

## Dual control of host actin polymerization by a *Legionella* effector pair

### Authors

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### SUPPLEMENTAL DATA LEGENDS

**Supplementary Data S1 – List of all cells, bacteria strains used in this study.**

**Supplementary Data S2 – List of plasmids used in this study.**

**Supplementary Data S3 – List of oligonucleotides used to realize mutant strains of *Legionella* as well as for cloning.**

**Supplementary Data S4 – Accession numbers of genomes used for evolutionary analyses.**

**Supplementary Data S5 – Fasta files of sequencing data of  $\Delta legK2$  and  $\Delta legK2/\Delta vipA$  *Legionella pneumophila* strains**

**Supplementary Data S6 – LegK2 and VipA mutations do not alter axenic growth of *L. pneumophila* Paris strains.** Axenic growth kinetics of *L. pneumophila* Paris WT,  $\Delta dotA$ ,  $\Delta legK2$ ,  $\Delta vipA$ ,  $\Delta legK2/\Delta vipA$ , mutant strains transformed with mCherry-expressing plasmids. AYE broth was seeded with  $0.1 \times 10^9$  bacteria/mL and then plated in a 96well-Greiner plate, with black wells and transparent flat bottom. OD<sub>600nm</sub> absorbance was measured every 30min at 595 nm on an Infinite M200 microplate reader (Tecan).

**Supplementary Data S7 – VipA and LegK2 are produced at the onset of the stationary phase in axenic medium at 30°C.** Luminescence emission from chromosomal translational fusions of *vipA::luc(A)* and *legK2::luc(B)* in *L. pneumophila* Paris strain grown in AYE medium at 30°C. Luminescence emission of VipA-Luc and LegK2-Luc fusion proteins is maximal at the onset of the stationary growth phase (growth at 30°C measured by OD<sub>600</sub>, dashed red line), with VipA-Luc slightly ahead of LegK2-Luc. The data shown are representative of 3 independent clones of each fusion.

**Supplementary Data S8 – Mass Spectrometry data following protein expression in *Legionella pneumophila* Paris WT at OD<sub>600nm</sub> 1, 2, 3, 4 and 5.** VipA accumulates in *L. pneumophila* up to OD<sub>600</sub> = 4. Three independent cultures of *L. pneumophila* Paris WT were grown in liquid medium AYE (at 30°C). At the desired OD<sub>600</sub> (1, 2, 3, 4 and 5), samples were collected, and their protein content was analysed by mass spectrometry after sample-specific labelling (see Material and methods for details). The graphs show the quantity detected at each OD<sub>600</sub> or as a ratio compared to the quantity at OD<sub>600</sub>. The RocC protein is used as a control as it is detected at the same range of quantity as VipA and its production during growth was previously studied by Western-blot<sup>23</sup>. Of note the RocC pattern of production detected by mass spectrometry corresponds to the one previously obtained by Western-blot<sup>23</sup>.

**Supplementary Data S9 - LegK2/VipA effector pairs are restricted and strongly conserved in the *L. pneumophila* species.** Phylogenetic tree representing the phylogenetic diversity of *Legionella* genomes displaying either LegK2 (blue green squares) or VipA genes (purple squares). The squares are filled when the genes have been found in corresponding genome and empty when genes have not been detected.

