Research Article

Serum Exosomal lncRNA AC007099.1 Regulates the Expression of Neuropeptide-Related FAP, as a Potential Biomarker for Hepatocarcinogenesis

Luqian Li, Fei Xiao, Guolian Liu, and Yufeng Chen

Department of Clinical Laboratory Medicine Maoming People's Hospital, China

Correspondence should be addressed to Yufeng Chen; yufengcheng@inu.edu.vn

Received 12 November 2021; Revised 24 December 2021; Accepted 8 January 2022; Published 10 February 2022

Academic Editor: Yuzhen Xu

Copyright © 2022 Luqian Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Neuropeptide-associated fibroblast activation protein (FAP) may be an important risk factor for neurovascular metastasis in hepatocellular carcinoma. Analysis of The Cancer Genome Atlas (TCGA) database showed that *FAP* mRNA was highly expressed in most human tumor tissues. The HPA database then verified that *FAP* was highly expressed in tumor tissues following protein translation. Survival analysis then showed that the level of *FAP* expression significantly affected the overall survival (OS), progress free interval (PFI), and disease specific survival (DSS) of patients with hepatocellular carcinoma. A high expression of *FAP* in tumor tissue is associated with poor patient prognosis. According to the results of spearman correlation, AC009099 and *FAP* were negatively correlated with miR-7152 expression, while AC009099 and *FAP* expression were positively correlated. The lncRNA AC007099.1, which may serve as a potential target for the treatment of hepatocellular carcinoma, was associated with liver cancer. AC007099.1/miR-7152/*FAP* was found to be associated with immune infiltration in patients with hepatocellular carcinoma. Enrichment analysis suggests that the AC009099/miR-7152/*FAP* ceRNA regulatory network is associated with neuropeptide functional pathways. In conclusion, a neuropeptide-related AC009099/miR-7152/*FAP* ceRNA regulatory network was constructed in this study.

1. Introduction

Liver cancer is one of the deadliest cancers in the world, and 70% to 90% of primary liver cancers are hepatocellular carcinomas. The incidence of the disease is increasing annually, with a higher prevalence in men than in women [1, 2]. At present, there is still a lack of effective treatments for hepatocellular carcinoma, except for physical and chemical treatments. These treatments include radiation, ablation, surgical resection, transplantation, and transcatheter arterial chemoembolization, while only a few expensive drugs have little effect [3, 4]. However, the specific mechanisms underlying the development of hepatocellular carcinoma have not been elucidated and need to be further explored [5].

Tumor microenvironment helps cancer cells evade immune surveillance and supports their proliferation, metastasis, and dissemination [6–8]. The heterogeneity of hepatocellular carcinoma cells then causes drug resistance

in hepatocellular carcinoma, which serves as a major challenge in treating hepatocellular carcinoma. The cancer microenvironment is generally stable and rarely progresses to drug-resistant subtypes. Therefore, it is much more favorable to select a component of the microenvironment as an antitumor target than a drug-resistant cancer cell.

Neuropeptides are emerging as therapeutic targets for the prevention of tumor-induced neurogenesis and immune escape [9]. Some studies have shown that various neuropeptides change tumor microenvironment during tumorigenesis, progression, and prognosis [10–12]. Fibroblast activation protein (FAP) is a dipeptidyl peptidase (DPP) and endopeptidase that is weakly expressed in normal adult human tissues but is greatly upregulated in activated mesenchymal cells of tumors [13]. A number of neuropeptides including Neuropeptide Y, B-type natriuretic peptide, substance P, and peptide YY have been found to be hydrolytic substrates of FAP [13]. Neuropeptide-related FAP has a

FIGURE 1: Upregulation and characterization of *FAP* expression in tumors. (a) Characterization of *FAP* expression in pancypotopenia. (b) Validation of *FAP* expression at the translational level by the HPA database (immunohistochemistry). (c) *FAP* copy number differences in TCGA-LIHC. (d–f) The correlation of survival time (OS) (d), progression-free survival (PFS) (e), and disease-specific survival (DSS) (f) with *FAP* expression was analyzed by Kaplan-Meier method for plotting survival curves, respectively. (g) Differences in *FAP* promoter methylation in normal and tumor groups of TCGA-LIHC. (h) cBioPortal-based demonstration of genomic alterations of *FAP* in TCGA-LIHC.

low expression in normal tissues but is elevated in inflammatory conditions [13]. Multiple studies have shown that *FAP* expression level is elevated in esophageal, colorectal, pancreatic, and breast cancers and has a role in promoting cancer progression [14]. We speculate that *FAP* may be an important risk factor for neurovascular metastasis of hepatoma.

