

Research Article

miR-34b/c rs4938723 T>C Decreases Neuroblastoma Risk: A Replication Study in the Hunan Children

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Received 3 May 2019; Revised 30 July 2019; Accepted 13 August 2019; Published 10 September 2019

Academic Editor: Kishore Chaudhry

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Neuroblastoma is the most common seen solid neural tumor in children less than age one. As mutation in the *miR-34b/c* gene is observed in several types of human malignancies, there likely to be similar events that contribute to the pathogenesis of neuroblastoma. We hypothesize that polymorphism in the *miR-34b/c* gene might predispose to neuroblastoma. Here, we conducted this replication study by genotyping rs4938723 T>C from *miR-34b/c* in Hunan children (162 subjects with neuroblastoma and 270 control subjects) and examined its effect on the risk of neuroblastoma. We determined such association using logistic regression, adjusted for age and gender. Relative to those with TT genotype, subjects with C allele had reduced neuroblastoma risk (TC vs. TT: adjusted OR = 0.46, 95%CI = 0.30-0.71; additive model: adjusted OR = 0.64, 95%CI = 0.47-0.88; TC/CC vs. TT: adjusted OR = 0.49, 95%CI = 0.33-0.73). Stratified analysis revealed that rs4938723 TC/CC carriers were less likely to develop neuroblastoma for patients in the subgroups of age ≤ 18 months, age > 18 months, females, males, tumors in retroperitoneal, tumors in other sites, and clinical stages II, III, IV, and III+IV. Our findings verified *miR-34b/c* rs4938723 C variant allele as a protective factor for the risk of neuroblastoma. Further investigation of how *miR-34b/c* rs4938723 T>C might modify neuroblastoma risk is warranted.

1. Introduction

Neuroblastoma is a childhood tumor that mainly derives from neural crest progenitor cells [1–3]. Despite representing about 8–10% of all pediatric cancer diagnoses, neuroblastoma disproportionately results in 12–15% of all childhood cancer-related mortality [4–6]. It is characterized by widely clinical heterogeneity, spans from spontaneous regression to therapy-refractory progression [7]. Another reflection of such heterogeneity was the contrasting survival rate of different subgroup patients [8, 9]. In patients with the low- and intermediate-risk neuroblastoma, the long-term survival rate is greater than 90% [10]. However, in patients with the high-risk neuroblastoma, less than 40% could finally survive [11, 12].

In the past decades, considerable progress has been made in understanding the genetic underpinnings of neuroblastoma. Exposed environmental factors of children and pregnant women were reported to predispose to neuroblastoma, but not finally defined [13, 14]. Mutations in *ALK* [15] and *PHOX2B* [16] were considered as two major causes of familial neuroblastoma. Other SNPs in genes including *LMO1* [17], *BARD1* [18], *TP53* [19], *LIN28B* [20], *HACE1* [20], *NEFL* [21], and *CDKN1B* [22] have more recently been identified to be associated with neuroblastoma predisposition. Moreover, the association of these SNPs to neuroblastoma risk has also been replicated in many other populations, especially the SNPs in the *BARD1* gene [23–25]. Taken together, however, all the current identified mutations still could not fully elucidate the etiology of neuroblastoma. We are still

on the way to fully reveal the genetic landscape of neuroblastoma. Identification of other somatic mutations will further clarify the mechanisms of neuroblastoma.

