

Research Article

Liver Assessment in Patients with Ataxia-Telangiectasia: Transient Elastography Detects Early Stages of Steatosis and Fibrosis

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Background. Ataxia-telangiectasia (A-T) is a rare autosomal-recessive multisystem disorder characterized by pronounced cerebellar ataxia, telangiectasia, cancer predisposition, and altered body composition. Liver diseases with steatosis, fibrosis, and hepatocellular carcinoma are frequent findings in older patients but sensitive noninvasive diagnostic tools are lacking. *Objectives*. To determine the sensitivity of transient elastography (TE) as a screening tool for early hepatic tissue changes and serum biomarkers for liver disease. *Methods*. Thirty-one A-T patients aged 2 to 25 years were examined prospectively from 2016–2018 by TE. In addition, we evaluated the diagnostic performance of liver biomarkers for steatosis and necroinflammatory activity (SteatoTest and ActiTest, Biopredictive, Paris) compared to TE. For calculation and comparison, patients were divided into two groups (<12, >12 years of age). *Results*. TE revealed steatosis in 2/21 (10%) younger patients compared to 9/10 (90%) older patients. Fibrosis was present in 3/10 (30%) older patients as assessed by TE. We found a significant correlation of steatosis with SteatoTest, alpha-fetoprotein (AFP), HbA1c, and triglycerides. Liver stiffness correlated significantly with SteatoTest, ActiTest, HbA1c, and triglycerides. *Conclusion*. Liver disease is a common finding in older A-T patients. TE is an objective measure to detect early stages of steatosis and fibrosis. SteatoTest and ActiTest are a good diagnostic assessment for steatosis and necroinflammatory activity in patients with A-T and confirmed the TE results.

1. Introduction

Ataxia-telangiectasia (A-T) is a rare, autosomal-recessive multisystem disorder characterized by progressive cerebellar ataxia, telangiectasia, immunodeficiency, and cancer predisposition [1–5]. Apart from the name-giving brain involvement, the disease also affects the lungs, endocrine system, and liver [6–13]. A-T-associated liver disease is an upcoming health issue that emerges in the second decade of life [7, 14]. Autopsy reports in A-T patients showed liver-specific pathological findings like nonalcoholic steatohepatitis (NASH), liver cirrhosis, and hepatocellular carcinoma (HCC) [7, 15–18].

Hepatopathy in A-T is usually mild and does not lead to a limitation of the synthesis or detoxification function of the liver [7]; however, up to 92.9% of older A-T patients are affected [14]. Liver disease certainly belongs to the complex phenotype of premature aging [19], which also includes insulin resistance (IR), diabetes mellitus type 2 [6, 20], and dyslipidemia and thus leads to an incomplete metabolic syndrome. All these factors naturally favor fatty remodeling of the liver with a consecutive increase in liver enzymes. In addition, a few severe cases of liver failure and HCC have been published as case reports on A-T patients [15, 17].

One of the central aims of modern hepatology is the search for noninvasive diagnostic procedures for

presymptomatic liver diseases in at-risk patient groups. In order to prevent severe courses and identify critical patients at an early stage, sensitive, fast, and minimally invasive screening tools are needed to monitor the course of the disease [21, 22]. Currently, a liver biopsy is the gold standard for assessing the severity of nonalcoholic fatty liver disease (NAFLD), NASH, and the stage of liver fibrosis. Nevertheless, limitations and complications include invasiveness, severe bleeding, sampling error, and pneumothorax [23].

Transient elastography (TE) is a precise, noninvasive method to determine the extent of fibrosis and the degree of fatty degeneration of the liver [24]. In contrast to biopsy, it is painless and noninvasive. In addition, it records a multiple of the parenchyma. The classification into fibrosis and steatosis stages is objective in comparison to normal sonography, as it is carried out using fixed cutoff values [22, 24, 25]. Measurements are reproducible and independent of the user [25, 26].

In this cross-sectional study, we prospectively evaluated liver assessment by TE and liver scores (FibroMax) for A-Tassociated liver disease to predict the extent of liver disease and identify at-risk patients for severe disease courses.

2. Patients and Methods

The current study is a prospective, cross-sectional, clinical, single-center trial.

2.1. Patients. From November 2016 to May 2018, 31 patients with a clinically and/or genetically confirmed diagnosis of A-T aged between two and 25 years were included in the study (Table 1). Malignancy and clinical and laboratory-associated infections were defined as the exclusion criteria.

The Ethics Committee of University Hospital Frankfurt approved the trial (Reference No. 504/15). The study was registered at clinicaltrials.gov (NCT03357978). One study visit was conducted. Written consent was obtained from all patients and/or caregivers. The study was conducted according to the ethical principles of the Declaration of Helsinki and regulatory requirements and the code of Good Clinical Practice.

We compared patients <12 years of age (group 1) with those \ge 12 years (group 2).

2.2. Transient Elastography. The examination was performed with FibroScan[®] (Echosens, Paris, France). The examination probe is formed by a vibration generator and an ultrasonic probe (3.5 MHz) aligned on the same axis. The vibration generator oscillates at a frequency of 50 Hz, which leads to shear waves in the liver tissue. The speed of propagation of this shear wave correlates directly with liver stiffness and therefore with the extent of fibrosis. The result of this liver stiffness measurement (LSM) is given in kilopascals (kPa) [25].

The interpretation of the measurement results was based on the limit values of a study on mixed hepatopathy by Fraquelli et al. [26]. A distinction was made between three

TABLE 1: Patients' characteristics.

