

## Research Article

# Liquiritigenin Protects Rats from Carbon Tetrachloride Induced Hepatic Injury through PGC-1 $\alpha$ Pathway

Yiping Zhang,<sup>1,2</sup> Yuanqiao He,<sup>3</sup> Hongbo Yu,<sup>1</sup> Fuying Ma,<sup>1</sup> Jianguo Wu,<sup>1</sup> and Xiaoyu Zhang<sup>1</sup>

<sup>1</sup>Key Laboratory of Molecular Biophysics of Ministry of Education, College of Life Science and Technology, Huazhong University of Science and Technology, Wuhan, Hubei 430074, China

<sup>2</sup>School of Basic Medical Science, Jiujiang University, Jiujiang, Jiangxi 332000, China

<sup>3</sup>Center of Laboratory Animal Science, Nanchang University, Nanchang, Jiangxi 30006, China

Correspondence should be addressed to Xiaoyu Zhang; leis428@yahoo.com

Received 15 April 2015; Revised 6 June 2015; Accepted 8 June 2015

Academic Editor: Yoshiji Ohta

Copyright © 2015 Yiping Zhang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The lack of effective treatment for liver cirrhosis and hepatocellular carcinomas imposes serious challenges to the healthcare system. Here, we investigated the efficacy and mechanism of liquiritigenin involved in preventing or retarding the progression of liver diseases in a rat model with chronic carbon tetrachloride (CCl<sub>4</sub>) exposure. Sprague Dawley rats were given CCl<sub>4</sub> and liquiritigenin alone or simultaneously for 8 weeks before liver was harvested to check histological changes by Hematoxylin and Eosin (H&E) staining, apoptosis by TUNEL assay, ROS by dihydroethidium staining, antioxidant enzyme activities and malondialdehyde using specific kits, and gene expression by quantitative real-time PCR and western blot. Chronic CCl<sub>4</sub> exposure caused profound changes in liver histology with extensive hepatocyte death (necrosis and apoptosis), fat accumulation, and infiltration of inflammatory cells, accompanied by depressed activities of antioxidant enzymes, increased oxidative stress, elevated expression of inflammation and fibrotic genes, and downregulation of PGC-1 $\alpha$ , NDI, and Bcl-x in rat liver. All these changes were abolished or alleviated by liquiritigenin. The results demonstrated that liquiritigenin is effective in protecting liver from injury or treating chronic liver diseases. The modulation of PGC-1 $\alpha$  and its downstream genes might play a critical role in relieving CCl<sub>4</sub>-induced hepatic pathogenesis by liquiritigenin.

## 1. Introduction

Primary liver cancer with the majority of the cases being hepatocellular carcinoma (HCC) is one of the most common malignancies and has become the second leading cause of cancer death worldwide [1, 2]. The dominant risk factor of HCC is liver cirrhosis, which most frequently resulted from chronic hepatitis B virus (HBV) or hepatitis C virus (HCV) infection [2, 3]. Liver cirrhosis on its own is another significant public health issue, with up to 10% prevalence in general population and over 750000 annual deaths [4]. Surgical resection of early stage HCC and liver cirrhosis is superior over transarterial chemoembolization and becoming increasingly popular [5]. However, efficient noninvasive interventions are still sought, especially for later stage HCC and cirrhosis.

As the major organ to metabolize and detoxify metabolites and xenobiotics, the liver is liable to the damage caused

by the hepatotoxicity of chemicals and oxidative stress [6]. Oxidative stress has been increasingly believed to play a crucial role in the pathogenesis of liver diseases [7]. While human and animal liver is capable of repairing damage with compensatory regeneration, it partially or completely loses its functions due to cirrhosis or cancer resulting from liver fibrosis as the injuries exceed its repairing capacity [8]. Carbon tetrachloride (CCl<sub>4</sub>) is widely used to establish rodent models of chronic liver injury as it is converted to highly reactive metabolites by the cytochrome P450 in liver [9], which in turn reduce antioxidant enzymes activity and eventually lead to membrane lipid peroxidation [10]. Therefore, reducing or eliminating reactive oxygen species (ROS) and free radicals would be an effective strategy for fighting hepatotoxicity. Peroxisome proliferator-activated receptor gamma, coactivator 1 alpha (PGC-1 $\alpha$ ) is a master regulator of energy metabolism and inflammation [11, 12];

it regulates a wide range of genes involved in mitochondria biogenesis and antioxidant.

Herbal medicine has been attracting increased interest in protecting and/or treating liver diseases [9, 13–15]. Liquiritigenin is a flavonone from *licorice*, the dried root of *Glycyrrhiza glabra*. It exhibits estrogenic [16], choleric [17], anti-inflammatory [18], and antitumor [19] effects. This study investigates the effect of liquiritigenin on CCl<sub>4</sub>-induced chronic liver injuries and the biochemical pathways involved.

## 2. Materials and Methods

**2.1. Animals.** All animal protocols were conformed to the ethic requirements stated in the Declaration of Helsinki and approved by the institutional animal care and usage committee of Huazhong University of Science and Technology. Sprague Dawley (SD) rats were purchased from the laboratory animal center of Huazhong University of Science and Technology and acclimatized to the facility for a week before experiments. The rats were randomly divided into 4 groups (15 rats in each group) and given food and water *ad libitum*. The control rats fed on normal chow only; CCl<sub>4</sub> group rats received 6 mg/kg CCl<sub>4</sub> subcutaneously 3 times a week; liquiritigenin group (Liqtn) rats received 10 mg/kg liquiritigenin ( $\geq 98\%$ , Cat number G02, Jiangxi Herbfine Hi-tech Co., Ltd., Nanchang, China) by gavages 3 times a week; and the CCl<sub>4</sub>/Liqtn group rats received 6 mg/kg CCl<sub>4</sub> subcutaneously and 10 mg/kg liquiritigenin by gavages 3 times a week. After 8 weeks of treatment, rats were sacrificed and the livers were harvested and treated for downstream studies.

**2.2. Pathology Observation.** The pathological changes of livers were observed after Hematoxylin and Eosin (H&E) staining. The slides were deparaffinized and hydrated, stained in alum haematoxylin for about 5 min, rinsed with running tap water, differentiated with 0.3% acid alcohol, rinsed with running tap water, stained with eosin for 2 min, and dehydrated, cleared, and mounted before being observed under a microscope by a pathologist.