MicroRNAs (miRNAs) are a class of nonprotein-coding, small, single-stranded RNAs with about 22 nucleotides [26]. miRNAs participate in transcriptional regulation through multiple mechanisms, including mRNA degradation, translational repression, or cleavage of mRNA [26–28]. In the past decade, more and more miRNAs are being identified that play vital regulatory roles in human disorders, including cancers. Mutations or single nucleotide polymorphisms (SNPs) in miRNA genes may alter the binding ability of miRNAs to their target mRNAs, thus resulting in diverse functional consequences and thereby possibly impact cancer susceptibility [29, 30]. rs4938723 T>C is located at the promoter region of *pri-miR-34b/c* [31]. Such T to C shift polymorphism might cause a disruption of GATA-X transcription factor binding capacity, which results in decreased *pri-miR-34b/c* expression [32]. Thus far, most studies have addressed the identification of *miR-34b/c* rs4938723 T>C in breast cancer [33], colorectal cancer [34], hepatocellular cancer [35], and nasopharyngeal carcinoma [36], whereas few studies focused on the role of *miR-34b/c* gene rs4938723 T>C in neuroblastoma risk. In our previous study conducted recently, we firstly found that rs4938723 T>C polymorphism was associated with a significantly decreased neuroblastoma risk [37]. Here, we further conducted a replication hospital-based case-control study aiming to verify the association between *miR-34b/c* rs4938723 T>C and neuroblastoma risk in Hunan children.

2. Materials and Methods

2.1. Study Subjects. Prior to analysis, the study protocols were approved by the Institutional Review Board of Hunan Children's Hospital. The current case-control study was carried out in Hunan Children's Hospital. A total of 162 cases were pathology-confirmed with neuroblastoma, and 270 controls with no prior history of neuroblastoma were randomly enrolled in the same area as cases. All guardians of participants provided written informed consent. The detailed information of selection criteria of study subjects was reported in our previous paper [38–40].

2.2. Genotyping. Genomic DNA was isolated from venous blood using a TIANamp Blood DNA Kit (TianGen Biotech Co. Ltd., Beijing, China). Genotype analysis of *miR-34b/c* gene rs4938723 T>C was undertaken using TaqMan SNP genotyping assay from Applied Biosystems [41–44]. Negative controls (with water) and duplicate test samples (10% of all the samples) were included in each 384-well plate. 100% concordant of genotypes in replicates were achieved.

2.3. Statistical Analysis. Tests for the Hardy-Weinberg equilibrium (HWE) were conducted for *miR-34b/c* rs4938723 T>C among control subjects with the use of the χ^2 test. Differences in demographic variables between case subjects and control subjects were analyzed using the two-sided χ^2 test. Neuroblastoma risk was determined as odds ratios (ORs)

and 95% confidence intervals (CIs), based on unconditional logistic regression adjusted for age and gender. A *P* value of <0.05 was used for statistical significance. The SAS release 9.1 (SAS Institute, Cary, NC) was used for statistical analyses.

3. Results

3.1. Association between *miR-34b/c* rs4938723 T>C and Neuroblastoma Susceptibility. A description of the demographic characteristics is provided in Supplemental Table 1. As shown in Table 1, genotype distributions of *miR-34b/c* rs4938723 T>C were compared between all cases and controls. The genotype for *miR-34b/c* rs4938723 T>C was in agreement with the HWE (HWE = 0.784) in the controls. Statistical analysis indicated that rs4938723 C variant allele was associated with decreased neuroblastoma risk (TC vs. TT: adjusted OR = 0.46, 95%CI = 0.30-0.71; additive model: adjusted OR = 0.64, 95%CI = 0.47-0.88; TC/CC vs. TT: adjusted OR = 0.49, 95%CI = 0.33-0.73).

3.2. Stratification Analysis. We further demonstrated whether the association between rs4938723 T>C genotype and neuroblastoma risk was modified by age, gender, tumor sites, and INSS stages (Table 2). We observed a significantly decreased risk of neuroblastoma for carriers of TC/CC genotype comparing with carriers of TT genotype in the subgroups of age \leq 18 months (adjusted OR = 0.35, 95%CI = 0.18-0.67), age > 18 months (adjusted OR = 0.55, 95%CI = 0.32-0.94), females (adjusted OR = 0.45, 95%CI = 0.24-0.82), and males (adjusted OR = 0.52, 95%CI = 0.29-0.90). Regarding sites of tumor origin, carriers of TC/CC genotype were less likely to have tumors in retroperitoneal (adjusted OR = 0.35, 95%CI = 0.20-0.60) and other sites (adjusted OR = 0.32, 95%CI = 0.11-0.94). We also found that the decreased risk of neuroblastoma associated with rs4938723 TC/CC genotypes was more pronounced among clinical stages II (adjusted OR = 0.35, 95%CI = 0.14-0.89), III (adjusted OR = 0.45, 95%CI = 0.25-0.83), IV (adjusted OR = 0.26, 95%CI = 0.12-0.60), and III+IV (adjusted OR = 0.38, 95%CI = 0.23-0.63).

