



Chemical Composition of Essential Oils from Aerial Parts of *Sisymbrium Irio* from Jordan

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Abstract: The volatile compounds obtained from *Sisymbrium irio* growing in Jordan has been analyzed by GC-MS. The major constituents are dioctyladipate (25.44%), *N*-(*n*-proyl) acetamide (14.77%), isopropyl isothiocyanate (11.55%), isobutyl isothiocyanate (6.75%), 3,7,11,15-tetramethyl-2-hexadecen-1-ol (6.52%), *cis*-8,11,14-eicosatrienoic acid (6.30%), heptacosane (3.89%), palmitic acid (3.45%), *n*-butyl isothiocyanate (2.85 %) and dimethoxyacetophenone (2.54 %).

Keywords: *Sisymbrium irio*, Cruciferae, Isopropyl isothiocyanate, Dioctyladipate.

Introduction

The cruciferae family is a large plant family that includes important food crops, herbs, ornamentals and weeds¹. Forty six genera of this family are distributed in Jordan, including the genus *Sisymbrium*. There are seven species belonging to this genus in Jordan². One of the most important species of this genus is *Sisymbrium irio*, which is grown in many parts of the world. The plant is a rich source of flavonoids^{3,4} and glucosinolates^{5, 6}. It has a sharp flavor and can be used in salads. The plant is used in folk medicine as a febrifuge, a stimulating poultice, treating asthma and for infections of the throat and chest⁷. In this paper we report on the isolation and analysis of the volatile compounds from the aerial parts of *Sisymbrium irio* collected from the north of Jordan.

Experimental

Aerial parts of *Sisymbrium irio* ("al-hwarineh or al-shalwa" in Arabic) were collected at the onset of flowering, from Irbid area (north of Jordan). Identification of the plant species was done by professor D. Al-Esawi (Department of Biological Sciences, Faculty of Science, University of Jordan).

Isolation of the volatile components

The fresh aerial parts of *Sisymbrium irio* (30.0 g) were hydrodistilled using a Clevenger-type apparatus to recover the essential oils for 4 h. The oils distillate was separate from trace of water by extraction three times with 3 mL of Ether and dried overnight over 3.0 g of anhydrous sodium sulfate.

GC-MS analysis

Gas chromatography - mass spectrometry (GC-MS) analyses were performed using Agilent 6890 series II-5973 mass spectrometer interfaced with a HP chemstation. The chromatographic conditions were as follow: column oven programe, 40 °C (10 min, isothermal) to 240 °C (10 min, isothermal) at 2 °C/min; the injector and detector temperatures were 270 and 280 °C, respectively. Helium was the carrier gas (flow rate 0.70 mL/min). A HP-5 MS capillary column (30×0.25 mm *i.d.*, 0.25 µm film thicknesses) was utilized. The actual temperatures in MS source reached approximately 180 °C. The ionization voltage was 70 eV. The volatile compounds were identified by comparing their MS patterns and retention times with those of known compounds published in the literature⁸.

Results and Discussion

Sisymbrium irio is a plant grown in north and middle Jordan. The oil was analyzed by GC-MS; to give the gas chromatogram shown in Figure 1. The chromatogram showed that the volatile oil contains the compounds listed in Table 1.