**2.3. Cell Apoptosis Assay.** The apoptosis of hepatic cells in rat livers was analyzed with a TUNEL (terminal deoxynucleotidyl transferase dUTP nick end labeling) kit from Boyotime (C1098, Shanghai, China) according manufacturer's instruction. Briefly, sections were fixed with 4% paraformaldehyde 30–60 min, washed with PBS twice for 10 min, and incubated with 0.1% Triton X-100 in PBS on ice for 2 min and then with 0.3% H<sub>2</sub>O<sub>2</sub> in methanol at room temperature for 20 min to quench endogenous peroxidase, which was followed by three washes with PBS. 50  $\mu$ L of Biotin labeling solution (including TdT enzyme and Biotin-dUTP) was added on top of the sections, which were incubated at 37°C for 60 min in a humidified chamber, washed once with PBS, incubated with 0.1–0.3 mL of labeling termination solution at room temperature for 10 min, and washed three times with PBS. The sections were then incubated with 50  $\mu$ L of Streptavidin-HRP working solution at room temperature for 30 min, washed with PBS for 3 times, incubated in

0.2–0.5 mL of DAB working solution at room temperature for 5–30 min, washed 3 times with PBS, and observed and photographed.

**2.4. Quantitative Real-Time Polymerase Chain Reaction (qPCR).** Total RNA was isolated from rat liver using Trizol reagent (Invitrogen, Shanghai, China) according supplier's protocol. The reverse transcription was done with 1  $\mu$ g of total RNA using PrimeScript 1st strand cDNA Synthesis Kit (Takara, Beijing, China). Quantitative real-time PCR was performed using SuperReal PreMix (SYBR Green) (FP204, Tiangen Biotech, Beijing, China) on ABI7300 (Applied Biosystems, Foster City, CA). The primers were AAGGATGGAGGCACGATTGG and GGGAACTTGATGATGGGCGA for MMP-2; TCGCCACCGGATTGAAGAAA and CTCGGGAAGGCACAGCAATA for RelA; CGGTTTCCCGTGCAATCAGT and ACACCGGGGACCAAATGATG for GSH-Px; GGAGCAAGGTCGCTTACAGA and TCCCACACATCAATCCCCAG for SOD2; AGTCCCATACACAACCGCAG and CCCTTGGGGTCATTTGGTGA for PGC-1 $\alpha$ ; AATTCCTCTGGCCTGCCTAC and ATCATAGGTCGGGAGGAGGT for Bcl-x; ATGGCCTTCCTCACCCTAGT and GGGTTGGGCGATAATAAAT for NADH dehydrogenase 1 (ND-1); and AACTCCCATTCCCTCCACCTT and GAGGGCCTCTCTTTGCTCT for GAPDH. The cycling program was 95°C 3 min followed by 40 cycles of 95°C 30 sec, 58°C 15 sec, and 68°C 30 sec. The relative mRNA levels were calculated using 2<sup>- $\Delta\Delta$ Ct</sup> method with GAPDH as the internal control.

**2.5. Western Blot.** Rat liver proteins (40  $\mu$ g) were resolved on 8% SDS-polyacrylamide gels and transferred onto PVDF membranes, which were blocked with 5% non-fat milk in PBST (0.5% Triton X-100 in PBS) for 30 min at room temperature and incubated with specified antibodies overnight at 4°C. The membranes were washed with PBST, incubated with horseradish peroxidase conjugated secondary antibodies at room temperature for 60 min, washed, and visualized with ECL reagent. First antibodies were RelA (112A1021) from Novus Biologicals (Littleton, CO), MMP2 (AF1488) from R&D Systems (Shanghai, China), PGC-1 $\alpha$  (SAB4200209) from Sigma-Aldrich (St. Louis, MO), Bcl-x (ab32370) from Abcam (Cambridge, MA), and  $\beta$ -Actin (20536-1-AP) from Proteintech (Wuhan, China).

**2.6. ROS Assayed with Dihydroethidium (DHE).** Hepatocytes were incubated with medium containing 10 mM DHE at 37°C in the dark for 15 min. The cells were washed twice with PBS before being observed under fluorescent microscope or analyzed by fluorescent activated cell sorting with band-pass filter at 585 nm. The mean fluorescence intensity was obtained from 10,000 cells.

**2.7. Lipid Peroxidation Assay.** The level of liver malondialdehyde (MDA) was measured using a kit (K739-100) from Bio-Vision (Milpitas, CA) according to manufacturer's protocol. Briefly, 10 mg of liver sample was homogenized in 300  $\mu$ L of MDA Lysis Buffer (with 3  $\mu$ L 100X BHT) and centrifuged



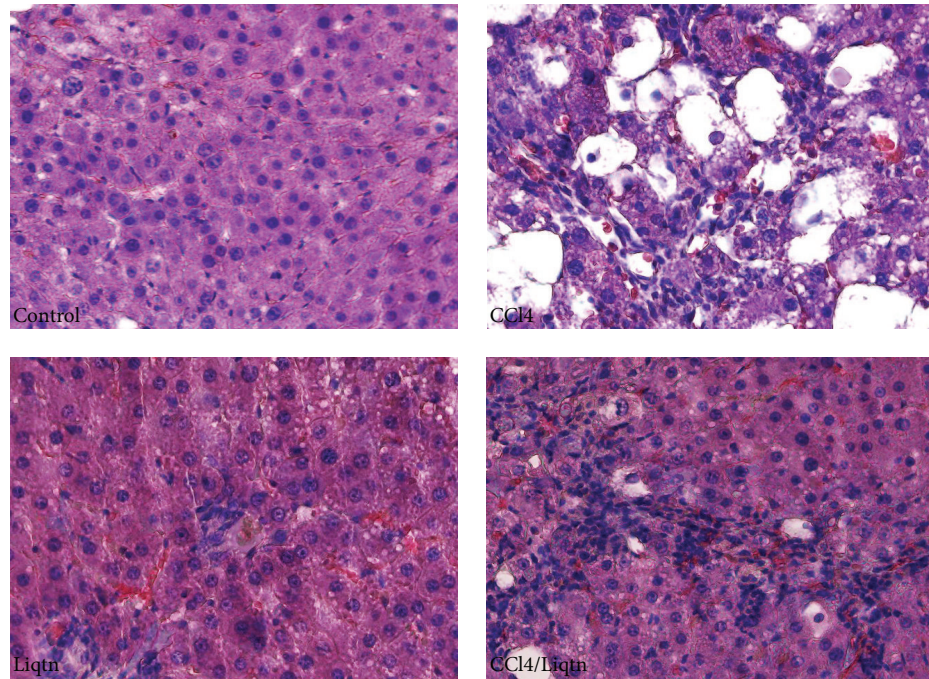


FIGURE 1: Liquiritigenin alleviated CCl<sub>4</sub> caused the histological destruction of rat livers. Rats were treated with CCl<sub>4</sub> and/or liquiritigenin for 8 weeks and liver sections were stained with Hematoxylin and Eosin. CCl<sub>4</sub> treated rats had severe liver histological abnormality with widespread hepatocyte death, fatty accumulation, and immune cell infiltration, which was largely relieved by liquiritigenin.