Recent studies have revealed that neuropeptides in a variety of tumor systems may be regulated by exosomes and that abnormalities in such regulation can affect neuroendocrine function [15–17]. Exosomes are membrane-bound vesicles that can transport biologically active molecules between cells [18]. In addition to regulating the microenvironment between cells and the immune system, exosomes also exchange and transmit genetic material in the intracellular space through “transport” [19–21]. In addition, exosomes are involved in the metastasis of tumor cells, angiogenesis, and the transformation of normal cells into tumor cells [22]. Tumor-derived exosomes contain many serological markers associated with cancer, such as miRNAs that can be used to detect early HCC. Exosomes secrete miRNAs and other substances that regulate the growth and metastasis of HCC. For example, the miRNA-21 secreted by HCC cells activates PDK1/AKT signalling in HSC, transforming HSC cells into CAFs. Other factors such as TGF- β further contribute to the development of cancer [23]. Furthermore, the binding of Toll-like receptors (TLRs) in immune cells to exosomes containing miRNA-21 and miR-29a activates the NF- κ B pathway in TLRs. This signaling cascade leads to the

secretion of a series of inflammatory factors that promote tumor growth and metastasis [24]. We propose a hypothesis that neuropeptide *FAP* in HCC may be regulated by exosomes in the tumor microenvironment.

The development of microarray and RNA sequencing technologies has facilitated further research into RNA and DNA. Such technologies have become an important part of biomedical research [25, 26]. The identification of liver cancer-specific molecular biomarkers further revealed the potential molecular mechanism of liver cancer, which is important for the early diagnosis, prognosis monitoring, and development of new molecular targeting drugs for this disease. In this study, differentially expressed genes (DEGs) of HCC were identified from the TCGA database. The functions and key genes obtained were then systematically analyzed as potential biomarkers during the development and prognosis of hepatocellular carcinoma.

2. Materials and Methods

2.1. Data Sources and Treatment of Differential Genes. The transcriptome sequencing data of the TCGA liver hepatocellular carcinoma (LIHC) dataset and the corresponding clinical data were downloaded from The Cancer Genome Atlas (TCGA) database [27]. The downloaded gene expression data and clinical data files were decompressed into text files, and the data was organized into processable expression matrix data using the R software program (version 3.6.3). The human gene annotation file was downloaded from the Ensembl website (<http://asia.ensembl.org/index>) and

FIGURE 2: Continued.

FIGURE 2: lncRNA AC007099.1 exosomes were defined as potential contributors to the upregulation of *FAP* expression in hepatocellular carcinoma. (a) A map of the distribution of all exosomes in the exoRBase liver cancer dataset. (b) Volcano map of differentially expressed mRNAs in healthy controls and liver cancer groups. (c) Volcano map of differentially expressed lncRNAs in healthy controls and liver cancer groups. (d) One of the 2549 upregulated lncRNAs in the TCGA-LIHC dataset was cooccurring in the exoRBase liver cancer exosome database ($\log_2 \text{FC} > 1$). (e) The 1333 potential target genes of AC007099.1 identified in the miRWalk dataset and 12 coregulated genes among 13 downregulated miRNAs in the TCGA-LIHC dataset ($\log_2 \text{FC} < -1$). (f) Constructed functional model of AC007099.1/hsa-miR-7152/*FAP* regulation.

converted to Gene ID in R for further analysis. Moreover, the Human Protein Atlas (HPA) (<https://www.proteinatlas.org/>) was used to observe and compare the expression levels of the identified key genes in normal and liver cancer tissues. LNCipedia was used to obtain the sequence of the lncRNA [28]. lncLocator was used to obtain subcellular localization of target lncRNAs [29].

