

Research Article

Diagnosis of Genital Tuberculosis in Infertile Women by Using the Composite Reference Standard

Riden Saxena^(b),¹ Kriti Shrinet^(b),^{2,3} Sachchida Nand Rai^(b),⁴ Kamal Singh,^{5,6} Shivi Jain,⁷ Shuchi Jain,¹ Deeksha Singh,¹ Shampa Anupurba^(b),⁵ and Madhu Jain^(b)

¹Department of Obstetrics & Gynecology, Institute of Medical Sciences, Banaras Hindu University, 221005, Varanasi, India ²School of Biotechnology, Banaras Hindu University, 221005, Varanasi, India

³School of Biotechnology, IFTM University, 244102, Moradabad, India

⁴Centre of Biotechnology, University of Allahabad, 211002, Prayagraj, India

⁵Department of Microbiology, Institute of Medical Sciences, Banaras Hindu University, 221005, Varanasi, India

⁶Virus Research & Diagnostic Laboratory, Department of Virology, Rajendra Memorial Research Institute of Medical Science, 800007, Patna, Bihar, India

⁷Department of Radiology, Institute of Medical Sciences, Banaras Hindu University, 221005, Varanasi, India

Correspondence should be addressed to Shampa Anupurba; shampa_anupurba@yahoo.co.in and Madhu Jain; drmadhujainbhu@gmail.com

Received 17 May 2022; Revised 13 July 2022; Accepted 29 July 2022; Published 16 August 2022

Academic Editor: Ioannis Kosmas

Copyright © 2022 Riden Saxena et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Female genital tuberculosis (FGTB) can be asymptomatic or even masquerade as other gynecological conditions. Conventional methods of FGTB diagnosis include various imaging, bacteriological, molecular, and pathological techniques that are only positive in a small percentage of patients, leaving many cases with undiagnosed condition. In the absence of a perfect diagnostic method, composite reference standards (CRSs) have been advocated in this diagnostic study. This study assesses the agreement between traditional diagnostic modalities using CRS and prevalent TB groups among different fallopian tube infertility manifestations. A total of 86 women with primary and secondary infertility were included in the study and subjected to bacteriological, pathological, and radiological examination for the diagnosis of FGTB. Results were evaluated statistically for concordance of the diagnostic tests to the CRS by sensitivity and specificity, while PPV and NPV were calculated for the performance of diagnostic tests of FGTB. We observed that 11.2% of women were found to be true positives by means of CRS. The positive findings by CRS were as follows: ultrasonography (13.9%), laparoscopy (14%), hysteroscopy (12%), GeneXpert (4.8%), culture (4.8%), polymerase chain reaction (4.8%), and histopathology (6.4%). GeneXpert and culture were found to have a perfect agreement with CRS. Hysterosalpingography, laparoscopy, and hysteroscopy have a fair agreement with CRS. Out of 43 women with tubal factor infertility, 6 women were found in the definitive TB group with mixed conditions of tubal manifestations. This study evaluates and demonstrates the reliability of the collective assessment of various diagnostic methods with CRS findings that help in identifying different TB groups of genital tuberculosis patients from all infertile patients by applying the criteria of CRS.

1. Introduction

FGTB is still a serious concern in low-income nations, causing substantial morbidity, particularly infertility at reproductive age [1]. Due to underreporting of cases, asymptomatic incidences, ambiguous symptomatology, and lack of effective diagnostics with high sensitivity, the exact prevalence of FGTB remains unknown [2, 3]. The reported incidence varies by country: 1% in US infertility clinics [4], 1% in Scandinavian countries [5], and 4–8% in Pakistan [6]. South Africa's share is 1% [7]. In different parts of India, the rates range from 16.1% to 19% [8]. The prevalence of female genital TB recently reported in India ranges from 45.1 cases per 100,000 women in the community-based research in the



FIGURE 1: Representative laparoscopic view and arrows showing caseous nodules in FGTB case.

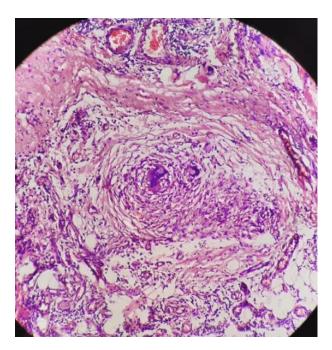


FIGURE 2: Representative granulomatous inflammation on histopathology showing well-formed granuloma with giant cell at center surrounded by epitheloid cells and lymphocytes, outermost surrounded by fibroblasts.

Andaman Islands to 48.5 percent among infertile women in north India [9, 10]. The pathophysiology of FGTB in infertile female patients may be considerably influenced by genetics, immunology, environment, and infection [11]. FGTB usually occurs secondary to pulmonary TB [12]. It causes a variety of nonspecific symptoms in women, ranging from infertility to irregular menstruation and pelvic pain [13]. FGTB is regarded as a chronic and asymptomatic or low symptomatic disease, so it may be difficult to diagnose in women with infertility [14]. Due to its paucibacillary nature,

