

# Common oxytocin receptor gene polymorphisms and the risk for preterm birth

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**Abstract.** Oxytocin is crucially involved in the onset and maintenance of labor. We investigated the association between oxytocin receptor gene polymorphisms and preterm birth. The presence of four common oxytocin receptor gene polymorphisms (rs2254298, rs53576, rs2228485 and rs237911) was evaluated in one hundred women with preterm birth and one hundred healthy women using restriction fragment length polymorphism genotyping. No association was found between the presence of any individual oxytocin receptor gene polymorphism and preterm birth. In haplotype analysis, the haplotype combination of rs2254298 A allele, rs2228485 C allele and rs237911 G allele was found to be significantly associated with an increased risk of preterm birth (OR = 3.2 [CI 1.04–9.8],  $p = 0.043$ ). In conclusion our findings suggest that a combination of three oxytocin receptor gene polymorphisms is associated with an increased risk for preterm birth. We propose further studies investigating the role of oxytocin receptor gene polymorphisms and preterm birth.

**Keywords:** Pregnancy, oxytocin receptor, single gene polymorphism, preterm birth

## 1. Introduction

Preterm birth (PTB) is a major health problem. It is the leading cause of perinatal mortality as well as neonatal and long-term morbidity in the industrialized world. In developed countries up to 11% of children are born preterm and several studies have reported increasing trends in PTB. Despite major progress in perinatal medicine throughout the last decades, PTB still accounts for 70% of neonatal deaths, up to 75% of neonatal morbidity and contributes to long-term neurocognitive deficits, pulmonary dysfunction and ophthalmologic disorders [29].

The mechanisms leading to PTB are complex and still not fully understood. Beside known risk factors, such as demographic characteristics, lifestyle and infection, genetic factors seem to play a crucial role in PTB [4,10,15,23,27]. One of the best predictors for preterm delivery is the women's history of a previous PTB [3]. Approximately 20% of women who deliver preterm subsequently have another PTB with the same partner; changing partners reduces the risk by one third [2,18]. Twin studies of pregnancy outcome estimate the heritability of PTB as 17% to 36% [4, 26]. Furthermore a large number of potential candidate genes associated with PTB and birth timing have been identified [22]. These studies have traditionally focused on one or a few candidate genes, chosen based on our current understanding of physiology. More recently, genome-wide linkage analyses in families with recurrent PTB and large-scale association studies have revealed further susceptibility genes for PTB [9,21,24].

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Oxytocin is crucially involved in the onset and maintenance of labor. It is a nine amino acid neuropeptide, which stimulates uterine contractions and lactation and modulates behavior and cognition [1,8,13,30]. The human oxytocin receptor (OXTR) is a polypeptide with seven transmembrane domains belonging to the class I G protein-coupled receptor family [8]. Oxytocin activates the OXTR, which is expressed in both the myometrium and the endometrium, as well as in diverse peripheral tissues and the central nervous system [8]. In the myometrium oxytocin activates phospholipase C, which increases the intracellular calcium concentration and thus intensifies uterine contractions. Mediated by activation of the OXTR, oxytocin additionally seems to assist cervical ripening by stimulating the synthesis of prostaglandins in the chorion, decidua, and amnion [7, 12].

The *OXTR* gene is located on chromosome 3p25.3. To date, several dozen single nucleotide polymorphisms (SNPs) have been identified within the *OXTR* gene. These SNPs are indicated in the National Center for Biotechnology Information database (dbSNP). However, little is known about their influence on human physiology. So far *OXTR* gene polymorphisms have mainly been investigated in a variety of psychiatric disorders and disease [13,19,30]. Additionally, although oxytocin is known to play a crucial role in parturition, until now, to the best of our knowledge, the association between *OXTR* gene polymorphisms and the risk of PTB has not been investigated. Therefore we evaluated the association of common *OXTR* single nucleotide polymorphisms (rs2254298, rs53576, rs2228485 and rs237911) and PTB.