2.2. Construction of Exosome-Neuropeptide ceRNA Network for Hepatocellular Carcinoma. DIANA-LncBase (<http://www.microrna.gr/LncBase>) is a database dedicated to documenting lncRNA-miRNA interactions. It includes all experimentally validated lncRNA-miRNA interactions in the literature. The DIANA-LncBase web tool (<http://www.microrna.gr/LncBase>) was used to predict potential target miRNAs for exosomal lncRNAs [30]. miRWalk is a comprehensive database of miRNA target genes (<http://mirwalk.umm.uni-heidelberg.de/>). miRWalk was used to predict potential miRNAs that might bind *FAP*. It not only records miRNA binding sites on the full-length sequences of genes but also combines them with predictions from 12 existing miRNA target prediction programs (DIANA-microTv4.0, DIANA-microT-CDS, miRanda-rel2010, mirBridge, miRDB4.0, miRmap, miRNAmap, doRiNA i.e., PicTar2, PITA RNA22v2, RNAhybrid2.1, and

TargetsCan6.2) to predict the binding information set for binding association [31].

2.3. Gene Set Enrichment Analysis (GSEA). GSEA was used to predict the pathway of *FAP* enrichment in hepatocellular carcinoma by GSEA [32, 33]. Patients with hepatocellular carcinoma were divided into high- and low-expression groups according to the median value of *FAP* expression. The c2.cp.kegg.v7.1.symbols.gmt functional dataset from MsigDB (The Molecular Signatures Database) was used as the reference gene set, and the normalized microarray matrix was analyzed. The number of random combinations was set to 1000.

2.4. Methylation and Expression Analysis. DNA methylation is an epigenetic difference. The human disease methylation databases DiseaseMeth (<http://bio-bigdata.hrbmu.edu.cn/diseasemeth/>), UALCAN (<http://ualcan.path.uab.edu/>), and MEXPRESS (<https://mexpress.be>) were used to assess the correlation between the methylation levels and the expression of *FAP* in hepatocellular carcinoma and normal tissues adjacent to the carcinoma.

2.5. Immune Cell Infiltration Analysis. Analysis of RNA-seq data from different subgroups of patients with liver cancer using the CIBERSORT algorithm was performed to

FIGURE 3: Correlation analysis of AC007099.1/miR-7152/FAP. (a) Expression pattern of lncRNA AC009099 distribution in subcellular. (b) Heat map of coexpressed genes of lncRNA AC009099, MIR7152, and FAP. (c, d) Correlation of lncRNA AC009099, MIR7152, and FAP was demonstrated using Person correlation analysis.

determine the relative proportions of 22 immune infiltrating cells as previous research [34, 35]. Spearman correlation analysis was performed for gene expression and immune cell content. A p value < 0.05 was considered a statistically significant difference.

2.6. Survival and Statistical Analysis. The results were statistically analyzed using the R software (version 3.6.0). Quantitative data were expressed as mean $\pm$ standard deviation, and a t -test was used for between-group analysis. Differences in the expression of the characteristic factors in the different groups were assessed using t -tests. Survival curves were plotted using the Kaplan-Meier method, and the overall survival time (OS), progression-free survival time (PFS), and disease-specific survival rate (DSS) were analyzed.

3. Results

3.1. Upregulation and Characterization of FAP Expression in Tumors. TCGA database analysis shows that the FAP gene is

highly expressed in most human tumor tissues (Figure 1(a)). The HAP database verified the expression of FAP at the translational level and found that the protein was highly expressed in tumor tissues (Figure 1(b)). There was a significant difference in the FAP copy number profile in TCGA-LIHC (Figure 1(c)). Results of the survival analysis showed that the expression levels of the FAP gene had a significant effect on the OS, PFS, and DSS of hepatocellular carcinoma patients. The prognostic indicators of survival, such as OS, PFS, and DSS, were worse in the group with high expression of the FAP gene, comparatively ($p < 0.05$) (Figures 1(d)–1(f)). Methylation analysis revealed that the FAP promoter was less methylated in the TCGA-LIHC, but the difference is not statistically significant (Figure 1(g)). Based on these results of cBioPortal, genomic alterations of FAP in TCGA-LIHC are shown in Figure 1(h). The mutation rate of FAP was only 0.8%, suggesting that mutations were not the main cause of the FAP expression changes. In conclusion, high expression of FAP in UVM tumor tissues was associated with poor

FIGURE 4: Immunological correlation analysis of AC007099.1/miR-7152/FAP and patients with hepatocellular carcinoma. (a) Expression of AC007099.1 in hepatocellular carcinoma correlates with immune cell infiltration. (b) Expression of miR-7152 in hepatocellular carcinoma correlates with immune cell infiltration. (c) Expression of FAP in hepatocellular carcinoma correlates with immune cell infiltration.

patient prognosis. Reduced promoter methylation in the FAP genome may be responsible for the upregulation of FAP expression.

