> <p>↓ cellular apoptosis</p> <p>↓ lipid peroxidation</p> | Hepatoprotective activity | He et al. [12] |
| APP/PS1 mice | Thonningianin A (10 μ M) compound from PCP leaves, stems, or flowers | <p>↑ in microglial cells triggered by A (1-42), NLRP3 inflammasome is degraded by autophagy</p> <p>↓ neural destruction</p> | Alzheimer's disease treatment | Zhou et al. [72] |
| Mice | PCP aqueous extracts 10.30 g/kg | <p>↓ oxidative stress caused by CYP2E1</p> <p>↑ oxidant defense mechanisms</p> <p>↑ HO⁻¹ and Nrf2 pathway</p> | Prevents ethanol-induced chronic liver damage | Cao et al. [62] |
| Rats | PC extract 150 and 300 mg/kg/day | <p>↓ HbA1c, TG, and TC</p> <p>↑ insulin</p> | Antihyperglycemic effects | Suna [63] |
| Zebra fish | PCP polyphenols | <p>↑ Keap1-Nrf2</p> | Neuro-protective | Sun et al. [73] |
| Zebra fish larvae | PCPE 25, 50, and 100 μ g/mL for 48 h | <p>↑ Keap1/Nrf2</p> <p>↓ mTOR/PI3K/Akt P2X7R blocking</p> | Hepatoprotective via antioxidation and autophagy | Zhao et al. [46, 47] |

TABLE 1: Continued.

| Study model/ animal type | Dose and extract type | Results | Treatment and overall effect | Reference |
|-------------------------------|---|---|---|---------------------|
| HEK293T cells B16F10 cells | 50, 100, or 200 $\mu\text{g}/\text{mL}$ Pc-EE for 24 h | ↓ tyrosinase ↓ melanin ↓ LC3B | Antiapoptotic, antiaging, anti-inflammatory, and antimelanogenic properties | Jeong et al. [7, 8] |
| Rat | 3 g/kg | ↑ cell proliferation inhibition ↑ scavenging ability | Antioxidant Antihepatocarcinoma | Lu et al. [74] |
| RAW264.7 cell with LPS | 15, 30, 60, and 120 $\mu\text{g}/\text{mL}$ | ↓ NO, TNF- α , and IL-1 β | Anti-inflammatory | Lin et al. [30] |
| Rat | 800 mg/kg | ↓ TG, TC, ALT, and AST | Antilipogenesis | Yuan and Ou [75] |
| HepG2 cells | 1, 10, and 100 μM | ↑ AMPK/SRIT1 ↑ PPAR- α | Antioxidant | Guo et al. [76] |
| Fish | 5.2 to 6.1 mg/L | ↓ Sobs, ACE, and Chao indices ↑ Bacteroides ↓ Actinobacteriota and Fusobacteriota | Anti-inflammatory | Ke et al. [44] |

PCE: Penthorum chinense extracts; HSCs: hematopoietic stem cells; LX-2: human hepatic stellate cell line; PI3K-Akt: protein kinase B; TGF- β : transforming growth factor; α -SMA: α -smooth muscle actin; NK cells: natural killer cells; CAT: catalase; GSH-Px: glutathione peroxidase; ROS: reactive oxygen species; Nrf2: nuclear factor erythroid 2-related factor; COX-2: cyclooxygenase 2; IL-6: interleukin-6; IL-1 β : interleukin-1 β ; NLRP3: Nod-like receptor protein 3; TNF- α : tumor necrosis factor- α ; SOD: superoxide dismutases; MDA: malondialdehyde; CYP2E1: cytochrome P450 2E1; HbA1c: serum glycosylated hemoglobin A1C; TG: triglyceride; TC: total cholesterol; AMPK: AMP-activated protein kinase; TG: total cholesterol; TG: triglyceride; NO: nitric oxide.

and downregulates the expression of Kelch-like ECH-associated protein 1 (Keap 1). PCP upregulates autophagy signaling pathways [1, 60].