## 2. Materials and methods

### 2.1. Patients

In the present retrospective case-control study 200 women with singleton pregnancies were included. Blood samples of 100 consecutive unrelated pregnant women who delivered preterm were included from the Biobank for normal and pathological pregnancies of the Department of Obstetrics and Gynecology, Medical University of Vienna General Hospital, Austria. This included women with spontaneous PTB due to preterm labor (29/100) and preterm premature rupture of membranes (PPROM) (71/100) but no cases where preterm delivery was indicated for medical reasons such as preeclampsia or fetal conditions such as intrauterine

growth restriction. Furthermore included cases did not demonstrate any known major fetal chromosomal and/or structural anomalies, significant maternal medical illness (e.g. chronic renal failure, congestive heart failure, connective tissue disorders) or other conditions such as polyhydramnios that increase the *a priori* risk for PTB. Preterm delivery was defined as delivery before 37 completed weeks of gestation. 100 women with at least one uncomplicated full term singleton pregnancy were asked to serve as controls. Gestational age was calculated from the first day of the last menstrual period and was confirmed by first-trimester ultrasound measurement of fetal crown-rump length. Clinical data were obtained from files at the Medical University of Vienna General Hospital. All patients were of Caucasian origin. Written informed consent was obtained from mothers before collection of biological materials. Approval was obtained by the institutional review board of the Medical University of Vienna (Reference EK Nr.385/2004 and EK Nr. 619/2006).

### 2.2. Genotyping

Genomic DNA was isolated from anticoagulated blood by using the QiAmp Blood Midi Kit, as described by the manufacturer (Quiagen, Hilden, Germany), and stored at  $-20^{\circ}\text{C}$ . The A/G polymorphism (rs2254298) in intron 3, the A/G polymorphism (rs53576) in intron 3, the C/T polymorphism (rs2228485) in exon 3 (amino acid change: N57N) and the A/G polymorphism (rs237911) in the 5'UTR of the *OXTR* gene were genotyped by using Restriction Fragment Length Polymorphisms.

The reaction was performed in a volume of 25  $\mu\text{l}$ , consisting of 12.5  $\mu\text{l}$  JumpStart<sup>TM</sup> REDTaq<sup>®</sup> ReadyMix<sup>TM</sup> PCR-Reaction Mix (Sigma, USA) and 40 ng of genomic DNA. The concentration of each primer was 20 pmol/ $\mu\text{l}$ . Cycle conditions were  $95^{\circ}\text{C}$  for 5 minutes; 32 cycles at  $95^{\circ}\text{C}$  for 30 seconds,  $60^{\circ}\text{C}$  for 40 seconds,  $72^{\circ}\text{C}$  for 40 seconds, and  $72^{\circ}\text{C}$  for 7 minutes (rs2254298); 5 minutes  $95^{\circ}\text{C}$ ; 32 cycles at  $95^{\circ}$  for 30 seconds,  $58^{\circ}\text{C}$  for 40 seconds,  $72^{\circ}\text{C}$  for 40 seconds, and  $72^{\circ}\text{C}$  for 7 minutes (rs53576); 5 minutes  $95^{\circ}\text{C}$ ; 31 cycles at  $95^{\circ}\text{C}$  for 30 seconds,  $65^{\circ}\text{C}$  for 40 seconds,  $72^{\circ}\text{C}$  for 40 seconds and finally 7 minutes at  $72^{\circ}\text{C}$  (rs2258485) and 5 minutes  $95^{\circ}\text{C}$ ; 32 cycles at  $95^{\circ}\text{C}$  for 30 seconds,  $63^{\circ}\text{C}$  for 40 seconds,  $72^{\circ}\text{C}$  for 40 seconds and 7 minutes at  $72^{\circ}\text{C}$  (rs237911). A 15  $\mu\text{l}$  of the PCR-product was digested over night using 5U of restriction enzyme. Fragments were separated by electrophoresis on an ethidium bromide-stained agarose gel (2%) and analyzed under UV light.

|           | Primer 5'-3'                                      | Product | RFLP   | Allele (bp)                |
|-----------|---|---------|--------|----------------------------|
| rs2254298 | TGAAAGCAGAGGTTGTGTGGACAGG<br>AACGCCACCCAGTTTCTTC  | 307bp   | BsrI   | A 9/163/34/101 G 9/163/135 |
| rs53576   | GCCCACCATGCTCTCCACATC<br>GCTGGACTCAGGAGGAATAGGGAC | 340bp   | BamH I | A 120/220 G 340            |
| rs2228485 | CTCATTGTCAGTGGCTCAGA<br>ATGAGCAGCAGCAGGTAGGT      | 340bp   | BsmI   | C 340 T 110/230            |
| rs237911  | CCCTTTACGGCTTGGCG<br>CCGCTCATTGTCAGTGGCTCAG       | 300bp   | AvaII  | A 300 G 100/200            |