Parameter	Group 1 $(n = 21)$	Group 2 $(n = 10)$	p value
Sex	9º/12ð	5\$/5ð	
Age (years)	6.5 ± 2.8	19.6 ± 3.5	< 0.0001
CAP (dB/m)	174.7 ± 45.1	302.2 ± 57.7	< 0.001
LSM (kPa)	4.6 ± 0.9	9 ± 6.9	< 0.001
AFP (ng/mL)	313.4 ± 267.2	540.8 ± 275.8	< 0.05
CRP (mg/dL)	0.2 ± 0.4	0.5 ± 0.8	n.s.
AST (U/L)	37.8 ± 7.9	49.8 ± 15.2	< 0.05
ALT (U/L)	25.1 ± 9.6	71.6 ± 25.8	< 0.001
GGT (U/L)	13.2 ± 4.5	123.7 ± 99.6	< 0.0001

Mean values \pm SD are shown; n.s. = not significant.

fibrosis stages: $F \ge 2$ = pronounced fibrosis, $F \ge 3$ = severe fibrosis, and F4 = cirrhosis.

At the same time, the controlled attenuation parameter (CAP) was measured using the same signals. The attenuation of the ultrasound signal (3.5 MHz) in the liver is measured in dB/m. The attenuation correlates with the degree of liver steatosis [27]. The different stages of steatosis were defined as follows: $S \ge 1 =$ steatosis in 11–33% of hepatocytes, $S \ge 2 =$ steatosis in 34–66% of hepatocytes, and S = 3 steatosis in 67–100% of hepatocytes. The cutoff values proposed by Karlas et al. in 2017 were used [28].

The examination was performed by an experienced physician in the supine position and maximum abduction of the right arm through a right intercostal space. In order to perform a standardized measurement, patients were asked to fast for at least 4 hours before the examination. A success rate of at least 60% or the interquartile range below 30% of the median measurement result was considered necessary.

The accuracy of the measurement may be reduced in obesity or ascites.

2.3. Liver Biomarkers and Liver Scores. FibroMax[®] (Bio-Predictive, Paris, France) is a noninvasive blood test for NAFLD screening that has been validated against liver biopsies [29] and is recommended by European guidelines [30]. FibroMax is composed of three different tests (SteatoTest, ActiTest, and FibroTest) for the assessment of steatosis, necroinflammatory activity, and fibrosis, respectively. The serum parameters such as α 2-macroglobulin, haptoglobin, apolipoprotein A1, bilirubin, gamma-glutamyl transferase (GGT), alanine aminotransferase (ALT), aspartate aminotransferase (AST), fasting glucose, cholesterol, and triglycerides, as well as gender, age, weight, and height, were recorded for the calculation of the liver score FibroMax.

The FibroMax data were calculated by BioPredictive (Paris, France) using a patented algorithm.

The FibroTest results were interpreted using the MET-AVIR score from F0 to F4: F0 = n0 fibrosis, F1 = portal fibrosis without septa, F2 = few septa, F3 = many septa without cirrhosis, and F4 = cirrhosis.

The steatosis test results were interpreted using the steatosis score from S0 to S4: S0 = no steatosis, S1 = mild steatosis, S2 = moderate steatosis, S3 = pronounced steatosis, and S4 = severe steatosis.

The ActiTest results were interpreted using the MET-AVIR score from A0 to A3: A0 = no necroinflammatory activity, A1 = mild activity, A2 = moderate activity, and A3 = severe activity. Data were interpreted according to Poynard et al. [31, 32].

In addition to liver biomarkers, alpha-fetoprotein (AFP), hemoglobin A1c (HbA1c), complete lipid profile (including cholesterol, high-density lipoprotein (HDL) and low-density lipoprotein (LDL) cholesterol, and triglycerides) and Creactive protein (CRP) as inflammatory markers were determined.

2.4. Statistical Analysis. For statistical analysis, GraphPad Prism 5.01 (GraphPad Software, Inc.) was used. Values are presented as arithmetic means with standard deviation (SD). For comparisons between the two study groups, the two-tailed Mann–Whitney *U* test was applied. Correlations were analyzed by Spearman's correlation coefficient. *p* values ≤ 0.05 were considered significant.

3. Results

In a study period of 19 months (November 2016 to May 2018), 31 patients with A-T were examined. Patients' characteristics are shown in Table 1. The age distribution ranged from 2 to 25 years (mean age: 10.7 years). The patients were divided into the two groups for evaluation. Twenty-one patients were <12 years (group 1), and ten patients were \geq 12 years (group 2). No patient had a history of infectious hepatitis or was taking hepatotoxic drugs regularly.

3.1. Transient Elastography. On average, 16.5 measurements were performed with a mean success rate of 74%. TE revealed steatosis in 2/21 (10%) of younger patients. In both cases, grade 2 steatosis was present. Fibrosis was not evident in any of younger patients. In comparison, steatosis was detectable in 9/10 (90%) (2 (20%) patients with grade 2 steatosis and 7 (70%) patients with grade 3 steatosis), and fibrosis was observed in 3/10 (30%, median age: 21 years) of older patients. Of these, one patient (10%, aged 20 years) had pronounced fibrosis and the other two patients (20%, aged between 21 and 25 years) had liver cirrhosis (fibrosis stage 4). These results are summarized in Supplementary Table 1. The corresponding LSM (group 1: 4.5 ± 0.93 kPa vs. group 2: 8.9 ± 6.9 kPa; p < 0.001) and CAP (group 1:174.7 ± 45.08 bD/ m vs. group 2: $302.2 \text{ dB/m} \pm 57.68 \text{ dB/m}; p < 0.001$) of the older group were significantly increased compared to those of the younger group (Figures 1(a) and 1(b)). Furthermore, a positive correlation of the values with age was shown (LSM: r = 0.59, p < 0.001; CAP: r = 0.82, p < 0.0001) (Figures 2(a) and 2(b)). The average age for steatosis in group 2 was 20.3 ± 2.7 years.