4. Discussion

In our present study, by examining the relationship between *miR-34b/c* rs4938723 T>C and neuroblastoma susceptibility, we identified *miR-34b/c* rs4938723 C allele to be significantly associated with decreased neuroblastoma susceptibility in Hunan children. Our finding, for the first time, implies that *miR-34b/c* rs4938723 T>C protects Chinese children from neuroblastoma risk in Hunan subjects.

miR-34b and *miR-34c* are submembers of the *miR-34* family which share a common primary transcript (*pri-miR-34b/c*) [45]. The *miR-34b/c* is located in human chromosome 11 [46, 47]. The biological role of *miR-34b/c* has been well documented in several types of cancers. Majid et al. [48] found that *miR-34b* inhibits prostate cancer through demethylation, active chromatin modifications, and AKT pathways. It is documented that *miR-34b/c* can target *TP53* and cooperate to suppress cell proliferation and adhesion-independent

TABLE 1: *miR34b/c* rs4938723 T>C polymorphism and neuroblastoma susceptibility.

Genotype	Cases (N = 162)	Controls (N = 270)	P ^a	Crude OR (95% CI)	P	Adjusted OR (95% CI) ^b	P ^b
rs4938723 T>C (HWE = 0.784)							
TT	100 (61.73)	117 (43.33)		1.00		1.00	
TC	47 (29.01)	123 (45.56)		0.45 (0.29-0.69)	0.0002	0.46 (0.30-0.71)	0.0004
CC	15 (9.26)	30 (11.11)		0.59 (0.30-1.15)	0.119	0.62 (0.32-1.23)	0.175
Additive			0.0008	0.62 (0.46-0.85)	0.0026	0.64 (0.47-0.88)	0.005
Dominant	62 (38.27)	153 (56.67)	0.0002	0.47 (0.32-0.71)	0.0002	0.49 (0.33-0.73)	0.0005
Recessive	147 (90.74)	240 (88.89)	0.542	0.82 (0.43-1.57)	0.542	0.86 (0.45-1.67)	0.665

OR: odds ratio; CI: confidence interval. ^a χ^2 test for genotype distributions between neuroblastoma cases and cancer-free controls. ^bAdjusted for age and gender.

TABLE 2: Association between *miR34b/c* rs4938723 T>C polymorphism and clinical parameters.