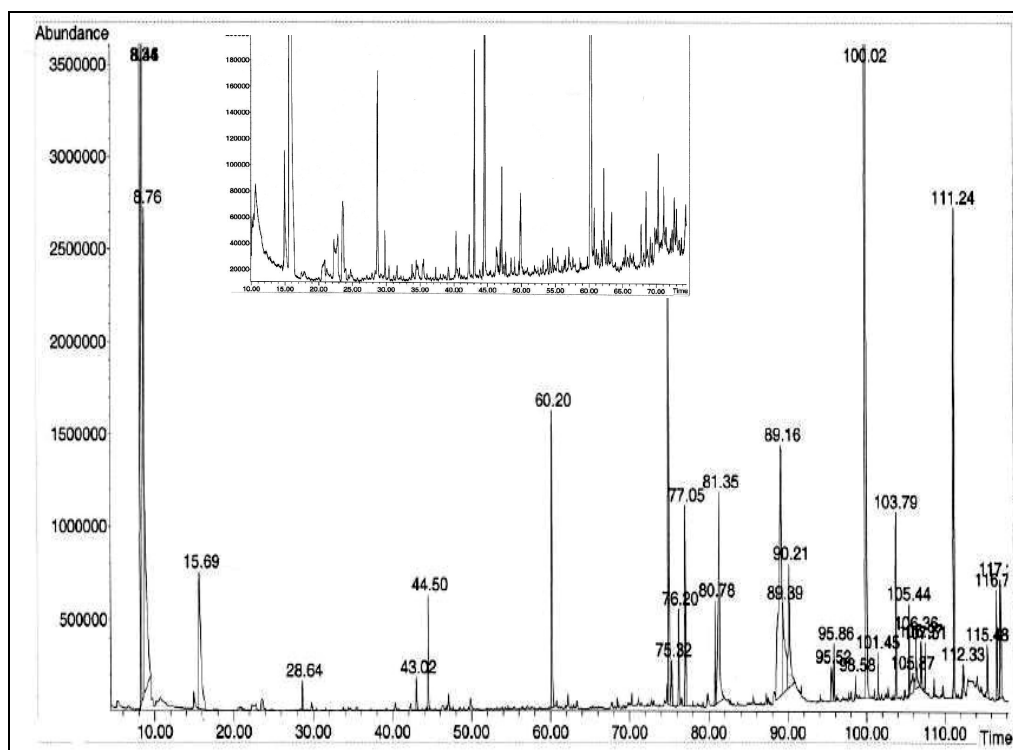


Figure 1. Gas chromatogram of volatile components of *Sisymbrium irio*.

Table 1. Composition and percentage of the essential oil from *Sisymbrium irio*.

| No. | Compound | M ⁺ | Retention time, min | Peak area % |
|-----|--|----------------|---------------------|-------------|
| 1 | Isopropyl isothiocyanate | 101 | 8.34 | 11.55 |
| 2 | <i>N</i> -(<i>n</i> -propyl)acetamide | 101 | 8.50 | 14.77 |
| 3 | Isobutyl isothiocyanate | 115 | 8.76 | 6.75 |
| 4 | 2 <i>E</i> -Hexenal | 98 | 9.65 | Trace |
| 5 | 3-Hexen-1-ol | 100 | 10.11 | Trace |
| 6 | Dimethyl sulphone | 94 | 14.92 | 0.10 |
| 7 | <i>n</i> - Butyl isothiocyanate | 115 | 15.69 | 2.85 |
| 8 | Tetra acetyl- <i>d</i> -xylopic nitrile | 343 | 20.87 | Trace |
| 9 | 3 <i>E</i> -Hexenoic acid | 114 | 22.80 | 0.05 |
| 10 | β -Terpinyl acetate | 136 | 23.56 | 0.12 |
| 11 | Nonanal | 142 | 29.79 | 0.07 |
| 12 | 4-(2,5-Dihydro-3-methoxy phenyl)butylamine | 181 | 30.37 | Trace |
| 13 | trans- α -Bisabolene epoxide | 220 | 30.39 | 0.05 |
| 14 | <i>p</i> -Anisaldehyde | 136 | 40.35 | 0.08 |
| 15 | Indole | 117 | 43.06 | 0.22 |
| 16 | <i>p</i> -Vinylguaiacol | 150 | 44.47 | 0.78 |
| 17 | Nicotine | 162 | 46.40 | 0.06 |
| 18 | 1,5,8-Trimethyl-1,2-dihydronaphthalene | 172 | 46.84 | Trace |
| 19 | 1,1,6-Trimethyl-1,2-dihydronaphthalene | 172 | 46.85 | Trace |
| 20 | 1,1,6-Trimethyl-1,2,3,4-tetrahydronaphthalene | 174 | 47.13 | 0.11 |
| 21 | <i>o</i> -Benzyl- <i>L</i> -serine | 195 | 47.69 | 0.06 |
| 22 | 3-Methyl indole | 131 | 48.98 | 0.05 |
| 23 | Isovanillin | 152 | 49.88 | 0.13 |
| 24 | 3',5'-Dimethoxyacetophenone | 180 | 60.20 | 2.54 |
| 25 | 1,2,3b,6,7,8,-Hexahdro-6,6-dimethyl cyclopenta[1,3]cyclopropa[1,2]cyclohepten-3(3 <i>H</i>)-one | 190 | 60.75 | Trace |
| 26 | 8-Isopropyl-1,2,3,7-tetramethylbicyclo[5,1,0] octa-5-en-2-one | 218 | 60.80 | 0.11 |
| 27 | Deoxysericealacone | 276 | 61.76 | Trace |
| 28 | 1,2-Dipalmitate glycerol | 568 | 61.84 | 0.07 |
| 29 | 5-Isopropenyl-2-methylcyclopent-1-enecarboxaldehyde | 150 | 62.16 | 0.16 |
| 30 | Tetrahydrospirilloxanthin | 600 | 62.60 | Trace |
| 31 | 4-(2,4,4-trimethyl-cyclohexa-1,5-dienyl)but-3-en-2-one | 190 | 63.31 | 0.05 |
| 32 | Methoxyeugenol | 194 | 67.71 | Trace |
| 33 | 13-Heptadecyn-1-ol | 252 | 68.49 | 0.06 |
| 34 | 2-(2-Methylpropylidene)-1 <i>H</i> -indene-1,3(2 <i>H</i>)-dione | 200 | 70.23 | 0.08 |
| 35 | 2,3,6-Trimethyl-1,4-naphthalenedione | 200 | 70.25 | Trace |
| 36 | 3,7,11,15-Tetramethyl-2-hexadecen-1-ol | 296 | 74.96 | 6.52 |
| 37 | 1-Eicosanol | 298 | 75.35 | 0.39 |