In addition, CAP and LSM correlated with ALT, AST, GGT, AFP, HbA1c, and triglycerides. There was a significant correlation of steatosis (CAP values) with ALT (r=0.77, p < 0.0001), AST (r=0.39, p<0.05), GGT (r=0.83, p < 0.0001), AFP (r=0.42, p<0.05), HbA1c (r=0.59, p < 0.01), and triglycerides (r=0.74, p<0.0001). LSM

3.2. Serum Biomarkers and Liver Scores. A complete data set for biomarkers was available for 30 patients. The results of FibroMax are shown in Table 2. Significantly lower values for group 1 concerning SteatoTest and ActiTest (SteatoTest: p< 0.0001; ActiTest: p < 0.001) were calculated, showing a normal function in group 1 and mild to moderate dysfunction in group 2. FibroTest did not show significant differences between the two groups, with normal to slightly elevated levels in all patients.

The results of SteatoTest indicated steatosis in 8/10 (80%) patients of group 2, whereas no patient in group 1 was affected. Three of ten (30%) older patients had mild steatosis, 2/10 (20%) had mild to moderate steatosis, 2/10 (20%) had moderate steatosis, and one patient (10%) had pronounced steatosis. SteatoTest had a significant correlation with age, LDL-HDL ratios, CRP, and necroinflammatory activity (ActiTest) (age: r = 0.74, p < 0.0001; LDL-HDL ratio: r = 0.79, p < 0.0001; CRP: r = 0.51, p < 0.01; and ActiTest: r = 0.89, p < 0.0001), as shown in Table 3.

The ActiTest for the assessment of necroinflammatory activity showed a minimal activity level of stage A0-1 in 3/20 (15%) patients of group 1 and a pathological result in 9/10 (90%) of older patients. One older patient had stage A0-1, 6/ 10 (60%) patients had stage A1-2, and 2/10 (20%) patients had stage A2, i.e., moderate necroinflammatory activity. ActiTest correlated significantly with triglycerides, CRP, CAP, and LSM, as shown in Figures 3(a)-3(d) and Table 3 (triglycerides: r=0.61, p < 0.001; CRP: r=0.41, p < 0.05; CAP: r=0.77, p < 0.0001; and LSM: r=0.53, p < 0.01). In addition, there was a significant correlation with age and the LDL-HDL ratio (age: r=0.8, p < 0.0001; LDL-HDL ratio: r=0.8, p < 0.0001).

FibroTest did not show a significant difference between the two patient groups. Six of twenty (30%) younger patients had fibrosis according to FibroTest. 4/20 (20%) patients had stage 0-1, one patient had stage 1-2, and one patient had stage 2. In the older group, 3/10 (30%) patients had stage 0-1 and 2/10 (20%) had stage 1-2 fibrosis. This means that a total of 5/10 (50%) patients were affected. The correlations of SteatoTest and ActiTest are shown in Table 3.

The number of patients in each category of SteatoTest, ActiTest, and FibroTest is shown in Supplementary Table 2.

3.3. Examination of Metabolic Biomarkers, Inflammation, and AFP. Table 4 shows lipid parameters and HbA1c. Four of ten older patients had type 2 diabetes. No difference was found between the two groups in total cholesterol. However, when broken down into HDL and LDL cholesterol, the older group showed significantly lower HDL cholesterol and significantly higher LDL cholesterol values (HDL cholesterol terol: $p \le 0.001$; LDL cholesterol: $p \le 0.05$).

The LDL cholesterol values of the two groups were within the normal range. HDL cholesterol was lower in 8/20 (40%) younger patients and in all older patients (100%).



FIGURE 1: (a) Steatosis assessed by TE. (b) Liver stiffness assessed by TE. Steatosis and liver stiffness are more pronounced in older patients. p < 0.001.



FIGURE 2: (a) Correlation of steatosis with age. (b) Correlation of liver stiffness with age. Steatosis and liver stiffness progress with age. p < 0.001.

	TABLE 2: Results	of FibroMax.	
Parameter	Group 1 $(n = 20)$	Group 2 $(n = 10)$	p value
SteatoTest	0.02 ± 0.02	0.47 ± 0.24	< 0.0001
ActiTest	0.1 ± 0.06	0.39 ± 0.16	< 0.001

Mean values ± SD are shown.

There was a significant difference in the LDL-HDL ratio between the two patient groups ($p \le 0.0001$). Triglycerides were significantly increased in group 2 (p < 0.0001). Five of ten (50%) older patients had values above the normal range. In group 1, the triglyceride values were all within the normal range. In addition, there was a significant correlation of triglycerides with age (r = 0.66, $p \le 0.0001$).

As expected, AFP was elevated in all patients. However, there was a significant correlation of AFP values with age (r=0.54, p<0.01).

For CRP, no significant difference between the two groups was found.

4. Discussion

A-T is a life-limiting systemic disease clinically characterized by neurodegeneration, radiosensitivity, increased risk of malignancy, immunodeficiency, failure to thrive, and

TABLE 3: Correlation of SteatoTest and ActiTest.

Parameter	Correlated parameter	R	<i>p</i> value
SteatoTest	Age	0.74	< 0.0001
	CAP	0.69	< 0.0001
	LSM	0.51	< 0.01
	Triglycerides	0.71	< 0.0001
	LDL/HDL	0.79	< 0.0001
	HbA1c	0.67	< 0.001
	ActiTest	0.89	< 0.0001
	CRP	0.51	< 0.01
ActiTest	Age	0.80	< 0.0001
	CAP	0.77	< 0.0001
	LSM	0.53	< 0.01
	Triglycerides	0.61	< 0.001
	LDL/HDL	0.80	< 0.0001
	CRP	0.41	< 0.05

hepatopathy [1, 3, 9–11, 14, 33]. To date, the clinical significance of liver disease is unclear, but up to over 90% of patients develop an elevation of liver enzymes with advancing age, which is associated with a high degree of fatty degeneration and sometimes fibrosis of the liver tissue [14].