the diagnostic dilemma of FGTB continues to be a challenge [15]. A timely diagnosis and effective treatment may prevent it. Bacterial cultures and PCR-based diagnostics are two instances of the more cutting-edge and effective diagnostic methods that are increasingly accessible for the detection of tuberculosis [16]. The sensitivity and specificity are significantly influenced by technical factors as well, including the use of appropriate controls, standard strains, adequate conditions, and the retesting of samples with suspicious positive results [17]. The present study is assessing the combined diagnostic modality for early detection of genital tuberculosis with accuracy. According to the TB guidelines, the diagnosis of FGTB should be made based on any one of the laparoscopic features typical for FGTB, any gynecological specimen positive for acid-fast smear, or positive for mycobacterium tuberculosis (MTB) on culture, any findings consistent with FGTB on histopathology [18]. In the CRS, we included culture, GeneXpert, PCR, histology, radiography, imaging, and history for the diagnosis of FGTB. TB will be confirmed if there are two of the following AFB microscopy/histopathology/consistent feature on USG and HSG/ laparoscopic or hysteroscopy features typical for FGTB or GeneXpert or culture positive in individuals with suspicion of FGTB. PCR or history with imaging features of FGTB will be considered in probable TB category, whereas only imaging features suggestive to FGTB will be grouped into possible TB group and patient tested negative for all tests will be considered as non-TB case. The primary objective of this prospective diagnostic accuracy study was to diagnose FGTB and assess TB in women with tubal factor infertility by means of CRS. We also evaluated the concordance of the diagnostic tests with respect to the CRS by sensitivity and specificity, while positive predictive value (PPV) and negative predictive value (NPV) were calculated for the performance of diagnostic tests of FGTB.

2. Materials and Methods

2.1. Study Design and Specimens. This study was conducted in the Department of Obstetrics and Gynecology and Department of Microbiology, Institute of Medical Science, Banaras Hindu University, India. The study was approved by the Ethics Committee [Dean/2018/EC/481]. During one year (December 2018 to December 2019) of research, a total of 86 women were included in the study with written consent. The unexplained and asymptomatic or low symptomatic infertility and general investigation for infertility helped to make suspicion for secondary tuberculosis. Post hoc sample size for two proportions was calculated. Only 62 women with primary (72%) and secondary (28%) infertility were selected and enrolled according to their clinical presentation. Exclusion criteria are as follows: women over the age of 45, with symptoms suggestive of pulmonary tuberculosis other than infertility, who had taken or were on a regimen of antituberculosis drugs, severe psychiatric dysfunctions, sexual disorders, infertility due to abnormality in ovulation, endocrine problems, pulmonary infections, multiple sclerosis or other autoimmune disorders, human immunodeficiency virus (HIV) and coinfections, diabetes,

USG		CRS	HSG		CRS
12/62	TB group $(n = 8)$	Non-TB group (n=54)	16/43	TB group $(n = 6)$	Non-TB group $(n = 37)$
Positive $(n = 12)$	6	6	Positive $(n = 16)$	5	11
Negative $(n = 50)$	2	48	Negative $(n = 27)$	1	26
Sensitivity	80.00%, (95% C	CI: 44.39% to 97.48%)	Sensitivity	85.71% (95% 0	CI: 42.13% to 99.64%)
Specificity	88.89%, (95% C	CI: 77.37% to 95.81%)	Specificity	70.27% (95% 0	CI: 53.02% to 87.13%)
PPV	57.14%, (95% C	CI: 37.10% to 75.09%)	PPV	35.29%, (95% (CI: 23.39% to 49.26%)
NPV	96.00%, (95% C	CI: 87.81% to 98.77%)	NPV	96.30%, (95% (CI: 80.71% to 99.38%)
Kappa value (95% CI)	Agreement	Level of agreement	Kappa value (95% CI)	Agreement	Level of agreement
0.52 (0.24 to 0.81)	72.74%	Moderate	0.31 (0.053 to 0.57)	59.22%	Fair

TABLE 1: Performance of the Imaging methods for the diagnosis of FGTB: sensitivity, specificity, and kappa agreement in comparison with composite reference standards.

PPV: positive predictive value; NPV: negative predictive value; TB group: TB-suspected patients; PCR: polymerase chain reaction; CRS: composite reference standard. For patients with suspicion of FGTB, diagnosis of TB was given if any two of culture/histopathology/radiological findings were positive.

TABLE 2: Performance of the Imaging endoscopic methods for the diagnosis of FGTB: sensitivity, specificity, and kappa agreement in comparison with composite reference standards.

Laparoscopy		CRS	Hysteroscopy		CRS
19/50	TB group $(n = 7)$	Non-TB group $(n = 43)$	15/50	TB group $(n = 6)$	Non-TB group $(n = 44)$
Positive $(n = 19)$	5	14	Positive $(n = 15)$	5	10
Negative $(n = 31)$	2	29	Negative $(n = 35)$	1	34
Sensitivity	77.78% (95% 0	CI: 39.99% to 97.19%)	Sensitivity	85.71% (95% 0	CI: 42.13% to 99.64%)
Specificity	67.44% (95% 0	CI: 51.46% to 80.92%)	Specificity	77.27% (95% 0	CI: 62.16% to 88.53%)
PPV	33.33% (95% (CI: 22.32% to 46.53%)	PPV	37.50% (95% 0	CI: 24.34% to 52.81%)
NPV	93.55% (95% (CI: 80.76% to 98.04%)	NPV	97.14% (95% 0	CI: 84.61% to 99.53%)
Kappa value (95% CI)	Agreement	Level of agreement	Kappa value (95% CI)	Agreement	Level of agreement
0.22 (-0.015 to 0.468)	58.64%	Fair	0.36 (0.09 to 0.64)	65.20%	Fair