#### SUPPLEMENTARY DATA

Strains		
Names	Genotypes	References
<b><i>Dictyostelium discoideum</i></b>		
DBS0236184	Calnexin-GFP	Müller-Taubenberger et al. (2001)
DBS0235534	Ax2-214	Watts & Ashworth, (1970)
<b><i>Acanthamoeba castellanii</i></b>		
Environmental isolate		
<b>Mammalian cells</b>		
HeLa cells		Inserm U1111, Lyon, France
U937	Human monocytes ATCC CRL1593.2	Sundstrom et al. (1976)
<b><i>Legionella pneumophila</i></b>		
WT	Virulent <i>L. pneumophila</i> strain Paris CIP 107629	Cazalet et al. (2004)
$\Delta dotA$	Paris <i>lpp2740::Km</i>	Cazalet et al. (2004)
$\Delta legK2$	Paris $\Delta lpp2076$ (scar-free deletion of <i>lpp2076</i> )	This study
$\Delta vipA$	Paris $\Delta lpp0457$ (scar-free deletion of <i>lpp0457</i> )	This study
$\Delta legK2\Delta vipA$	Paris $\Delta lpp2076 \Delta lpp0457$ (scar-free deletion of <i>lpp2076</i> and <i>lpp0457</i> )	This study
<i>legK2::luc</i>		This study
<i>vipA::luc</i>		This study
<i>legK2::gfp</i>		This study
<b><i>Escherichia coli</i></b>		
DH5 $\alpha$	<i>endA1 hsdR17 supE4, thi-1 recA1 gyrArelA, <math>\Delta lac</math></i>	Laboratory
BL21	<i>B dcmompThsdS(rB-mB-) gal</i>	Laboratory
XL1 Blue	<i>endA1 gyrA96(nalR) thi-1 recA1 relA1 lac glnV44 F'[/Tn10 proAB+ lacIq<math>\Delta</math>(lacZ)M15] hsdR17(rK- mK+)</i>	Stratagene

#### SUPPLEMENTARY DATA S1

Plasmids		
Names	Characteristics	References
Donor vectors for Gateway cloning		
pDONR <sup>TM</sup> 207	Donor Gateway vector	Invitrogen
pDONR <sup>TM</sup> 207- <i>vipA</i>	Donor Gateway vector with insertion of lpp0457 gene	This study
Mammalian cells expression vectors for Gateway cloning		
pDEST27	Expression Gateway vector for mammalian cells allowing overexpression of GST-tagged protein	Invitrogen
pDEST27- <i>legK2</i>	pDEST27 with insertion of lpp2076 gene for expression of GST-LegK2 fusion protein	This study
pDEST27- <i>legK2</i> <sub>K112M</sub>	pDEST27 with insertion of lpp2076 gene for expression of GST-LegK2 <sub>K112M</sub> dead kinase-mutant fusion protein	This study
pCI-Neo3Flag	Expression Gateway vector for mammalian cells allowing overexpression of Flag-tagged protein	Invitrogen
pCI-Neo3Flag- <i>ArpC1b</i>	pCI-Neo3Flag with insertion of ARPC1B human gene for expression of Flag-ARPC1B fusion protein	(Michard et al. 2015)
peGFP-Nterm	Expression Gateway vector for mammalian cells allowing overexpression of Nterminal GFP-tagged protein	INSERM U1111, Lyon, France
peGFP-Cterm	Expression Gateway vector for mammalian cells allowing overexpression of Nterminal GFP-tagged protein	INSERM U1111, Lyon, France
peGFP-N- <i>vipA</i>	peGFP-Nterm with insertion of lpp0457 gene for expression of VipA-GFP fusion protein	This study
peGFP-C- <i>vipA</i>	peGFP-Cterm with insertion of lpp0457 gene for expression of GFP-VipA fusion protein	This study
Bacteria expression vectors		
pXDC50	Expression plasmid for <i>L. pneumophila</i> allowing expression of mCherry fluorescent protein	(Hervet et al. 2011)
pXDC50-lpp2076	Expression plasmid for <i>L. pneumophila</i> allowing inducible expression of mCherry fluorescent protein and expression of LegK2 under control of its own promoter	This study
pXDC61	Expression plasmid for <i>L. pneumophila</i> allowing overexpression of $\beta$ -Lactamase	Charpentier et al. (2009)
pXDC61- <i>legK2</i>	Expression plasmid for <i>L. pneumophila</i> allowing overexpression of $\beta$ -Lactamase-LegK2 fusion protein	This study
pXDC61- <i>vipA</i>	Expression plasmid for <i>L. pneumophila</i> allowing overexpression of $\beta$ -Lactamase-VipA fusion protein	This study
pGEX- <i>legK2</i>	Overproduction and <i>in vitro</i> purification of GST-LegK2	This study
pQE30- <i>vipA</i>	Overproduction and <i>in vitro</i> purification of GST-LegK2	This study