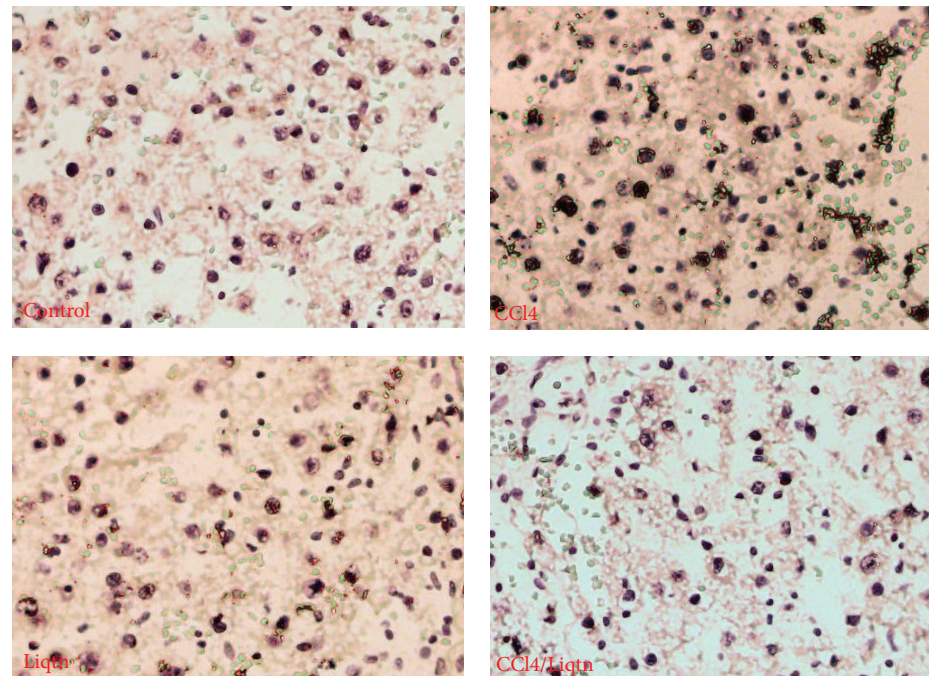


FIGURE 2: Liquiritigenin inhibited hepatocyte apoptosis in CCl<sub>4</sub> treated rat livers. The apoptotic cells in rat liver sections were detected with a commercial TUNEL kit. The results showed increased apoptosis in rat liver exposed to CCl<sub>4</sub> while liquiritigenin protected the liver cells from CCl<sub>4</sub>-induced apoptosis.