Administration of PCP extract in broiler chicken in aflatoxin B1-induced liver damage and oxidative stress can be reduced significantly. Aflatoxin poses a great threat to the poultry industry. PCP enhances growth performance, immunoglobulin level, and oxidative capability and reversing oxidative stress and pathological lesions in the liver; PCP administered to AFB1-affected birds lessens the negative effects; also, apoptosis was reversed [25, 26]. Figure 3 is a schematic diagram illustrating the therapeutic effects and underlying mechanism of action PCP in different animals. Table 1 summarizes the studies with PCP and biological effects in different animals.

3.6. Immunomodulatory Effects. Traditional Chinese medicines are recognized as balancing methodology to modern medication and most promising and a safe alternative therapeutic alongside proven the immunomodulation agents in clinical practice [64, 65]. Immunomodulators promote the defense mechanism against the pathogens including viruses, thus supporting the immunity and homeostasis and could be an effective way to prevent the infections and pathogenesis. Several chronic diseases, allergies, viral infections, immune disorders, metabolic diseases, cancer, and inflammations are associated with immune system. Flavonoids, sterol compounds, polysaccharides, carotenoids, and terpenoids are important phytochemicals with well-known chemical structures and significant immunomodulating properties [66]. PCP contains the main types of components that are polyphenols, flavonoids, coumarins, lignans, organic acids, and sterols. It is proved that polysaccharides (galactose, arabinose, galacturonic acid, rhamnose, and glucose are polysaccharides) in PCP are involved in the immunoregulation of H₂₂ tumor-bearing mice. In vitro research indicates that PCP-4 inhibits the growth of xenograft tumors by

safeguarding immune organs, enhancing immune cell activity, and encouraging apoptosis [67]. PCP in common carp (*Cyprinus carpio*) improved the gut microbiota population and finally improved the intestinal immunity [44].

Gallic acid from PCPE is investigated for the immunomodulatory effects in immunosuppressed Swiss albino mice (cyclophosphamide and cisplatin). Gallic acid with different dosage (100, 200, and 400 mg/kg) regimens was orally administered for seven days. The results show that gallic acid could be used as an adjuvant immunosuppressive drug to decrease the adverse effects of immunosuppressive agents on the immune system [68, 69].

4. Conclusion and Future Directions

This updated review critically explained that PCP contains a wide range of medicinal important phytochemicals like flavonoids, phenylpropanoid, polyphenol, and organic acids. The bioactive component of PCP shows medicinal properties like antimicrobial, hepato, neuroprotection, anti-inflammatory, and strong immunoregulatory effects. Most of effects are mainly concerned with the regulation of apoptosis, mitochondrial, AMPK, autophagy, TLR4, Keap1-Nrf2, NF- κ B, p38-MAPK, IRT3-TGF- β , and Nrf2 signaling pathways. In poultry, it can be used as feed additive for health-promoting effects including liver tonic (detoxification of xenobiotics and repairing of liver), gut microflora for boosting immunity/or antimicrobial effect, and washing out of infectious microbes. In large animals, it protects different metabolic injuries improving gut health. In the future, it may be used in lactating animals for improving milk yield and activation of beneficial ruminal microbes' population. It is suggested that various industries (pharmaceutical, nutraceutical, food, and cosmetic) should increase the production capabilities due to variety of beneficial effects of PCP on animal health.