To the best of our knowledge, the present study is the first prospective trial addressing noninvasive procedures to characterize liver disease in A-T and relate outcomes of TE to



FIGURE 3: (a) Correlation of ActiTest with triglycerides, (b) correlation of ActiTest with CRP, (c) correlation of ActiTest with CAP, and (d) correlation of ActiTest with LSM.

Parameter	Group 1 (<i>n</i> = 21)	Group 2 (<i>n</i> = 10)	p value
Cholesterol (mg/dL)	178.1 ± 36.3	190.9 ± 32.4	n.s.
HDL (mg/dL)	62.3 ± 20.4	35.1 ± 6.4	< 0.001
LDL (mg/dL)	106.5 ± 23.5	123.8 ± 25.1	< 0.05
LDL-HDL ratio	1.9 ± 0.6	3.7 ± 1	< 0.0001
Triglycerides (mg/ dL)	66.5 ± 34.3	200.4 ± 98.8	< 0.0001
Diabetes type 2	0	4 (40%)	
HbA1C (%Hb)	4.8 ± 0.4	5.7 ± 0.6	< 0.0001

TABLE 4: Metabolic biomarkers.

Mean values ± SD are shown.

liver biomarkers and liver scores. Our results demonstrate for the first time that 10% of younger patients as opposed to 90% of older patients have liver steatosis. In line with this finding, higher degrees of steatosis were present in the older group. Pronounced fibrosis was found in 3/10 (30%) older patients. In two patients, cirrhosis was already present according to TE. We found a significantly higher necroinflammatory activity (ActiTest) in the older patient group. Necroinflammation can be defined as the immune response to necrosis [34]. Therefore, ActiTest is a good marker for the progression of liver disease to steatosis and apoptosis with increased necroinflammatory activity. The presence of necroinflammatory activity, which also correlates with the CRP value, indicates NASH, which according to our data mainly affects older patients. In summary, SteatoTest and ActiTest are a suitable diagnostic assessment for steatosis and necroinflammatory activity in patients with A-T and have confirmed the TE results.

FibroTest did not show a significant difference between the two patient groups or a significant correlation with age or the results of TE, most likely since FibroMax is not licensed below the age of 14 years due to the physiological increase of some of the serum markers used for calculation.

We were also able to show a significant correlation of AFP with steatosis (CAP). However, the mechanism has not been elucidated so far. AFP is mainly known as a tumor marker for HCC. Serum AFP may also be elevated by germ cell tumors, viral hepatitis, liver fibrosis, and neurodegenerative diseases such as A-T [35-37]. Among other mechanisms, tumor suppressor p53 acts as a repressor on the AFP gene during development and regeneration of the liver [38–40]. Via the reduced activation of p53 due to the absence of the ataxia-telangiectasia-mutated (ATM) kinase, an increased expression of AFP could thus occur in A-T [41]. A mutation of p53 is also frequently found in HCC [42], which could be a possible explanation for an increase in AFP. In contrast to CAP, however, no significant correlation of LSM with AFP was found. The missing correlation of AFP with the LSM could thus be explained by a loss of functional liver tissue in cirrhosis.

Dyslipidemia is common in older A-T patients [6, 8]. While triglycerides and LDL cholesterol were significantly higher, HDL cholesterol was significantly lower in the older patient group. In addition, there was a significant correlation of triglycerides with age. The association between elevated liver enzymes and dyslipidemia has been described before [6]. We could also show a significant correlation of HbA1c and triglycerides with CAP and LSM, emphasizing the effects of metabolic risk factors for liver tissue remodeling. Diabetic metabolism leads to fatty liver remodeling due to hyperglycemia and hypertriglyceridemia and thus increases the risk of NASH. In addition, the described alterations in cholesterol and triglyceride levels in patients indicate an increased atherosclerotic risk profile [43]. They could also play an important role in the development of steatosis, through the accumulation of fat in the liver [44]. The increased influx of free fatty acids to the liver leads to an increased synthesis of triglycerides and very low-density lipoprotein (VLDL), as well as a reduced synthesis of HDL cholesterol [45].

ATM is induced by the accumulation of fat and thus elevates oxidative stress in the liver [46] and acts as an activator of p53, which in turn activates the p53 upregulated modulator of apoptosis (PUMA) [47]. PUMA is a crucial player in steatosis and apoptosis in hepatocytes [48]. Since hepatocyte apoptosis correlates with the severity of NASH and the stage of fibrosis [49], it can be assumed that steatosis related apoptosis is partly responsible for the progression of liver disease [46]. In the absence of ATM, alternative signaling pathways must be activated, which ultimately cause fibrotic remodeling and death of functional liver tissue.

Fibrosis results from connective tissue remodeling due to chronic inflammation of the liver tissue [50, 51], e.g., in response to steatosis and apoptosis [46]. The progression of fibrosis to cirrhosis of the liver then appears to develop through repetitive phases of inflammation and the subsequent reparative immune response [52]. This leads to a loss of functional liver tissue, which is replaced by scar tissue [50, 53].

ATM can be activated directly by oxidative stress, even independently by double-strand breaks in DNA [54]. The absence of ATM leads to low antioxidant capacity [55]. As a result, macromolecules, lipids, and DNA are exposed to permanent oxidative stress and damage it causes. Similar results have been shown for NAFLD and in particular NASH, where oxidative stress and lipid peroxidation are also elevated [56]. For example, hepatocytes with excess fat are particularly susceptible to oxidative stress and DNA damage [57]. This may be related to the fact that saturated free fatty acids promote the formation of ROS, thereby inducing apoptosis and inflammation [58]. It is also known that oxidative stress increases with the severity of liver disease [57]. Oxidative stress and resulting inflammation appear to play a major role in the development and progression of liver disease. A link between hepatopathy in A-T and the already known increased oxidative stress levels in patients therefore seems very likely.