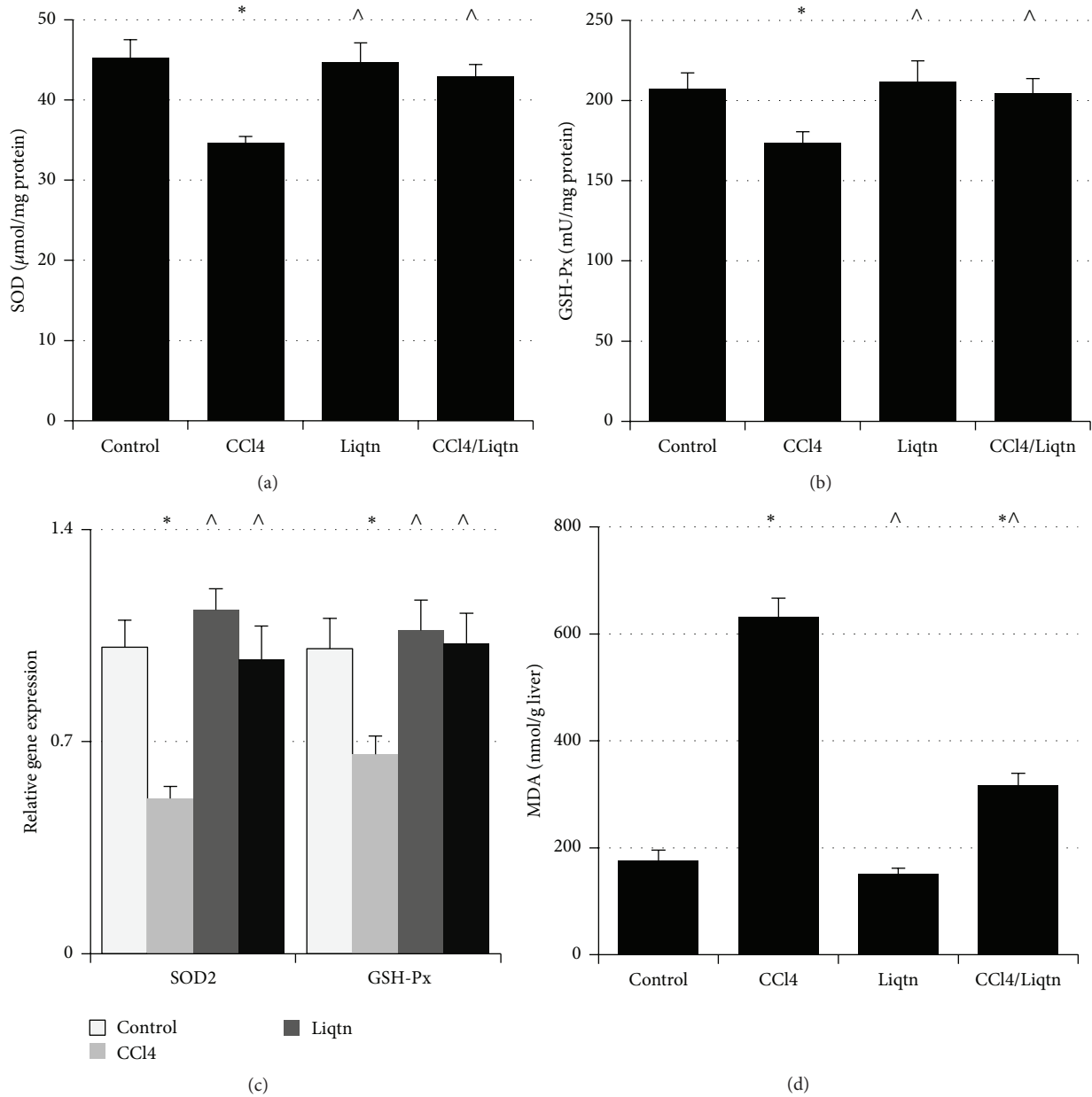


FIGURE 3: CCl<sub>4</sub>-elicited oxidative stress was effectively inhibited by liquiritigenin. The enzymatic activities of liver SOD (a) and GSH-Px (b) were analyzed with commercially available assay kits. (c) The mRNA levels of SOD2 and GPx in livers of CCl<sub>4</sub> and/or liquiritigenin treated rats were assessed by quantitative real-time PCR. (d) The MDA (lipid peroxidation) level was analyzed using an assay kit. \**P* < 0.05 compared to control rats; ^*P* < 0.05 compared to CCl<sub>4</sub> treated rats.

10 min at 13000 g at 4°C. 200 μL of the supernatant was added to an Eppendorf tube. MDA standard curve was set according to instruction to make 0, 4, 8, 12, 16, and 20 nmol standard. 600 μL of TBA reagent was added to each tube containing sample or standard, incubated at 95°C for 60 min, and cooled down on ice for 10 min. 200 μL reaction mixture from each tube was transferred into a 96-well microplate and read at a wavelength of 532 nm on a microplate reader (BioTek, Beijing, China).

**2.8. Activities of Superoxide Dismutase Activity (SOD) and Glutathione Peroxidase (GSH-Px).** The activities of SOD

(ab65354) and GSH-Px (ab102530) were analyzed using assay kits from Abcam (Cambridge, MA) according to manufacturer's instructions.

For GSH-Px activity measurement, liver tissues (10 mg) were washed with cold PBS and homogenized in 200 μL cold Assay Buffer with a Dounce homogenizer on ice for about 15 passes and centrifuged at 10000 g for 15 min at 4°C and the supernatant was collected into a fresh tube. Assay plates were set up with standards, proper controls, and samples. 40 μL of freshly made reaction mix was added to samples and controls and incubated at room temperature for 25 min. Then 10 μL cumene hydroperoxide solution was

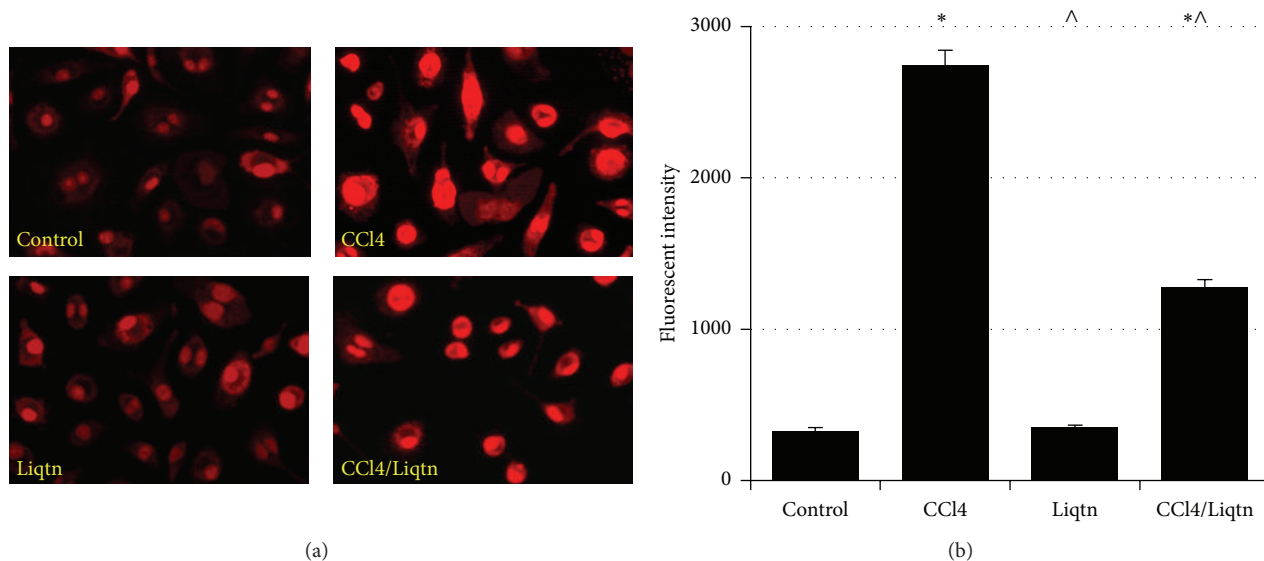


FIGURE 4: Liquiritigenin mitigated CCl<sub>4</sub>-induced ROS. The cellular ROS in rat livers treated with CCl<sub>4</sub> and/or liquiritigenin was stained with dihydroethidium followed by fluorescence microscopy or flow cytometry. \* $P < 0.05$  compared to control rats; ^ $P < 0.05$  compared to CCl<sub>4</sub> treated rats.

added to each well, mixed, and read at 340 nm on a plate reader (BioTek). The plates were incubated in the dark at 25°C for 5 min and read again at 340 nm.

For SOD assay, 10 mg of liver tissue was washed in cold PBS to remove red blood cells and homogenized in cold 0.1 M Tris/HCl, pH 7.4 (containing 0.5% Triton X-100, 5 mM  $\beta$ -ME, and 0.1 mg/mL PMSF) and centrifuged for 5 min at 4°C at 14000 g, and then supernatant was transferred into a clean tube. The standards, controls, and samples were added to 96-well assay plates; then, 200  $\mu$ L of WST working solution was added to all wells and 20  $\mu$ L of Enzyme Working Solution was added to wells containing samples and proper controls only according to manufacturer's instruction. The plates were mixed and incubated 20 min at 37°C before being read on a microplate reader (BioTek) at 450 nm.

