

Research Article

Antiangiogenic Effect of Ethanol Extract of *Vigna angularis* via Inhibition of Phosphorylation of VEGFR2, Erk, and Akt

Oh Sung Kwon,¹ Myoung Seok Jeong,² Bonglee Kim,² and Sung-Hoon Kim²

¹Department of East West Medical Science, Graduate School of East West Medical Science, Kyung Hee University, Suwon 446-701, Republic of Korea

²College of Korean Medicine, Kyung Hee University, Seoul 130-701, Republic of Korea

Correspondence should be addressed to Sung-Hoon Kim; sungkim7@khu.ac.kr

Received 19 June 2015; Revised 3 August 2015; Accepted 4 August 2015

Academic Editor: Shun-Wan Chan

Copyright © 2015 Oh Sung Kwon et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Though dietary azuki bean (*Vigna angularis*) seed containing antioxidant proanthocyanidins was known to have multibiological activities including antioxidant, hypotensive, anti-inflammatory, and immunomodulatory activities, the antiangiogenic activity of ethanol extract of *Vigna angularis* (EVA) was never reported so far. In the present study, the antiangiogenic mechanism of EVA was examined in human umbilical vein endothelial cells (HUVECs). EVA showed weak cytotoxicity in HUVECs, while it significantly suppressed the VEGF induced proliferation of HUVECs. Consistently, wound healing assay revealed that EVA inhibited the VEGF induced migration of HUVECs. Also, EVA abrogated the VEGF induced tube formation of HUVECs in a concentration dependent fashion. Furthermore, Matrigel plug assay showed that EVA significantly reduced the hemoglobin level of Matrigel plug in mice compared to untreated control. Of note, EVA effectively attenuated the phosphorylation of VEGFR2, Erk, and Akt in VEGF-treated HUVECs. Overall, our findings suggest that EVA inhibits angiogenesis in VEGF-treated HUVECs via inhibition of phosphorylation of VEGFR2, ERK, and Akt.

1. Introduction

Angiogenesis is well known as a phenomenon that makes new blood vessels from the preexisting vasculature [1]. The aberrant angiogenesis has role in a variety of diseases such as cancer [2, 3], psoriasis [4], arthritis [5], obesity [6], endometriosis [7], atherosclerosis [8], hypertension [9], diabetic retinopathy [10], and age-related macular degeneration [11]. Many of angiogenesis inhibitors have been developed for cancer treatment and prevention [12, 13], since Judah Folkman first claimed the critical role of angiogenesis for tumor growth [14].

Vascular endothelial growth factor (VEGF) binds to VEGF receptors (VEGFR), VEGFR1, VEGFR2, and VEGFR3, during angiogenesis [15]. Among these, VEGFR2 (KDR/Flk-1) has most important role in angiogenesis, including transduction of angiogenic signals after binding VEGF [16]. VEGFR2 became dimerized after the binding [17], promoting the activation of extracellular signal-regulated kinases

(ERKs), phosphatidylinositol 3-kinase (PI3K), and Akt [18–20].

Recently angiogenesis inhibitors [21–24] from natural products are attractive, though not powerful, with less side effects such as fetal development, bleeding, blood clot, hypertension, and protein in urine [25, 26]. Our team also reported potent antiangiogenic materials from natural products, such as STB-HO [27], beta-sitosterol [28], cryptotanshinone [29], and emodin [30].

Vigna angularis that has been usually used as dietary azuki bean was known to have antioxidative [31], hepatoprotective [32], antimicrobial [33], osteogenic [34], anti-inflammatory [9, 35], hypotensive [36, 37], antimetastatic [38], and immunomodulatory [39] activities. Until now, there is no evidence on the antiangiogenic activity of *Vigna angularis*. We studied the antiangiogenic mechanism of ethanol extract of *Vigna angularis* (EVA) in VEGF-treated human umbilical vein endothelial cells (HUVECs) using cytotoxicity assay,

proliferation assay, wound healing migration assay, tube formation assay, Matrigel plug assay, and Western blotting.

2. Materials and Methods

2.1. Preparation of Ethanol Extract of *Vigna angularis*. Azuki bean (*Vigna angularis*) was purchased from Hanil Herbal Company (Seoul, Republic of Korea), identified by Professor Namin Baek, Kyung Hee University, and stored at Cancer Preventive Material Development Research Center (CPMDRC), Kyung Hee University, Republic of Korea, with voucher specimen number (2012-05-07). Azuki bean (600 g) was soaked in 95% ethanol for one week at room temperature. The EtOH solution was then filtered, evaporated under reduced pressure, and lyophilized to obtain 14.87 g of ethanol extract of *Vigna angularis* (EVA yield = 2.47%). The extract was dissolved in DMSO.

2.2. Cell Culture. Human umbilical vein endothelial cells (HUVECs, passages between 4 and 6) were obtained from fresh human umbilical cord veins after collagenase treatment as described previously [22, 40]. The cells were maintained in M199 (Invitrogen, Carlsbad, CA) with 20% fetal bovine serum (FBS), 5 U/mL heparin and 3 ng/mL basic fibroblast growth factor (R&D Systems, Minneapolis, MN), and 100 U/mL of antibiotic-antimycotic in 0.1% gelatin coated flasks. The cells were cultured at 37°C in a humidified atmosphere containing 5% CO₂.

2.3. Cytotoxicity Assay. Cytotoxicity induced by EVA was evaluated in HUVECs by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay. The cells were seeded onto microplates (1 × 10⁴ cells/well) and treated with 0, 10, 20, 30, 40, and 50 µg/mL of EVA for 48 h. The cells were treated with MTT solution (1 mg/mL) for 2 h. DMSO was added to cells for 2 h. Optical density (OD) of the solution was measured using a microplate reader (Molecular Devices Co., Sunnyvale, CA) at 570 nm. Cell viability was calculated as a percentage of viable cells in EVA-treated group versus untreated control by the following equation. Cell viability (%) = [OD (EVA) – OD (Blank)]/[OD (Control) – OD (Blank)] × 100.

2.4. Proliferation Assay. Cell proliferation was determined using a 5-bromo-2'-deoxyuridine (BrdU) colorimetric assay kit (Roche, Mannheim, Germany). Briefly, HUVECs (5 × 10³ cells/well) were seeded into 0.1% gelatin coated microplates and incubated in a humidified atmosphere containing 5% CO₂ at 37°C. After starvation for 6 h in M199 containing 5% heat-inactivated FBS, the cells were treated with various concentrations of EVA (10, 20, and 30 µg/mL) in the presence or absence of VEGF (50 ng/mL) and incubated at 37°C for 48 h. 10 µL of BrdU (100 µg/mL) was added; the cells were incubated at 37°C for 6 h. The cells were fixed with anti-BrdU and measured by the substrate reaction. 25 µL of 1 M H₂SO₄ was added to stop the reaction and the absorbance was calculated by using an ELISA reader (Molecular Devices Co., USA) (450 nm with 690 nm correction).