The type of liver involvement in A-T is not yet defined, as the published cases with severe portal hypertension causing ascites and varices had no evidence of cirrhosis at a liver biopsy [15, 16, 18]. It seems that whether these complications are to be attributed to steatosis/NASH leading to portal hypertension in the absence of cirrhosis [59] or to vascular liver disease/porto-sinusoidal vascular disorder (PSVD) is still open for discussion [60]. In this view, clinicians have to consider the value of a liver biopsy, especially in a research context where little is known about liver disease in A-T (i.e., the diagnosis of PSVD can be histological). The rate of adverse events in the modern era is negligible, and the information one can obtain is precious.

The study has some limitations. Due to the size of the study population and the monocentric study design, no general statements can be made. There are no corresponding ultrasound reports that can be compared with the TE results. This is a major limit, not only because it could have shown how liver disease (i.e., fibrosis staging) would have been underestimated by ultrasound only, but also because signs of portal hypertension (i.e., splenomegaly, dilated portal vein, and collaterals) could have been detected and correlated with the absence/presence of liver cirrhosis at TE. In addition, the application of spleen stiffness would have been useful to address more specifically the severity of portal hypertension, as is complementary to LSM [61], and the spleen stiffness measurement (SSM)/LSM ratio could be informative in these cases [62].

Additionally, the examination methods used in this study are not yet established practice in pediatrics, so there are no universally accepted cutoff values for children, but several studies have highlighted the value of TE with CAP [63–66]. The proposed cutoff values are largely consistent with those for adults, which were used in this study. Also FibroTest could show its diagnostic value in some pediatric studies and could distinguish between severe fibrosis and absence of fibrosis [67]. To the best of our knowledge, there are no pediatric studies on FibroMax to date.

Due to the mildness of liver disease, validation of TE by biopsy (gold standard) was not performed. Although CAP is not recommended as a standard tool for stratification of hepatic steatosis [68], there are data showing a good correlation with other steatosis markers, and our data also support this. Another limitation is the lack of a healthy control group to compare the results. As only one TE measurement was performed in each patient, no interobserver comparison is possible. For detection of steatosis by TE, larger cohorts and multicenter studies are needed in order to validate the clinical application.

5. Conclusion

Premature aging, low-grade inflammation, oxidative stress, dyslipidemia, and IR as common features of A-T contribute to the development of NAFLD. Liver disease in the context of A-T should be monitored regularly in order to prevent long-term consequences, such as NASH, cirrhosis, and HCC. TE and liver scores are a well reproducible, noninvasive method detecting early stages of liver disease. SteatoTest and ActiTest are useful to assess steatosis and necroinflammatory activity in patients with A-T and have confirmed the TE results.

Abbreviations:

A-T:	Ataxia-telangiectasia
NASH:	Nonalcoholic steatohepatitis
BMI:	Body mass index
HCC:	Hepatocellular carcinoma
IR:	Insulin resistance
NAFLD:	Nonalcoholic fatty liver disease
HbA1c:	Hemoglobin A1c
TE:	Transient elastography
CF:	Cystic fibrosis
CFLD:	Cystic fibrosis liver disease
CRP:	C-reactive protein
LSM:	Liver stiffness measurement
CAP:	Controlled attenuation parameter
GGT:	Gamma-glutamyl transferase
ALT:	Alanine aminotransferase
AST:	Aspartate aminotransferase
AFP:	Alpha-fetoprotein
HDL:	High-density lipoprotein
LDL:	Low-density lipoprotein
SD:	Standard deviation
ATM:	Ataxia-telangiectasia-mutated
VLDL:	Very low-density lipoprotein
PUMA:	P53 upregulated modulator of apoptosis
PSVD:	Porto-sinusoidal vascular disorder.

Data Availability

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Additional Points

At a Glance Commentary. Scientific knowledge on the subject: The majority of older A-T patients suffer from liver disease. This is the first prospective measurement of transient elastography in a larger A-T cohort. What This Study Adds to the Field. Transient elastography is a new, noninvasive, and reproducible technique that measures liver stiffness and detects early stages of NASH and cirrhosis in A-T patients.

Ethical Approval

The Ethics Committee of University Hospital Frankfurt approved the trial (Reference No. 504/15). The study was registered at clinicaltrials.gov (NCT03357978). The study was conducted according to the ethical principles of the Declaration of Helsinki and regulatory requirements and the code of Good Clinical Practice.

Consent

Written consent was obtained from all patients and/or caregivers.

Disclosure

H. Donath and S. Wölke are the co-first authors.

Conflicts of Interest

The authors declare no conflicts of interest.

Authors' Contributions

HD, SW, UH, SZ, JT, RS, and RPD designed the study, collected the data, and interpreted and carried out statistical analysis. HD, SW, UH, and SZ made patient visits. VK performed TE. TP performed the FibroMax calculation. HD and SZ wrote the manuscript. All authors have read, revised, and approved the final manuscript. H. Donath and S. Wölke equally contributed to this work.

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Supplementary Materials

Supplementary Table 1: an overview of the distribution of fibrosis and steatosis stages detected by TE. Supplementary Table 2: the distribution of the stage of liver involvement measured by ActiTest, SteatoTest, and FibroTest. (*Supplementary Materials*)

