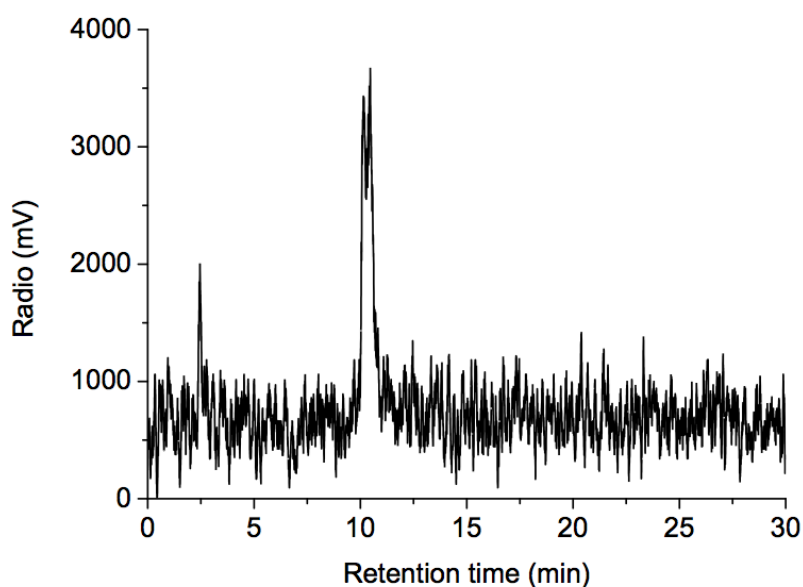
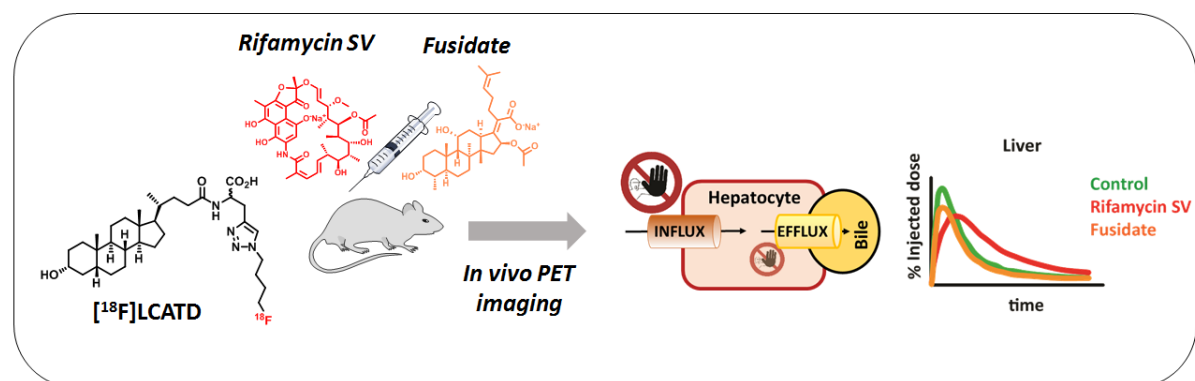