Table 1  
Patients' characteristics

| Characteristics                     | Women with preterm birth    | full term pregnancy         | P-value  | OR (95%CI)      |
|-------------------------------------|-----------------------------|-----------------------------|----------|-----------------|
| Total number of patients            | 100                         | 100                         | —        | —               |
| Maternal age (years)                | 28.9 (6.4) <sup>2</sup>     | 28.9 (6.4) <sup>2</sup>     | 0.9      | —               |
| History of previous preterm birth   | 20                          | 0                           | < 0.0001 | 2.3 (1.9–2.6)   |
| Gestational age at delivery (weeks) | 29.5 (3.3) <sup>2</sup>     | 39.5 (1.1) <sup>2</sup>     | < 0.001  | —               |
| Birth weight (grams)                | 1356.3 (605.0) <sup>2</sup> | 3379.9 (465.7) <sup>2</sup> | < 0.001  | —               |
| First pregnancy                     | 32                          | 46                          | 0.06     | 0.5 (0.3–0.9)   |
| Delivery mode                       |                             |                             |          |                 |
| Caesarean section                   | 84                          | 30                          | < 0.0001 | 12.3 (6.2–24.3) |
| Spontaneous delivery                | 16                          | 70                          |          |                 |
| Antenatal bacterial vaginosis       | 9                           | 2                           | 0.06     | 4.8 (1.0–23.0)  |
| LLETZ conisation                    | 3                           | 1                           | 0.6      | 3.1 (0.3–29.9)  |
| Smoking                             | 11                          | 1                           | 0.005    | 12.2 (1.5–96.7) |

<sup>1</sup>Odds Ratio (95% Confidence Interval); <sup>2</sup>Mean (Standard deviation); <sup>3</sup>Student's t-test; <sup>4</sup>Chi-square test.

### 2.3. Statistical analysis

After testing for normality using Kolmogorov-Smirnov test, values are given as medians (interquartile range [IQR]) or means (standard deviation [SD]) where appropriate. Hardy-Weinberg Equilibrium was tested by chi square tests comparing expected and observed genotype frequencies. Groups were compared using Student's t-tests and chi-square tests where appropriate. Statistics have been performed accordingly. P-values and Odds Ratios (OR) with 95% Confidence Intervals (95% CI) were calculated major/major vs. major/minor and minor/minor allele, i.e. major versus presence of at least one minor allele. Haplotype frequencies were estimated by using SAS/Genetics software. A logistic regression model was estimated to assess the combined effect of rs2254298, rs2228485 and rs237911 haplotypes on the risk for PTB. The haplotype rs2254298 G allele – rs2228485 T allele – rs237911 A allele was set as reference haplotype. Haplotypes occurring with a frequency of four percent or less were pooled. P-values < 0.05 were considered statistically significant. They have not been corrected for multiple testing. Statistical software SPSS for Windows (SPSS 11.0, SPSS Inc., Chicago, IL) and SAS System (Version 9.1 SAS Institute Inc., Cary, NC) were used for statistical analysis. The linkage disequilibrium (LD) plot was generated with Haploview (Version 4.2 Broad Institute, Cambridge, MA).

### 3. Results

Patient characteristics are shown in Table 1. For rs53576, 100% prevalence of the GG genotype was found in both groups and therefore this gene polymorphism was not included in further statistical analysis. Distributions of the gene polymorphisms rs2254298 ( $p = 0.9$  and  $p = 0.5$ ), rs2228485 ( $p = 0.8$  and  $p = 1.0$ ) and rs237911 ( $p = 1.0$  and  $p = 0.9$ ) in women with PTB and controls were in Hardy-Weinberg equilibrium, respectively. A LD blot for cases is shown in Fig. 1. No significant association was found between the investigated OXTR genotypes and PTB in univariate analysis (Table 2). The main risk factors for PTB were history of PTB (OR 6.0[95% confidence interval (CI) 4.0–9.0]) and smoking (OR 12.2[95%CI 1.5–96.7]). Other well-known risk factors such as antenatal bacterial vaginosis (OR 4.8[95%CI 1.0–23.0]) and history of LLETZ (OR 3.1[95%CI 0.3–29.9]) were not associated with preterm birth in our sample (Table 1). Additionally, haplotype analysis was performed to identify risk combinations of the three OXTR gene polymorphisms and preterm birth. The haplotype combination rs2254298 A allele, rs2228485 C allele, and rs237911 G allele was found to be significantly associated with an increased risk of preterm birth (OR = 3.5 [CI 1.1–11.0],  $p = 0.03$ , Table 3).