References

- E. Boder and R. P. Sedgwick, "Ataxia-telangiectasia; a familial syndrome of progressive cerebellar ataxia, oculocutaneous telangiectasia and frequent pulmonary infection," *Pediatrics*, vol. 21, no. 4, pp. 526–554, 1958.
- [2] M. Ambrose and R. A. Gatti, "Pathogenesis of ataxia-telangiectasia: the next generation of ATM functions," *Blood*, vol. 121, no. 20, pp. 4036–4045, 2013.
- [3] M. F. Lavin, "Ataxia-telangiectasia: from a rare disorder to a paradigm for cell signalling and cancer," *Nature Reviews Molecular Cell Biology*, vol. 9, no. 10, pp. 759–769, 2008.
- [4] R. Micol, L. Ben Slama, F. Suarez et al., "Morbidity and mortality from ataxia-telangiectasia are associated with ATM genotype," *The Journal of Allergy and Clinical Immunology*, vol. 128, no. 2, pp. 382–389.e1, 2011.
- [5] C. Rothblum-Oviatt, J. Wright, M. A. Lefton-Greif, S. A. McGrath-Morrow, T. O. Crawford, and H. M. Lederman, "Ataxia telangiectasia: a review," *Orphanet Journal of Rare Diseases*, vol. 11, no. 1, p. 159, 2016.
- [6] A. Nissenkorn, Y. Levy-Shraga, Y. Banet-Levi, A. Lahad, I. Sarouk, and D. Modan-Moses, "Endocrine abnormalities in ataxia telangiectasia: findings from a national cohort," *Pediatric Research*, vol. 79, no. 6, pp. 889–894, 2016.
- [7] B. Weiss, A. Krauthammer, M. Soudack et al., "Liver disease in pediatric patients with ataxia telangiectasia: a novel report," *Journal of Pediatric Gastroenterology and Nutrition*, vol. 62, no. 4, pp. 550–555, 2016.
- [8] T. L. Paulino, M. N. Rafael, S. Hix et al., "Is age a risk factor for liver disease and metabolic alterations in ataxia Telangiectasia patients?" *Orphanet Journal of Rare Diseases*, vol. 12, no. 1, p. 136, 2017.
- [9] S. Voss, J. Pietzner, F. Hoche et al., "Growth retardation and growth hormone deficiency in patients with Ataxia telangiectasia," *Growth Factors*, vol. 32, no. 3-4, pp. 123–129, 2014.

- [10] H. Pommerening, S. van Dullemen, M. Kieslich, R. Schubert, S. Zielen, and S. Voss, "Body composition, muscle strength and hormonal status in patients with ataxia telangiectasia: a cohort study," *Orphanet Journal of Rare Diseases*, vol. 10, no. 1, p. 155, 2015.
- [11] S. Woelke, H. Pommerening, M. Kieslich, R. Schubert, and S. Zielen, "Growth hormone treatment in patients with ataxia telangiectasia," *Growth Factors*, vol. 35, no. 2-3, pp. 125–130, 2017.
- [12] S. A. Schroeder and S. Zielen, "Infections of the respiratory system in patients with ataxia-telangiectasia," *Pediatric Pulmonology*, vol. 49, no. 4, pp. 389–399, 2014.
- [13] L. J. Ross, S. Capra, B. Baguley et al., "Nutritional status of patients with ataxia-telangiectasia: a case for early and ongoing nutrition support and intervention," *Journal of Paediatrics and Child Health*, vol. 51, no. 8, pp. 802–807, 2015.
- [14] H. Donath, S. Woelke, M. Theis et al., "Progressive liver disease in patients with ataxia telangiectasia," *Front Pediatr*, vol. 7, p. 458, 2019.
- [15] L. H. Pillsbury, S. M. Peters, R. L. Wientzen, T. Phillips, Z. L. Papadopoulou, and J. A. Bellanti, "Ataxia telangiectasia: immunologically mediated renal and hepatic failure," *Annals* of Allergy, vol. 55, no. 4, pp. 539–8, 1985.
- [16] M. Casaril, G. B. Gabrielli, F. Capra, and G. C. Falezza, "Atassia-teleangiectasia. Descrizione di un caso con emorragie cerebrali multiple e cirrosi epatica," *Minerva Medica*, vol. 73, no. 34, pp. 2183–2188, 1982.
- [17] M. M. Patil and S. V. Patil, "Ataxia telangiectasia with hepatocellular carcinoma," *Indian Pediatrics*, vol. 46, no. 6, p. 546, 2009.
- [18] T. Caballero, M. Caba-Molina, J. Salmerón, and M. Gómez-Morales, "Nonalcoholic steatohepatitis in a patient with ataxia-telangiectasia," *Case Reports Hepatol*, vol. 201412 pages, Article ID 761250, 2014.
- [19] Y. Shiloh and H. M. Lederman, "Ataxia-telangiectasia (A-T): an emerging dimension of premature ageing," *Ageing Research Reviews*, vol. 33, pp. 76–88, 2017.
- [20] H. Donath, U. Hess, M. Kieslich et al., "Diabetes in patients with ataxia telangiectasia: a national cohort study," *Front Pediatr*, vol. 8, p. 317, 2020.
- [21] H. Morikawa and N. Kawada, "Non-invasive diagnosis of liver fibrosis," *Clin J Gastroenterol*, vol. 4, no. 5, pp. 283–291, 2011.
- [22] G. L.-H. Wong, "Update of liver fibrosis and steatosis with transient elastography (Fibroscan)," *Gastroenterology Report*, vol. 1, no. 1, pp. 19–26, 2013.
- [23] A. A. Bravo, S. G. Sheth, and S. Chopra, "Liver biopsy," New England Journal of Medicine, vol. 344, no. 7, pp. 495–500, 2001.
- [24] L. Castera, X. Forns, and A. Alberti, "Non-invasive evaluation of liver fibrosis using transient elastography," *Journal of Hepatology*, vol. 48, no. 5, pp. 835–847, 2008.
- [25] L. Sandrin, B. Fourquet, J.-M. Hasquenoph et al., "Transient elastography: a new noninvasive method for assessment of hepatic fibrosis," *Ultrasound in Medicine and Biology*, vol. 29, no. 12, pp. 1705–1713, 2003.
- [26] M. Fraquelli, C. Rigamonti, G. Casazza et al., "Reproducibility of transient elastography in the evaluation of liver fibrosis in patients with chronic liver disease," *Gut*, vol. 56, no. 7, pp. 968–973, 2007.
- [27] M. Sasso, M. Beaugrand, V. de Ledinghen et al., "Controlled attenuation parameter (CAP): a novel VCTE[™] guided ultrasonic attenuation measurement for the evaluation of hepatic steatosis: preliminary studyand validation in a cohort of patients with chronic liver disease from various causes,"

Ultrasound in Medicine and Biology, vol. 36, no. 11, pp. 1825–1835, 2010.