## SUPPORTING INFORMATION

### Graphical Abstract

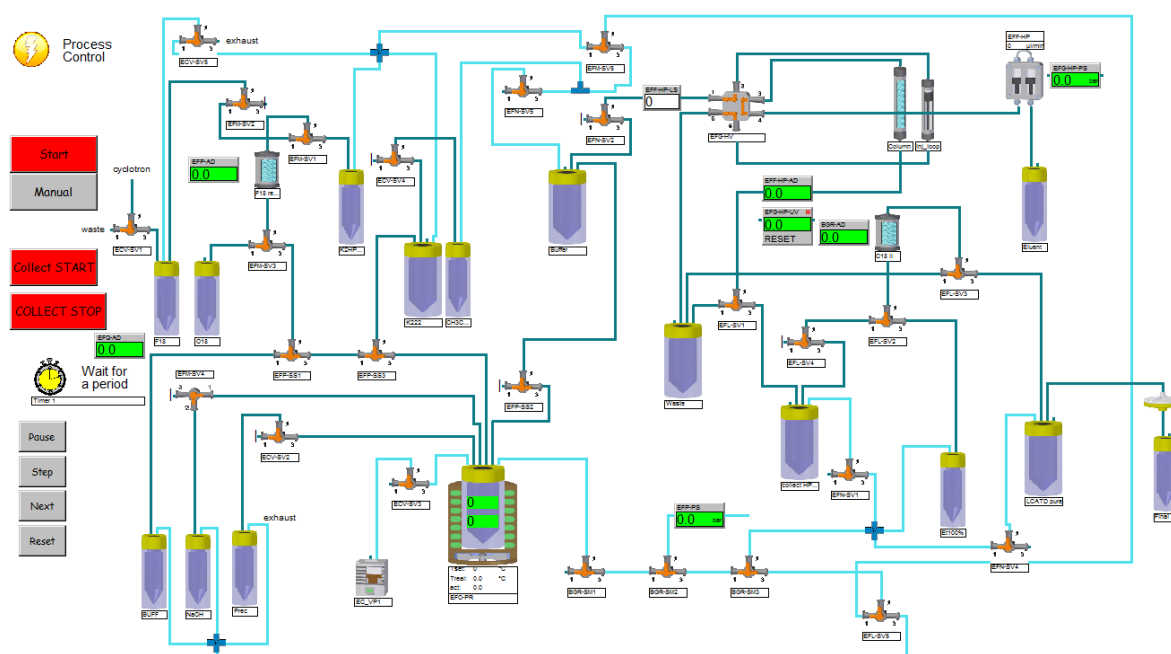


**Figure S1.** Radio- HPLC profile of the bile extract of a representative control rat. The peaks at 10.08 minutes correspond to the two  $[^{18}\text{F}]$ LCATD diastereoisomers.

### Radiosynthesis of $[^{18}\text{F}]$ LCATD

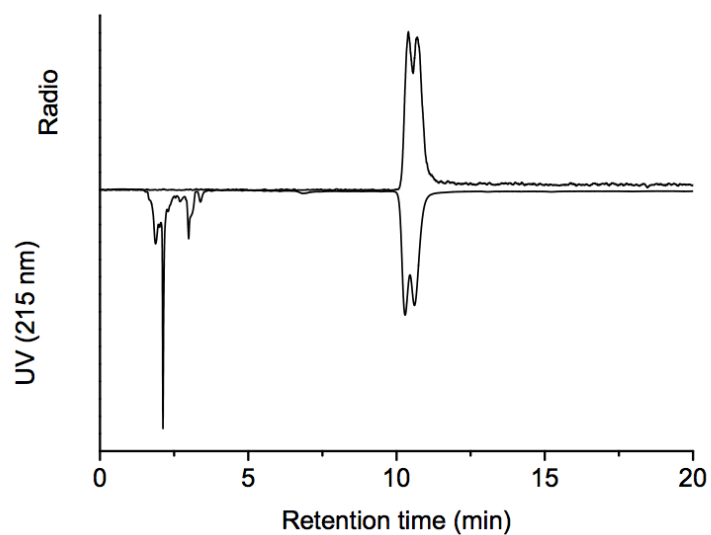
Radiosynthesis, purification and formulation steps were performed in an Eckert & Ziegler ModularLab system equipped with semipreparative HPLC.  $[^{18}\text{F}]$ Fluoride ( $\sim 16$  GBq) was delivered from the cyclotron into a stock vial and then separated from the  $[^{18}\text{O}]\text{H}_2\text{O}$  by means of a Chromafix PS- $\text{HCO}_3^-$  ion exchange cartridge (Waters).  $[^{18}\text{F}]$ Fluoride was then eluted with the solution of  $\text{K}_2\text{HPO}_4$  (5 mg in 0.5 mL of sterile water) and collected in the reactor, followed by the solution of Kryptofix K222 (15 mg in 1 mL of anhydrous  $\text{CH}_3\text{CN}$ ). A first drying cycle was accomplished by heating at  $95^\circ\text{C}$  under both vacuum and helium flow (400 s) and then under vacuum only (150 s) before delivering an additional amount of anhydrous  $\text{CH}_3\text{CN}$ . The drying process was repeated and the solution of mesylate precursor (prepared according to *Bioorg. Med. Chem.* **2017**, 25, 963-976)