3.2. *lncRNA AC007099.1 Exosomes Were Defined as Potential Contributors to the Upregulation of FAP Expression in Hepatocellular Carcinoma.* Based on the

FIGURE 5: Continued.

FIGURE 5: AC007099.1/miR-7152/FAP is associated with the prognosis of patients with hepatocellular carcinoma. (a–c) Relationship between lncRNA AC009099 and overall survival time (OS)/progression-free survival time (PFS)/disease-specific survival rate (DSS). (d) Histogram of the relationship between miR-7152-5p expression and hepatocarcinogenesis. (e) Expression of lncRNA AC009099 in hepatocellular carcinoma and normal tissues adjacent to the carcinoma. (f) Expression of miR-7152-5p in hepatocellular carcinoma and normal tissues adjacent to the carcinoma. (g) ROC curves were used to demonstrate the discriminatory ability of AC0070099, FAP, and miR7152 against hepatocellular carcinoma.

exoRBase dataset, all detected lncRNAs in liver cancer exosomes are shown in Figure 2(a). Figure 2(a) shows that lncRNA AC007099.1 is mainly expressed in bile, suggesting that lncRNA AC007099.1 may be associated with the process of bile secretion. Volcano plots of differentially expressed mRNA and lncRNA in healthy control and liver cancer groups are shown in Figures 2(b) and 2(c). AC007099 was also present among the 2549 upregulated lncRNAs in the TCGA-LIHC datasets of the exoRBase liver cancer exosome database (Figure 2(d)). The 1333 potential target genes of AC007099 identified in the miRWalk dataset and the 13 downregulated miRNAs in the TCGA-LIHC

dataset were intersected to obtain 12 common downregulated genes ($\log_2\text{FC} < -1$) (Figure 2(e)). Among them, miR-7152 was regulated by AC007099. Accordingly, we constructed a model of the AC007099/hsa-miR-7152/FAP regulatory network (Figure 2(f)).

3.3. Correlation Analysis and Prognostic Analysis of AC007099.1/miR-7152/FAP. The expression pattern of lncRNA AC009099.1 in the subcellular distribution suggested that it was enriched in the cytoplasm, nucleus, cytosol, and exosome (Figure 3(a)). As predicted, AC009099.1 and FAP were negatively correlated with miR-

FIGURE 6: GSEA: (a) GSEA of FAP in hepatocellular carcinoma; (b) GSEA of miR-7152 gene in hepatocellular carcinoma; (c) GSEA of liver cancer lncRNA AC009099.

7152 expression. AC009099.1 was positively correlated with the FAP expression (Figures 3(b)–3(d)). High expression of AC009099.1 was correlated with good OS and DSS, however, not with PFI (Figures 3(a)–3(c)). Hsa-miR-7152 and AC009099.1 were highly expressed in normal and tumor tissues, respectively (Figures 3(d)–3(f)). The ROC curves suggest that miR-7152 (AUC = 0.568), FAP (AUC = 0.813), and AC007099 (AUC = 0.732) are good predictors of hepatocarcinogenesis (Figure 3(g)).

3.4. AC007099.1/miR-7152/FAP Is Associated with Immune Infiltration in Patients with Hepatocellular Carcinoma. The AC007099.1/miR-7152/FAP pathway was found to be associated with immune infiltrations in patients with hepatocellular carcinoma. High expression of AC007099.1 was positively correlated with the infiltration of CD56 bright NK cells, iDC, and T helper cells. It was negatively correlated with the infiltration of Treg, CD56 dim NK cells, and Tgd (Figure 4(a)). High expression of miR-7152 was positively

correlated with the infiltration of Th17 cells. It was negatively correlated with the infiltration of Macrophages, Th1 cells, B cells, DC, Tem, and T cells (Figure 4(b)). The high expression of FAP was positively correlated with the infiltration of most immune cells (Figure 4(c)). Therefore, the potential AC007099.1/miR-7152/FAP pathway was also found to be associated with immune infiltration in patients with hepatocellular carcinoma (Figure 5).