Variables	Cases/controls		OR (95% CI)	P	AOR (95% CI) ^a	P ^a
	TT No. (%)	TC/CC No. (%)				
Age (month)						
≤18	42/36 (25.9)/(13.3)	27/66 (16.7)/(24.4)	0.35 (0.19-0.66)	0.001	0.35 (0.18-0.67)	0.002
>18	58/81 (35.8)/(30.0)	35/87 (21.6)/(32.2)	0.56 (0.34-0.94)	0.029	0.55 (0.32-0.94)	0.027
Gender						
Females	49/52 (30.2)/(19.2)	30/77 (18.5)/(28.5)	0.41 (0.23-0.74)	0.003	0.45 (0.24-0.82)	0.010
Males	51/65 (31.5)/(24.1)	32/76 (19.7)/(28.1)	0.54 (0.31-0.93)	0.027	0.52 (0.29-0.90)	0.020
Sites of origin						
Adrenal gland	17/117 (10.5)/(43.3)	14/153 (8.64)/(56.7)	0.63 (0.30-1.33)	0.225	0.65 (0.31-1.38)	0.265
Retroperitoneal	55/117 (33.9)/(43.3)	23/153 (14.2)/(56.7)	0.32 (0.19-0.55)	<0.0001	0.35 (0.20-0.60)	0.0002
Mediastinum	16/117 (9.87)/(43.3)	20/153 (12.3)/(56.7)	0.96 (0.48-1.93)	0.900	0.96 (0.48-1.95)	0.917
Others	12/117 (7.41)/(43.3)	5/153 (3.08)/(56.7)	0.32 (0.11-0.93)	0.036	0.32 (0.11-0.94)	0.039
Clinical stages						
I	23/117 (14.2)/(43.3)	25/153 (15.4)/(56.7)	0.83 (0.45-1.54)	0.556	0.86 (0.46-1.61)	0.645
II	15/117 (9.26)/(43.3)	7/153 (4.32)/(56.7)	0.36 (0.14-0.90)	0.030	0.35 (0.14-0.89)	0.027
III	34/117 (21.0)/(43.3)	20/153 (12.3)/(56.7)	0.45 (0.25-0.82)	0.009	0.45 (0.25-0.83)	0.010
IV	28/117 (17.3)/(43.3)	9/153 (5.55)/(56.7)	0.25 (0.11-0.54)	0.0005	0.26 (0.12-0.60)	0.001
4s	0/117 (0.00)/(43.3)	1/153 (0.006)/(56.7)	—	—	—	—
I+II+4s	38/117 (23.4)/(43.3)	32/153 (19.7)/(56.7)	0.64 (0.38-1.09)	0.103	0.65 (0.38-1.10)	0.110
III+IV	62/117 (38.3)/(43.3)	29/153 (17.9)/(56.7)	0.36 (0.22-0.59)	<0.0001	0.38 (0.23-0.63)	0.0002

OR: odds ratio; CI: confidence interval; AOR: adjusted odds ratio. ^aAdjusted for age and gender, omitting the corresponding stratify factor.

growth [49]. Findings from Wong et al. [50] offer a new insight into the tumor suppressor role of *miR-34b/c* in myeloma. However, there still lacks research regarding the role of *miR-34b/c* in neuroblastoma. The rs4938723 T>C polymorphism, locates within the CpG island of *pri-miR-34b/c*, was intensively investigated. In a study conducted in a Chinese population by Liu et al. [51], they found that CC and TC+CC genotypes of *pri-miR-34b/c* rs4938723 contribute to a higher susceptibility of hepatocellular carcinoma when compared with the TT genotype, respectively. Hashemi et al. [52] found that *miR-34b/c* rs4938723 C allele was correlated with a decreased risk of acute lymphoblastic leukemia, in a sample of an Iranian population with 110 children with acute lymphoblastic leukemia and 120 healthy children. However, Zhu et al. [53] failed to obtain a relationship between *miR-34b/c* rs4938723 and esophageal squamous cell

carcinoma risk, in 248 Kazakh patients with esophageal squamous cell carcinoma and 300 frequency-matched control subjects. Polymorphisms may exert distinct genetic effects on the susceptibility of cancer, depending on different cancer types, ethnicities, and regions.

In 2017, we performed a first case-control study regarding *miR-34b/c* rs4938723 T>C and neuroblastoma susceptibility in Chinese children, including 393 cases and 812 controls [37]. We firstly provided an evidence that *miR-34b/c* rs4938723 T>C displayed a protective role from neuroblastoma. However, such evidence needs further validation. Herein, we further verified the protective role of *miR-34b/c* rs4938723 T>C in neuroblastoma risk in another sample of Chinese. Such protective role could also be seen in other cancer types, such as colorectal cancer [34], gastric cancer [54], and esophageal cancer [55]. Other genetic, environmental

factors, and gene-environment interaction may cooperatively determine the protective role of *miR-34b/c* rs4938723 T>C in neuroblastoma risk [56, 57].

Strengths of the current study also accompany some limitations. First, statistic power may be compromised as the sample size is not large enough. Second, as a hospital-based case-control study, inclusion of the nonrepresentative subjects in this study may result in inherent selection bias. Third, conclusions obtained here lack generalizability as subjects are all genetic Chinese descent. Therefore, cautions should be taken if the current conclusion is extrapolated to other populations. Fourth, the selected SNP was based on prior knowledge of potentially functional SNPs. Other important tagging SNPs within the *miR-34b/c* gene may be omitted. Last, environment factors and gene-environment interactions could not be assessed in the current study, with the absence of environmental data.