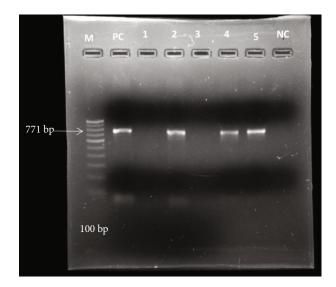


FIGURE 3: Gel image of amplified PCR product of MPT64 gene. M: marker 100 bp; PC: positive control (H37Rv); lanes 2, 4, and 5: positive band for Mycobacterium tuberculosis (*mpt64* gene); lanes 1 and 3: negative for *mpt64* gene; and NC: negative control (PCR grade water).

malnutrition, and other medical disorders like hypertension and peritoneal adhesions due to previous abdominal surgery. Control samples: 62 ETBs samples were selected from fertile women coming for medical termination of pregnancy in the family planning department.

2.2. Imaging Data Collection. All of the patients were subjected to a thorough clinical imaging examination such as USG, HSG, hysteroscopy, and laparoscopy. HSG was not done on a regular basis, but the results were recorded whenever it was done from the outside. When feasible, diagnostic video laparoscopy and visual hysteroscopy were performed.

2.2.1. Ultrasonography (USG). All 62 women were investigated for the presence of loculated ascites, bilateral, predominantly solid adnexal masses with scattered small calcification, thickened peritoneum, thickened omentum, and endometrial involvement on high resolution abdominal and transvaginal USG alerted to the possibility of genital tract TB [19].

2.3. Hysterosalpingography (HSG). Out of 62 women, 43 women had HSG findings of tubal factor infertility. TB manifested in various forms in HSG and nonspecific changes like tubal occlusion, tubal dilatation, diverticular outpouching (salpingitis isthmic nodosa), irregular contour, and hydrosalpinx to specific patterns like pipestem tube, cotton wool plug, cobblestone tube, golf club tube, leopard skin tube, and beaded tube. In the presence of synechiae, tubal

GeneXpert	CRS		Culture	CRS	
3/62	TB group $(n = 3)$	Non-TB group $(n = 59)$	3/62	TB group $(n = 3)$	Non-TB group $(n = 59)$
Positive $(n = 3)$	3	0	Positive $(n = 3)$	3	0
Negative $(n = 59)$	0	59	Negative $(n = 59)$	0	59
Sensitivity	100.00% (95% CI: 29.24% to 100.00%)		Sensitivity	100.00% (95% CI: 29.24% to 100.00%)	
Specificity	100.00% (95% CI: 93.94% to 100.00%)		Specificity	100.00% (95% CI: 93.94% to 100.00%)	
PPV	100.00%		PPV	100.00%	
NPV	100.00%		NPV	100.00%	
Kappa value (95% CI)	Agreement	Level of agreement	Kappa value (95% CI)	Agreement	Level of agreement
1.0 (1.0 to 1.0)	90.79%	Perfect	1.0 (1.0 to 1.0)	90.79%	Perfect

TABLE 3: Performance of the bacteriology for the diagnosis of FGTB: sensitivity, specificity, and kappa agreement in comparison with composite reference standards.

TABLE 4: Performance of the PCR and HPE for the diagnosis of FGTB: sensitivity, specificity, and kappa agreement in comparison with composite reference standards.

PCR		CRS	HPE		CRS
5/62	TB group $(n = 3)$	Non-TB group $(n = 59)$	4/62	TB group $(n = 4)$	Non-TB group $(n = 58)$
Positive $(n = 5)$	3	2	Positive $(n = 4)$	3	1
Negative $(n = 57)$	0	57	Negative $(n = 58)$	1	57
Sensitivity	100.00% (95% CI: 29.24% to 100.00%)		Sensitivity	75% (95% CI: 19.41% to 99.37%)	
Specificity	96.61% (95% 0	CI: 88.29% to 99.59%)	Specificity	98.28% (95% 0	CI: 90.76% to 99.96%)
PPV	60.00% (95% 0	CI: 27.75% to 85.42%)	PPV	75.00% (95% 0	CI: 28.29% to 95.78%)
NPV	100.00%		NPV	99.28% (95% CI: 91.26% to 99.68%)	
Kappa value (95% CI)	Agreement	Level of agreement	Kappa value (95% CI)	Agreement	Level of agreement
0.73 (0.38 to 1.0)	87.88%	Substantial	0.73 (0.37 to 1.0)	87.93%	Substantial

blockage in the transition zone between the isthmus and the ampulla, calcified lymph nodes, multiple constrictions, and adnexal calcifications that are irregular, linear, or nodular, TB will be highly suspected. Special features such as collarstud abscess, T-shaped uterus, and pseudounicornuate uterus, as well as nonspecific features such as synechiae formation, uterine contour distortion, obliteration of the uterine cavity, and venous and lymphatic intravasations, may be seen as a consequence of tuberculosis. Tubal manifestations were categorized into definitive TB, probable TB, possible TB, and non-TB groups [20].