## SUPPLEMENTARY DATA S2

Primers		
Names	Sequences	Description
1_vipA fwd	GGGGACAAGTTTGTACAAAAAAGCAGGCTTAatgcctatcagtaatgcc ttt	vipA insertion in pDONR207 vector for peGFP-C-VipA
2_vipA rev	GGGGACCACTTTGTACAAGAAAGCTGGGTTgagatttttttttcgacggt agtg	
3_vipA fwd	GGGGACCACTTTGTACAAGAAAGCTGGGTTgagatttttttttcgacggt agtg	vipA insertion in pDONR207 vector for peGFP-N-VipA
4_vipA rev	GGGGACCACTTTGTACAAGAAAGCTGGGTTgagatttttttttcgacggt agtg	
5_legK2 fwd	attggggaagcgggtaccgggtttattacataaattgaagga	legK2 insertion in pXDC61 vector in XmaI site
6_legK2 rev	tatctagaggatccccgggttttagcttgggcctcgcat	
7_vipA fwd	cattggggaagcgggtaccggcctatcagtaatgccttcttaa	vipA insertion in pXDC61 vector in XmaI site
8_vipA rev	tatctagaggatccccggctagagatttttttttcgacggta	
9_P1 vipA	TTCAACTCATCCTAGCATTG	vipA inactivation in <i>Legionella</i> by KanMazF technique
10_P2 vipA	CAGCAATATGATATTCTGGC	
11_P3 vipA	GGCCCAATTCGCCCTATAGTGAGTCGATGGATGGTCAAGCATTAT C	
12_P4 vipA	GGGTTTGCTCGGGTCGGTGGCATATGGTGCTCTATGATACTGACA TC	
13_P5 vipA	GATGTCAGTACATAGAGCACATGGATGGTCAAGCATTATC	
14_P6 vipA	GATAATGCTTGACCATCCATGTGCTCTATGATACTGACATC	legK2 inactivation in <i>Legionella</i> by KanMazF technique
19_P1 legK2	TTACCAATGTAATGAGACATCG	
20_P2 legK2	GGCCCAATTCGCCCTATAGTGAGTCGtcttgaggtagaggtgttcc	
21_P3 legK2	GGGTTTGCTCGGGTCGGTGGCATATGCTGCAAATCAAGATAAGCA ACC	
22_P4 legK2	ATAGCATGCACAACCTTTGATATCAAGCATCC	
23_P5 legK2	GGTTGCTTATCTTGATTTGCAGtcttgaggtagaggtgttcc	
24_P6 legK2	GGAACAACCTCTACCTCAAGACTGCAAATCAAGATAAGCAACC	
vipA linker REV	GCTGCCCCACCTCCGAGATTTTTTTTTTCGACGGTAGTG	Insertion of gene <i>luc</i> for translational fusion by kanMazF technique
luc REV	CAATTTGGGCTTTCCGCCCTTC	
pLLA01_TRAD FW	GGAGGTGGGGGCAGCAGATCCGCCAAAAAC ATAAAGAAAGG	
lpp0457 trad after luc	GAAGGGCGGAAAGCCCAAATTGTAGTTCAGCCAATTTCTGTTC	
legK2linker REV	GCTGCCCCACCTCCCTTGGGCCTCGCATCAATGAAG	
legK2 fusion luc after	GAAGGGCGGAAAGCCCAAATTGTAAAAAACGTCACTGAAGTGTT AG	Insertion of gene <i>sfGFP</i> for translational fusion by kanMazF technique
SD ATG lpp2076	CATAATGCTCTCTATAAGCC	
SD ATG lpp2076 SfGFP	GGCTTATAGGAGAGCATTATGTCTAAAGGTGAAGAACTGTTC	
Comp start lpp2076-end sfGFP	CTTCAAATTTATGTAATAAACTTTGTAGAGCTCATCCATGCC	
start lpp2076	GTTTATTACATAAATTTGAAG	
25_Fwd pGEX	GGGCCCCTGGAACAGAAC	Construction of pGEX-legK2 plasmid by PCR SLIC
26_Rev pGEX	CTGGGATCCCCGAATTCCC	
27_Fwd legK2 pGEX	GTTCTGTTCCAGGGGCCGttttattacataaattgaaggaac	
28_Rev legK2 pGEX	GGGAATTCGGGGATCCAGttagcttgggcctcgcatc	Construction of pQE30-vipA plasmid by BamHI digestion SLIC
31_Fwd vipA pQE30	GCTCGCATGCGGATCctagagatttttttttcgacggtag	
32_Rev vipA pQE30	TCACCATCACGGATCccctatcagtaatgccttcttaagtt	