Abbreviations

| | |
|--------------------|---|
| ALP: | Alkaline phosphatase |
| ALT: | Alanine aminotransferase |
| AST: | Aspartate transaminase |
| ATGL: | Triglyceride lipase |
| CAT: | Catalase |
| CCl ₄ : | Carbon tetrachloride |
| DPPH: | 2,2-Diphenyl-1-picrylhydrazyl |
| GIT: | Gastrointestinal tract |
| GSH: | Glutathione |
| HG: | High glucose |
| HUVEC: | Human umbilical vein endothelial cells |
| IL: | Interleukin |
| LDLR: | Low-density lipoprotein receptor |
| LPS: | Lipopolysaccharide |
| MRSA: | Methicillin resistance <i>S. aureus</i> |
| MDA: | Malonaldehyde |
| NLRP3: | Nod-like receptor protein 3 |
| PCP: | <i>Penthorum chinense Pursh</i> |
| PCPC: | <i>Penthorum chinense Pursh</i> compound |
| PCPE: | <i>Penthorum chinense Pursh</i> extract |
| PCSK9: | Proprotein convertase subtilisin/kexin type 9 |
| ROS: | Reactive oxygen species |
| TNF- α : | Tumor necrosis factor- α |
| ThA: | Thoningianin A (TA) |
| TCM: | Traditional Chinese medicines |
| NO: | Nitric oxide |
| VIPR1: | Vasoactive intestinal polypeptide receptor 1. |

Data Availability

The data supporting this review paper are from previously reported studies and datasets, which have been cited.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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References

- [1] A. Wang, M. Li, H. Huang et al., "A review of *Penthorum chinense Pursh* for hepatoprotection: traditional use, phytochemistry, pharmacology, toxicology and clinical trials," *Journal of Ethnopharmacology*, vol. 251, article 112569, 2020.
- [2] A. Wang, L. Lin, and Y. Wang, "Traditional Chinese herbal medicine *Penthorum chinense Pursh*: a phytochemical and pharmacological review," *The American Journal of Chinese Medicine*, vol. 43, no. 4, pp. 601–620, 2015.
- [3] Y. Sun, L. He, W. Wang et al., "Polyphenols from *Penthorum chinense Pursh*. Attenuates high glucose-induced vascular inflammation through directly interacting with Keap1 protein," *Journal of Ethnopharmacology*, vol. 268, article 113617, 2021.
- [4] D. Huang, Y. Jiang, W. Chen, F. Yao, and L. Sun, "Polyphenols with anti-proliferative activities from *Penthorum chinense Pursh*," *Molecules*, vol. 19, no. 8, pp. 11045–11055, 2014.
- [5] X. Sun, A. Wu, B. Y. K. Law et al., "The active components derived from *Penthorum chinense Pursh* protect against oxidative-stress-induced vascular injury via autophagy induction," *Free Radical Biology and Medicine*, vol. 146, pp. 160–180, 2020.
- [6] Y. C. Du, L. Lai, H. Zhang et al., "Kaempferol from: *Penthorum chinense Pursh* suppresses HMGB1/TLR4/NF- κ B signaling and NLRP3 inflammasome activation in acetaminophen-induced hepatotoxicity," *Food & Function*, vol. 11, no. 9, pp. 7925–7934, 2020.