2.3.1. Endoscopy. Out of 62 women, 50 women were undergone laparoscopy and hysteroscopy. For the diagnosis of FGTB, microcaseations, and micropolyps, fibrosed ostia, synechia bands, narrow cavity/T-shape cavity, and Asherman's syndrome are considered as diagnostic classification by hysteroscopy. During hysteroscopy, the color of the endometrium, the opening of the endometrial glands, and any TB features such as tubercles, shaggy regions, and intrauterine adhesions were all thoroughly examined. The entire pelvic and abdominal cavity, including the fallopian tubes, uterus, ovaries, Douglas pouch, uterovesical pouch, liver, peritoneum, intestines, and gall bladder, were thoroughly examined for any tuberculous lesions such as tubercles, shaggy areas, pyosalpinx, hydrosalpinx, beading of tubes, pelvic, abdominal or perihepatic adhesions, ovarian tuberculosis, tube patency, and all other abnormalities carefully studied by laparoscopy. Sacculated tubes, convoluted, fluidfilled vesicles, yellow discoloration of mesosalpinx, hydrosalpinx, lead pipe appearance, encysted fluid collection, tuboovarian mass, pyosalpinx, various grades of pelvic adhesions, and miliary tubercles appearances are considered as diagnostic classification by laparoscopy (Figure 1) [21].

2.4. Processing of Endometrial Tissue Biopsy. Endometrial biopsies (EMBs) from all 62 women were aspirated using Karman cannula no. 4, between the 20th and 25th day of menstruation in the mini operation theatre of the hospital. In the BSL-3 laboratory, each EMB sample was centrifuged for 20 minutes at 6,000 rpm in a tube containing sterile normal saline, and the pellet (about 1 mL) was transferred to a 1.5-mL Eppendorf tube containing fine glass beads up to one-third of the Eppendorf tube's capacity. For 1 minute, the material was homogenized in a tissue lyser (Bertin Technologies Pvt. Ltd) [22]. Each homogenized tissue sample was divided into three parts: GeneXpert, PCR, and culture. The colony grew on Lowenstein Jensen (L-J) media were again subjected to AFB staining and PCR.

2.5. GeneXpert MTB/RIF Assay. One ml of homogenized EMB was mixed with 2.0 ml of GeneXpert sample reagent. For 30 seconds, the mixture was vortexed. After allowing the sample to stand for 15 minutes at room temperature,

TABLE 5: Findings of fallopian tube TB suspected infertility patients and clinical assessment of patients on the basis of composite reference standard (CRS) criteria.

Hysterosalpingogram	Definitive TB groups	Probable TB groups	Possible TB groups	Non-TB groups	Total
<i>n</i> = 43	CRS or GeneXpert or culture	PCR or history + imaging suggestive for FGTB	Imaging suggestive to FGTB	Negative for all tests	
Calcifications	_	1	2	1	4
Tubal outline irregular	1	1	2	_	4
Tubal occlusion	2	2	3	1	8
Tubal dilation	2	—	2	_	4
Peritubal adhesion	_	1	2	—	3
Calcifications+ tubal outline irregular	_	1	_	1	2
Calcifications+ tubal outline irregular + tubal occlusion	_	1	2	_	3
Tubal occlusion+ peritubal adhesion	_	1	1	_	2
Tubal occlusion+ tubal outline irregular	_	_	1	1	2
Tubal occlusion+ calcifications+ peritubal adhesion	_	—	1	—	1
Tubal occlusion+ tubal dilation	1	1	2		4
Tubal outline irregular+ peritubal adhesion		_	1	1	2
Tubal dilation+ calcifications (hydrosalpinx)	_	—	1	1	2
Normal spills	—	1	1	—	2
Total	6	10	21	6	43

2 ml of the mixed sample was transferred to the test cartridge. The cartridge was inserted into the GeneXpert instrument (Cepheid). Within 2 hours, the results were reported as affirmative or negative, as well as sensitivity to the rifampicin (RIF) resistance determining region of the *rpoB* gene using molecular beacons [23].

2.6. DNA Extraction. DNA isolation was carried out from the homogenized EMB using the cetyl trimethylammonium bromide chloroform (CTAB-chloroform) method with slight modifications in BSL-3 Lab [24]. Thermo Scientific NanoDrop 2000 was used to analyze the quality and quantity of isolated DNA.

2.7. Polymerase Chain Reaction. We chose a highly conserved and restricted region of the MPT64 (771 bp) gene encoded by the regions of difference₂ (RD₂). Forward and reverse sequences (5'-3') of the primers were ACCGAA CACTCATTTCCGC and CTACTCCCGGAGGAATTTCG, respectively. Reaction conditions were as follows: initial denaturation at 95°C for 5 min, 30 cycles of 95°C for 30 s, 59°C for 45 s, 72°C for 45 s, and 30 cycles of 95°C for 30 s, 59°C for 45 s, 72°C for 45 s, and 10-minute final elongation phase at 72°C [25].

2.8. Solid Culture. About $100 \,\mu$ l homogenized EMB was inoculated on the L-J medium slant in a bottle and left on a horizontal plain until the inocula were absorbed. The cul-

ture bottles were incubated at 37°C. The inoculated bottles were inspected after 24 hours, 48 hours, and then once a week for the next eight weeks. AFB staining was performed from a colony grown on L-J media [26].

2.9. Histopathological Examination. The biopsy specimens were cut into paraffin-embedded tissue slices and fixed in 10% formalin, hematoxylin, and eosin stains and were used to stain the sections. Caseating granuloma is indicated in samples for the diagnosis of genital tract TB, along with epithelioid cells, giant cells, fibrosis, and lymphocyte proliferation coupled with caseous necrosis (Figure 2) [22].