2.5. Wound Healing Migration Assay. The migratory activity of HUVECs was tested by a wound healing assay. The cells (4 × 10⁵ cells/2 mL) were exposed to various concentrations of EVA (10 and 20 µg/mL) in a 6-well plate in the presence or absence of VEGF (50 ng/mL) and incubated in a humidified atmosphere containing 5% CO₂ at 37°C. The plates were scratched with a 200 µL pipet tip and washed with PBS for 3 times. Then, cells were treated with EVA extracts (0, 10, and 20 µg/mL) in media. The cells were fixed and stained with Diff-Quick after 24 h, and randomly chosen fields were photographed with a fluorescence microscope (AXIO observer A1, Zeiss, Germany). The migrated cells were calculated.

2.6. Tube Formation Assay. Tube formation assay was conducted as described previously [41]. 250 µL of Matrigel (Becton Dickinson Labware, Bedford, MA) was added to 24-well plates and incubated at 37°C for 1 h. HUVECs (1 × 10⁵ cells/well) were treated with EVA (0, 10, and 20 µg/mL) in the presence or absence of VEGF (50 ng/mL). And cells were treated with LY294002 (20 µM), PD98059 (50 µM), EVA (20 µg/mL), and VEGF (50 ng/mL). Randomly chosen fields were photographed with Axiovert S 100 light microscope (Carl Zeiss, USA) at 100x magnification after 18 h. Tube network was measured by using NIH Scion image program.

2.7. Matrigel Plug Assay. Six-week-old C57BL/6 mice (Chungang animal expt. Co., Seoul, Republic of Korea) were given subcutaneous injection of 0.5 mL of growth factor reduced Matrigel containing VEGF (50 ng/mL) and heparin (10 U/mL). EVA extracts (40 µg/mL in PBS) were orally administrated (300 µg/kg) after 7 days of Matrigel injection (three mice per group). Ten days later, the mice were sacrificed, and the matrigel plugs were detached. To indirectly quantify the formation of new blood vessel, the amount of hemoglobin (Hb) was calculated using the hemoglobin assay kit (YD Diagnostics, Republic of Korea). The concentration of Hb was measured from standard curve made from the known amount of Hb provided according to the manufacturer's protocol.

2.8. Western Blotting. After treatment of EVA (0, 10, or 20 µg/mL) in a presence of VEGF (50 ng/mL), HUVECs were lysed with RIPA buffer (50 mM Tris-HCl, 150 mM NaCl, 1% NP-40, 0.25% sodium deoxycholic acid, 1 M EDTA, 1 mM Na₃VO₄, 1 mM NaF, and protease inhibitors cocktail, pH 7.4). Samples were quantified by using a Bio-Rad DC protein assay kit II (Bio-Rad, Hercules, CA, USA), separated by electrophoresis on 8 to 15% SDS-PAGE gel and electrotransferred onto a Hybond ECL transfer membrane (Amersham Pharmacia, Piscataway, NJ, USA). After blocking with 3–5% nonfat skim milk, the membrane was incubated with antibodies for p-Akt, Akt, VEGFR2 (FLK1), p-Erk, and Erk (Cell Signaling Technology, Danvers, MA) and β-actin (Sigma Aldrich Co., St. Louis, MO). Blots were exposed to horseradish peroxidase- (HRP-) conjugated secondary anti-mouse or rabbit antibodies (AbDserotec, Kidlington, UK). Protein expression was measured by using enhanced

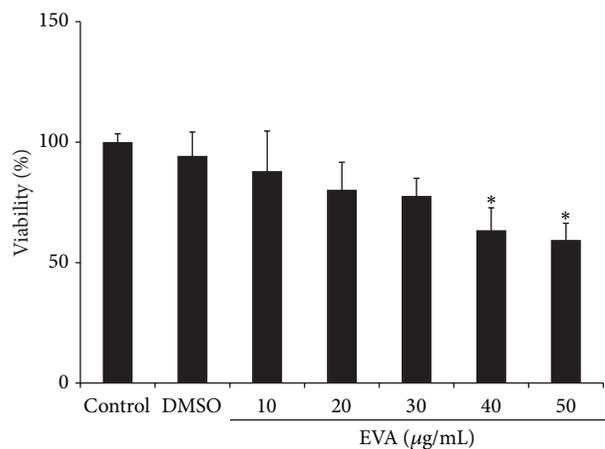


FIGURE 1: EVA exerts weak cytotoxicity against HUVECs. Cytotoxic effect of EVA was evaluated in HUVECs by MTT assay. The cells were seeded onto 96-well microplates at a density of 1×10^4 cells/well and treated with various concentrations of EVA (0, 10, 20, 30, 40, and 50 $\mu\text{g/mL}$) for 48 h. Data are presented as means \pm SD. * $p < 0.05$ versus DMSO-treated group.

chemiluminescence (ECL) system (Amersham Pharmacia, Piscataway, NJ, USA).

2.9. Statistical Analyses. All data were expressed as mean \pm SD (standard deviation). The statistically significant differences between untreated control and EVA-treated cells were calculated by ANOVA test followed by a post hoc analysis (Tukey t - or Dunnetts multiple comparison test) using Prism software 5 (GraphPad Software, Inc., San Diego, CA, USA).

3. Results

3.1. EVA Shows Weak Cytotoxicity in HUVECs and Suppressed VEGF Induced Proliferation of HUVECs. Cytotoxicity of EVA was evaluated in HUVECs using MTT assay. As shown in Figure 1, EVA decreased the viability of HUVECs to $\sim 60\%$ at the concentration of 50 μM for 48 h. EVA treatment showed cytotoxicity in VEGF-treated HUVECs in a concentration dependent manner (Figure 2).

3.2. EVA Significantly Inhibits the VEGF Induced Migration and Tube Formation in HUVECs. It was well known that endothelial cells migrate in tandem using adhesion molecules such as integrins and form loops to become a full-fledged vessel lumen like tube formation [42]. To check the physiological angiogenic formation, wound healing assay and tube formation assay were carried out in HUVECs.

As shown in Figures 3(a) and 3(b), EVA significantly reduced the number of tube forming endothelial cells, whereas tube-like formation by endothelial cells was enhanced by VEGF in HUVECs. Similarly, EVA significantly suppressed the VEGF induced migratory activity of HUVECs, while the gap physically made by tip was narrowed via repair through migratory activity of VEGF-treated HUVECs (Figures 4(a) and 4(b)).

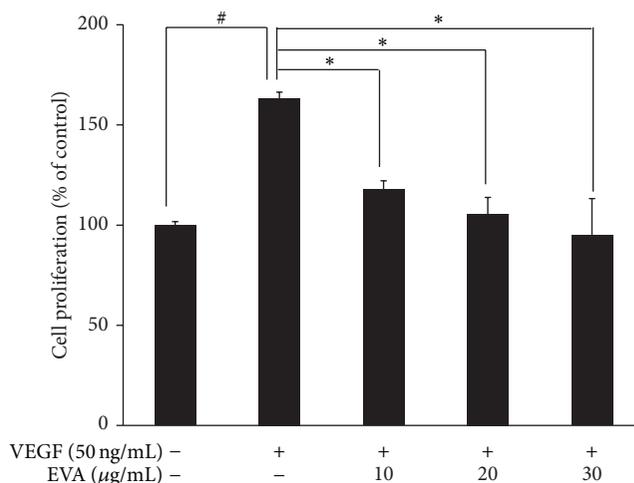


FIGURE 2: EVA inhibits VEGF induced proliferation in HUVECs. Cell proliferation was determined using a BrdU colorimetric assay kit. Cells (5×10^3 cells/well) were seeded onto 96-well plates and incubated for 24 h. After 6 h starvation, the cells were exposed to various concentrations of EVA (10, 20, and 30 $\mu\text{g/mL}$) in the presence or absence of VEGF (50 ng/mL) and incubated for 48 h at 37°C. Data are presented as means \pm SD. ** $p < 0.05$ versus VEGF only treated group.