**2.9. Statistical Analysis.** The data was presented as mean  $\pm$  standard deviation. The differences between treatment groups and control were analyzed with one-way analysis of variance. It was considered statistically significant if a  $P$  value was less than 0.05.

### 3. Results

**3.1. Liquiritigenin Alleviated CCl<sub>4</sub>-Induced Hepatic Injuries.** CCl<sub>4</sub> treatment caused the loss of regular liver structure seen in control rat with widespread necrosis of hepatocytes, fatty accumulation, and significant lymphocytes infiltration. Liquiritigenin treatment significantly reduced the severity of CCl<sub>4</sub>-induced hepatic damages with much less necrosis of hepatocytes and few diffused fatty changes (Figure 1). Meanwhile, the number of apoptotic hepatocytes was markedly

increased by CCl<sub>4</sub> treatment, which was suppressed by liquiritigenin (Figure 2).

**3.2. Liquiritigenin Relieved CCl<sub>4</sub> Caused Oxidative Stress.** Chronic CCl<sub>4</sub> exposure significantly decreased the activities of superoxide dismutase (Figure 3(a)) and glutathione peroxidase (Figure 3(b)) as well as their mRNA levels (Figure 3(c)) in rat liver. The SOD and GSH-Px activities in the livers of rats exposed to chronic CCl<sub>4</sub> were reduced by 23.5% and 16.3% compared to control rats, respectively (Figures 3(a) and 3(b)). Liquiritigenin treatment abolished CCl<sub>4</sub>-induced reduction of SOD (Figure 3(a)) and GSH-Px (Figure 3(b)) activities and their expression levels (Figure 3(c)). The SOD activity of rat livers treated with both liquiritigenin and CCl<sub>4</sub> recovered from 34.6  $\mu$ mol/mg protein in CCl<sub>4</sub>-treated rats to 42.9  $\mu$ mol/mg protein ( $P < 0.05$ ), which was similar to that of control rats (45.2  $\mu$ mol/mg protein) (Figure 3(a)). The activity of GSH-Px of liquiritigenin/CCl<sub>4</sub> rat livers was 17.9% higher than that of CCl<sub>4</sub> rat livers ( $P < 0.05$ , Figure 3(b)). The liver mRNA levels of SOD2 and GSH-Px of CCl<sub>4</sub> treated rats were 57.3% and 65.8% of those of the control, which were improved to 97.1% and 102.3% of the control in rats that received liquiritigenin while being exposed to CCl<sub>4</sub> (Figure 3(c)).

The lipid peroxidation levels were analyzed by measuring hepatic malondialdehyde levels in rats treated with CCl<sub>4</sub> and/or liquiritigenin. Chronic CCl<sub>4</sub> exposure caused an about 3.5-fold increase of MDA in rat livers (Figure 3(d)). Liquiritigenin treatment lowered liver MDA levels by almost 50% in CCl<sub>4</sub>-exposed rats (Figure 3(d)).

Meantime, the reactive oxygen species level of hepatocytes from CCl<sub>4</sub> treated rats was more than 8-fold higher than that of control rat hepatocytes. Liquiritigenin alone did not