- [7] D. Jeong, J. Lee, S. H. Park et al., "Antiphotaging and Antimelanogenic Effects of *Penthorum chinense Pursh* Ethanol Extract due to Antioxidant- and Autophagy-Inducing Properties," *Oxidative Medicine and Cellular Longevity*, vol. 2019, Article ID 9679731, 14 pages, 2019.
- [8] D. Jeong, S. H. Park, M. H. Kim et al., "Anti-melanogenic effects of ethanol extracts of the leaves and roots of *Patrinia villosa* (thunb.) Juss through their inhibition of CREB and induction of ERK and autophagy," *Molecules*, vol. 25, no. 22, p. 5375, 2020.
- [9] W. W. Zhao, W. W. Guo, J. F. Guo, X. Wang, X. Q. Chen, and X. Wu, "Three new flavonoids from *Penthorum chinense Pursh* and their docking studies," *Natural Product Research*, vol. 35, no. 1, pp. 49–56, 2021.
- [10] H. Feng, Z. M. Wang, G. Y. Dong, and Z. Wu, "Studies on chemical constituents from *Penthorum chinense Pursh*," *Zhongguo Zhong Yao Za Zhi*, vol. 26, no. 4, pp. 260–262, 2001.
- [11] W.-W. Guo, F. Qiu, X.-Q. Chen, Y.-Y. Ba, X. Wang, and X. Wu, "In-vivo absorption of pinocembrin-7-O- β -D-glucoside in rats and its in-vitro biotransformation," *Scientific Reports*, vol. 6, no. 1, article 29340, 2016.
- [12] L. He, S. Zhang, C. Luo et al., "Functional teas from the stems of *Penthorum chinense Pursh*: phenolic constituents, antioxidant and hepatoprotective activity," *Plant Foods for Human Nutrition*, vol. 74, no. 1, pp. 83–90, 2019.
- [13] C. Liu, J. Z. Liao, and P. Y. Li, "Traditional Chinese herbal extracts inducing autophagy as a novel approach in therapy of nonalcoholic fatty liver disease," *World Journal of Gastroenterology*, vol. 23, no. 11, pp. 1964–1973, 2017.
- [14] C. Zhou, L. J. Liu, J. Zhuang et al., "A systems biology-based approach to uncovering molecular mechanisms underlying effects of traditional Chinese medicine Qingdai in chronic myelogenous leukemia, involving integration of network pharmacology and molecular docking technology," *Medical Science Monitor*, vol. 24, pp. 4305–4316, 2018.
- [15] F. Zhou, A. Wang, D. Li, Y. Wang, and L. Lin, "Pinocembrin from *Penthorum chinense Pursh* suppresses hepatic stellate cells activation through a unified SIRT3-TGF- β -Smad signaling pathway," *Toxicology and Applied Pharmacology*, vol. 341, pp. 38–50, 2018.
- [16] J. Yin, W. Ren, B. Wei et al., "Characterization of chemical composition and prebiotic effect of a dietary medicinal plant *Penthorum chinense Pursh*," *Food Chemistry*, vol. 319, p. 126568, 2020.
- [17] Z. Du, D. Huang, P. Shi et al., "Integrated Chemical Interpretation and Network Pharmacology Analysis to Reveal the