5. Conclusions

In conclusion, here, we provided the possibility of *miR-34b/c* rs4938723 T>C in predicting neuroblastoma risk. Our study serves as a basis for future replication studies in independent populations or for functional studies of *miR-34b/c* rs4938723 T>C in neuroblastoma risk.

Abbreviations

miRNA: MicroRNA
 SNP: Single nucleotide polymorphism
 HWE: Hardy-Weinberg equilibrium
 OR: Odds ratio
 CI: Confidence interval.

Data Availability

The data used to support the findings of this study are available from the corresponding authors upon request.

Conflicts of Interest

The authors declared that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Authors' Contributions

All authors contributed significantly to this work. YL, JL, ZX, YX, JH, and ZL performed the research study and collected the data; JH analyzed the data; JH and ZL designed the research study; YL, ZJZ, and HZ wrote the paper; JH prepared all the tables. All authors reviewed the manuscript. In addition, all authors have read and approved the manuscript.

Acknowledgments

This study was supported by grants from the Pearl River S and T Nova Program of Guangzhou (No: 201710010086), the Hunan Provincial Natural Science Foundation Project

(No: 2018JJ2210), and the Hunan Provincial Key Laboratory of Pediatric Emergency Medicine (No: 2018TP1028).

Supplementary Materials

Supplemental Table 1: frequency distribution of selected characteristics in neuroblastoma cases and cancer-free controls for Hunan children. (*Supplementary Materials*)

References

- [1] M. Capasso and S. J. Diskin, "Genetics and genomics of neuroblastoma," *Cancer Treatment and Research*, vol. 155, pp. 65–84, 2010.
- [2] M. Schwab, F. Westermann, B. Hero, and F. Berthold, "Neuroblastoma: biology and molecular and chromosomal pathology," *The Lancet Oncology*, vol. 4, no. 8, pp. 472–480, 2003.
- [3] K. Durinck and F. Speleman, "Epigenetic regulation of neuroblastoma development," *Cell and Tissue Research*, vol. 372, no. 2, pp. 309–324, 2018.
- [4] N.-K. V. Cheung and M. A. Dyer, "Neuroblastoma: developmental biology, cancer genomics and immunotherapy," *Nature Reviews. Cancer*, vol. 13, no. 6, pp. 397–411, 2013.
- [5] M. R. Esposito, S. Aveic, A. Seydel, and G. P. Tonini, "Neuroblastoma treatment in the post-genomic era," *Journal of Biomedical Science*, vol. 24, no. 1, p. 14, 2017.
- [6] M. S. Irwin and J. R. Park, "Neuroblastoma: paradigm for precision medicine," *Pediatric Clinics of North America*, vol. 62, no. 1, pp. 225–256, 2015.
- [7] J. M. Maris, M. D. Hogarty, R. Bagatell, and S. L. Cohn, "Neuroblastoma," *The Lancet*, vol. 369, no. 9579, pp. 2106–2120, 2007.
- [8] K. K. Matthay, J. M. Maris, G. Schleiermacher et al., "Neuroblastoma," *Nature Reviews. Disease Primers*, vol. 2, no. 1, article 16078, 2016.
- [9] F. Westermann and M. Schwab, "Genetic parameters of neuroblastomas," *Cancer Letters*, vol. 184, no. 2, pp. 127–147, 2002.
- [10] G. Schleiermacher, V. Mosseri, W. B. London et al., "Segmental chromosomal alterations have prognostic impact in neuroblastoma: a report from the INRG project," *British Journal of Cancer*, vol. 107, no. 8, pp. 1418–1422, 2012.
- [11] J. M. Maris, "Recent advances in neuroblastoma," *The New England Journal of Medicine*, vol. 362, no. 23, pp. 2202–2211, 2010.
- [12] A. T. Look, F. A. Hayes, J. J. Shuster et al., "Clinical relevance of tumor cell ploidy and N-myc gene amplification in childhood neuroblastoma: a Pediatric Oncology Group study," *Journal of Clinical Oncology*, vol. 9, no. 4, pp. 581–591, 1991.
- [13] M. N. Cook, A. F. Olshan, H. A. Guess et al., "Maternal medication use and neuroblastoma in offspring," *American Journal of Epidemiology*, vol. 159, no. 8, pp. 721–731, 2004.
- [14] F. Menegaux, A. F. Olshan, J. P. Neglia, B. H. Pollock, and M. L. Bondy, "Day care, childhood infections, and risk of neuroblastoma," *American Journal of Epidemiology*, vol. 159, no. 9, pp. 843–851, 2004.
- [15] S. Ogawa, J. Takita, M. Sanada, and Y. Hayashi, "Oncogenic mutations of ALK in neuroblastoma," *Cancer Science*, vol. 102, no. 2, pp. 302–308, 2011.
- [16] D. Trochet, F. Bourdeaut, I. Janoueix-Lerosey et al., "Germline mutations of the paired-like homeobox 2B (PHOX2B) gene in