3.3. EVA Significantly Reduces Hemoglobin Content in VEGF Induced Angiogenesis of Matrigel Plugs of C57BL/6 Mice. To confirm the antiangiogenic activity of EVA, Matrigel plug assay was carried out. EVA (300 $\mu\text{g/kg}$) extract was administered orally to C57BL/6 mice for 7 days before growth factor reduced Matrigel was subcutaneously injected into right flank of mice. As shown in Figures 5(a) and 5(b), EVA significantly reduced hemoglobin content in VEGF induced angiogenesis of Matrigel plugs of C57BL/6 mice, while dark red color was shown in Matrigel plugs of untreated control group.

3.4. EVA Effectively Attenuates the Phosphorylation of VEGFR2, Erk, and Akt in VEGF-Treated HUVECs. Since Dr. Judah Folkman suggested the importance of tumor angiogenesis, a variety of angiogenesis related factors such as VEGF, bFGF, PDGF, angiopoietin, and transforming growth factor- β (TGF- β) were studied for years [1, 43]. To elucidate antiangiogenic molecular mechanism of EVA, Western blot analysis was performed in VEGF-treated HUVECs. As shown in Figure 6, EVA effectively attenuated the phosphorylation of VEGFR2 and Erk and Akt in VEGF-treated HUVECs.

3.5. Akt, Erk Inhibitors, and EVA Attenuate Tube Formation Enhanced by VEGF in HUVECs. To investigate the roles of Erk and Akt in angiogenesis, tube formation assay was conducted with HUVECs in the presence of LY294002 (Akt inhibitor) or PD98059 (Erk inhibitor). As shown in Figures 7(a) and 7(b), Akt, Erk inhibitors, and EVA treatments inhibited tube formation enhanced by VEGF. Particularly, LY294002 and EVA significantly attenuated the tube formation.

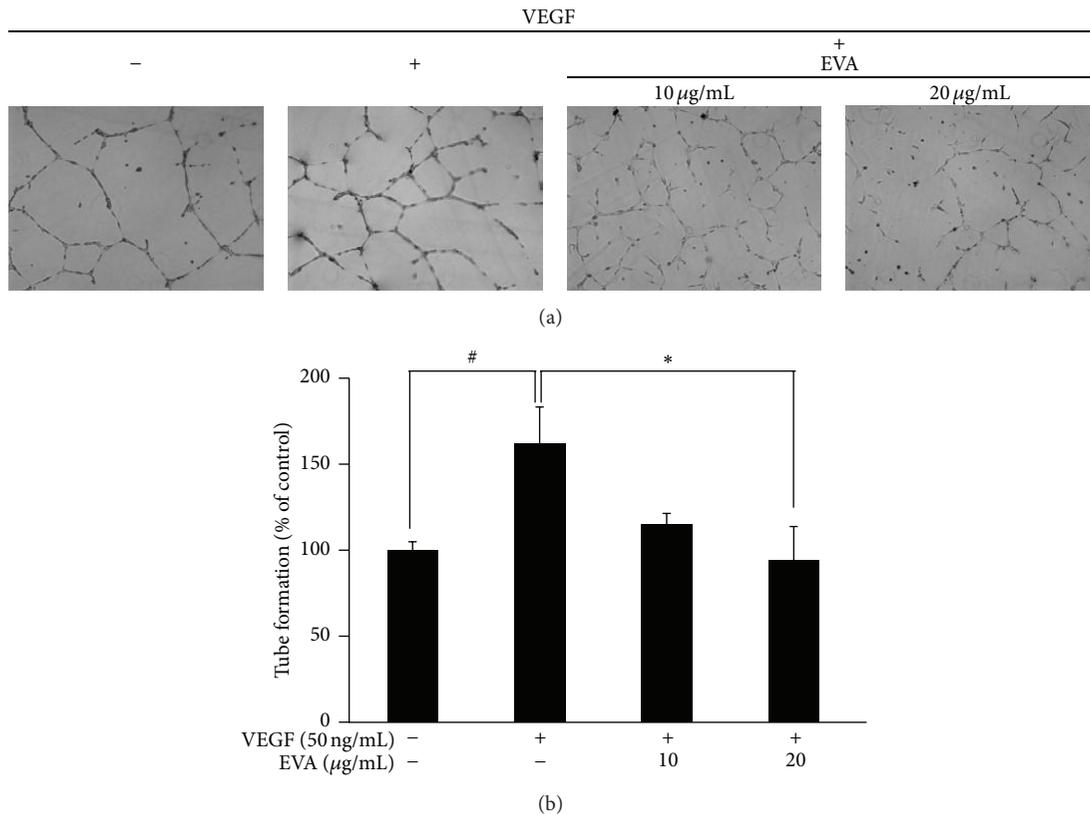


FIGURE 3: EVA abrogates VEGF induced tube formation in HUVECs. (a) Matrigel (250 μL) was added to 24-well plates and allowed to solidify for 1 h at 37°C. HUVECs (1×10^5 cells/well) were treated with 10 and 20 $\mu\text{g}/\text{mL}$ of EVA in the presence or absence of VEGF (50 ng/mL) and incubated at 37°C. After 18 h, randomly chosen fields were photographed under an Axiovert S 100 light microscope. (b) Bar graph represents the quantification of tube formation of untreated control or VEGF-treated control and EVA-treated groups. Data are presented as means \pm SD. $^{* \#} p < 0.05$ versus VEGF-treated group.

4. Discussion

Azuki bean (*Vigna angularis*) has been used in east Asian food, and its coat of seed was known to contain water-soluble anthocyanins, catechins, and flavonols [44]. Despite multibiological activities of azuki bean, its antiangiogenic activity was never reported until now. So, this study elucidates the antiangiogenic mechanism of azuki bean in VEGF-treated HUVECs.

Though EVA exerted weak cytotoxicity in HUVECs, EVA significantly suppressed the VEGF induced proliferation of HUVECs at nontoxic concentrations, implying that EVA has antiangiogenic potential by blocking pathological angiogenic process via VEGF induced proliferation of endothelial cells, since these endothelial cells become motile and invasive and protrude filopodia, which drives new vessel formation [14, 25, 42].

Also, EVA inhibited VEGF induced migration of HUVECs in wound healing assay and suppressed VEGF induced tube formation of HUVECs in a concentration dependent manner in tube formation assay, strongly indicating that EVA could inhibit the motility and tube formation of proliferative endothelial cells.