3.5. Enrichment Analysis of AC007099.1/miR-7152/FAP. AC007099.1, miR-7152, and FAP were investigated for potential specific signaling pathways and potential molecular mechanisms (NOM $p < 0.05$, FDR $q < 0.05$). Normalized enrichment scores (NES) were calculated. GSEA results showed that AC007099.1 was enriched in GPCR ligand binding and class-A/1 rhodopsin-like receptors (Figure 6(a)). miR-7152 was enriched in G-alphaI signaling events (Figure 6(b)). FAP was enriched in signaling by receptor tyrosine kinase and G-alphaI signaling events (Figure 6(c)). The above results suggest that AC007099.1, miR-7152, and FAP may be associated with neuropeptide-related functional pathways.

4. Discussion

FAP was specifically expressed in the liver, and its low expression levels in hepatocellular carcinoma may be implicated in the poor prognosis of the disease. The AC009099/miR-7152/FAP ceRNA regulatory network was constructed in this study. The protein encoded by the human FAP gene is a single-channel transmembrane protein containing 760 amino acids, a short cytoplasmic amino-terminal (6 amino acids), a transmembrane region (20 amino acids), and a large extracellular domain [6, 7]. Previous studies have shown that FAP plays an important role in predicting tumor aggressiveness. Its frequent expression in malignant tumors suggests that FAP-targeted therapy may be a very attractive strategy [36]. Currently, much research has been devoted to FAP as a protease in the tumor microenvironment. Since FAP is expressed in most epithelial cancers and sarcomas, it has a wide range of applications in cancer [37, 38]. The survival analysis results showed that FAP gene expression levels significantly affected the OS, PFS, and DSS in patients with hepatocellular carcinoma. An observed high expression of FAP in UVM tumor tissues was associated with poor patient prognosis and reduced promoter methylation in the FAP gene. FAP hydrolyses a variety of neuropeptides and does serious work in maintaining normal neuroendocrine function [39]. The results of the enrichment analysis suggest that AC007099.1, miR-7152, and FAP may be associated with neuropeptide-related functional pathways. Thus, the AC009099/miR-7152/FAP ceRNA regulatory network is thought to be functionally relevant to neuropeptides.

The lncRNA AC007099.1 was found to be differentially expressed in hepatocellular carcinoma exosomes. miRNA is an important risk marker for HCC and liver injury [40, 41]. When lncRNAs and mRNAs have the same microRNA response element (MRE), there is competition between them

for the same kind of miRNA. In other words, the level of lncRNA expression in the cell directly affects the number of miRNAs that can be bound by the corresponding mRNAs; that is, lncRNAs indirectly regulate the expression level of mRNAs through the bridge of MRE and thus regulate cellular functions. miR-7152 was found to be possibly regulated by AC009099. AC009099 and FAP expression levels were negatively correlated with miR-7152 expression, while AC009099 and FAP expression were positively correlated [42]. AC007099.1 and FAP were highly expressed while miR-7152 expression was decreased in hepatocellular carcinoma. Therefore, an AC009099/miR-7152/FAP ceRNA regulatory network was constructed in this study. Moreover, AC009099, miR-7152, and FAP were all found to be associated with infiltration of numerous immune cells in this study. The present study revealed that the expression of AC007099.1, miR-7152, and FAP in hepatocellular carcinoma could be used as a novel independent prognostic marker based on their observed roles during poor prognosis.

Based on their expression levels, potential targets for hepatocellular carcinoma treatment may include AC007099, FAP, and miR-7152. AC007099.1 and FAP were highly expressed, while miR-7152 expression was reduced in hepatocellular carcinoma. Hence, AC007099.1 may regulate the expression of miR-7152 through adsorption, thus promoting the proliferation, migration, and invasion of hepatocellular carcinoma cells. However, the bioinformatics analysis was only based on the TCGA database, which has some limitations. These hepatocellular carcinoma-related signaling pathways and key genes need further validation by various tests using molecular biological techniques and experimental data.