Table 2  
Genotype and allele frequencies of *OXTR* gene polymorphisms in women with preterm birth and women with full term pregnancy

|                  | Women with preterm birth | Women with full term pregnancy | P-value           | OR (95% CI)                |
|------------------|--------------------------|--------------------------------|-------------------|----------------------------|
| <i>rs2254298</i> |                          |                                |                   |                            |
| G/G              | 70 (70.0%)               | 82 (82.0%)                     | 0.07 <sup>1</sup> | 2.0 (1.0–3.8) <sup>1</sup> |
| G/A              | 28 (28.0%)               | 15 (15.0%)                     |                   |                            |
| A/A              | 2 (2.0%)                 | 3 (3.0%)                       |                   |                            |
| <i>Alleles</i>   |                          |                                |                   |                            |
| G                | 168 (84.0%)              | 179 (89.5%)                    | 0.1               | 1.6 (0.9–2.9)              |
| A                | 32 (16.0%)               | 21 (10.5%)                     |                   |                            |
| <i>rs2228485</i> |                          |                                |                   |                            |
| T/T              | 54 (54.0%)               | 64 (64.6%)                     | 0.1 <sup>1</sup>  | 1.6 (0.9–2.8) <sup>1</sup> |
| T/C              | 41 (41.0%)               | 31 (31.3%)                     |                   |                            |
| C/C              | 5 (5.0%)                 | 4 (4.0%)                       |                   |                            |
| <i>Alleles</i>   |                          |                                |                   |                            |
| T                | 149 (74.5%)              | 159 (80.3%)                    | 0.2               | 1.4 (0.9–2.2)              |
| C                | 51 (25.5%)               | 39 (19.7%)                     |                   |                            |
| <i>rs237911</i>  |                          |                                |                   |                            |
| G/G              | 76 (76.0%)               | 79 (75.0%)                     | 0.7 <sup>1</sup>  | 1.2 (0.6–2.3) <sup>1</sup> |
| G/A              | 22 (22.0%)               | 18 (22.0%)                     |                   |                            |
| A/A              | 2 (2.0%)                 | 3 (3.0%)                       |                   |                            |
| <i>Alleles</i>   |                          |                                |                   |                            |
| G                | 174 (87.0%)              | 176 (88.0%)                    | 0.9               | 1.1 (0.6–2.0)              |
| A                | 26 (13.0%)               | 24 (12.0%)                     |                   |                            |

<sup>1</sup> P-value and Odds Ratio (95% Confidence Interval) were calculated major/major vs. major/minor and minor/minor allele.

Table 3

Haplotype analysis of the three *OXTR* gene polymorphisms in women with full term pregnancy (controls) and women with preterm birth (cases)

| Haplotypes (rs2254298-rs2228485-rs237911) | Cases (%) | Controls (%) | P-value | OR (95%CI)        |
|---|-----------|--------------|---------|-------------------|
| G-T-G (reference)                         | 65.7      | 70.8         |         |                   |
| A-C-G                                     | 7.0       | 2.4          | 0.03    | 3.5 (1.1–11.0)    |
| A-T-G                                     | 8.7       | 7.5          | 0.5     | 1.4 (0.6–3.3)     |
| G-C-A                                     | 12.7      | 9.8          | 0.1     | 1.7 (0.9–3.5)     |
| G-C-G                                     | 5.6       | 7.2          | 0.7     | 0.8 (0.4–1.9)     |
| others                                    | 0.3       | 2.3          | 0.2     | 0.1 (< 0.001–3.2) |

#### 4. Discussion

In the present study we evaluated the association between four common *OXTR* gene polymorphisms and risk of PTB. We were unable to demonstrate an association between any of four common individual *OXTR* gene polymorphisms and PTB. Nonetheless, the haplotype rs2254298 A allele- rs2228485 C allele- rs237911 G allele was found to be associated with an increased risk of PTB.

Oxytocin, exerting its effects via the *OXTR*, plays a crucial role in parturition. Around the onset of labor, uterine sensitivity to oxytocin markedly increases, which is associated with both an upregulation of *OXTR* mRNA levels and a strong increase in the density of myometrial oxytocin receptors [6,14]. Therefore in modern obstetric practice, oxytocin is widely used to induce and augment contractions during labor [28]. Furthermore, associations between several maternal gene polymorphisms, such as tumor necrosis factor, interleukin (IL)-4, IL-1 $\beta$  receptor antago-

nist, matrix metalloproteinase 9,  $\beta$ 2-adrenergic receptor, vascular endothelial growth factor and for Factor V Leiden [11] and PTB have been described. Consequently, we speculated that *OXTR* gene polymorphisms might also play a role in PTB and to the best of our knowledge are the first to have investigated association of *OXTR* single nucleotide polymorphisms rs2254298, rs53576, rs2228485, rs237911 and PTB.