To confirm *in vivo* antiangiogenic activity of EVA, Matrigel plug assay was performed in C57BL6 mice orally administered by EVA extracts for 1 week earlier. Here, we observed less red color in Matrigel plugs isolated from mice treated by EVA, while black red color was shown in those of untreated control group. The treatment of EVA significantly reduced the hemoglobin level of Matrigel plug in mice compared to untreated control, demonstrating antiangiogenic activity of EVA. However, it requires further *in vivo* angiogenesis studies using Matrigel plug assay via direct EVA treatment in the increased number of mice or CAM assay in the near future.

It was well documented that several molecules were involved in angiogenesis process. Angiogenesis is processed via the interaction of the VEGF family of proangiogenic cytokines and their receptors. The VEGF family includes VEGFA, VEGFB, VEGFC, VEGFD, VEGFE, and placental growth factor (PGF) with three receptors including VEGFR1 (Fms-like tyrosine kinase 1/Flt-1), VEGFR2 (Flk-1/KDR), and VEGFR3 (Flt-4) [20, 45, 46]. Also, VEGFR2 expression is typically limited to vessel endothelial cells, whereas VEGFR1 can be usually found in bone-marrow derived progenitors [47]. Furthermore, there was evidence that Akt and Erk mediate angiogenesis in endothelial cells [48–53]. sc-221226, an

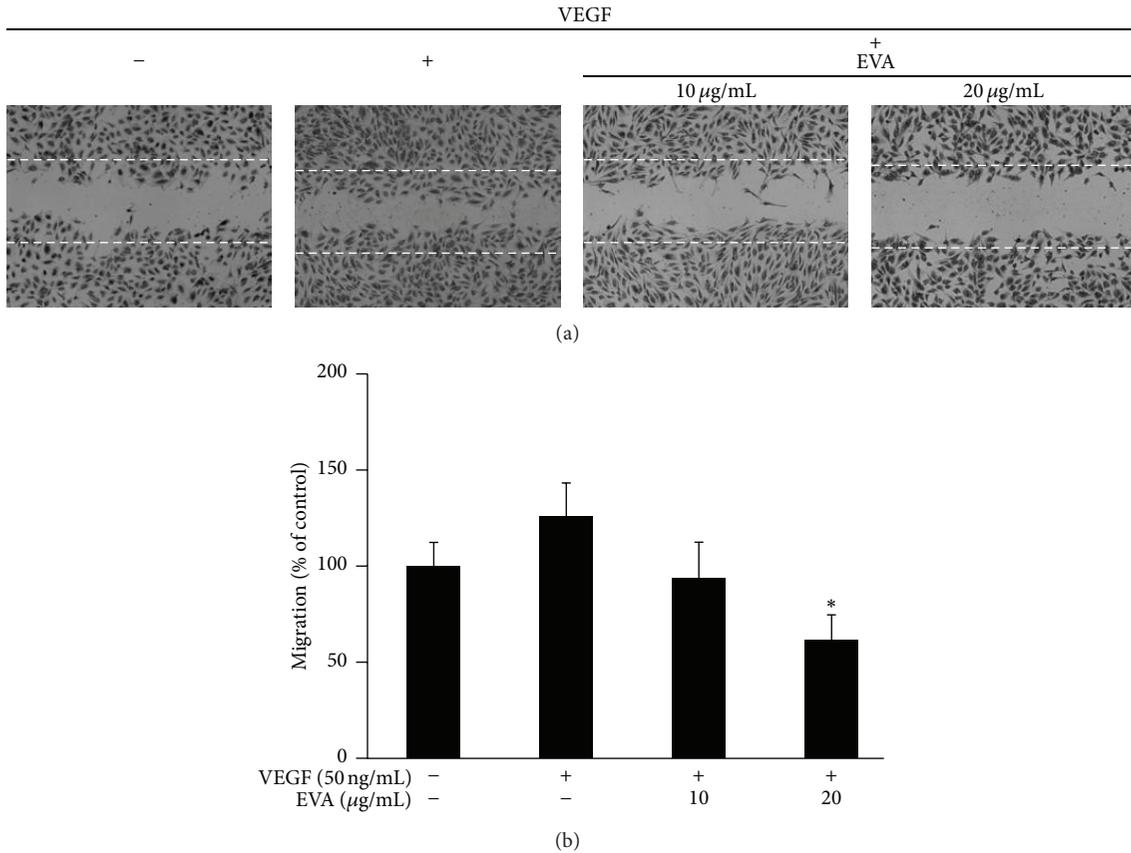


FIGURE 4: EVA suppresses VEGF induced migration in HUVECs by wound healing assay. (a) The cells (4×10^5 cells/2 mL) were treated with 0, 10, and 20 $\mu\text{g/mL}$ of EVA in a 6-well plate in the presence of VEGF (50 ng/mL) and incubated at 37°C. The number of cells migrated into the scratched area was calculated. (b) Bar graph represents the quantification of migrated cells of untreated control or VEGF-treated control and EVA-treated groups. Data are presented as means \pm SD. * $p < 0.05$ versus VEGF only treated group.

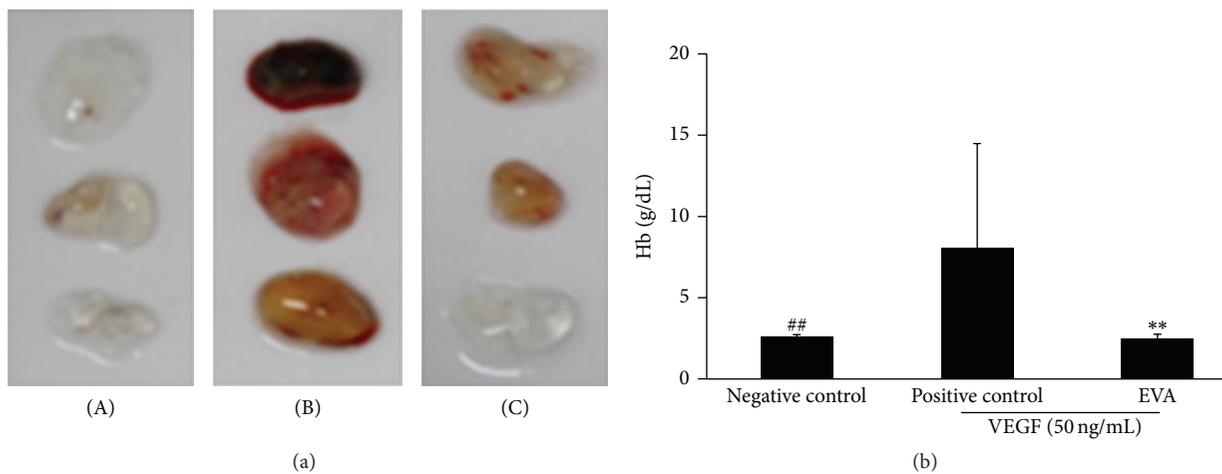


FIGURE 5: EVA significantly reduces hemoglobin content in VEGF induced angiogenesis in Matrigel plugs of mice. Six-week-old C57BL/6 mice were given EVA (300 $\mu\text{g/kg}$) extracts via oral administration for 7 days before growth factor reduced Matrigel with VEGF (25 ng) and heparin (5 U) was subcutaneously injected into right flank of C57BL/6. Ten days later, Matrigel plugs were removed. (a) The amount of hemoglobin (Hb) was measured using the hemoglobin assay kit. ((A), (B), and (C) represent negative control, positive control, and EVA-treated group, resp.) (b) Bar graph represents the quantification of Hb concentration from each of groups. Data are presented as means \pm SD. **,# $p < 0.01$ versus positive control group.