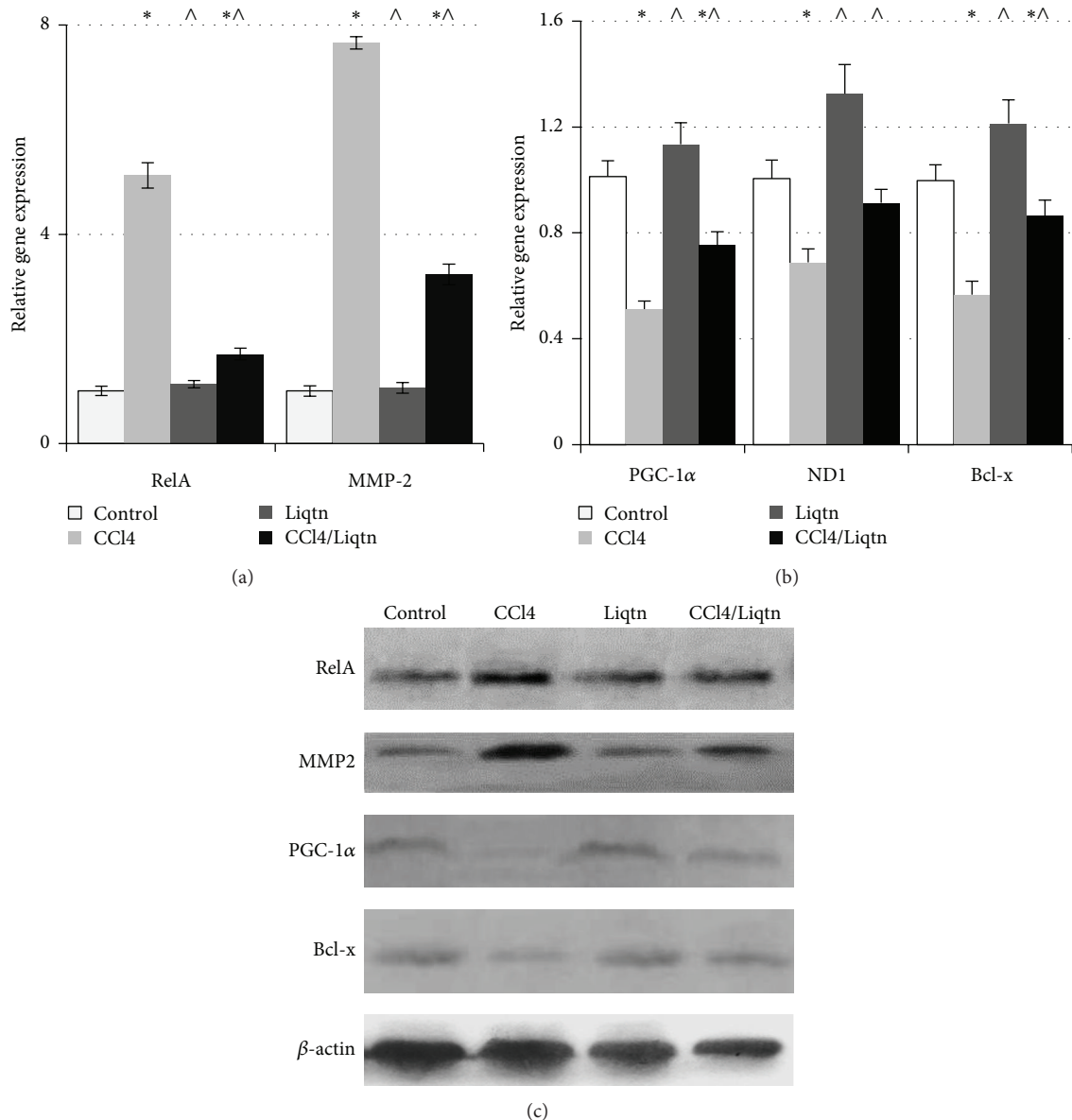


FIGURE 5: Liquiritigenin overturned the abnormal gene expression caused by CCl<sub>4</sub>. The expression of examined genes was analyzed by quantitative real-time PCR ((a) and (b)) and western blot (c). \* $P < 0.05$  compared to control rats; <sup>^</sup> $P < 0.05$  compared to CCl<sub>4</sub> treated rats.

have effects on the ROS level of rat hepatocytes, but it reduced ROS levels by more than 53% compared to hepatocytes from rats exposed to CCl<sub>4</sub> only (Figure 4).

**3.3. Liquiritigenin Reduced CCl<sub>4</sub>-Induced Hepatic Inflammatory and Fibrotic Responses.** The mRNA levels of RelA and MMP2 were increased by more than 5- and 7.5-fold, respectively, over control rats by CCl<sub>4</sub> treatment, which was reduced by about 66% and 58% by liquiritigenin (Figure 5(a)). Consequently, the protein levels of RelA and MMP2 were significantly increased in livers of CCl<sub>4</sub> treated rats and the increase was attenuated by liquiritigenin (Figure 5(c)).

**3.4. Mitochondrial Function Was Restored by Liquiritigenin.** After long-term CCl<sub>4</sub> treatment, the transcript levels of

PGC-1 $\alpha$ , ND1, and Bcl-x of rat liver were reduced by 30 to 50% compared to control rats. In the liver of rats that received liquiritigenin on top of CCl<sub>4</sub>, the mRNA levels of PGC-1 $\alpha$ , ND1, and Bcl-x were more than 30–50% higher than those of CCl<sub>4</sub>-treated rats ( $P < 0.05$ ) (Figure 5(b)). The hepatic PGC-1 $\alpha$ , ND1, and Bcl-x protein levels were also substantially reduced by chronic CCl<sub>4</sub> exposure and the reduction was alleviated by liquiritigenin (Figure 5(c)).

## 4. Discussion

Liquiritigenin showed promising effects in treating chronic liver injuries and preventing liver cirrhosis and hepatocellular carcinoma. After 8 weeks of CCl<sub>4</sub> exposure, the expression levels of PGC-1 $\alpha$ , Bcl-x, and ND-1 were significantly decreased whereas those of NF- $\kappa$ B and MMP-2 were

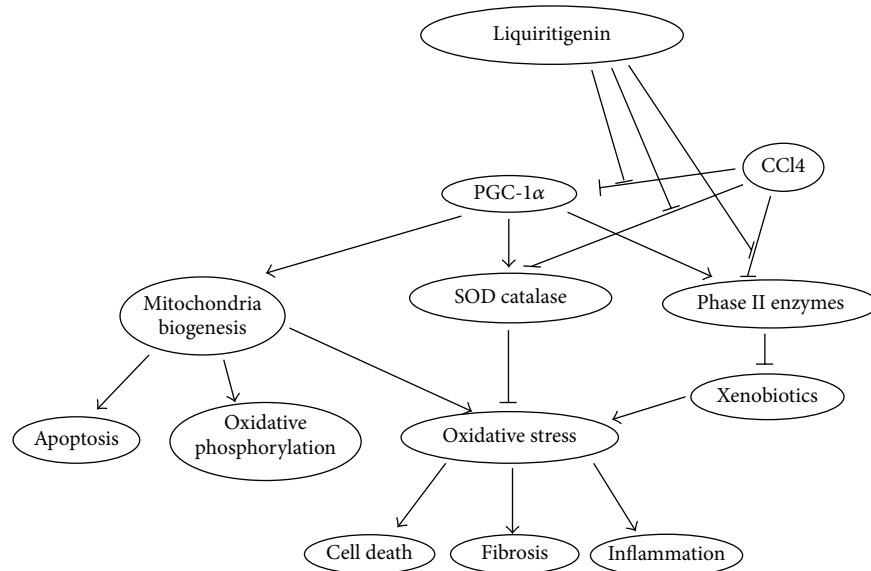


FIGURE 6: A proposed model on how liquiritigenin alleviates CCl<sub>4</sub>-induced liver injuries. CCl<sub>4</sub> inhibits the expression of PGC-1 $\alpha$ , indirectly inhibits the expression of genes involved in mitochondrial function, antioxidant, and phase II detoxification, and directly inhibits the activities of antioxidant enzymes and phase II enzymes, which leads to oxidative stress, cell death inflammation, and fibrosis. Liquiritigenin prevents or reverse CCl<sub>4</sub>-induced liver injuries by derepressing PGC-1 $\alpha$  expression and relieving the inhibition of antioxidant enzyme and phase II enzyme activities, leading to normalized expression of genes involved in mitochondrial function, antioxidant, and phase II detoxification, reduced oxidative stress, cell death inflammation, and fibrosis.

markedly increased, which led to heightened oxidative stress, increased cell death, and destruction of liver structure. Treatment with liquiritigenin essentially prevented above-mentioned pathological genes with better preserved liver histology, decreased ROS and MDA levels, muted cell death, and suppressed inflammatory and fibrotic responses.