5. Conclusions

In conclusion, exploring the expression, regulatory mechanism, and prognostic value of FAP in hepatocellular carcinoma will be beneficial to the clinical treatment of patients with hepatocellular carcinoma and provide a new set of potential molecular targets for the treatment of hepatocellular carcinoma. In this study, a neuropeptide-related AC009099/miR-7152/FAP ceRNA regulatory network was constructed.

Abbreviations

TCGA: The Cancer Genome Atlas
 FAP: Fibroblast activation protein
 HPA: The Human Protein Atlas
 OS: Overall survival
 PFI: Progress free interval
 DSS: Disease specific survival
 HCC: Hepatocellular carcinomas
 MRE: MicroRNA response element.

Data Availability

The transcriptome sequencing data of the TCGA-LIHC dataset and the corresponding clinical data were

downloaded from The Cancer Genome Atlas (TCGA) database. The human gene annotation file was downloaded from the Ensemble website (<http://asia.ensembl.org/index>).

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Luqian Li and Fei Xiao contributed equally to this work.

Acknowledgments

We thank Bullet Edits Limited for the linguistic editing and proofreading of the manuscript. The work is supported by the High-level Hospital Construction Research Project of Maoming People's Hospital.

References

- [1] I. Takayanagi and Y. Kasuya, "Effects of some drugs on the circular muscle of the isolated lower oesophagus," *The Journal of Pharmacy and Pharmacology*, vol. 29, no. 9, pp. 559-560, 1977.
- [2] A. Jemal, K. D. Miller, J. Ma et al., "Higher lung cancer incidence in young women than young men in the United States," *The New England Journal of Medicine*, vol. 378, pp. 1999-2009, 2018.
- [3] A. Singh, S. Shafi, T. Upadhyay, A. K. Najmi, K. Kohli, and F. H. Pottoo, "Insights into nanotherapeutic strategies as an impending approach to liver cancer treatment," *Current Topics in Medicinal Chemistry*, vol. 20, pp. 1839-1854, 2020.
- [4] J. Chen, X. Song, H. Zhang, Y. Qu, and J. Miao, "Sophorolipid produced from the new yeast strain *Wickerhamiella domercqiae* induces apoptosis in H7402 human liver cancer cells," *Applied Microbiology and Biotechnology*, vol. 72, pp. 52-59, 2006.
- [5] H. Yang, F. Xiong, X. Wei, Y. Yang, M. N. MA, and R. Zhou, "Overexpression of LAPTM4B-35 promotes growth and metastasis of hepatocellular carcinoma in vitro and in vivo," *Cancer Letters*, vol. 294, pp. 236-244, 2010.
- [6] M. J. Scanlan, B. K. Raj, B. Calvo et al., "Molecular cloning of fibroblast activation protein alpha, a member of the serine protease family selectively expressed in stromal fibroblasts of epithelial cancers," *Proceedings of the National Academy of Sciences*, vol. 91, pp. 5657-5661, 1994.
- [7] L. A. Goldstein, G. Ghersi, M. L. Pineiro-Sanchez et al., "Molecular cloning of seprase: a serine integral membrane protease from human melanoma," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1361, no. 1, pp. 11-19, 1997.
- [8] H. Qian, Y. Zhang, B. Wu et al., "Structure and function of HECT E3 ubiquitin ligases and their role in oxidative stress," *Journal of Translational Internal Medicine*, vol. 8, pp. 71-79, 2020.
- [9] R. D. Cervantes-Villagrana, D. Albores-García, A. R. Cervantes-Villagrana, and S. J. García-Acevez, "Tumor-induced neurogenesis and immune evasion as targets of innovative anti-cancer therapies," *Signal Transduction and Targeted Therapy*, vol. 5, p. 99, 2020.