2.10. Statistical Analysis. Sensitivity, specificity, PPV, and NPV were determined by comparing diagnostic test results with CRS. For the agreement analysis, the Kappa chi-square test was used. Significant p value of 0.05 was considered. Analysis was done by online tools such as Medcalc and GraphPad Prism 8.

3. Results

3.1. Performance of Imaging Methods. By using the USG method, out of 62 women, 12 (19.3%) were found positive for TB. Furthermore, out of 43 women, 16 (37%) were found positive by HSG (Table 1). There were findings of FGTB on laparoscopy and hysteroscopy in 19 (38%) and 15 (30%) of total women, respectively (Table 2). However, when USG,

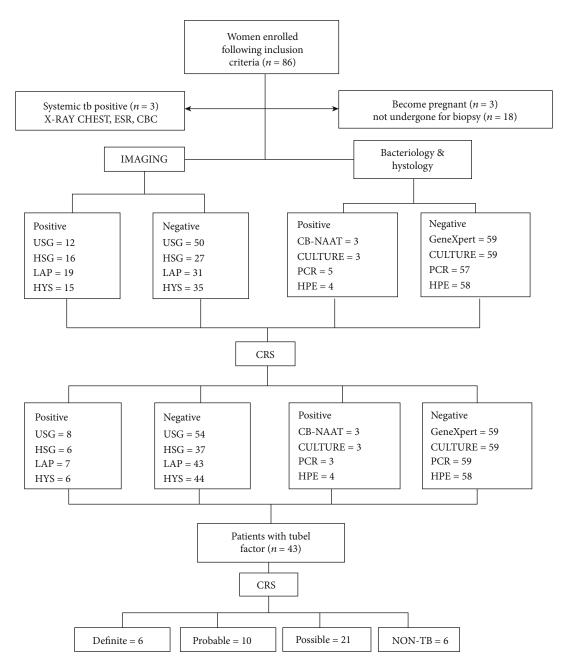


FIGURE 4: Diagnostic algorithm for FGTB. ESR: erythrocyte sedimentation rate; CBC: complete blood count; USG: ultrasonography; HSG: hysterosalpingograpgy; LAP: laparoscopy; HYS: hysteroscopy; PCR: polymerase chain reaction; HPE: histopathology; and CRS: composite reference standard.

HSG, laparoscopy, and hysteroscopy were compared with CRS, 8 (12.9%), 6 (9.6%), 7 (14%), and 6 (12%) were found in the TB group, respectively.

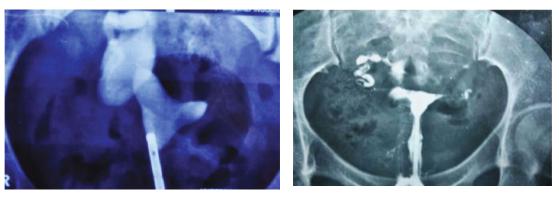
3.2. Performance of Bacteriology Methods. Out of 62 samples, 3 (4.8%) samples were found to be positive by GeneXpert and culture method. Only one sample was found to be rifampicin resistant by GeneXpert. Further, 5 isolates were also found positive by PCR (Figure 3). However, when these results were compared with CRS, similar 3 samples are found in the confirmed TB group, whereas 2 samples were found in probable TB group and 57 were in non-TB group, respectively, as shown in Table 3.

3.3. Performance of Histopathology Methods. Of 62 samples, 4 (6.4%) samples were found to be positive by HPE. However, when compared with CRS, 4 samples were found in confirmed TB group, and 58 were in non-TB group (Table 4).

3.4. Agreement of Imaging, Bacteriology, and Histopathological Results with CRS. When the kappa (k) value was calculated, GeneXpert and culture showed perfect

Patients no.	Methods	Findings
OBG19RS 14	USG, HSG, hysteroscopy, GeneXpert, culture, PCR	Diffused endometrial border, dilated tube, hydrosalpinx, GeneXpert positive, culture positive, PCR positive
OBG19RS 23	USG, HSG, laparoscopy, hysteroscopy, GeneXpert, culture, PCR	Beaded tubes, intrauterine adhesion, GeneXpert positive, culture positive, PCR positive with rifampicin resistant, caseous nodules
OBG19RS 38	History, USG, HSG, laparoscopy, hysteroscopy, GeneXpert, culture, PCR	Localized peritoneal spill with tubal occlusion, GeneXpert positive, culture positive, PCR positive, tubercular nodules
OBG19RS 40	History, USG, HSG, laparoscopy, hysteroscopy	Bilateral tubal dilation, pyosalpinx
OBG19RS 44	USG, HSG, laparoscopy, HPE	Tubal occlusion and dilation, cornual block
OBG19RS 49	USG, HSG, laparoscopy, hysteroscopy, HPE	Outline irregular, visualized endometrial disease-like tubercles
OBG19RS 58	History, USG, laparoscopy, hysteroscopy, HPE	Intrauterine adhesions, heterogeneous endometrium with irregular surface, epithelial granulomatous nodules
OBG19RS 59	USG, laparoscopy, HPE	Endometritis, lesions on uterus like tubercles, caseous nodules

TABLE 6: Clinical findings in patients of genital tuberculosis by using CRS.