- Anti-Liver Fibrosis Effect of *Penthorum chinense*,” *Frontiers in Pharmacology*, vol. 1231, 2022.
- [18] Y. Jiang, M. Zhong, H. Zhan et al., “Integrated strategy of network pharmacology, molecular docking, HPLC-DAD and mice model for exploring active ingredients and pharmacological mechanisms of *Penthorum chinense* Pursh against alcoholic liver injury,” *Journal of Ethnopharmacology*, vol. 298, article 115589, 2022.
- [19] W. Guo, Y. Jiang, X. Chen et al., “Identification and quantitation of major phenolic compounds from *Penthorum chinense* Pursh. by HPLC with tandem mass spectrometry and HPLC with diode array detection,” *Journal of Separation Science*, vol. 38, no. 16, pp. 2789–2796, 2015.
- [20] Z.-L. Sun, Y.-Z. Zhang, F. Zhang et al., “Quality assessment of *Penthorum chinense* Pursh through multicomponent qualification and fingerprint, chemometric, and antihepatocarcinoma analyses,” *Food & Function*, vol. 9, no. 7, pp. 3807–3814, 2018.
- [21] D. Huang, X. Wang, L. Sun, W. Chen, and L. Sun, “Two new phenylpropanoids from *Penthorum chinense* Pursh,” *Phytochemistry Letters*, vol. 28, pp. 84–87, 2018.
- [22] D. Huang, Z. Dong, L. Sun, W. Chen, and L. Sun, “Two neolignans from *Penthorum chinense* and their antiproliferative activities,” *Natural Product Research*, vol. 34, no. 11, pp. 1515–1520, 2020.
- [23] F. Nabi, M. A. Arain, F. Hassan et al., “Nutraceutical role of selenium nanoparticles in poultry nutrition: a review,” *World’s Poultry Science Journal*, vol. 76, no. 3, pp. 459–471, 2020.
- [24] Y. Hu, S. Wang, A. Wang, L. Lin, M. Chen, and Y. Wang, “Antioxidant and hepatoprotective effect of *Penthorum chinense* Pursh extract against *t*-BHP-induced liver damage in L02 cells,” *Molecules*, vol. 20, no. 4, pp. 6443–6453, 2015.
- [25] F. Nabi, M. A. Arain, Z. A. Bhutto et al., “Effect of early feeding of L-arginine and L-threonine on hatchability and post-hatch performance of broiler chicken,” *Tropical Animal Health and Production*, vol. 54, no. 6, p. 380, 2022.
- [26] F. Nabi, W. Tao, R. Ye et al., “*Penthorum chinense* Pursh Extract Alleviates Aflatoxin B1-Induced Liver Injury and Oxidative Stress through Mitochondrial Pathways in Broilers,” *Frontiers in Veterinary Science*, vol. 9, article 822259, 2022.
- [27] L. Liu, X. Luo, M. Zou, L. Zhang, M. Yin, and X. Zhang, “Macroporous resin-assisted enrichment and isolation of antioxidant and cytotoxic phenolics from *Penthorum chinense*,” *Revista Brasileira de Farmacognosia*, vol. 31, no. 6, pp. 854–858, 2021.
- [28] M. Wang, X.-J. Zhang, R. Feng et al., “Hepatoprotective properties of *Penthorum chinense* Pursh against carbon tetrachloride-induced acute liver injury in mice,” *Chinese Medicine*, vol. 12, no. 1, p. 32, 2017.
- [29] L. Fu, J. Yuan, J. Huang et al., “Optimal extraction and antioxidant activities of flavonoids from *Penthorum chinense* Pursh,” *Journal of Food Measurement and Characterization*, vol. 13, no. 3, pp. 2253–2264, 2019.
- [30] L. M. Lin, L. J. Zhao, J. Deng et al., “Enzymatic Extraction, Purification, and Characterization of Polysaccharides from *Penthorum chinense* Pursh: Natural Antioxidant and Anti-Inflammatory,” *BioMed Research International*, vol. 2018, Article ID 3486864, 13 pages, 2018.
- [31] Z. Xu, B. Wang, L. Fu et al., “Optimization extraction, purification and antioxidant activities of polysaccharides from *Penthorum chinense* Pursh,” *International Journal of Food Engineering*, vol. 15, no. 3-4, 2019.