- [10] M. Körner, B. Waser, and J. C. Reubi, "Neuropeptide Y receptors in renal cell carcinomas and nephroblastomas," *International Journal of Cancer*, vol. 115, pp. 734-741, 2005.
- [11] L. J. Helman, P. S. Cohen, S. D. Averbuch, M. J. Cooper, H. R. Keiser, and M. A. Israel, "Neuropeptide Y expression distinguishes malignant from benign pheochromocytoma," *Journal of Clinical Oncology*, vol. 7, pp. 1720-1725, 1989.
- [12] A. Kowalczyk, A. Wiecek, E. Franek, and F. Kokot, "Plasma concentration of leptin, neuropeptide Y and tumor necrosis factor alpha in patients with cancers, before and after radio- and chemotherapy," *Polskie Archiwum Medycyny Wewnętrznej*, vol. 106, pp. 657-668, 2001.
- [13] P. F. Wong, M. G. Gall, W. W. Bachovchin, M. C. GW, F. M. Keane, and M. D. Gorrell, "Neuropeptide Y is a physiological substrate of fibroblast activation protein: enzyme kinetics in blood plasma and expression of Y2R and Y5R in human liver cirrhosis and hepatocellular carcinoma," *Peptides*, vol. 75, pp. 80-95, 2016.
- [14] A. Kasprzak and A. Adamek, "The neuropeptide system and colorectal cancer liver metastases: mechanisms and management," *International Journal of Molecular Sciences*, vol. 21, no. 10, p. 3494, 2020.
- [15] N. Christou, S. Blondy, V. David et al., "Neurotensin pathway in digestive cancers and clinical applications: an overview," *Cell Death & Disease*, vol. 11, p. 1027, 2020.
- [16] D. M. Stranford, M. E. Hung, E. S. Gargus, R. N. Shah, and J. N. Leonard, "A systematic evaluation of factors affecting extracellular vesicle uptake by breast cancer cells," *Tissue Engineering. Part A*, vol. 23, pp. 1274-1282, 2017.
- [17] L. Muñoz-Moreno, M. J. Carmena, A. V. Schally, J. C. Prieto, and A. M. Bajo, "Stimulation of neuroendocrine differentiation in prostate cancer cells by GHRH and its blockade by GHRH antagonists," *Investigational New Drugs*, vol. 38, pp. 746-754, 2020.
- [18] Q. Han, H. Zhao, Y. Jiang, C. Yin, and J. Zhang, "HCC-derived exosomes: critical player and target for cancer immune escape," *Cell*, vol. 8, p. 558, 2019.
- [19] C. Roma-Rodrigues, L. R. Raposo, R. Cabral, F. Paradinha, P. V. Baptista, and A. R. Fernandes, "Tumor microenvironment modulation via gold nanoparticles targeting malicious exosomes: implications for cancer diagnostics and therapy," *International Journal of Molecular Sciences*, vol. 18, no. 1, p. 162, 2017.
- [20] G. Chen, A. C. Huang, W. Zhang et al., "Exosomal PD-L1 contributes to immunosuppression and is associated with anti-PD-1 response," *Nature*, vol. 560, pp. 382-386, 2018.
- [21] S. Li, Z. Lin, X. Jiang, and X. Yu, "Exosomal cargo-loading and synthetic exosome-mimics as potential therapeutic tools," *Acta Pharmacologica Sinica*, vol. 39, pp. 542-551, 2018.
- [22] X. Li and X. Zeng, "Shikonin suppresses progression and epithelial-mesenchymal transition in hepatocellular carcinoma (HCC) cells by modulating miR-106b/SMAD7/TGF- β signaling pathway," *Cell Biology International*, vol. 44, pp. 467-476, 2020.
- [23] Y. Zhou, H. Ren, B. Dai et al., "Hepatocellular carcinoma-derived exosomal miRNA-21 contributes to tumor progression by converting hepatocyte stellate cells to cancer-associated fibroblasts," *Journal of Experimental & Clinical Cancer Research*, vol. 37, p. 324, 2018.