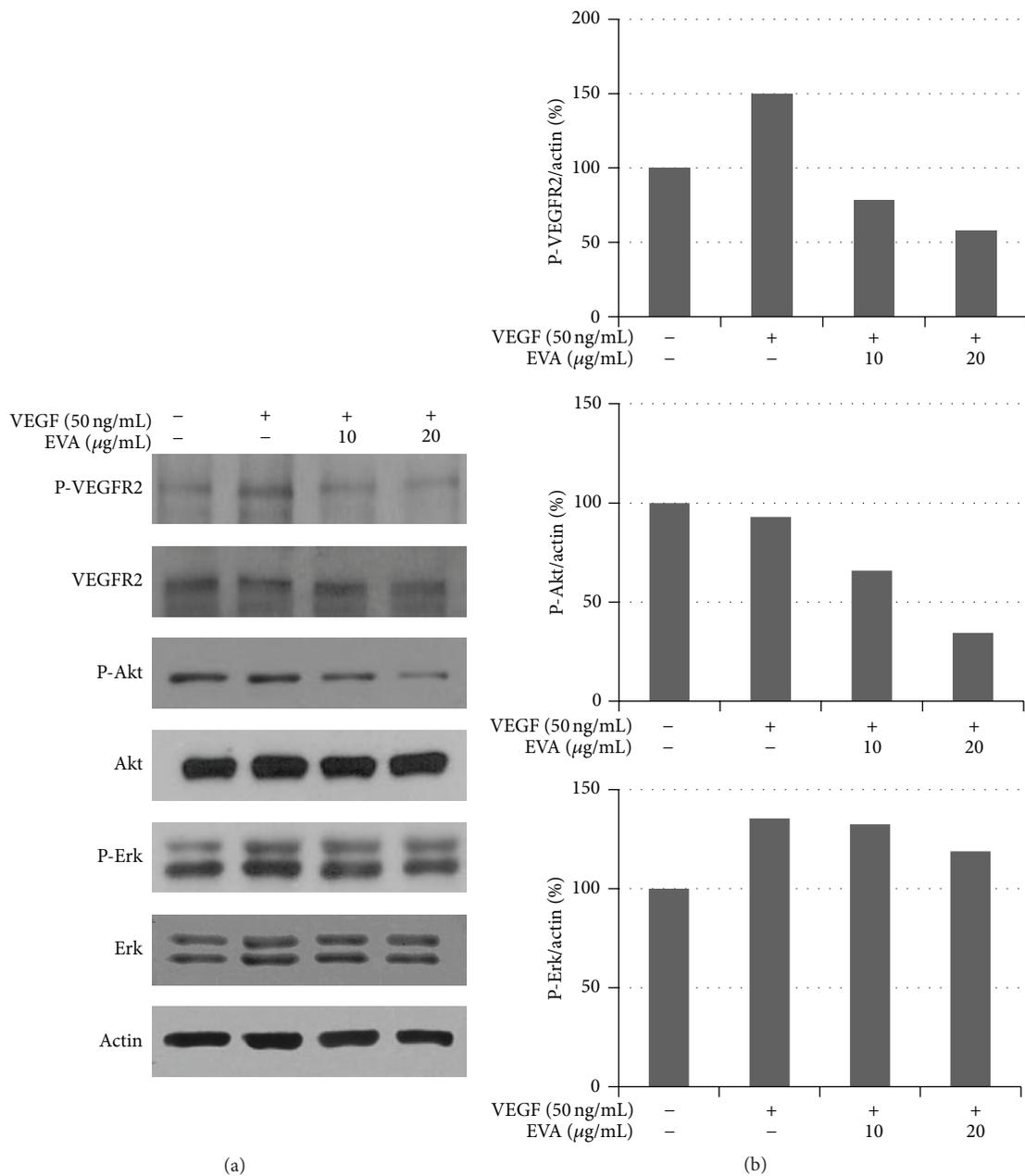


FIGURE 6: EVA effectively attenuates the phosphorylation of VEGFR2, Erk, and Akt in VEGF-treated HUVECs. Cells were treated with 10 or 20 $\mu\text{g}/\text{mL}$ of EVA in the presence of VEGF for 20 h and lysed in RIPA buffer. (a) The cell lysates were prepared and subjected to Western blotting for p-VEGFR2, VEGFR2, p-Akt, Akt, p-Erk, Erk, and β -actin. (b) Bar graphs represent fold change between actin and p-VEGFR2, p-Akt, or p-Erk groups.

Akt inhibitor, attenuated sphingosine-1-phosphate induced angiogenesis in endothelial progenitor cells [54]. Also, low molecular weight fucoidan inhibited angiogenesis *in vitro* and *in vivo* evidenced by attenuation of tube formation of hypoxic HUVECs and suppressing Akt activity reduced hypoxia-induced VEGF secretion in T24 cells, supporting the involvement of Akt in the induction of HIF-1 α and VEGF and angiogenesis [55]. Epidermal growth factor-like domain 7 promotes angiogenesis via ERK signaling pathway

in endothelial cells [56]. And fractalkine, a large cytokine protein of 373 amino acids, stimulates angiogenesis by activation of ERK and Akt [57]. These results support our conclusions. The result of Western blotting revealed that EVA effectively attenuated the phosphorylation of VEGFR2, Erk, and Akt in VEGF-treated HUVECs, implying that EVA inhibited angiogenesis in VEGF-treated HUVECs via suppression of VEGFR2, Akt, and Erk. Also, LY294002 (Akt inhibitor) showed similar effect on VEGF-enhanced tube formation,