- neuroblastoma,” *American Journal of Human Genetics*, vol. 74, no. 4, pp. 761–764, 2004.
- [17] D. A. Oldridge, A. C. Wood, N. Weichert-Leahey et al., “Genetic predisposition to neuroblastoma mediated by a LMO1 super-enhancer polymorphism,” *Nature*, vol. 528, no. 7582, pp. 418–421, 2015.
- [18] M. Capasso, M. Devoto, C. Hou et al., “Common variations in BARD1 influence susceptibility to high-risk neuroblastoma,” *Nature Genetics*, vol. 41, no. 6, pp. 718–723, 2009.
- [19] S. J. Diskin, M. Capasso, M. Diamond et al., “Rare variants in TP53 and susceptibility to neuroblastoma,” *JNCI: Journal of the National Cancer Institute*, vol. 106, no. 4, article dju047, 2014.
- [20] S. J. Diskin, M. Capasso, R. W. Schnepf et al., “Common variation at 6q16 within HACE1 and LIN28B influences susceptibility to neuroblastoma,” *Nature Genetics*, vol. 44, no. 10, pp. 1126–1130, 2012.
- [21] M. Capasso, S. Diskin, F. Cimmino et al., “Common genetic variants in NEFL influence gene expression and neuroblastoma risk,” *Cancer Research*, vol. 74, no. 23, pp. 6913–6924, 2014.
- [22] M. Capasso, L. D. McDaniel, F. Cimmino et al., “The functional variant rs34330 of CDKN1B is associated with risk of neuroblastoma,” *Journal of Cellular and Molecular Medicine*, vol. 21, no. 12, pp. 3224–3230, 2017.
- [23] F. Cimmino, M. Avitabile, S. J. Diskin et al., “Fine mapping of 2q35 high-risk neuroblastoma locus reveals independent functional risk variants and suggests full-length BARD1 as tumor-suppressor,” *International Journal of Cancer*, vol. 143, no. 11, pp. 2828–2837, 2018.
- [24] M. Capasso, S. J. Diskin, F. Totaro et al., “Replication of GWAS-identified neuroblastoma risk loci strengthens the role of BARD1 and affirms the cumulative effect of genetic variations on disease susceptibility,” *Carcinogenesis*, vol. 34, no. 3, pp. 605–611, 2013.
- [25] J. Shi, Y. Yu, Y. Jin et al., “Functional polymorphisms in BARD1 association with neuroblastoma in a regional Han Chinese population,” *Journal of Cancer*, vol. 10, no. 10, pp. 2153–2160, 2019.
- [26] G. C. Shukla, J. Singh, and S. Barik, “MicroRNAs: processing, maturation, target recognition and regulatory functions,” *Molecular and Cellular Pharmacology*, vol. 3, no. 3, pp. 83–92, 2011.
- [27] D. P. Bartel, “MicroRNAs: target recognition and regulatory functions,” *Cell*, vol. 136, no. 2, pp. 215–233, 2009.
- [28] K. K. Farh, A. Grimson, C. Jan et al., “The widespread impact of mammalian microRNAs on mRNA repression and evolution,” *Science*, vol. 310, no. 5755, pp. 1817–1821, 2005.
- [29] Z. Hu, J. Chen, T. Tian et al., “Genetic variants of miRNA sequences and non-small cell lung cancer survival,” *The Journal of Clinical Investigation*, vol. 118, no. 7, pp. 2600–2608, 2008.
- [30] B. M. Ryan, “microRNAs in cancer susceptibility,” *Advances in Cancer Research*, vol. 135, pp. 151–171, 2017.
- [31] Y. Xu, L. Liu, J. Liu et al., “A potentially functional polymorphism in the promoter region of miR-34b/c is associated with an increased risk for primary hepatocellular carcinoma,” *International Journal of Cancer*, vol. 128, no. 2, pp. 412–417, 2011.
- [32] P. Bossard and K. S. Zaret, “GATA transcription factors as potentiators of gut endoderm differentiation,” *Development*, vol. 125, no. 24, pp. 4909–4917, 1998.