- [24] X. Yu, M. Odenthal, and J. Fries, "Exosomes as miRNA carriers: formation–function–future," *International Journal of Molecular Sciences*, vol. 17, p. 2028, 2016.
- [25] S. C. Sealfon and T. T. Chu, "RNA and DNA Microarrays," in *Biological Microarrays* vol. 671, A. Khademhosseini, K.-Y. Suh, and M. Zourob, Eds., pp. 3–34, Humana Press, 2011.
- [26] R. Stark, M. Grzelak, and J. Hadfield, "RNA sequencing: the teenage years," *Nature Reviews. Genetics*, vol. 20, pp. 631–656, 2019.
- [27] K. Tomczak, P. Czerwińska, and M. Wizniewicz, "Review The Cancer Genome Atlas (TCGA): an immeasurable source of knowledge," *Contemporary Oncology*, vol. 19, no. 1A, pp. 68–77, 2015.
- [28] P. J. Volders, J. Anckaert, K. Verheggen et al., "LNCipedia 5: towards a reference set of human long non-coding RNAs," *Nucleic Acids Research*, vol. 47, pp. D135–D139, 2019.
- [29] Z. Cao, X. Pan, Y. Yang, Y. Huang, and H.-B. Shen, "The lncLocator: a subcellular localization predictor for long non-coding RNAs based on a stacked ensemble classifier," *Bioinformatics*, vol. 34, pp. 2185–2194, 2018.
- [30] M. D. Paraskevopoulou, I. S. Vlachos, D. Karagkouni et al., "DIANA-LncBase v2: indexing microRNA targets on non-coding transcripts," *Nucleic Acids Research*, vol. 44, pp. D231–D238, 2016.
- [31] C. Sticht, C. De La Torre, A. Parveen, and N. Gretz, "miRWalk: an online resource for prediction of microRNA binding sites," *PLoS One*, vol. 13, article e0206239, 2018.
- [32] S. Canzler and J. Hackermüller, "multiGSEA: a GSEA-based pathway enrichment analysis for multi-omics data," *BMC Bioinformatics*, vol. 21, p. 561, 2020.
- [33] Y. Chen, C. Chen, X. Xiao, Z. Huang, X. Huang, and W. Yao, "TNF- α Induces Neutrophil Apoptosis Delay and Promotes Intestinal Ischemia- Reperfusion-Induced Lung Injury through Activating JNK/FoxO3a Pathway," *Oxidative Medicine and Cellular Longevity*, vol. 2021, Article ID 8302831, 13 pages, 2021.
- [34] A. M. Newman, C. L. Liu, M. R. Green et al., "Robust enumeration of cell subsets from tissue expression profiles," *Nature Methods*, vol. 12, pp. 453–457, 2015.
- [35] X. Kang, Y. Chen, B. Yi et al., "An integrative microenvironment approach for laryngeal carcinoma: the role of immune/methylation/autophagy signatures on disease clinical prognosis and single-cell genotypes," *Journal of Cancer*, vol. 12, pp. 4148–4171, 2021.
- [36] P. Mhawech-Fauceglia, D. Wang, D. Samrao et al., "Clinical implications of marker expression of carcinoma-associated fibroblasts (CAFs) in patients with epithelial ovarian carcinoma after treatment with neoadjuvant chemotherapy," *Cancer Microenvironment*, vol. 7, pp. 33–39, 2014.
- [37] C. Egger, C. Cannel, C. Gérard et al., "Effects of the fibroblast activation protein inhibitor, PT100, in a murine model of pulmonary fibrosis," *European Journal of Pharmacology*, vol. 809, pp. 64–72, 2017.
- [38] Z. Wen, Q. Liu, J. Wu et al., "Fibroblast activation protein α -positive pancreatic stellate cells promote the migration and invasion of pancreatic cancer by CXCL1-mediated Akt phosphorylation," *Annals of Translational Medicine*, vol. 7, pp. 532–532, 2019.
- [39] F. M. Keane, N. A. Nadvi, T.-W. Yao, and M. D. Gorrell, "Neuropeptide Y, B-type natriuretic peptide, substance P and peptide YY are novel substrates of fibroblast activation protein- α ," *The FEBS Journal*, vol. 278, pp. 1316–1332, 2011.
- [40] X. Wang, Y. He, B. Mackowiak, and B. Gao, "MicroRNAs as regulators, biomarkers and therapeutic targets in liver diseases," *Gut*, vol. 70, pp. 784–795, 2021.
- [41] A. A. Mohamed, A. A. Omar, R. R. El-Awady et al., "MiR-155 and MiR-665 role as potential non-invasive biomarkers for hepatocellular carcinoma in Egyptian patients with chronic hepatitis C virus infection," *Journal of Translational Internal Medicine*, vol. 8, pp. 32–40, 2020.
- [42] K. Yata, L. B. Beder, S. Tamagawa et al., "MicroRNA expression profiles of cancer stem cells in head and neck squamous cell carcinoma," *International Journal of Oncology*, vol. 47, pp. 1249–1256, 2015.