(a)

(b)



FIGURE 5: (a) HSG showing right hydrosalpinx and left cornual block; (b) HSG showing irregular uterine cavity with a localized peritoneal spill on the right and tubal occlusion on left; (c) HSG showing irregular and deformed uterine cavity with mild right hydrosalpinx; and (d) HSG showing normal uterine cavity and fallopian tubes without any peritoneal spill.

agreement with CRS, each with a value of 1.0, whereas the agreement of PCR (k = 0.73) and HPE (k = 0.73) was found to be substantial with CRS. USG (k = 0.52) showed moderate agreement with CRS. However, HSG (k = 0.31), laparoscopy (k = 0.22), and hysteroscopy (k = 0.36) showed fair agreement with the CRS, respectively (Tables 1 and 2).

3.5. *Tuberculosis in Women with Tubal Factors*. A total of 43 were found to have tubal factor infertility. Out of these, only 6 (13.9%) women were characterized in the definitive TB category, whereas 10 (23.2%) samples were found in the probable TB group and 21 (48.8%) samples were categorized in the possible TB group. Among all women, tubal occlusion

and tubal dilation are the most prevalent conditions for TB infection, but tubal conditions fall into probable TB and possible TB cannot be ruled out as negative TB. Three samples from the probable TB group had dual conditions (calcifications+ tubal outline irregular, tubal occlusion + peritubal adhesion, and tubal occlusion+ tubal dilation), whereas only one sample with triple conditions (calcifications + tubal outline irregular + tubal occlusion) falls into the same group of TB. One woman found definitive TB with dual tubal abnormal conditions (tubal occlusion + tubal dilation). Two women were found with normal spillage conditions. Only 6 women were categorized in non-TB group (Table 5).

4. Discussion

According to the guidelines for extra-pulmonary TB for India, criteria should be used to make a diagnosis of FGTB [18]. But problems arise when the presentation is variable, and a high level of clinical suspicion is necessary to make the diagnosis. Around 11% of individuals report having no symptoms other than infertility, and these patients require a diagnostic workup to rule out all prevalent causes of infertility [27].

The presence of AFB on microscopy, culture, or histopathological evidence of TB granuloma provides a definitive diagnosis, but it is only positive in a small number of cases and forces the use of additional modalities such as PCR, hysteroscopy, or laparoscopy findings to make a timely diagnosis for early treatment. However, there is no gold standard approach to detecting FGTB, so in this study, we shared the experience and reliability of our proposed criteria of CRS in the diagnosis of FGTB (Figure 4). We compared a mix of microbiological, histological, molecular, and radiological methods and history of TB. Kappa is a statistical coefficient that evaluates the degree of agreement between two raters (judges) who classify things into mutually exclusive groups [28].

In our study, the agreement between conventional investigation and CRS corresponds to the collective diagnostic model. In GeneXpert, whereas culture has perfect agreement, PCR and HPE have substantial agreement with CRS. Most of the time, endometrial biopsy does not contain sufficient numbers of bacilli for AFB or culture investigation [29]. Several investigations have been conducted in the past using endometrial tissue as well as tissues from other organs, but due to low pick-up rates (1-18%) of bacilli or most probably as a result of the monthly shedding of the endometrium's superficial layers, identification of TB was very poor [22, 30, 31].

In the imaging, USG shows moderate agreement with CRS, whereas HSG, laparoscopy, and hysteroscopy each have fair agreement, respectively. The sensitivity of the imaging and pathology tests ranged from 75.0% to 85.71%, whereas specificity was found to range from 70.27% to 98.28%. On the other hand, GeneXpert and culture have 100% sensitivity and specificity, but PCR has 100% sensitivity and 96.61% specificity. The three women who tested positive for GeneXpert also tested positive for culture and PCR which indicates active TB. Of the 5 PCR-positive samples, 3 samples were categorized as definitive TB, and 2 were in the probable TB category (Tables 5 & 6). One patient

(OBG19RS23) with tubal blockage went under all diagnostic tests, and the findings in the fallopian tubes were beaded tubes and intrauterine adhesion. She was found to be GeneXpert positive with rifampicin resistance, and culture positive, PCR positive, and caseous nodules were found in HPE (Table 6). This trend indicates the presence of active bacilli in the patients. Our study is comparable to a recent study conducted by Sethi et al. [3], where PCR was found 22.39% positive and HPE 2.99%. However, no cases were found culture positive. Our findings suggest high reliability on our criteria of CRS, as clinicians can be quick to identify FGTB patients in order to avoid irreversible damage.

In high prevalence countries, mycobacterium culture and histopathology facilities are inadequate [32]. In that situation, the infection is often detected during HSG for the first time in any of the infertility investigations [33]. Furthermore, HSG is still the gold standard for tubal lumen assessment [20] and is a useful method for the diagnosis of female genital TB [34]. Genital TB causes a range of HSG appearances, from nonspecific to specific findings (Figure 5). According to Chavhan [34] and Afzali [14], the appearance of calcified lymph nodes in the pelvic or along the length of the fallopian tubes may confirm a diagnosis of tuberculosis. In our study, no women with calcified fallopian tubes were found to have definitive TB. But other samples which lie in the probable and possible groups cannot be ruled out as non-TB group. In tubal outline, caseous ulceration of the tubal mucosa results in an irregular, ragged, or diverticular appearance of the tubal lumen contour. Tubal occlusion (18%) is the most common finding by HSG in TB conditions in our study. Scarring can cause several constrictions along the course of the fallopian tube, giving it a "beaded" pattern. Each condition of tubal occlusion from all tubal blockage cases was also found to be prevalent (46.51%) in our study. We found HSG findings with tubal occlusion and dilation were the most prevalent conditions of FGTB in women with tubal factor infertility. The previous history is not much helpful factor to include in the model as only three women with a history of TB out of 11 women were found to be FGTB positive by CRS. The remaining 8 women with history and suffering from tubal blockage with imaging consistency were categorized in probable TB. Overall, out of 62 patients, 8 (12.9%) patients were found TB positive, in which 6 (9.6%) women with tubal blockage and 2 asymptomatic women were found to be TB positive by using CRS (Table 6). By using HSG, the typical radiographic features of genital TB are reliable indicators. All 62 samples (control) from the patients without mycobacterial infections were found to be negative for both PCR and culture as expected. The PCR assay cannot differentiate live and dead bacteria; hence, it is recommended only for new and active cases [35]. We could only include EMB samples in our study as EMB curettage is less invasive. The number of samples is less as it is a one-year study.