#### SUPPLEMENTARY DATA S3

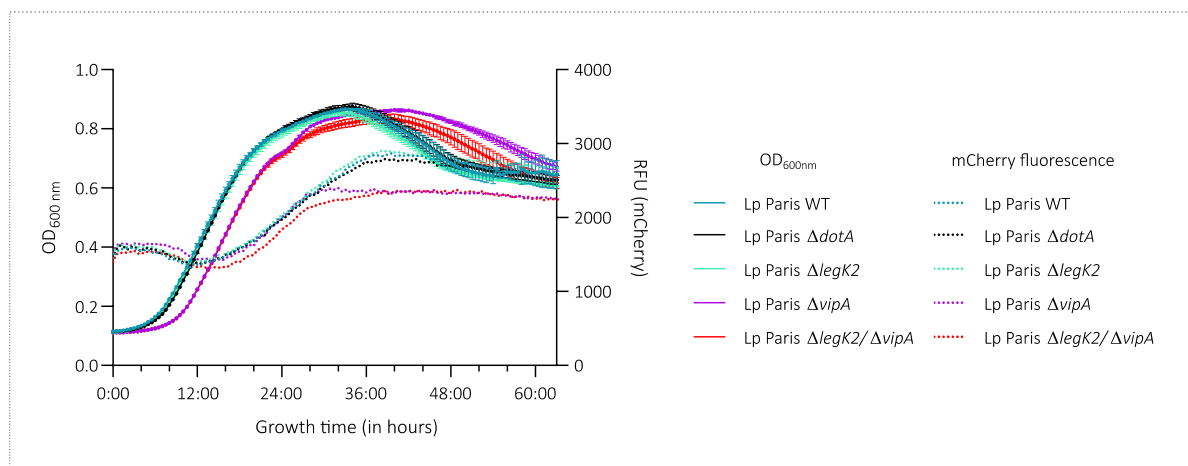
Accession #	Strain names	Accession #	Strain names
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GCF001467645.1	<i>L. geestiana</i> ATCC49504	GCF900187095.1	<i>L. waltersii</i> NCTC13017
GCF001468165.1	<i>L. spiritensis</i> Mt.St.Helens-9	GCF000770585.1	<i>L. norrlandica</i> LEGN
GCF900186965.1	<i>L. spiritensis</i> NCTC11990	GCF001582625.1	<i>L. pneumophila</i> FS_4_1103abu
GCF001467615.1	<i>L. erythra</i> SE-32A-C8	GCF000586195.1	<i>L. pneumophila</i> subsp. fraseri ATCC35251
GCF001468125.1	<i>L. rubrilucens</i> WA-270A-C2	GCF000586315.1	<i>L. pneumophila</i> subsp. fraseri ATCC33156
GCF001467505.1	<i>L. birminghamensis</i> CDC#1407-AL-14	GCF001639045.1	<i>L. pneumophila</i> PtVFX/2014
GCF001467975.1	<i>L. quinlivanii</i> CDC#1442-AUS-E	GCF001582535.1	<i>L. pneumophila</i> SZ069
GCF001467025.1	<i>L. brunensis</i> ATCC43878	GCF001886835.1	<i>L. pneumophila</i> subsp. fraseri Dallas 1E
GCF000953655.1	<i>L. hackeliae</i> ATCC35250	GCF001582295.1	<i>L. pneumophila</i> SZ099
GCF001467705.1	<i>L. hackeliae</i> 798-PA-H	GCF001886795.1	<i>L. pneumophila</i> subsp. fraseri Detroit-1
GCF002240035.1	<i>L. clemsonensis</i> CDC-D5610	GCF001583645.1	<i>L. pneumophila</i> SZ026
GCF001691475.1	<i>L. jamestowniensis</i> 974010_12	GCF001600905.1	<i>L. pneumophila</i> NY24
GCF001467745.1	<i>L. jamestowniensis</i> JA-26-G1-E2	GCF000586255.1	<i>L. pneumophila</i> subsp. pascullei ATCC33737
GCF900187355.1	<i>L. Lansingensis</i> NCTC12830	GCF001582645.1	<i>L. pneumophila</i> WD_4_1102b-36
GCF001467795.1	<i>L. Lansingensis</i> ATCC49751	GCF000953935.1	<i>L. pneumophila</i> Ymt294
GCF001467765.1	<i>L. jordanis</i> BL-540	GCF001582405.1	<i>L. pneumophila</i> FS_10_1101a-3
GCF001648675.1	<i>L. jordanis</i> ATCC33623	GCF000092625.1	<i>L. pneumophila</i> 2300/99 Alcoy
GCF000308315.1	<i>L. tunisiensis</i> LegM	GCF000092545.1	<i>L. pneumophila</i> str. Corby