- [33] J. T. Bensen, C. K. Tse, S. J. Nyante, J. S. Barnholtz-Sloan, S. R. Cole, and R. C. Millikan, “Association of germline microRNA SNPs in pre-miRNA flanking region and breast cancer risk and survival: the Carolina Breast Cancer Study,” *Cancer Causes & Control*, vol. 24, no. 6, pp. 1099–1109, 2013.
- [34] L. B. Gao, L. J. Li, X. M. Pan et al., “A genetic variant in the promoter region of miR-34b/c is associated with a reduced risk of colorectal cancer,” *Biological Chemistry*, vol. 394, no. 3, pp. 415–420, 2013.
- [35] Y. Han, R. Pu, X. Han et al., “Associations of pri-miR-34b/c and pre-miR-196a2 polymorphisms and their multiplicative interactions with hepatitis B virus mutations with hepatocellular carcinoma risk,” *PLoS One*, vol. 8, no. 3, article e58564, 2013.
- [36] L. Li, J. Wu, X. Sima et al., “Interactions of miR-34b/c and TP53 polymorphisms on the risk of nasopharyngeal carcinoma,” *Tumour Biology*, vol. 34, no. 3, pp. 1919–1923, 2013.
- [37] J. He, Y. Zou, X. Liu et al., “Association of common genetic variants in pre-microRNAs and neuroblastoma susceptibility: a two-center study in Chinese children,” *Molecular Therapy - Nucleic Acids*, vol. 11, pp. 1–8, 2018.
- [38] J. He, F. Wang, J. Zhu et al., “The TP53 gene rs1042522 C>G polymorphism and neuroblastoma risk in Chinese children,” *Aging*, vol. 9, no. 3, pp. 852–859, 2017.
- [39] J. He, Y. Zou, T. Wang et al., “Genetic variations of GWAS-identified genes and neuroblastoma susceptibility: a replication study in southern Chinese children,” *Translational Oncology*, vol. 10, no. 6, pp. 936–941, 2017.
- [40] Z. J. Zhuo, W. Liu, J. Zhang et al., “Functional polymorphisms at ERCC1/XPF genes confer neuroblastoma risk in Chinese children,” *eBioMedicine*, vol. 30, pp. 113–119, 2018.
- [41] J. Chang, J. Tian, Y. Yang et al., “A rare missense variant in TCF7L2 associates with colorectal cancer risk by interacting with a GWAS-identified regulatory variant in the MYC enhancer,” *Cancer Research*, vol. 78, no. 17, pp. 5164–5172, 2018.
- [42] J. Chang, J. Tian, Y. Zhu et al., “Exome-wide analysis identifies three low-frequency missense variants associated with pancreatic cancer risk in Chinese populations,” *Nature Communications*, vol. 9, no. 1, article 3688, 2018.
- [43] J. Chang, R. Zhong, J. Tian et al., “Exome-wide analyses identify low-frequency variant in CYP26B1 and additional coding variants associated with esophageal squamous cell carcinoma,” *Nature Genetics*, vol. 50, no. 3, pp. 338–343, 2018.
- [44] J. Li, J. Chang, J. Tian et al., “A rare variant P507L in TPP1 interrupts TPP1-TIN2 interaction, influences telomere length, and confers colorectal cancer risk in Chinese population,” *Cancer Epidemiology, Biomarkers & Prevention*, vol. 27, no. 9, pp. 1029–1035, 2018.
- [45] M. Rokavec, H. Li, L. Jiang, and H. Hermeking, “The p53/miR-34 axis in development and disease,” *Journal of Molecular Cell Biology*, vol. 6, no. 3, pp. 214–230, 2014.
- [46] T. C. Chang, E. A. Wentzel, O. A. Kent et al., “Transactivation of miR-34a by p53 broadly influences gene expression and promotes apoptosis,” *Molecular Cell*, vol. 26, no. 5, pp. 745–752, 2007.
- [47] L. He, X. He, L. P. Lim et al., “A microRNA component of the p53 tumour suppressor network,” *Nature*, vol. 447, no. 7148, pp. 1130–1134, 2007.
- [48] S. Majid, A. A. Dar, S. Saini et al., “miRNA-34b inhibits prostate cancer through demethylation, active chromatin modifications,