- [32] M. A. Gyamfi and Y. Aniya, “Antioxidant properties of Thoningianin A, isolated from the African medicinal herb, *Thonningia sanguinea*,” *Biochemical Pharmacology*, vol. 63, no. 9, pp. 1725–1737, 2002.
- [33] S. I. Abdelwahab, S. Mohan, M. A. Abdulla et al., “The methanolic extract of *Boesenbergia rotunda* (L.) Mansf. and its major compound pinostrobin induces anti-ulcerogenic property *in vivo*: possible involvement of indirect antioxidant action,” *Journal of Ethnopharmacology*, vol. 137, no. 2, pp. 963–970, 2011.
- [34] Y. Lu, Y. Du, X. Qin et al., “Comprehensive evaluation of effective polyphenols in apple leaves and their combinatory antioxidant and neuroprotective activities,” *Industrial Crops and Products*, vol. 129, pp. 242–252, 2019.
- [35] J. Y. Seo, R. P. Pandey, J. Lee, J. K. Sohng, W. Namkung, and Y. I. Park, “Quercetin 3-*O*-xyloside ameliorates acute pancreatitis *in vitro* via the reduction of ER stress and enhancement of apoptosis,” *Phytomedicine*, vol. 55, pp. 40–49, 2019.
- [36] Y. Ma, Y. Liu, A. Sun et al., “Intestinal absorption and neuroprotective effects of kaempferol-3-*O* -rutinoside,” *RSC Advances*, vol. 7, no. 50, pp. 31408–31416, 2017.
- [37] S. K. Wong, K. Y. Chin, and S. Ima-Nirwana, “The osteoprotective effects of kaempferol: the evidence from *in vivo* and *in vitro* studies,” *Drug Design, Development and Therapy*, vol. 13, pp. 3497–3514, 2019.
- [38] L. M. Coussens and Z. Werb, “Inflammation and cancer,” *Nature*, vol. 420, no. 6917, pp. 860–867, 2002.
- [39] A. Mantovani, “Molecular pathways linking inflammation and cancer,” *Current Molecular Medicine*, vol. 10, no. 4, pp. 369–373, 2010.
- [40] M. Khatami, “Inflammation aging, and cancer: tumoricidal versus tumorigenesis of immunity: a common denominator mapping chronic diseases,” *Cell Biochemistry and Biophysics*, vol. 55, no. 2, pp. 55–79, 2009.
- [41] C. L. Chen and D. D. Zhang, “Anti-inflammatory effects of 81 Chinese herb extracts and their correlation with the characteristics of traditional Chinese medicine,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2014, Article ID 985176, 8 pages, 2014.
- [42] Y. Chen, T. Li, P. Tan et al., “Kaempferol from *Penthorum chinense* Pursh attenuates hepatic ischemia/reperfusion injury by suppressing oxidative stress and inflammation through activation of the Nrf2/HO-1 signaling pathway,” *Frontiers in Pharmacology*, vol. 13, article 857015, 2022.
- [43] H.-S. Chae, P. Pel, J. Cho et al., “Identification of neolignans with PCSK9 downregulatory and LDLR upregulatory activities from *Penthorum chinense* and the potential in cholesterol uptake by transcriptional regulation of LDLR via SREBP2,” *Journal of Ethnopharmacology*, vol. 278, article 114265, 2021.
- [44] F. Ke, P. Xie, Y. Yang et al., “Effects of nisin, cecropin, and *Penthorum chinense* Pursh on the intestinal microbiome of common carp (*Cyprinus carpio*),” *Frontiers in Nutrition*, vol. 8, article 729437, 2021.
- [45] W. Tao, X. Yue, R. Ye et al., “Hepatoprotective Effect of the *Penthorum chinense* Pursh Extract against the CCl₄-Induced Acute Liver Injury via NF- κ B and p38-MAPK PATHWAYS in Dogs,” *Animals*, vol. 12, no. 5, p. 569, 2022.
- [46] X. Zhao, M. Zhou, Y. Deng et al., “*Penthorum chinense* Pursh protects liver from alcohol-induced steatosis in zebrafish by mechanisms including inhibition of oxidative stress and increase in autophagy,” 2020.