GCF001467625.1	<i>L. feeleeii</i> WO-44C	GCF000823425.1	<i>L. pneumophila</i> 12_4117
GCF001648615.1	<i>L. feeleeii</i> ATCC35072	GCF000586095.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> ATCC43703
GCF000621525.1	<i>L. fairfieldensis</i> ATCC49588	GCF001582385.1	<i>L. pneumophila</i> SH135
GCF000756695.1	<i>L. massiliensis</i>	GCF900063795.1	<i>L. pneumophila</i> 2531STDY5467313
GCF000756815.1	<i>L. massiliensis</i> LegA	GCF000586075.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> ATCC43736
GCF001467585.1	<i>L. drozanskii</i> ATCC700990	GCF001592705.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> Toronto-2005
GCF001467895.1	<i>L. nautarum</i> ATCC49506	GCF900119765.1	<i>L. pneumophila</i> ST62
GCF900167045.1	<i>L. maceachernii</i> ATCC35300	GCF900053665.1	<i>L. pneumophila</i> 2531STDY5467288
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GCF001467055.1	<i>L. adelaidensis</i> 1762-AUS-E	GCF001583655.1	<i>L. pneumophila</i> SH003
GCF001467825.1	<i>L. londiniensis</i> ATCC 49505	GCF900060375.1	<i>L. pneumophila</i> 2531STDY5467394
GCF000512715.1	<i>L. oakridgensis</i> RV-2-2007	GCF000695015.1	<i>L. pneumophila</i> TUM 13948
GCF001648605.1	<i>L. oakridgensis</i> ATCC 33761	GCF001582555.1	<i>L. pneumophila</i> TL-12
GCF001467785.1	<i>L. israelensis</i> Bercovier 4	GCF000586235.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> ATCC33823
GCF000162755.2	<i>L. drancourtii</i> LLAP12	GCF000950745.1	<i>L. pneumophila</i> Twr292
GCF001465875.1	<i>L. saoudiensis</i> LH-SWC	GCF001582225.1	<i>L. pneumophila</i> ATCC35096
GCF001467695.1	<i>L. gratiana</i> Lyon 8420412	GCF000306865.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> Lorraine
GCF001467545.1	<i>L. cincinnatiensis</i> CDC#72-OH-14	GCF900050185.1	<i>L. pneumophila</i> 2531STDY5467306
GCF001468135.1	<i>L. santicrucis</i> SC-63-C7	GCF001582215.1	<i>L. pneumophila</i> ATCC43130