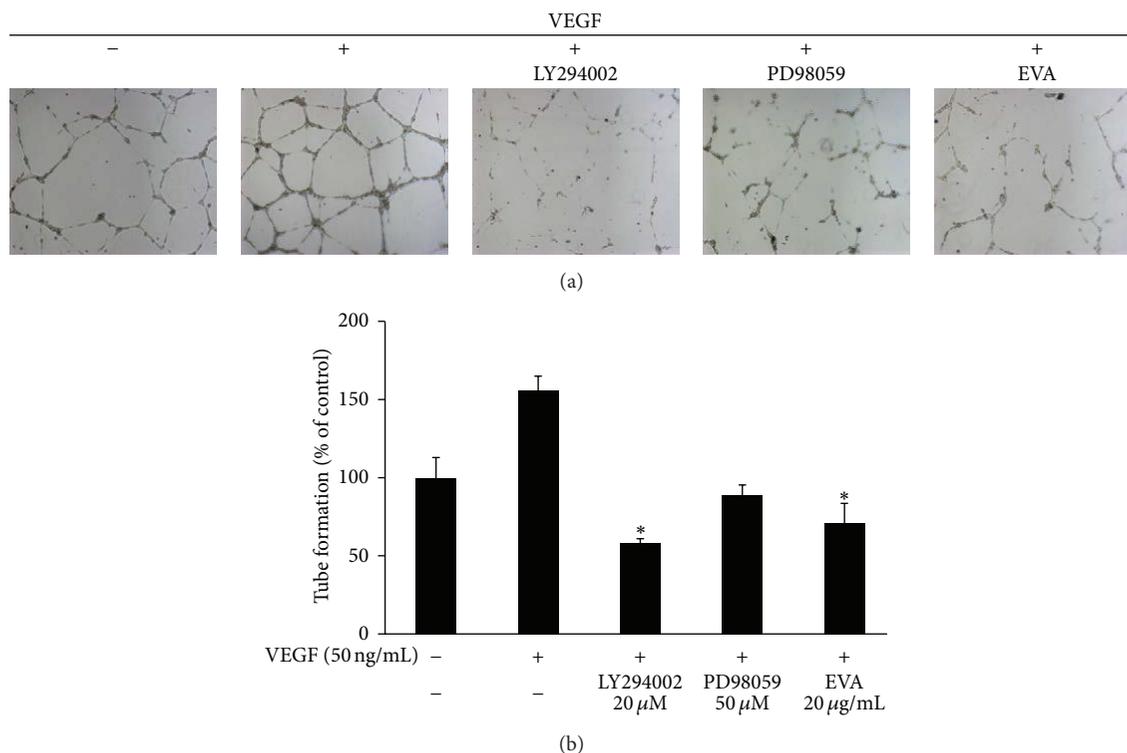


FIGURE 7: Akt, Erk inhibitors, and EVA attenuate tube formation enhanced by VEGF in HUVECs. (a) Matrigel (250 μ L) was added to 24-well plates and EVA (20 μ g/mL), LY294002 (Akt inhibitor, 20 μ M), and PD98059 (Erk inhibitor, 50 μ M) were treated to HUVECs (1×10^5 cells/well) in the presence of VEGF (50 ng/mL) and incubated at 37°C. After 18 h, randomly chosen fields were photographed under an Axiovert S 100 light microscope. (b) Bar graph represents the quantification of tube formation of each group. Data are presented as means \pm SD. * $p < 0.05$ versus VEGF-treated group.

indicating that inhibitory effect of EVA on angiogenesis in HUVECs could be by inhibition of Akt. In addition, azuki bean was reported to contain four furanylmethyl glycosides, angularides A–D, one ent-kaurane diterpene glycoside, angularin A, and four triterpenoid saponins, angulasaponins A–D, cyanidin and (+)-catechin, proanthocyanidins, digalactosyl ononitol, and gibberellin 2-oxidases [36, 44, 58–60]. Among them, anthocyanidin was well known to exert antiangiogenic activity in endothelial cells [61, 62].

In summary, EVA significantly suppressed the VEGF induced proliferation, migration, tube formation, and angiogenesis in HUVECs, and the molecular mechanism is the phosphorylation inhibition of VEGFR2 and Erk and Akt in VEGF-treated HUVECs. Overall, our findings suggest that EVA inhibits angiogenesis in VEGF-treated HUVECs via phosphorylation inhibition of VEGFR2 and Erk and Akt. Our study suggests that EVA is a promising natural agent which has antiangiogenic activity in HUVECs. However, further study is required to find out a leading antiangiogenic constituent from EVA and its potency for clinical application as a functional food.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Authors' Contribution

Oh Sung Kwon, Myoung Seok Jeong, and Bonglee Kim are co-first authors.

Acknowledgment

This work was supported by The Korea Science and Engineering Foundation (KOSEF) grant funded by the Korean government (MEST) (nos. 2011-0006220 and 0013640).

References

- [1] J. Folkman and M. Klagsbrun, "Angiogenic factors," *Science*, vol. 235, no. 4787, pp. 442–447, 1987.
- [2] H.-J. Lee, N.-J. Seo, S.-J. Jeong et al., "Oral administration of penta-O-galloyl-beta-d-glucose suppresses triple-negative breast cancer xenograft growth and metastasis in strong association with JAK1-STAT3 inhibition," *Carcinogenesis*, vol. 32, no. 6, pp. 804–811, 2011.
- [3] A. Bikfalvi, "Tumor angiogenesis," *Bulletin du Cancer*, vol. 94, no. 7, supplement, pp. F193–F198, 2007.
- [4] A. K. Weidemann, A. A. Crawshaw, E. Byrne, and H. S. Young, "Vascular endothelial growth factor inhibitors: investigational therapies for the treatment of psoriasis," *Clinical, Cosmetic and Investigational Dermatology*, vol. 6, pp. 233–244, 2013.