5. Conclusion

Although composite reference standard can minimize the degree of bias, they cannot entirely eliminate it since a

combination of imperfect tests is unlikely to generate a composite standard with perfect sensitivity and specificity. In our study, bacteriology had a perfect and substantial agreement with CRS, so we should collect a sufficient endometrial sample using the right approach, preferably laparoscopic or hysteroscopy guided, from a highly suspicious site. The combination of imaging tests with CRS has a better scope as their agreement is fair with CRS. Due to differential causes or earlier damage of fallopian tubes by TB in infertile women, HSG manifestations have high numbers of false positivity, so high suspicion for each patient is required. Our study demonstrates that, while still using microbiological, histological, and radiological techniques, the composite reference standard incorporates many least invasive diagnostic modalities and is conducive to conclude FGTB.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors thank C&DST lab staff and the hospital staff involved for their voluntary participation. The authors would like to acknowledge UGC-Research SRF Fellowship for awarding fellowship to Mr. Riden Saxena (Ref. No-R/ Dev.(SRF-JRF)'R'A/C/Medicine/IMS/58668/2018) and Dr. D.S. Kothari Postdoctoral Scheme for awarding the fellowship to Dr. Sachchida Nand Rai (Ref. No-F.4-2/2006 (BSR)/BL/19-20/0032).

References

- N. Gupta, J. B. Sharma, S. Mittal, N. Singh, R. Misra, and M. Kukreja, "Genital tuberculosis in Indian infertility patients," *International Journal of Gynecology & Obstetrics*, vol. 97, no. 2, pp. 135–138, 2007.
- [2] I. K. Neonakis, D. A. Spandidos, and E. Petinaki, "Female genital tuberculosis: a review," *Scandinavian Journal of Infectious Diseases*, vol. 43, no. 8, pp. 564–572, 2011.
- [3] A. Sethi, B. Bajaj, D. Nair, D. Pachauri, M. Gupta, and A. Mahajan, "Comparison of conventional methods with newer diagnostic modalities to detect genital tuberculosis in infertile women," *J. Obstet. Gynecol. India*, pp. 1–7, 2022.
- [4] G. Schaefer, "Female genital tuberculosis," *Clinical Obstetrics and Gynecology*, vol. 19, no. 1, pp. 223–239, 1976.
- [5] V. Falk, K. Ludviksson, and G. Agren, "Genital tuberculosis in women: analysis of 187 newly diagnosed cases from 47 Swedish hospitals during the ten-year period 1968 to 1977," *American Journal of Obstetrics and Gynecology*, vol. 138, no. 7, pp. 974–977, 1980.
- [6] S. Shahzad, "Investigation of the prevalence of female genital tract tuberculosis and its relation to female infertility: an observational analytical study," *Iran. J. Reprod. Med.*, vol. 10, no. 6, pp. 581–588, 2012.