GCF000621685.1	<i>L. sainthelensi</i> ATCC35248	GCF001601075.1	<i>L. pneumophila</i> NY27
GCF001468105.1	<i>L. sainthelensi</i> Mt.St.Helens-4	GCF900062515.1	<i>L. pneumophila</i> 2531STDY5467417
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GCF000176095.1	<i>L. longbeachae</i> D-4968	GCF000586355.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> ATCC33154
GCF000701265.1	<i>L. wadsworthii</i> ATCC33877	GCF001582565.1	<i>L. pneumophila</i> ATCC33154
GCF001467945.1	<i>L. parisiensis</i> PF-209-C-C2	GCF900057235.1	<i>L. pneumophila</i> 2531STDY5467316
GCF001736145.1	<i>L. parisiensis</i> DSM 19216	GCF900092465.1	<i>L. pneumophila</i> Lpm7613
GCF001468035.1	<i>L. tucsonensis</i> ATCC49180	GCF001753125.1	<i>L. pneumophila</i> C11_O
GCF000333755.1	<i>L. anisa</i> Linanissette	GCF001752705.1	<i>L. pneumophila</i> E10_P
GCF002082905.1	<i>L. anisa</i> FDAARGOS_200	GCF001753265.1	<i>L. pneumophila</i> E2_N
GCF001468065.1	<i>L. steigerwaltii</i> SC-18-C9	GCF000008485.1	<i>L. pneumophila</i> subsp. <i>pneumophila</i> str. Philadelphia 1
GCF000621385.1	<i>L. cherrii</i> DSM 19213	GCF001601055.1	<i>L. pneumophila</i> NY17
GCF001467035.1	<i>L. cherrii</i> ORW	GCF900058585.1	<i>L. pneumophila</i> 2532STDY5467522
GCF001468005.1	<i>L. steelei</i> IMVS3376	GCF900062435.1	<i>L. pneumophila</i> 2532STDY5467504
GCF000953135.1	<i>L. fallonii</i> LLAP-10	GCF900062395.1	<i>L. pneumophila</i> 2532STDY5467530
GCF000373765.1	<i>L. shakespearei</i> DSM 23087	GCF000823645.1	<i>L. pneumophila</i> 12_4904
GCF001468025.1	<i>L. shakespearei</i> ATCC49655	GCF900055205.1	<i>L. pneumophila</i> 2532STDY5467502
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GCF000423305.1	<i>L. moravica</i> DSM 19234	GCF900053395.1	<i>L. pneumophila</i> 2532STDY5467482
GCF001467865.1	<i>L. moravica</i> ATCC43877	GCF000826165.1	<i>C. burnetii</i> Cb171_QLYMPHOMA
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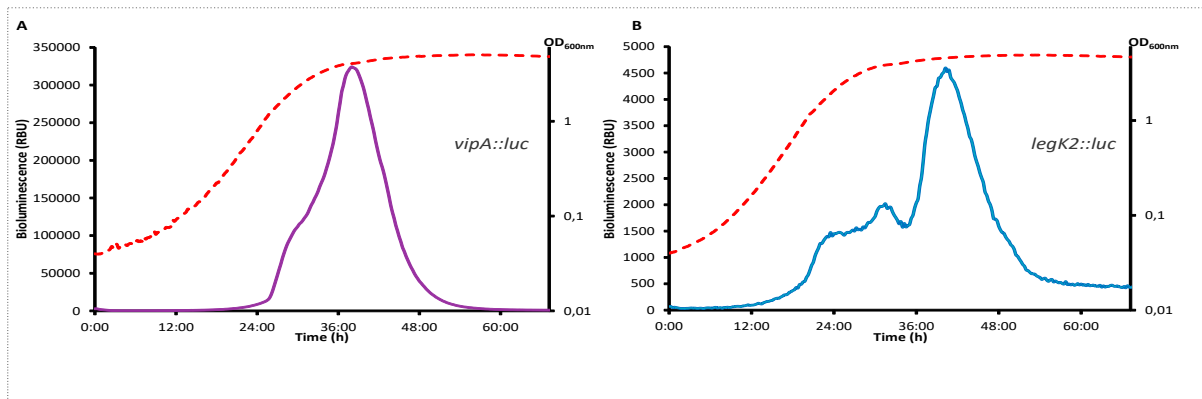
## SUPPLEMENTARY DATA S4

Fasta files of sequencing data of  $\Delta legK2$  and  $\Delta legK2/\Delta vipA$  *Legionella pneumophila* strains - RNA-seq and sequencing data have been deposited in European Nucleotide Archive database at EMBL-EBI (<https://www.ebi.ac.uk/ena>) under accession number PRJEB62121.

## Supplementary Data S5



## SUPPLEMENTARY DATA S6



## SUPPLEMENTARY DATA S7

■ LegK2

