

Association of Epstein-Barr virus with human mammary carcinoma. Pros and cons

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The Epstein-Barr virus (EBV) is associated with the development of different malignancies. In the last few years, EBV has been detected in a subset of breast tumors. The EBV genome was detected by PCR and Southern-blot analysis and identification of the infected cells was determined using different *in situ* methods. EBV has detected more frequently in steroid hormone receptors negative tumors, in high histological SBR grade tumors and furthermore, the EBV genome was also observed in metastatic lymph nodes, along with EBV detection in the primary tumor. Opposing results are discussed.

1. Introduction

The Epstein-Barr virus (EBV) is associated with the development of different malignancies such as Burkitt's lymphoma, nasopharyngeal carcinoma (NPC) [11] and carcinomas in various organs [3,10,14]. In the last few years, EBV has been detected in a subset of breast tumors and reported in different studies [1,7,8]. To this aim, different approaches were used. The EBV genome was detected by PCR and Southern-blot analysis and identification of the infected cells was determined using various *in situ* techniques.

2. Investigation of the presence of the EBV genome by PCR in breast cancer, healthy tissue and lymph node metastasis

In three separate studies, PCR techniques allowed the detection of the virus genome in 20–51% of in-

vasive breast carcinoma of different histological types whereas no or only rare positive cases was obtained with healthy tissue [1,7,8]. These results suggest that EBV is restricted to the tumor.

However, a study performed with samples collected over a period of 30 years failed to detect the EBV genome in 34 breast carcinomas (including 16 medullary carcinoma) [4]. Negative results could be due to differences in the preparation of samples (fixation, storing) over such a long period of time.

Investigation of lymph nodes with metastasis suggested that EBV could already be present in the tumor cells prior to their migration [1].

3. EBV presence and breast cancer prognostic factors

We also observed a statistically significant relationship between EBV presence and several poor prognostic factors for breast carcinomas, such as, steroid hormone receptors negative, high histological SBR grade, and association with axillary node invasion [1]. Findings which suggest that the infection by EBV may be related to a high metastatic potential of the tumor.

4. Breast cancer and EBV: Identification of the infected cells

The direct detection of EBV products had to be addressed. *In situ* hybridization (ISH) with EBER-1 (Epstein-Barr virus encoded small RNA1) probes, a very sensitive method for the detection of EBV in infected cells, is widely used because of the high EBER RNA copy number. EBER-1 were identified on frozen sections, in a fraction of malignant cells in six different breast tumors [7] while in other studies using paraffin sections, EBER-1 transcripts could not be detected [1,2,5,8]. Technical problems (related to tissue fixation, probe penetration in breast tissue) may be responsible for the negative results. In addition, the

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regulation of transcription of EBERs remains poorly understood and their high expression in infected cells might not be universal. NPC which exhibits varying degrees of differentiation lacks EBERs expression in some areas [9]. Moreover Takeuchi et al. [12] did not observe any EBER-1 expression in some EBV-positive NPC cases. Contrary to the other EBV associated diseases, it appears that EBERs ISH is not the best method for identifying infected cells in breast cancers.

Immunohistochemical studies were performed on 60 invasive breast cancers collected and processed 11 to 20 years ago [2]. The authors did not detect the latent membrane protein 1 (LMP-1) although, in a previous study, [8] a distinct staining for LMP-1 was observed in scattered epithelial cells in several of the tumor sections examined. The fact that few cells were stained, the low sensitivity of immunochemistry and the epidemiologic difference of the samples analyzed could explain the divergence between these two results. The Epstein-Barr nuclear antigen 2 (EBNA-2) was not detected in the breast carcinoma samples [2]. This result is coherent with all *in vivo* studies which show that EBNA2 is not expressed in EBV associated tumors. This antigen is only expressed in lymphoproliferative disorders and lymphomas of immunodeficient patients [11].

EBNA1 is essential for the maintenance of the viral episome in infected cells and is constantly expressed in all EBV infections [11]. EBNA-1, has been detected in breast cancer tumor cells by immunohistochemistry [1, 6]. Monoclonal antibodies distinctly showed nuclear staining of many epithelial tumor cells while normal cells (including lymphocytes) were not labeled. The fact that only a fraction of tumor cells were found to be EBNA-1 positive in breast cancer could reflect low expression or low accessibility of the protein to staining in some cells. Alternatively, at this stage of the disease, the virus may have been lost in a fraction of those cells.

RT-PCR analysis of cDNAs encoding the EBNA1 protein could be detected in a series of EBV positive infiltrating breast cancers and not in EBV negative samples (Boualaga and Joab, unpublished results). EBNA-1 is able to induce malignancies in transgenic mice by a mechanism which is not yet understood [13]. The expression of EBNA-1 in breast tumors might be important in the transformation phenomenon.