(5 mg in 1 mL of anhydrous CH<sub>3</sub>CN) was then delivered into the reactor containing the [<sup>18</sup>F]KF-K222 dry complex. The mixture was heated at 90 °C for 15 min. The reactor was cooled to 40 °C before adding the sodium hydroxide solution (0.5 mL, 2 M in 50% aqueous methanol) which was allowed to react for 10 min. After the hydrolysis step, the crude reaction mixture was neutralized with 2M aqueous HCl (0.5 mL) and pumped into a vial containing the HPLC eluent (PBS/CH<sub>3</sub>CN 60:40, 1.5 mL). The mixture (about 3 mL) was then loaded in the 5 mL RP-HPLC loop and then injected in the Phenomenex Luna C18 column (5 µm, 250 × 10 mm, 100 Å), which was eluted with PBS/CH<sub>3</sub>CN 60:40 (5 mL min<sup>-1</sup> flow). The peak corresponding to the desired product was collected in a vial containing 50 mL of distilled water. The diluted solution of pure [<sup>18</sup>F]LCATD was then flushed through a C18 SPE cartridge Oasis® HLB Plus (Waters) in order to trap the tracer. The cartridge was then eluted with absolute ethanol (0.8 mL) and the resulting [<sup>18</sup>F]LCATD was passed through a sterile filter and finally delivered into a vial, placed in a second hot cell, containing 7.2 mL of sterile PBS. Up to 2 GBq of injectable solution of [<sup>18</sup>F]LCATD were obtained (8 mL, up to 250 MBq mL<sup>-1</sup>). The total synthesis, purification and formulation time was about 90 min. The tracer was obtained in 25 ± 5% (n = 5) decay corrected radiochemical yield. HPLC analysis of the formulated product confirmed the identity (co-injection with cold reference material) and the purity of the tracer (RCP > 99%).

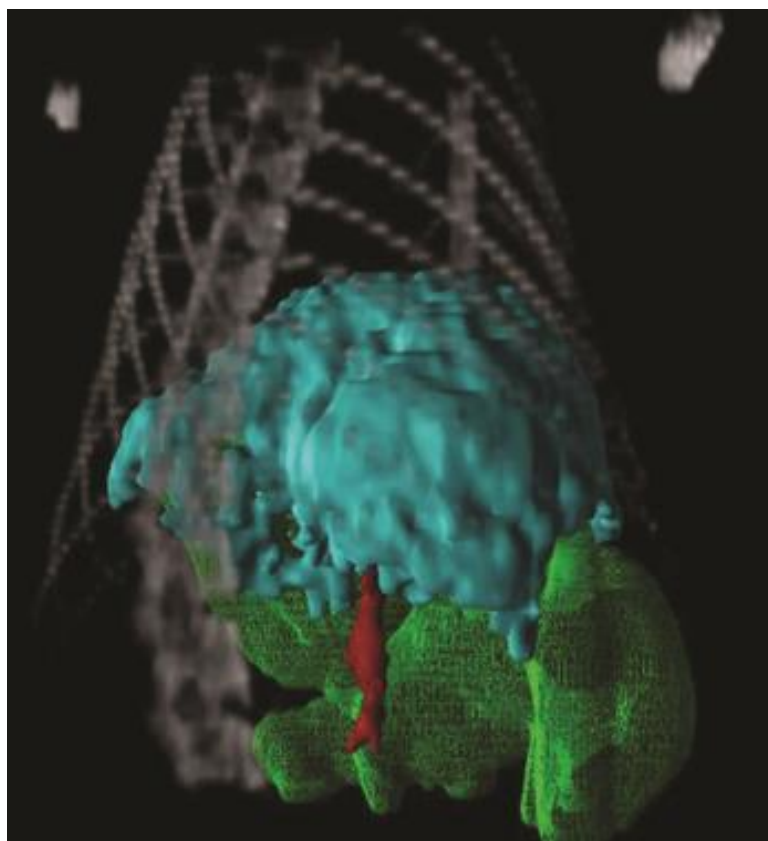


**Figure S2. Configuration of Eckert & Ziegler module for production of [<sup>18</sup>F]LCATD**

Co-injection of formulated [<sup>18</sup>F]**LACTD** and cold **LCATD** reference compound.



**Figure S3.** Co-injection of formulated [ $^{18}\text{F}$ ]LCATD and cold LCATD reference ( $t_R = 10.35$  and  $10.66$  min).



**FIGURE S4:** ROIs for the quantification of the activity in the blood (abdominal aorta, in red), liver (in cyan), and bile (gastrointestinal tract, green).

**Supplementary movie (see the attached file).** Rotating view of ROIs for the quantification of the activity in the blood (abdominal aorta, in red), liver (in cyan), and bile (gastrointestinal tract, green).