- [5] X. Kong, Y. Zhang, C. Liu et al., "Anti-angiogenic effect of trip-tolide in rheumatoid arthritis by targeting angiogenic cascade," *PLoS ONE*, vol. 8, no. 10, Article ID e77513, 2013.
- [6] A. A. Joharapurkar, N. A. Dhansha, and M. R. Jain, "Inhibition of the methionine aminopeptidase 2 enzyme for the treatment of obesity," *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 7, pp. 73–84, 2014.
- [7] D. Djokovic and C. Calhaz-Jorge, "Angiogenesis as a therapeutic target in endometriosis," *Acta Medica Portuguesa*, vol. 27, no. 4, pp. 489–497, 2014.
- [8] K. Demuth, I. Myara, and N. Moatti, "Biology of the endothelial cell and atherogenesis," *Annales de Biologie Clinique*, vol. 53, no. 4, pp. 171–191, 1995.
- [9] Y. Mukai and S. Sato, "Polyphenol-containing azuki bean (*Vigna angularis*) seed coats attenuate vascular oxidative stress and inflammation in spontaneously hypertensive rats," *Journal of Nutritional Biochemistry*, vol. 22, no. 1, pp. 16–21, 2011.
- [10] Y. Yang, K. Yang, Y. Li et al., "Decursin inhibited proliferation and angiogenesis of endothelial cells to suppress diabetic retinopathy via VEGFR2," *Molecular and Cellular Endocrinology*, vol. 378, no. 1–2, pp. 46–52, 2013.
- [11] H. Wang, J. Guo, X. Z. West et al., "Detection and biological activities of carboxyethylpyrrole ethanolamine phospholipids (CEP-EPs)," *Chemical Research in Toxicology*, vol. 27, no. 12, pp. 2015–2022, 2014.
- [12] B. Al-Husein, M. Abdalla, M. Trepte, D. L. DeRemer, and P. R. Somanath, "Antiangiogenic therapy for cancer: an update," *Pharmacotherapy*, vol. 32, no. 12, pp. 1095–1111, 2012.
- [13] C. A. Staton, N. J. Brown, and M. W. R. Reed, "Current status and future prospects for anti-angiogenic therapies in cancer," *Expert Opinion on Drug Discovery*, vol. 4, no. 9, pp. 961–979, 2009.
- [14] J. Folkman, "What is the role of endothelial cells in angiogenesis?" *Laboratory Investigation*, vol. 51, no. 6, pp. 601–604, 1984.
- [15] H. J. Cui, A. L. Yang, H. J. Zhou et al., "Buyang huanwu decoction promotes angiogenesis via vascular endothelial growth factor receptor-2 activation through the PI3K/Akt pathway in a mouse model of intracerebral hemorrhage," *BMC Complementary and Alternative Medicine*, vol. 15, p. 91, 2015.
- [16] M. Shibuya and L. Claesson-Welsh, "Signal transduction by VEGF receptors in regulation of angiogenesis and lymphangiogenesis," *Experimental Cell Research*, vol. 312, no. 5, pp. 549–560, 2006.
- [17] R. Roskoski Jr., "VEGF receptor protein-tyrosine kinases: structure and regulation," *Biochemical and Biophysical Research Communications*, vol. 375, no. 3, pp. 287–291, 2008.
- [18] I. Zachary and G. Gliki, "Signaling transduction mechanisms mediating biological actions of the vascular endothelial growth factor family," *Cardiovascular Research*, vol. 49, no. 3, pp. 568–581, 2001.
- [19] L. Claesson-Welsh and M. Welsh, "VEGFA and tumour angiogenesis," *Journal of Internal Medicine*, vol. 273, no. 2, pp. 114–127, 2013.
- [20] N. Ferrara, H.-P. Gerber, and J. LeCouter, "The biology of VEGF and its receptors," *Nature Medicine*, vol. 9, no. 6, pp. 669–676, 2003.
- [21] T. P. Hamsa and G. Kuttan, "Antiangiogenic activity of berberine is mediated through the downregulation of hypoxia-inducible factor-1, VEGF, and proinflammatory mediators," *Drug and Chemical Toxicology*, vol. 35, no. 1, pp. 57–70, 2012.
- [22] J.-E. Huh, E.-O. Lee, M.-S. Kim et al., "Penta-O-galloyl-beta-D-glucose suppresses tumor growth via inhibition of angiogenesis and stimulation of apoptosis: roles of cyclooxygenase-2 and mitogen-activated protein kinase pathways," *Carcinogenesis*, vol. 26, no. 8, pp. 1436–1445, 2005.
- [23] L.-L. Wang, J.-J. Li, Z.-B. Zheng, H.-Y. Liu, G.-J. Du, and S. Li, "Antitumor activities of a novel indolin-2-ketone compound, Z24: more potent inhibition on bFGF-induced angiogenesis and bcl-2 over-expressing cancer cells," *European Journal of Pharmacology*, vol. 502, no. 1–2, pp. 1–10, 2004.
- [24] F. Zhang, Z. Zhang, L. Chen et al., "Curcumin attenuates angiogenesis in liver fibrosis and inhibits angiogenic properties of hepatic stellate cells," *Journal of Cellular and Molecular Medicine*, vol. 18, no. 7, pp. 1392–1406, 2014.
- [25] R. J. Young and M. W. R. Reed, "Anti-angiogenic therapy: concept to clinic," *Microcirculation*, vol. 19, no. 2, pp. 115–125, 2012.
- [26] L. S. Wood, "Management of vascular endothelial growth factor and multikinase inhibitor side effects," *Clinical Journal of Oncology Nursing*, vol. 13, no. 6, pp. 13–18, 2009.
- [27] S.-Y. Cho, H.-J. Lee, S.-M. Cho, B. Kim, Y. K. Jung, and S.-H. Kim, "Particled *Mica*, STB-HO has chemopreventive potential via G1 arrest, and inhibition of proliferation and vascular endothelial growth factor receptor 2 in HCT colorectal cancer cells," *BMC Complementary and Alternative Medicine*, vol. 13, article 189, 2013.
- [28] S. H. Sook, H.-J. Lee, J.-H. Kim et al., "Reactive oxygen species-mediated activation of AMP-activated protein kinase and c-jun N-terminal kinase plays a critical role in beta-sitosterol-induced apoptosis in multiple myeloma U266 cells," *Phytotherapy Research*, vol. 28, no. 3, pp. 387–394, 2014.
- [29] H.-J. Lee, D.-B. Jung, E. J. Sohn et al., "Inhibition of hypoxia inducible factor alpha and astrocyte-elevated gene-1 mediates cryptotanshinone exerted antitumor activity in hypoxic PC-3 cells," *Evidence-based Complementary and Alternative Medicine*, vol. 2012, Article ID 390957, 13 pages, 2012.
- [30] M.-K. Ha, Y. H. Song, S.-J. Jeong et al., "Emodin inhibits proinflammatory responses and inactivates histone deacetylase 1 in hypoxic rheumatoid synoviocytes," *Biological and Pharmaceutical Bulletin*, vol. 34, no. 9, pp. 1432–1437, 2011.
- [31] Y. Yao, X. Cheng, S. Wang, L. Wang, and G. Ren, "Influence of altitudinal variation on the antioxidant and antidiabetic potential of azuki bean (*Vigna angularis*)," *International Journal of Food Sciences and Nutrition*, vol. 63, no. 1, pp. 117–124, 2012.
- [32] K.-H. Han, M. Fukushima, K. Ohba et al., "Hepatoprotective effects of the water extract from adzuki bean hulls on acetaminophen-induced damage in rat liver," *Journal of Nutritional Science and Vitaminology*, vol. 50, no. 5, pp. 380–383, 2004.
- [33] Y. Hori, S. Sato, and A. Hatai, "Antibacterial activity of plant extracts from azuki beans (*Vigna angularis*) in vitro," *Phytotherapy Research*, vol. 20, no. 2, pp. 162–164, 2006.
- [34] T. Itoh, M. Nakamura, H. Nakamichi, M. Ando, Y. Tsukamasa, and Y. Furuichi, "Regulation of the differentiation of osteoblasts and osteoclasts by a hot-water extract of adzuki beans (*Vigna angularis*)," *Bioscience, Biotechnology, and Biochemistry*, vol. 78, no. 1, pp. 92–99, 2014.
- [35] Y. Jiang, K.-W. Zeng, B. David, and G. Massiot, "Constituents of *Vigna angularis* and their *in vitro* anti-inflammatory activity," *Phytochemistry*, vol. 107, pp. 111–118, 2014.
- [36] Y. Mukai and S. Sato, "Polyphenol-containing azuki bean (*Vigna angularis*) extract attenuates blood pressure elevation and modulates nitric oxide synthase and caveolin-1 expressions in rats