Despite the crucial role of the *OXTR* in the onset and maintenance of labor, we were not able to ascertain a role of any individual of the investigated *OXTR* single nucleotide polymorphisms in PTB. This may partly be due to the fact that despite the fact that these *OXTR* gene polymorphisms have been investigated in a variety of diseases, their biological effects and exact influence on the oxytocin mediated pathways have not been clarified [13,19,30]. Thus, individual polymorphisms may not have any effect and influence on labor at all. Furthermore, the observed genotype distributions vary significantly between populations and thus possibly different causes for PTB. Therefore, we cannot rule out that

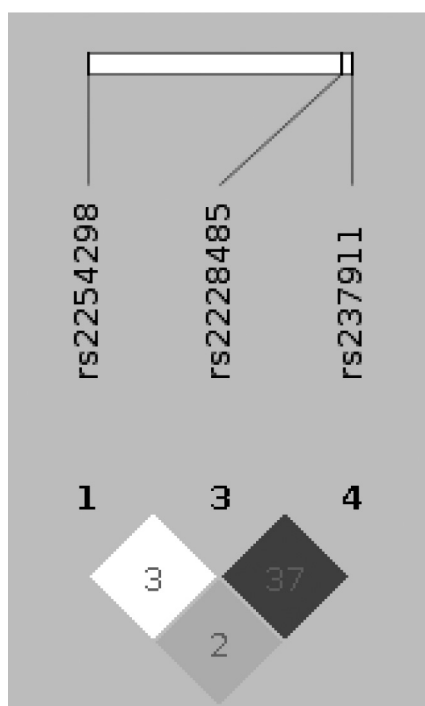


Fig. 1. Linkage disequilibrium (LD) plots of three *OXTR* gene polymorphisms in women with preterm birth. The names and relative positions of the polymorphisms are shown at the top. Pairwise  $D' \times 100$  values are shown in the squares. Darker colors indicate higher LD.

by studying a more homogenous population, such as Caucasian women with preterm labor only, a significant association with individual *OXTR* gene polymorphisms might be found.

Since presence of any individual *OXTR* gene polymorphism could not be associated with PTB we further sought to investigate their combined effect [4]. Thus, we performed haplotype analysis to identify possible *OXTR* risk combinations for an increased risk of PTB. The haplotype combination of rs2254298 A allele, rs2228485 C allele, and rs237911 G allele was significantly associated with an increased risk of preterm birth. This seems interesting, as SNPs rs2254298 and rs2228485 have been previously associated with positive and negative affect [13]. Consequently it has been hypothesized that human affect regulation may be mediated by Oxytocin via its effect on the hypothalamic-pituitary-adrenal axis. On a possible role of this pathway in the pathogenesis of PTB can still only be speculated since the underlying biological mechanisms are still not fully understood.

Our study has several limitations. First of all, it was not designed to investigate the association between *OX-*

*TR* gene polymorphism and Oxytocin serum levels or *OXTR* expression. Moreover, the included cases and controls reflect a highly selected group of patients as all have been recruited at a single tertiary care center. However, as expected, we found that maternal smoking and history of PTB were the main risk factors for PTB in cases. This is in accordance with the present literature [3,5,16,17,20,25]. Conversely, bacterial vaginosis and LLETZ conisation were not associated with PTB in our study, although they represent established risk factors for PTB. This might be attributed to the limited number of patients included in the present study. Additionally, genome-wide studies are far superior to studies such as ours looking at selected gene polymorphisms to gain a better etiologic and biologic understanding of the hereditary components of PTB [22]. Recently, genome-wide linkage analyses and association studies performed in Finnish families with recurrent PTB have revealed insulin-like growth factor 1 and follicle hormone stimulating hormone receptor genes as susceptibility genes for PTB [9,21]. Furthermore, as only maternal blood samples were available we did not take into account the possible role of fetal genes in our study. A large genetic association study of candidate genes involved in adverse pregnancy outcome has recently accentuated that both maternal and fetal DNA variants are associated with spontaneous preterm delivery [24]. Also, the ideal study genetic study of PTB would take the intricate interactions of environmental factors and genetic predispositions into account. However, given the current methodology and knowledge of biology, the ideal genetic study of PTB may not yet be possible.

To conclude, oxytocin is a crucial factor in parturition and PTB seems to be strongly affected by hereditary factors. To the best of our knowledge, this is the first study to investigate the association between four common *OXTR* gene polymorphisms and the risk of PTB in Caucasian women. We were not able to ascertain an association between any individual investigated *OXTR* gene polymorphisms and PTB. However we found that the haplotype combination of rs2254298 A allele, mutant rs2228485 C allele, and rs237911 G allele was significantly associated with an increased risk of PTB. We propose further studies investigating the role of *OXTR* gene polymorphisms and PTB.

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