Although the involvement of antioxidant and phase II enzymes is the common theme, many pathways have been implicated in the protective effects of various compounds against CCl<sub>4</sub>-induced liver injuries [8–10, 20–22]. Ursolic acid alleviated the inhibition of nuclear factor E2-related factor 2 (Nrf2) by CCl<sub>4</sub> and restored the expression of Nrf2 downstream antioxidative genes to protect against liver fibrosis [20]. Nrf2 was shown to inhibit transforming growth factor- (TGF-)  $\beta$ /Smad signaling and plasminogen activator inhibitor- (PAI-) 1 expression, which led to inhibition of fibrosis [23]. Paraoxonases (PON) 1 and 3, which inhibit lipoprotein oxidation and atherosclerosis, and other antioxidant enzymes were involved in CCl<sub>4</sub>-induced liver injury in rats [21]. Ribosomal S-6 kinase acting downstream of mitogen-activated protein kinase (MAPK) pathway/extracellular signal-regulated kinase (ERK1/2) signal pathway to phosphorylate C/EBP-beta in hepatic stellate cells was shown to play a critical role in the progression of CCl<sub>4</sub>-induced liver fibrosis [24]. A variety of pathways have been shown to be involved in liver injuries leading to cirrhosis and hepatocellular carcinoma. As the master regulator of bioenergetic homeostasis and responses to oxidative stress, PGC-1 $\alpha$  was shown to mediate the effects of liquiritigenin to relieve CCl<sub>4</sub> induced chronic liver injury in rat (Figure 6).

PGC-1 $\alpha$  is a transcriptional coactivator that was first identified as a regulator of mitochondrial biogenesis and function

and regulates the genes involved in energy metabolism [25]. En ensuing studies have vastly expanded the functions of PGC-1 $\alpha$ . PGC-1 $\alpha$  (and PGC-1 $\beta$ ) regulated macrophage polarization towards M2 and inhibited the expression of M1 proinflammatory cytokines to reduce inflammation during low-intensity exercise [26]. PGC-1 $\alpha$  promoted physiological increase of mitochondrial content to protect L6 myoblasts against H<sub>2</sub>O<sub>2</sub> induced apoptosis [26]. While promoting mitochondrial biogenesis, PGC-1 $\alpha$  activated the expression of antioxidant enzymes including SOD2 and thioredoxin (Trx2) [27]. Moreover, activation of PGC-1 $\alpha$  (and PGC-1 $\beta$ ) mediated mitochondrial biogenesis was at the center of cytoprotective effects of many small molecules [28, 29]. Carbon monoxide/heme oxygenase-1 system worked through NF-E2-related factor-2/PGC-1 $\alpha$ /nuclear respiratory factor 1 to induce mitochondrial biogenesis that led to pro-survival and anti-inflammatory effects during *Staphylococcus aureus* sepsis in mice [29]. It was clear that PGC-1 $\alpha$  might play a central role in liver injuries leading to cirrhosis and liver cancers (Figure 6). CCl<sub>4</sub> (and other insults) inhibited the expression of PGC-1 $\alpha$ , which in turn repressed the expression of antioxidant enzymes including SOD2 and GSH-Px, mitochondrially encoded oxidative phosphorylation genes including ND1, and mitochondrial antiapoptosis genes including Bcl-x. The resulting oxidative stress caused inflammation, cell necrosis, and fibrosis. Without effective intervention, these pathological changes would lead to the end stage diseases liver cirrhosis and/or hepatocellular carcinomas. Liquiritigenin effectively blocked the pathological effects of CCl<sub>4</sub> starting right from derepression of PGC-1 $\alpha$  expression, which recovered the expression levels of antioxidant enzymes, mitochondrial function regulators, inflammatory and fibrogenetic

genes, and apoptosis regulators. These changes led to reduced oxidative stress, cell death, and liver structure destruction during chronic CCl<sub>4</sub> exposure.

In conclusion, liquiritigenin showed strong efficacy in inhibiting chronic exposure of CCl<sub>4</sub> caused hepatic injuries through PGC-1 $\alpha$  mediated derepression of antioxidant enzymes, oxidative metabolism genes, and mitochondrial pathway antiapoptotic genes. Moreover, it also inhibited inflammatory and fibrogenetic responses in rat liver upon CCl<sub>4</sub> exposure.

## Conflict of Interests

The authors declare that there is no conflict of interests.

## Acknowledgments

The authors are grateful to the Centre of Analysis and Test of Huazhong University of Science and Technology for LC-MSn and NMR spectra analysis. This work was supported by the Fundamental Research Funds for the Central Universities (HUST: 2012QN064), the National High Technology Research and Development Program of China (863 Program) (no. 2012AA101807), Chinese National Science Foundation (81360364), and Jiangxi Provincial Department of Education Science and Technology Grant (GJJ13738).