- and AKT pathways,” *Clinical Cancer Research*, vol. 19, no. 1, pp. 73–84, 2013.
- [49] D. C. Corney, A. Flesken-Nikitin, A. K. Godwin, W. Wang, and A. Y. Nikitin, “MicroRNA-34b and microRNA-34c are targets of p53 and cooperate in control of cell proliferation and adhesion-independent growth,” *Cancer Research*, vol. 67, no. 18, pp. 8433–8438, 2007.
- [50] K. Y. Wong, R. L. H. Yim, C. C. So, D. Y. Jin, R. Liang, and C. S. Chim, “Epigenetic inactivation of the MIR34B/C in multiple myeloma,” *Blood*, vol. 118, no. 22, pp. 5901–5904, 2011.
- [51] C. J. Liu, X. W. Ma, X. J. Zhang, and S. Q. Shen, “pri-miR-34b/c rs4938723 polymorphism is associated with hepatocellular carcinoma risk: a case-control study in a Chinese population,” *International Journal of Molecular Epidemiology and Genetics*, vol. 8, no. 1, pp. 1–7, 2017.
- [52] M. Hashemi, G. Bahari, M. Naderi, S. Sadeghi-Bojd, and M. Taheri, “Pri-miR-34b/c rs4938723 polymorphism is associated with the risk of childhood acute lymphoblastic leukemia,” *Cancer Genetics*, vol. 209, no. 11, pp. 493–496, 2016.
- [53] J. Zhu, L. Yang, W. You et al., “Genetic variation in miR-100 rs1834306 is associated with decreased risk for esophageal squamous cell carcinoma in Kazakh patients in Northwest China,” *International Journal of Clinical and Experimental Pathology*, vol. 8, no. 6, pp. 7332–7340, 2015.
- [54] C. Yang, X. Ma, D. Liu et al., “Promoter polymorphisms of miR-34b/c are associated with risk of gastric cancer in a Chinese population,” *Tumour Biology*, vol. 35, no. 12, pp. 12545–12554, 2014.
- [55] J. Yin, X. Wang, L. Zheng et al., “Hsa-miR-34b/c rs4938723 T>C and hsa-miR-423 rs6505162 C>A polymorphisms are associated with the risk of esophageal cancer in a Chinese population,” *PLoS One*, vol. 8, no. 11, article e80570, 2013.
- [56] J. Tian, J. Chang, J. Gong et al., “Systematic functional interrogation of genes in GWAS loci identified ATF1 as a key driver in colorectal cancer modulated by a promoter-enhancer interaction,” *American Journal of Human Genetics*, vol. 105, no. 1, pp. 29–47, 2019.
- [57] S. Mei, J. Ke, J. Tian et al., “A functional variant in the boundary of a topological association domain is associated with pancreatic cancer risk,” *Molecular Carcinogenesis*, vol. 58, no. 10, pp. 1855–1862, 2019.



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