- [7] K. Margolis, P. A. B. Wranz, T. F. Kruger, J. J. Joubert, and H. J. Odendaal, Genital tuberculosis at Tygerberg Hospital: Prevalence, Clinical Presentation and Diagnosis, 1992.
- [8] S. N. Tripathy and S. N. Tripathy, "Infertility and pregnancy outcome in female genital tuberculosis," *International Journal* of Gynecology & Obstetrics, vol. 76, no. 2, pp. 159–163, 2002.
- [9] R. Parvez, A. P. Sugunan, P. Vijayachari et al., "Prevalence of female genital tuberculosis, its risk factors and associated clinical features among the women of Andaman Islands, India: a community-based study," *Public Health*, vol. 148, pp. 56–62, 2017.
- [10] N. Singh, G. Sumana, and S. Mittal, "Genital tuberculosis: a leading cause for infertility in women seeking assisted conception in North India," *Archives of Gynecology and Obstetrics*, vol. 278, no. 4, pp. 325–327, 2008.
- [11] V. Bhanothu, V. Lakshmi, J. P. Theophilus, R. Rozati, P. Badhini, and B. Vijayalaxmi, "Investigation of toll-like receptor-2 (2258G/A) and interferon gamma (+ 874T/A) gene polymorphisms among infertile women with female genital tuberculosis," *PLoS One*, vol. 10, no. 6, article e0130273, 2015.
- [12] S. J. F. Ara, S. Ahmed, A. A. Saleh, M. M. A. Molla, S. Chowdhury, and S. Anwar, "Endometrial cytokine expression from clinically suspected genital tuberculosis patients at tertiary care hospitals in Dhaka," *J. Clin. Tuberc. Other Mycobact. Dis.*, vol. 27, article 100301, 2022.
- [13] G. A. Grace, D. B. Devaleenal, and M. Natrajan, "Genital tuberculosis in females," *The Indian Journal of Medical Research*, vol. 145, no. 4, pp. 425–436, 2017.
- [14] N. Afzali, F. Ahmadi, and F. Akhbari, "Various hysterosalpingography findings of female genital tuberculosis: a case series," *Iran. J. Reprod. Med.*, vol. 11, no. 6, pp. 519–524, 2013.
- [15] S. N. Tripathy and H. P. SNTripathy, "Diagnostic Dilemma in Genital Tuberculosis," *Practical Guide in Infertility*, vol. 142, 2018.
- [16] Y. Wang, R. Shao, C. He, and L. Chen, "Emerging progress on diagnosis and treatment of female genital tuberculosis," *The Journal of International Medical Research*, vol. 49, no. 5, p. 03000605211014999, 2021.
- [17] V. Bhanothu, J. P. Theophilus, and R. Rozati, "Use of endoovarian tissue biopsy and pelvic aspirated fluid for the diagnosis of female genital tuberculosis by conventional versus molecular methods," *PLoS One*, vol. 9, no. 5, article e98005, 2014.
- [18] I. Convenors, Initiative of Central TB Division Ministry of Health and Family Welfare, Government of India Index-TB Guidelines-Guidelines on Extra-Pulmonary Tuberculosis for India, 2016.
- [19] J. B. Sharma, S. Dharmendra, S. Agarwal, and E. Sharma, "Genital tuberculosis and infertility," *Fertil. Sci. Res.*, vol. 3, no. 1, p. 6, 2016.
- [20] S. Merchant, A. Bharati, and P. Badhe, "Female genital tract tuberculosis," *J. Women's Imaging*, vol. 6, no. 4, pp. 146–152, 2004.
- [21] J. B. Sharma, "Tuberculosis and obstetric and gynecological practice," *Progress in obstetrics and gynecology*, vol. 18, pp. 395–427, 2008.
- [22] R. B. P. Thangappah, C. N. Paramasivan, and S. Narayanan, "Evaluating PCR, culture & histopathology in the diagnosis of female genital tuberculosis," *The Indian Journal of Medical Research*, vol. 134, no. 1, p. 40, 2011.
- [23] S. Selfegna and A. Alelign, "Detection of Mycobacterium tuberculosis and Rifampicin Resistance Using GeneXpert MTB/RIF Assay at Enat Hospital, Central Ethiopia," *Tuberculosis Research and Treatment*, vol. 2022, 2022.

- [24] S. Honore-Bouakline, J. P. Vincensini, V. Giacuzzo, P. H. Lagrange, and J. L. Herrmann, "Rapid diagnosis of extrapulmonary tuberculosis by PCR: impact of sample preparation and DNA extraction," *Journal of Clinical Microbiology*, vol. 41, no. 6, pp. 2323–2329, 2003.
- [25] K. Singh, R. Kumari, R. Tripathi, A. Gupta, and S. Anupurba, "Mutation in MPT64 gene influencing diagnostic accuracy of SD bioline assay (capilia)," *BMC Infectious Diseases*, vol. 19, no. 1, pp. 1–6, 2019.
- [26] "Guidelines: Central TB Division," https://tbcindia.gov.in/ index1.php?lang=1&level=1&sublinkid=4571&lid=3176, (accessed Mar. 23, 2021).
- [27] "Home:: Central TB Division," https://tbcindia.gov.in/, (accessed Aug. 16, 2021).
- [28] M. L. McHugh, "Interrater reliability: the kappa statistic," *Biochem. medica*, vol. 22, no. 3, pp. 276–282, 2012.
- [29] A. M. Sutherland, "The changing pattern of tuberculosis of the female genital tract. A thirty year survey," *Archives of Gynecol*ogy, vol. 234, no. 2, pp. 95–101, 1983.
- [30] G. Goel, R. Khatuja, G. Radhakrishnan, R. Agarwal, S. Agarwal, and I. Kaur, "Role of newer methods of diagnosing genital tuberculosis in infertile women," *Indian Journal of Pathology & Microbiology*, vol. 56, no. 2, pp. 155–157, 2013.
- [31] G. Shrivastava, T. Bajpai, G. S. Bhatambare, and K. B. Patel, "Genital tuberculosis: comparative study of the diagnostic modalities," *J. Hum. Reprod. Sci.*, vol. 7, no. 1, pp. 30–33, 2014.
- [32] R. Figueroa-Damian, I. Martinez-Velazco, R. Villagrana-Zesati, and J. L. Arredondo-Garcia, "Tuberculosis of the female reproductive tract: effect on function," *International Journal of Fertility and Menopausal Studies*, vol. 41, no. 4, pp. 430–436, 1996.
- [33] P. D. Hoeprich and M. C. Jordan, *Infectious diseasesa modern* treatise of infectious processes, 1989.
- [34] G. B. Chavhan, P. Hira, K. Rathod et al., "Female genital tuberculosis: hysterosalpingographic appearances," *The British Journal of Radiology*, vol. 77, no. 914, pp. 164–169, 2004.
- [35] N. Malhotra, U. B. Singh, V. Iyer, P. Gupta, and N. Chandhiok, "Role of laparoscopy in the diagnosis of genital TB in infertile females in the era of molecular tests," *Journal of Minimally Invasive Gynecology*, vol. 27, no. 7, pp. 1538–1544, 2020.