Contd...

| | | | | |
|----|--|-----|--------|-------|
| 38 | Oleic acid | 282 | 76.56 | 0.67 |
| 39 | Palmitic acid | 256 | 81.34 | 3.45 |
| 40 | <i>cis</i> -8,11,14-Eicosatrienoic acid | 306 | 89.16 | 6.30 |
| 41 | 12-Methyl- <i>E,E</i> - 2,13-octadecadien-1-ol | 280 | 89.39 | 2.04 |
| 42 | Stearic acid | 284 | 90.21 | 2.07 |
| 43 | Docasane | 310 | 95.86 | 0.36 |
| 44 | Eicosanoic acid | 312 | 98.57 | 0.15 |
| 45 | Diocetyl adipate | 370 | 100.00 | 25.44 |
| 46 | Octacosane | 394 | 103.78 | 1.25 |
| 47 | 1-Hexacosene | 364 | 105.87 | 0.40 |
| 48 | Docasanoic | 340 | 106.38 | 0.60 |
| 49 | Nonacosane | 408 | 107.51 | 0.33 |
| 50 | Heptacosane | 380 | 111.26 | 3.89 |
| 51 | Squalene | 410 | 116.71 | 0.99 |
| 52 | Octadecanal | 268 | 117.23 | 1.10 |
| 53 | Dotriacontane | 450 | 118.70 | 0.06 |

*Trace : < 0.01

The oil contained 53 components, representing 97.48% of the total oil. The main constituents of the oil were dioctyladipate (25.44%), *N*-(*n*-proyl) acetamide (14.77%), isopropyl isothiocyanat (11.55%), isobutyl isothiocyanate (6.75%), 3,7,11,15-tetramethyl-2-hexadecen-1-ol (6.52%), *cis*-8,11,14-eicosatrienoic acid (6.30%), heptacosane (3.89%), palmitic acid (3.45%), *n*-butyl isothiocyanate (2.85%) and dimethoxyacetophenone (2.54%). A classification of the constituents of the *sisymbrium irio* L. oil is given in Table 2. The oil included seven acids and two esters (38.80%), 11 nitrogen and sulfur containing compounds (36.41%), fifteen terpenoids (8.19%), six aliphatic hydrocarbons (6.29%), five aromatic compounds (3.53%), four fatty alcohols (2.49%) and three of other compounds (1.17%).

Table 2. Classification of the constituents of the *Sisymbrium irio*.

| No. | Component | Peak area, % |
|-----|--|--------------|
| 1 | Acids | 13.29 |
| 2 | Esters | 25.51 |
| 3 | Aliphatic hydrocarbons | 6.29 |
| 4 | Aromatic compounds | 3.53 |
| 5 | Terpenoids | 8.19 |
| 6 | Fatty alcohols | 2.49 |
| 7 | Nitrogen containing compounds | 15.16 |
| 8 | Sulfur containing compounds | 0.10 |
| 9 | (Nitrogen + sulfur) containing compounds | 21.15 |
| 10 | Others | 1.17 |

Conclusion

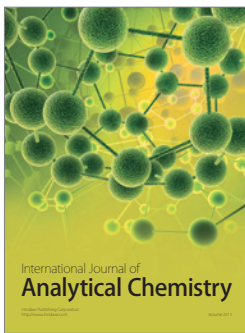
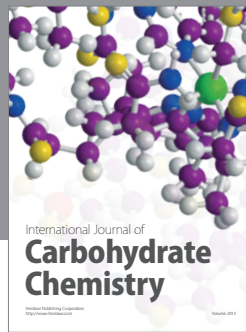
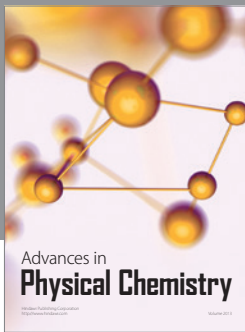
Many crucifers such as *Sisymbrium irio* contain glucosinolates which are precursors of many volatile compounds, in particular nitriles and isothiocyanates. Isothiocyanates, which are strongly odorous and pungent compounds, are formed by the action of the enzyme thioglucoside glucohydrolase (myrosinase) on the glucosinolate when the plant tissue is disrupted^{9,10}. In our case, intact plant material was subjected to an isolation procedure so that myrosinase action was restrained; hydrodistillation gave predominantly Isothiocyanates degradation products as observed in the chemical composition of the oil.

Acknowledgement

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