5. Conclusion

The discrepancies between results need to be resolved since an association of EBV with breast cancer

have potential relevance to its early detection, treatment and even prevention. This implies the need for more studies. As positive results are more conclusive than negative ones, it would appear that, PCR of comparable efficiency should be performed on frozen material with an appropriate single copy gene of the cellular genome as a control. Optimized RT-PCR conditions should be used for detection of EBV transcripts. *In situ* methods have to be developed to confirm the already published work. The proportion of EBV-infected breast tumor cells, would have to be confirmed by sensitive techniques (ie *in situ* PCR). Real time PCR on microdissected tumors cells would be a valuable tool for the determination of the number of EBV genome copies per cell.

The question of EBV as an etiologic factor remains to be answered. However, it could still be a useful prognostic marker or provide molecular targets for therapy.

Note added in proof

While this review was in press, Fina et al. (Br J Cancer **84** (2001) 783–790) described the presence of EBV in a large subset of breast cancer. Moreover, EBER ISH was found to be positive in a fraction of tumor cells.

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References

- [1] M. Bonnet, J.M. Guinebretiere, E. Kremmer, V. Grunewald, E. Benhamou, G. Contesso and I. Joab, Detection of Epstein-Barr virus in invasive breast cancers, *J. Natl. Cancer Inst.* **91** (1999), 1376–1381.
- [2] J.S. Chu, C.C. Chen and K.J. Chang, In situ detection of Epstein-Barr virus in breast cancer, *Cancer Lett.* **124** (1998), 53–57.
- [3] I.W. Dimery, J.S. Lee, M. Blick, G. Pearson, G. Spitzer and W.K. Hong, Association of the Epstein-Barr virus with lymphoepithelioma of the thymus, *Cancer* **61** (1988), 2475–2480.
- [4] M.J. Gaffey, H.F. Frierson, S.E. Mills, J.C. Boyd, R.J. Zarbo, J.F. Simpson, L.K. Gross and L.M. Weiss, Identification of lymphocyte subpopulation and their significance, *Modern. Pathol* **6** (1993), 721–728.

- [5] S.L. Glaser, R.F. Ambinder, J.A. Digiuseppe, P.L. Hornross and J.L. Hsu, Absence of Epstein-Barr virus EBER-1 transcripts in an epidemiologically diverse group of breast cancers, *Int J Cancer* **75** (1998), 555–558.
- [6] F.A. Grasser, P.G. Murray, E. Kremmer, K. Klein, K. Remberger and W. Feiden et al., Monoclonal antibodies directed against the Epstein-Barr virus-encoded nuclear antigen 1 (EBNA1): Immunohistologic detection of EBNA1 in the malignant cells of Hodgkin's disease, *Blood* **84** (1994), 3792–3798.
- [7] L.G. Labrecque, D.M. Barnes, I.S. Fentiman and B.E. Griffin, Epstein-Barr virus in epithelial cell tumors: A breast cancer study, *Cancer Res* **55** (1995), 39–45.
- [8] Y.A. Luqmani and S. Shousha, Presence of Epstein-Barr virus in breast carcinoma, *Int J Oncol* **6** (1995), 899–903.
- [9] R. Pathmanathan, U. Prasad, G. Chandrika, R. Sadler, K. Flynn and N. Raabtraub, Undifferentiated, nonkeratinizing, and squamous cell carcinoma of the nasopharynx: Variants of Epstein-Barr virus-infected neoplasia, *Am J Pathol* **146** (1995), 1355–1367.
- [10] N. Raab-Traub, P. Rajadurai, K. Flynn and A.P. Lanier, Epstein-Barr virus infection in carcinoma of the salivary gland, *J Virol* **65** (1991), 7032–7036.
- [11] A.B. Rickinson and E. Kieff, Epstein-Barr virus, in: *Virology*, B.N. Fields, D.M. Knipe AND A.L. Howley Et, eds, Lippincott-Raven Press, Philadelphia, 1996, pp. 2397–2446.
- [12] H. Takeuchi, R. Kobayashi, M. Hasegawa and K. Hirai, Detection of latent Epstein-Barr virus (EBV) DNA in paraffin sections of nasopharyngeal carcinomas expressing no EBV-encoded small RNAs using in situ PCR, *Arch Virol* **142** (1997), 1743–1756.
- [13] J.B. Wilson, J.L. Bell and A.J. Levine, Expression of Epstein-Barr virus nuclear antigen-1 induces B cell neoplasia in transgenic mice, *EMBO J* **15** (1996), 3117–3126.
- [14] M.P. Wong, L.P. Chung, S.T. Yuen, S.Y. Leung, S.Y. Chan and E. Wang et al., In situ detection of Epstein-Barr virus in non small cell lung carcinomas, *J Pathol* **177** (1995), 233–240.



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