- [47] X. Zhao, L. Li, M. Zhou et al., "An overview of the mechanism of *Penthorum chinense* Pursh on alcoholic fatty liver," *Evidence-based Complementary and Alternative Medicine*, vol. 2020, Article ID 4875764, 13 pages, 2020.
- [48] T. Ma, H. Zhang, T. Li et al., "Protective effect of pinocembrin from *Penthorum chinense* Pursh on hepatic ischemia reperfusion injury via regulating HMGB1/TLR4 signal pathway," *Phytotherapy Research*, vol. 37, no. 1, pp. 181–194, 2022.
- [49] B. Ding, Q. Ding, S. Zhang et al., "Characterization of the anti-*Staphylococcus aureus* fraction from *Penthorum chinense* Pursh stems," *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, p. 219, 2019.
- [50] W. Tao, Z. Li, F. Nabi, Y. Hu, Z. Hu, and J. Liu, "*Penthorum chinense* Pursh Compound Ameliorates AFB1-Induced Oxidative Stress and Apoptosis via Modulation of Mitochondrial Pathways in Broiler Chicken Kidneys," *Frontiers in Veterinary Science*, vol. 8, article 750937, 2021.
- [51] A. Rasul, F. M. Millimouno, W. Ali Eltayb, M. Ali, J. Li, and X. Li, "Pinocembrin: a novel natural compound with versatile pharmacological and biological activities," *BioMed Research International*, vol. 2013, Article ID 379850, 9 pages, 2013.
- [52] W. Bao, K. Li, S. Rong et al., "Curcumin alleviates ethanol-induced hepatocytes oxidative damage involving heme oxygenase-1 induction," *Journal of Ethnopharmacology*, vol. 128, no. 2, pp. 549–553, 2010.
- [53] M. Pulido-Moran, J. Moreno-Fernandez, C. Ramirez-Tortosa, and M. Ramirez-Tortosa, "Curcumin and health," *Molecules*, vol. 21, no. 3, p. 264, 2016.
- [54] A. Ullah, S. Munir, S. L. Badshah et al., "Important flavonoids and their role as a therapeutic agent," *Molecules*, vol. 25, no. 22, p. 5243, 2020.
- [55] R. M. Lamuela-Raventós, A. I. Romero-Pérez, C. Andrés-Lacueva, and A. Tornero, "Review: health effects of cocoa flavonoids," *Food Science and Technology International*, vol. 11, pp. 159–176, 2016.
- [56] S. H. Wang, Y. L. Hu, and T. X. Liu, "Plant distribution and pharmacological activity of flavonoids," *Traditional Medicine Research*, vol. 4, no. 5, pp. 269–287, 2020.
- [57] Q. H. Zeng, X. W. Zhang, X. L. Xu et al., "Antioxidant and anticomplement functions of flavonoids extracted from *Penthorum chinense* Pursh," *Food & Function*, vol. 4, no. 12, pp. 1811–1818, 2013.
- [58] C. Dai, E. Tian, Z. Hao et al., "Aflatoxin B1 Toxicity and Protective Effects of Curcumin: Molecular Mechanisms and Clinical Implications," *Antioxidants*, vol. 11, no. 10, p. 2031, 2022.
- [59] X. Li, W. Zhao, M. Xiao et al., "*Penthorum chinense* Pursh. extract attenuates non-alcoholic fatty liver disease by regulating gut microbiota and bile acid metabolism in mice," *Journal of Ethnopharmacology*, vol. 294, article 115333, 2022.
- [60] Z. Wang, K. Jiang, Q. Ding, S. Liu, X. Dou, and B. Ding, "A comparison of the biological activities of ethyl acetate fractions from the stems and leaves of *Penthorum chinense* Pursh," *Frontiers of Agricultural Science and Engineering*, vol. 7, no. 4, pp. 505–512, 2020.
- [61] L. Kang, S. Yang, Y. Peng, J. Dai, and X. Ying, "Research on Extraction Process of Gallic Acid from *Penthorum chinense* Pursh by Aqueous Ethanol," *Green and Sustainable Chemistry*, vol. 5, no. 2, pp. 63–69, 2015.
- [62] Y.-W. Cao, Y. Jiang, D.-Y. Zhang et al., "Protective effects of *Penthorum chinense* Pursh against chronic ethanol-induced liver injury in mice," *Journal of Ethnopharmacology*, vol. 161, pp. 92–98, 2015.