## References

- [1] F. X. Bosch, J. Ribes, M. Díaz, and R. Cléries, "Primary liver cancer: worldwide incidence and trends," *Gastroenterology*, vol. 127, no. 5, supplement 1, pp. S5–S16, 2004.
- [2] D. Y.-K. But, C.-L. Lai, and M.-F. Yuen, "Natural history of hepatitis-related hepatocellular carcinoma," *World Journal of Gastroenterology*, vol. 14, no. 11, pp. 1652–1656, 2008.
- [3] H. B. El-Serag, "Epidemiology of viral hepatitis and hepatocellular carcinoma," *Gastroenterology*, vol. 142, no. 6, pp. 1264–1273, 2012.
- [4] F. Imani, M. Motavaf, S. Safari, and S. M. Alavian, "The therapeutic use of analgesics in patients with liver cirrhosis: a literature review and evidence-based recommendations," *Hepatitis Monthly*, vol. 14, no. 10, Article ID e23539, 2014.
- [5] T. Kapitanov, U. P. Neumann, and M. Schmeding, "Hepatocellular carcinoma in liver cirrhosis: surgical resection versus transarterial chemoembolization—a meta-analysis," *Gastroenterology Research and Practice*, vol. 2015, Article ID 696120, 8 pages, 2015.
- [6] H. Jaeschke, G. J. Gores, A. I. Cederbaum, J. A. Hinson, D. Pessayre, and J. J. Lemasters, "Mechanisms of hepatotoxicity," *Toxicological Sciences*, vol. 65, no. 2, pp. 166–176, 2002.
- [7] R. Siegel, J. Ma, Z. Zou, and A. Jemal, "Cancer statistics, 2014," *CA—A Cancer Journal for Clinicians*, vol. 64, no. 1, pp. 9–29, 2014.
- [8] D. E. Semenov, N. A. Zhukova, E. P. Ivanova et al., "Hepatoprotective properties of betulonic acid amide and heptral in toxic liver injury induced by carbon tetrachloride in combination with ethanol," *Bulletin of Experimental Biology and Medicine*, vol. 158, no. 3, pp. 336–341, 2015.
- [9] Y. S. El-Sayed, M. A. Lebda, M. Hassinin, and S. A. Neoman, "Chicory (*Cichorium intybus* L.) root extract regulates the oxidative status and antioxidant gene transcripts in CCl<sub>4</sub>-induced hepatotoxicity," *PLOS ONE*, vol. 10, no. 3, Article ID e0121549, 2015.
- [10] M. Kanter, O. Coskun, and M. Budancamanak, "Hepatoprotective effects of *Nigella sativa* L and *Urtica dioica* L on lipid peroxidation, antioxidant enzyme systems and liver enzymes in carbon tetrachloride-treated rats," *World Journal of Gastroenterology*, vol. 11, no. 42, pp. 6684–6688, 2005.
- [11] J. Shoag and Z. Arany, "Regulation of hypoxia-inducible genes by PGC-1 $\alpha$ ," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 30, no. 4, pp. 662–666, 2010.
- [12] P. S. Eisele and C. Handschin, "Functional crosstalk of PGC-1 coactivators and inflammation in skeletal muscle pathophysiology," *Seminars in Immunopathology*, vol. 36, no. 1, pp. 27–53, 2014.
- [13] K. M. Ashry, Y. S. El-Sayed, R. M. Khamiss, and I. M. El-Ashmawy, "Oxidative stress and immunotoxic effects of lead and their amelioration with myrrh (*Commiphora molmol*) emulsion," *Food and Chemical Toxicology*, vol. 48, no. 1, pp. 236–241, 2010.
- [14] H. Niranjana Murthy, V. S. Dandin, and K. Yoeup Paek, "Hepatoprotective activity of ginsenosides from *Panax ginseng* adventitious roots against carbon tetrachloride treated hepatic injury in rats," *Journal of Ethnopharmacology*, vol. 158, pp. 442–446, 2014.
- [15] B. Brai, R. Adisa, and A. Odetola, "Hepatoprotective properties of aqueous leaf extract of *Persea Americana*, Mill (Lauraceae) 'avocado' against CCl<sub>4</sub>-induced damage in rats," *African Journal of Traditional, Complementary and Alternative Medicines*, vol. 11, no. 2, pp. 237–244, 2014.
- [16] J. E. Mersereau, N. Levy, R. E. Staub et al., "Liquiritigenin is a plant-derived highly selective estrogen receptor beta agonist," *Molecular and Cellular Endocrinology*, vol. 283, no. 1-2, pp. 49–57, 2008.
- [17] Y. W. Kim, H. E. Kang, M. G. Lee et al., "Liquiritigenin, a flavonoid aglycone from licorice, has a choleric effect and the ability to induce hepatic transporters and phase-II enzymes," *The American Journal of Physiology—Gastrointestinal and Liver Physiology*, vol. 296, no. 2, pp. G372–G381, 2009.
- [18] J. K. Min, C. H. Lee, S. E. Jang et al., "Liquiritigenin ameliorates TNBS-induced colitis in mice," *Journal of Gastroenterology and Hepatology*, vol. 30, no. 5, pp. 858–865, 2015.
- [19] D. Wang, J. Lu, Y. Liu et al., "Liquiritigenin induces tumor cell death through mitogen-activated protein kinase- (MPAKs-) mediated pathway in hepatocellular carcinoma cells," *BioMed Research International*, vol. 2014, Article ID 965316, 11 pages, 2014.
- [20] J. Q. Ma, J. Ding, L. Zhang, and C. M. Liu, "Protective effects of ursolic acid in an experimental model of liver fibrosis through Nrf2/ARE pathway," *Clinics and Research in Hepatology and Gastroenterology*, vol. 39, no. 2, pp. 188–197, 2015.
- [21] M. M. Hafez, O. A. Al-Shabanah, N. O. Al-Harbi et al., "Association between paraoxonases gene expression and oxidative stress in hepatotoxicity induced by CCl<sub>4</sub>," *Oxidative Medicine and Cellular Longevity*, vol. 2014, Article ID 893212, 12 pages, 2014.
- [22] R. A. Khan, M. R. Khan, and S. Sahreen, "CCl<sub>4</sub>-induced hepatotoxicity: protective effect of rutin on p53, CYP2E1 and the antioxidative status in rat," *BMC Complementary and Alternative Medicine*, vol. 12, article 178, 2012.
- [23] C. J. Oh, J.-Y. Kim, A.-K. Min et al., "Sulforaphane attenuates hepatic fibrosis via NF-E2-related factor 2-mediated inhibition



- of transforming growth factor- $\beta$ /Smad signaling," *Free Radical Biology and Medicine*, vol. 52, no. 3, pp. 671–682, 2012.
- [24] M. Buck and M. Chojkier, "A ribosomal S-6 kinase-mediated signal to C/EBP- $\beta$  is critical for the development of liver fibrosis," *PLoS ONE*, vol. 2, no. 12, Article ID e1372, 2007.
- [25] Z. Wu, P. Puigserver, U. Andersson et al., "Mechanisms controlling mitochondrial biogenesis and respiration through the thermogenic coactivator PGC-1," *Cell*, vol. 98, no. 1, pp. 115–124, 1999.
- [26] G. Yakeu, L. Butcher, S. Isa et al., "Low-intensity exercise enhances expression of markers of alternative activation in circulating leukocytes: roles of PPAR $\gamma$  and Th2 cytokines," *Atherosclerosis*, vol. 212, no. 2, pp. 668–673, 2010.
- [27] Z. Lu, X. Xu, X. Hu et al., "PGC-1 $\alpha$  regulates expression of myocardial mitochondrial antioxidants and myocardial oxidative stress after chronic systolic overload," *Antioxidants & Redox Signaling*, vol. 13, no. 7, pp. 1011–1022, 2010.
- [28] R. D. Brose, G. Shin, M. C. McGuinness et al., "Activation of the stress proteome as a mechanism for small molecule therapeutics," *Human Molecular Genetics*, vol. 21, no. 19, pp. 4237–4252, 2012.
- [29] N. C. MacGarvey, H. B. Suliman, R. R. Bartz et al., "Activation of mitochondrial biogenesis by heme oxygenase-1-mediated NF-E2-related factor-2 induction rescues mice from lethal *Staphylococcus aureus* sepsis," *American Journal of Respiratory and Critical Care Medicine*, vol. 185, no. 8, pp. 851–861, 2012.



**Hindawi**  
Submit your manuscripts at  
<http://www.hindawi.com>