- [28] T. Karlas, D. Petroff, M. Sasso et al., "Individual patient data meta-analysis of controlled attenuation parameter (CAP) technology for assessing steatosis," *Journal of Hepatology*, vol. 66, no. 5, pp. 1022–1030, 2017.
- [29] M. Munteanu, V. Ratziu, R. Morra, D. Messous, F. Imbert-Bismut, and T. Poynard, "Noninvasive biomarkers for the screening of fibrosis, steatosis and steatohepatitis in patients with metabolic risk factors: FibroTest-FibroMax experience," *J Gastrointestin Liver Dis*, vol. 17, no. 2, pp. 187–191, 2008.
- [30] European Association for the Study of the Liver Easl, "EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease," *Journal of Hep-atology*, vol. 64, no. 6, pp. 1388–1402, 2016.
- [31] T. Poynard, V. Ratziu, S. Naveau et al., "The diagnostic value of biomarkers (SteatoTest) for the prediction of liver steatosis," *Comparative Hepatology*, vol. 4, no. 1, 10 pages, 2005.
- [32] T. Poynard, F. Imbert-Bismut, M. Munteanu et al., "Overview of the diagnostic value of biochemical markers of liver fibrosis (FibroTest, HCV FibroSure) and necrosis (ActiTest) in patients with chronic hepatitis C," *Comparative Hepatology*, vol. 3, no. 1, p. 8, 2004.
- [33] M. Ehlayel, A. Soliman, and V. De Sanctis, "Linear growth and endocrine function in children with ataxia telangiectasia," *Indian Journal of Endocrinology and Metabolism*, vol. 18, no. 7, pp. S93–S96, 2014.
- [34] M. Sarhan, W. G. Land, W. Tonnus, C. P. Hugo, and A. Linkermann, "Origin and consequences of necroinflammation," *Physiological Reviews*, vol. 98, no. 2, pp. 727– 780, 2018.
- [35] C. Sauzay, A. Petit, A.-M. Bourgeois et al., "Alphafoetoprotein (AFP): a multi-purpose marker in hepatocellular carcinoma," *Clinica Chimica Acta*, vol. 463, pp. 39–44, 2016.
- [36] K. Nakao and T. Ichikawa, "Recent topics on α-fetoprotein," *Hepatology Research*, vol. 43, no. 8, pp. 820–825, 2013.
- [37] J. H. Schieving, M. de Vries, J. M. G. van Vugt et al., "Alphafetoprotein, a fascinating protein and biomarker in neurology," *European Journal of Paediatric Neurology*, vol. 18, no. 3, pp. 243–248, 2014.
- [38] R. Cui, T. T. Nguyen, J. H. Taube, S. A. Stratton, M. H. Feuerman, and M. C. Barton, "Family members p53 and p73 act together in chromatin modification and direct repression of alpha-fetoprotein transcription," *Journal of Biological Chemistry*, vol. 280, no. 47, pp. 39152–39160, 2005.
- [39] T. T. Nguyen, K. Cho, S. A. Stratton, and M. C. Barton, "Transcription factor interactions and chromatin modifications associated with p53-mediated, developmental repression of the alpha-fetoprotein gene," *Molecular and Cellular Biology*, vol. 25, no. 6, pp. 2147–2157, 2005.
- [40] D. S. Wilkinson, S. K. Ogden, S. A. Stratton et al., "A direct intersection between p53 and transforming growth factor beta pathways targets chromatin modification and transcription repression of the alpha-fetoprotein gene," *Molecular and Cellular Biology*, vol. 25, no. 3, pp. 1200–1212, 2005.
- [41] A. Stray-Pedersen, A. L. Borresen-Dale, E. Paus, C. R. Lindman, T. Burgers, and T. G. Abrahamsen, "Alpha fetoprotein is increasing with age in ataxia-telangiectasia," *European Journal of Paediatric Neurology*, vol. 11, no. 6, pp. 375–380, 2007.
- [42] S.-Y. Peng, W. J. Chen, P.-L. Lai, Y.-M. Jeng, J.-C. Sheu, and H.-C. Hsu, "High alpha-fetoprotein level correlates with high stage, early recurrence and poor prognosis of hepatocellular