- [63] D. Huang, Y. Jiang, W. Chen, F. Yao, G. Huang, and L. Sun, "Evaluation of hypoglycemic effects of polyphenols and extracts from *Penthorum chinense*," *Journal of Ethnopharmacology*, vol. 163, pp. 256–263, 2015.
- [64] F. Nabi, M. Shahzad, J. Liu et al., "Hsp90 inhibitor celastrol reinstates growth plate angiogenesis in thiram-induced tibial dyschondroplasia," *Avian Pathology*, vol. 45, no. 2, pp. 187–193, 2016.
- [65] R. V. Nugraha, H. Ridwansyah, M. Ghozali, A. F. Khairani, and N. Atik, "Traditional herbal medicine candidates as complementary treatments for COVID-19: a review of their mechanisms, pros and cons," *Evidence-Based Complementary and Alternative Medicine*, vol. 2020, Article ID 2560645, 12 pages, 2020.
- [66] C. C. Wen, H. M. Chen, and N. S. Yang, "Developing phyto-compounds from medicinal plants as immunomodulators," *Advances in Botanical Research*, vol. 62, pp. 197–272, 2012.
- [67] Y. Chen, P. Chen, H. Liu, Y. Zhang, and X. Zhang, "*Penthorum chinense* Pursh polysaccharide induces a mitochondrial-dependent apoptosis of H22 cells and activation of immunoregulation in H22 tumor-bearing mice," *International Journal of Biological Macromolecules*, vol. 224, pp. 510–522, 2022.
- [68] S. Shruthi, K. K. Vijayalaxmi, and K. B. Shenoy, "Immunomodulatory Effects of Gallic Acid against Cyclophosphamide- and Cisplatin-induced Immunosuppression in Swiss Albino Mice," *Indian Journal of Pharmaceutical Sciences*, vol. 80, no. 1, pp. 150–160, 2018.
- [69] B. C. L. Chan, L. F. Li, S. Q. Hu et al., "Gallic acid is the major active component of cortex moutan in inhibiting immune maturation of human monocyte-derived dendritic cells," *Molecules*, vol. 20, no. 9, pp. 16388–16403, 2015.
- [70] Q. Ding, Z. Jin, J. Dong et al., "Bioactivity evaluation of pinocembrin derivatives From *Penthorum chinense* Pursh stems," *Natural Product Communications*, vol. 14, no. 9, 2019.
- [71] J. Hu, L. Zheng, W. Tang, Y. Xu, Q. Fan, and H. Xie, "Hypoglycemic Effects of Extracts of *Penthorum chinense* Pursh in High Fat Diet and Streptozotocin-induced Type 2 Diabetic Rats," *Modern Food Science & Technology*, vol. 36, no. 2, pp. 25–31, 2020.
- [72] X.-G. Zhou, W.-Q. Qiu, L. Yu et al., "Targeting Microglial Autophagic Degradation of the NLRP3 Inflammasome for Identification of Thonningianin A in Alzheimer's Disease," *Inflammation and Regeneration*, vol. 42, no. 1, p. 25, 2022.
- [73] Y. Sun, L. He, W. Wang et al., "Activation of Atg7-dependent autophagy by a novel inhibitor of the Keap1-Nrf2 protein-protein interaction from *Penthorum chinense* Pursh. attenuates 6-hydroxydopamine-induced ferroptosis in zebrafish and dopaminergic neurons," *Food & Function*, vol. 13, no. 14, pp. 7885–7900, 2022.
- [74] Q. Lu, M. H. Jiang, J. G. Jiang, R. F. Zhang, and M. W. Zhang, "Isolation and identification of compounds from *Penthorum chinense* Pursh with antioxidant and antihepatocarcinoma properties," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 44, pp. 11097–11103, 2012.
- [75] Y. Yuan and X. Ou, "Research on acute toxicity of *Penthorum chinense* Pursh. Total flavonoids and therapeutic effect on AFL rats," *Medicinal Plant*, vol. 9, no. 1, pp. 56–59, 2018.
- [76] W. W. Guo, X. Wang, X. Q. Chen et al., "Flavonones from *Penthorum chinense* ameliorate hepatic steatosis by activating the SIRT1/AMPK pathway in HepG2 cells," *International Journal of Molecular Sciences*, vol. 19, no. 9, p. 2555, 2018.