- with hypertension," *Nutrition, Metabolism and Cardiovascular Diseases*, vol. 19, no. 7, pp. 491–497, 2009.
- [37] S. Sato, Y. Mukai, J. Yamate et al., "Effect of polyphenol-containing azuki bean (*Vigna angularis*) extract on blood pressure elevation and macrophage infiltration in the heart and kidney of spontaneously hypertensive rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 35, no. 1, pp. 43–49, 2008.
- [38] T. Itoh, H. Umekawa, and Y. Furuichi, "Potential ability of hot water adzuki (*Vigna angularis*) extracts to inhibit the adhesion, invasion, and metastasis of murine B16 melanoma cells," *Bio-science, Biotechnology and Biochemistry*, vol. 69, no. 3, pp. 448–454, 2005.
- [39] H.-H. Kim, S.-W. Kim, D.-S. Kim, H.-M. Oh, M.-C. Rho, and S.-H. Kim, "Vigna angularis inhibits mast cell-mediated allergic inflammation," *International Journal of Molecular Medicine*, vol. 32, no. 3, pp. 736–742, 2013.
- [40] E. A. Jaffe, R. L. Nachman, C. G. Becker, and C. R. Minick, "Culture of human endothelial cells derived from umbilical veins. Identification by morphologic and immunologic criteria," *The Journal of Clinical Investigation*, vol. 52, no. 11, pp. 2745–2756, 1973.
- [41] S.-J. Lee, H. M. Lee, S.-T. Ji, S.-R. Lee, W. Mar, and Y. S. Gho, "1,2,3,4,6-Penta-O-galloyl-beta-D-glucose blocks endothelial cell growth and tube formation through inhibition of VEGF binding to VEGF receptor," *Cancer Letters*, vol. 208, no. 1, pp. 89–94, 2004.
- [42] G. Tobelem, "Endothelial cell growth: biology and pharmacology in relation to angiogenesis," *Blood Coagulation & Fibrinolysis*, vol. 1, no. 6, pp. 703–705, 1990.
- [43] S. De Falco, "Antiangiogenesis therapy: an update after the first decade," *Korean Journal of Internal Medicine*, vol. 29, no. 1, pp. 1–11, 2014.
- [44] U. Takahama, R. Yamauchi, and S. Hirota, "Isolation and characterization of a cyanidin-catechin pigment from adzuki bean (*Vigna angularis*)," *Food Chemistry*, vol. 141, no. 1, pp. 282–288, 2013.
- [45] N. Ferrara and T. Davis-Smyth, "The biology of vascular endothelial growth factor," *Endocrine Reviews*, vol. 18, no. 1, pp. 4–25, 1997.
- [46] B. Sitohy, J. A. Nagy, and H. F. Dvorak, "Anti-VEGF/VEGFR therapy for cancer: reassessing the target," *Cancer Research*, vol. 72, no. 8, pp. 1909–1914, 2012.
- [47] M. Kowanetz and N. Ferrara, "Vascular endothelial growth factor signaling pathways: therapeutic perspective," *Clinical Cancer Research*, vol. 12, no. 17, pp. 5018–5022, 2006.
- [48] A. Jozkowicz, A. Nigisch, B. Winter, G. Weigel, I. Hukb, and J. Dulaka, "15-deoxy-delta12,14-prostaglandin-J2 inhibits expression of eNOS in human endothelial cells," *Prostaglandins & Other Lipid Mediators*, vol. 74, no. 1–4, pp. 11–28, 2004.
- [49] J. Kroll and J. Waltenberger, "VEGF-A induces expression of eNOS and iNOS in endothelial cells via VEGF receptor-2 (KDR)," *Biochemical and Biophysical Research Communications*, vol. 252, no. 3, pp. 743–746, 1998.
- [50] Z.-Z. Zheng and Z.-X. Liu, "Activation of the phosphatidylinositol 3-kinase/protein kinase Akt pathway mediates CD151-induced endothelial cell proliferation and cell migration," *International Journal of Biochemistry and Cell Biology*, vol. 39, no. 2, pp. 340–348, 2007.
- [51] J. Ma, H. Sawai, N. Ochi et al., "PTEN regulate angiogenesis through PI3K/Akt/VEGF signaling pathway in human pancreatic cancer cells," *Molecular and Cellular Biochemistry*, vol. 331, no. 1-2, pp. 161–171, 2009.
- [52] W. Liu, S. A. Ahmad, N. Reinmuth et al., "Endothelial cell survival and apoptosis in the tumor vasculature," *Apoptosis*, vol. 5, no. 4, pp. 323–328, 2000.
- [53] A. Franovic, C. E. Holterman, J. Payette, and S. Lee, "Human cancers converge at the HIF-2 α oncogenic axis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 50, pp. 21306–21311, 2009.
- [54] H. Wang, K. Y. Cai, W. Li, and H. Huang, "Sphingosine-1-phosphate induces the migration and angiogenesis of EPCs through the Akt signaling pathway via sphingosine-1-phosphate receptor 3/platelet-derived growth factor receptor-beta," *Cellular and Molecular Biology Letters*, 2015.
- [55] M. C. Chen, W. L. Hsu, P. A. Hwang, and T. C. Chou, "Low molecular weight fucoidan inhibits tumor angiogenesis through downregulation of HIF-1/VEGF signaling under hypoxia," *Marine Drugs*, vol. 13, no. 7, pp. 4436–4451, 2015.
- [56] Y. Cao and Q. Lyu, "Epidermal growth factor-like domain 7 promotes endothelial cell migration and angiogenesis by activating ERK signaling pathway," *Zhonghua Bing Li Xue Za Zhi*, vol. 44, no. 4, pp. 270–273, 2015.
- [57] S.-J. Lee, S. Namkoong, Y.-M. Kim et al., "Fractalkine stimulates angiogenesis by activating the Raf-1/MEK/ERK- and PI3K/Akt/eNOS-dependent signal pathways," *The American Journal of Physiology—Heart and Circulatory Physiology*, vol. 291, no. 6, pp. H2836–H2846, 2006.
- [58] T. Kaneta, T. Kakimoto, and H. Shibaoka, "Gibberellin A₃ causes a decrease in the accumulation of mRNA for ACC oxidase and in the activity of the enzyme in azuki bean (*Vigna angularis*) epicotyls," *Plant and Cell Physiology*, vol. 38, no. 10, pp. 1135–1141, 1997.
- [59] S.-H. Park, M. Nakajima, M. Sakane, Z.-J. Xu, K. Tomioka, and I. Yamaguchi, "Gibberellin 2-oxidases from seedlings of adzuki bean (*Vigna angularis*) show high gibberellin-binding activity in the presence of 2-oxoglutarate and Co²⁺," *Bioscience, Biotechnology and Biochemistry*, vol. 69, no. 8, pp. 1498–1507, 2005.
- [60] E. Yanase, K. Nishimoto, Y. O. Kamatari, and S.-I. Nakatsuka, "Isolation of a new xanthylum-related pigment from adzuki beans *Vigna angularis*," *Bioscience, Biotechnology and Biochemistry*, vol. 76, no. 8, pp. 1571–1572, 2012.
- [61] M.-K. Kang, S. S. Lim, J.-Y. Lee, K. M. Yeo, and Y.-H. Kang, "Anthocyanin-rich purple corn extract inhibit diabetes-associated glomerular angiogenesis," *PLoS ONE*, vol. 8, no. 11, Article ID e79823, 2013.
- [62] K. Matsubara, T. Kaneyuki, T. Miyake, and M. Mori, "Antiangiogenic activity of nasunin, an antioxidant anthocyanin, in eggplant peels," *Journal of Agricultural and Food Chemistry*, vol. 53, no. 16, pp. 6272–6275, 2005.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