- [43] I. G. A. Andrade, B. T. Costa-Carvalho, R. da Silva et al., "Risk of atherosclerosis in patients with ataxia telangiectasia," *Annals of Nutrition and Metabolism*, vol. 66, no. 4, pp. 196– 201, 2015.
- [44] K. Cusi, "Role of insulin resistance and lipotoxicity in nonalcoholic steatohepatitis," *Clinics in Liver Disease*, vol. 13, no. 4, pp. 545–563, 2009.
- [45] N. Li, J. Fu, D. P. Koonen, J. A. Kuivenhoven, H. Snieder, and M. H. Hofker, "Are hypertriglyceridemia and low HDL causal factors in the development of insulin resistance?" *Atherosclerosis*, vol. 233, no. 1, pp. 130–138, 2014.
- [46] E. K. Daugherity, G. Balmus, A. Al Saei et al., "The DNA damage checkpoint protein ATM promotes hepatocellular apoptosis and fibrosis in a mouse model of non-alcoholic fatty liver disease," *Cell Cycle*, vol. 11, no. 10, pp. 1918–1928, 2012.
- [47] A. Villunger, E. M. Michalak, L. Coultas et al., "p53- and druginduced apoptotic responses mediated by BH3-only proteins puma and noxa," *Science*, vol. 302, no. 5647, pp. 1036–1038, 2003.
- [48] S. C. Cazanave, J. L. Mott, N. A. Elmi et al., "JNK1-dependent PUMA expression contributes to hepatocyte lipoapoptosis," *Journal of Biological Chemistry*, vol. 284, no. 39, pp. 26591– 26602, 2009.
- [49] A. E. Feldstein, A. Canbay, P. Angulo et al., "Hepatocyte apoptosis and fas expression are prominent features of human nonalcoholic steatohepatitis," *Gastroenterology*, vol. 125, no. 2, pp. 437–443, 2003.
- [50] D. Schuppan, M. Ruehl, R. Somasundaram, and E. G. Hahn, "Matrix as a modulator of hepatic fibrogenesis," *Seminars in Liver Disease*, vol. 21, no. 03, pp. 351–372, 2001.
- [51] R. Bataller and D. A. Brenner, "Liver fibrosis," *Journal of Clinical Investigation*, vol. 115, no. 2, pp. 209–218, 2005.
- [52] D. Schuppan, R. Surabattula, and X. Y. Wang, "Determinants of fibrosis progression and regression in NASH," *Journal of Hepatology*, vol. 68, no. 2, pp. 238–250, 2018.
- [53] H. Popper and S. Udenfriend, "Hepatic fibrosis," *The American Journal of Medicine*, vol. 49, no. 5, pp. 707–721, 1970.
- [54] Z. Guo, S. Kozlov, M. F. Lavin, M. D. Person, and T. T. Paull, "ATM activation by oxidative stress," *Science*, vol. 330, no. 6003, pp. 517–521, 2010.
- [55] J. Reichenbach, R. Schubert, C. Schwan, K. Müller, H. J. Böhles, and S. Zielen, "Anti-oxidative capacity in patients with ataxia telangiectasia," *Clinical and Experimental Immunology*, vol. 117, no. 3, pp. 535–539, 2001.
- [56] M. Parola and G. Robino, "Oxidative stress-related molecules and liver fibrosis," *Journal of Hepatology*, vol. 35, no. 2, pp. 297–306, 2001.
- [57] R. N. Hardwick, C. D. Fisher, M. J. Canet, A. D. Lake, and N. J. Cherrington, "Diversity in antioxidant response enzymes in progressive stages of human nonalcoholic fatty liver disease," *Drug Metabolism and Disposition*, vol. 38, no. 12, pp. 2293–2301, 2010.
- [58] Y. Noguchi, J. D. Young, J. O. Aleman, M. E. Hansen, J. K. Kelleher, and G. Stephanopoulos, "Tracking cellular metabolomics in lipoapoptosis- and steatosis-developing liver cells," *Molecular BioSystems*, vol. 7, no. 5, pp. 1409–1419, 2011.
- [59] G. Baffy and J. Bosch, "Overlooked subclinical portal hypertension in non-cirrhotic NAFLD: is it real and how to measure it?" *Journal of Hepatology*, vol. 76, no. 2, pp. 458–463, 2022.

- [60] A. De Gottardi, C. Sempoux, and A. Berzigotti, "Porto-sinusoidal vascular disorder," *Journal of Hepatology*, vol. 77, no. 4, pp. 1124–1135, 2022.
- [61] E. Dajti, F. Ravaioli, G. Marasco et al., "A combined baveno VII and spleen stiffness algorithm to improve the noninvasive diagnosis of clinically significant portal hypertension in patients with compensated advanced chronic liver disease," *American Journal of Gastroenterology*, vol. 117, no. 11, pp. 1825–1833, 2022.
- [62] J. Ferreira-Silva, R. Gaspar, R. Liberal, H. Cardoso, and G. Macedo, "Splenic-hepatic elastography index is useful in differentiating between porto-sinusoidal vascular disease and cirrhosis in patients with portal hypertension," *Digestive and Liver Disease*, vol. 55, no. 1, pp. 75–80, 2023.
- [63] B. R. Chen and C. Q. Pan, "Non-invasive assessment of fibrosis and steatosis in pediatric non-alcoholic fatty liver disease," *Clinics and Research in Hepatology and Gastroenterology*, vol. 46, no. 1, Article ID 101755, 2022.
- [64] J. Y. Hwang, H. M. Yoon, J. R. Kim et al., "Diagnostic performance of transient elastography for liver fibrosis in children: a systematic review and meta-analysis," *American Journal of Roentgenology*, vol. 211, no. 5, pp. W257–W266, 2018.
- [65] X. Gong, T. Zhu, X. Peng, D. Xing, and M. Zhang, "Diagnostic accuracy of transient elastography and two-dimensional shear wave elastography for staging liver fibrosis in children or adolescents: a systematic review and meta-analysis," *Current medical imaging*, vol. 18, 2022.
- [66] S. Jia, Y. Zhao, J. Liu et al., "Magnetic resonance imagingproton density fat fraction vs. Transient elastographycontrolled attenuation parameter in diagnosing nonalcoholic fatty liver disease in children and adolescents: a meta-analysis of diagnostic accuracy," *Front Pediatr*, vol. 9, Article ID 784221, 2021.
- [67] J. Flores-Calderón, S. Morán-Villota, G. Ramón-García et al., "Non-invasive markers of liver fibrosis in chronic liver disease in a group of Mexican children. A multicenter study," *Annals* of *Hepatology*, vol. 11, no. 3, pp. 364–368, 2012.
- [68] A. Berzigotti, E. Tsochatzis, J. Boursier et al., "Electronic address: easloffice@easloffice.eu; Clinical Practice Guideline Panel; Chair:; EASL Governing Board representative:; Panel members:. EASL Clinical Practice Guidelines on non-invasive tests for evaluation of liver disease severity and prognosis -2021 update," *Journal of Hepatology*, vol. 75, no. 3, pp. 659– 689, 2021.