

Research Article

Phytochemical Analysis Using UPLC-MSⁿ Combined with Network Pharmacology Approaches to Explore the Biomarkers for the Quality Control of the Anticancer Tannin Fraction of *Phyllanthus emblica* L. Habitat in Nepal

Lingfang Wu,^{1,2,3} Qiunan Zhang,¹ Wenyi Liang,¹ Yongben Ma,^{2,3} Liying Niu ,^{2,3} and Lanzhen Zhang ¹

¹School of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing 100102, China

²Hebei TCM Formula Granule Engineering and Technology Research Center, Hebei University of Chinese Medicine, Shijiazhuang 050091, China

³Hebei TCM Quality Evaluation & Standardization Engineering Research Center, Shijiazhuang 050091, China

Correspondence should be addressed to Living Niu; niuliyingyy@163.com and Lanzhen Zhang; zhanglanzhen01@126.com

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Phyllanthus emblica L. is widely used in traditional Tibetan medicine for its therapeutic effects on treating liver, kidney, and bladder problems. We have reported that the tannin fraction has a good anti-hepatocellular carcinoma effect, but its active ingredients are not clear. This study was to find the active ingredients of the tannin fraction using UPLC-MSⁿ and network pharmacology. First of all, the UPLC-MSⁿ method was employed to obtain high-resolution mass spectra of different components, and 110 compounds were obtained. Then a network pharmacology method was used to find biomarkers for quality control. Network pharmacology results showed that gallic acid, punicalagin A, punicalagin B, methyl gallate, geraniin, corilagin, chebulinic acid, chebulagic acid, and ellagic acid should be the biomarkers of the tannin fraction. Furthermore, 9 components were detected in the serum, which also proved that they could be biomarkers, because we generally believe that the ingredients which are absorbed into the blood are effective. In the end, a simple method for simultaneously determining the contents of the 9 compounds was constructed by HPLC-DAD. This research established a new method to find biomarkers of traditional Chinese medicine. This is of great significance to improving the quality standards of Tibetan medicine.

1. Introduction

Traditional Tibetan medicine has evolved from 2,300 years ago and still plays an important role in protecting human health. It is a vital part of traditional Chinese medicine. It can draw extensive attention for its mysterious nature and good effectiveness. *Phllanthus emblica* L. is widely used in traditional Tibetan medicine due to its numerous pharmacological applications in chronic diseases (for example, hypertension, hepatitis, blood stasis, and pharyngitis) [1–4]. It is an edible fruit indigenous to Southeast Asia and has been considered as a potent functional food. It is increasingly recognized that food and diet can maintain health and reduce the risk of chronic diseases. *Phllanthus emblica* L. exhibits several biological effects, antioxidant [5, 6], anti-age [7], anticancer [8], anti-car-diovascular diseases [9], anti-diabetes [10], anti-inflamma-tory [6, 11], anti-microbial [12], anti-diarrheal [13], immune-modulating [14], hepato- and gastroprotective activities [15], analgesic activities [16], and so on. Also, hydrolysable tannins may be effective substances [17–20].

As part of our phytochemical investigation of medicinal plants for the discovery of new bioactive natural products, we have already reported the chemical constituents [17, 18] isolated from *Phyllanthus emblica* L., and the tannin fraction has good antitumor activity [19, 20]. We also established the stable preparation processes of the tannin fraction of *Phyllanthus emblica* L. However, most of the chemicals in the tannin fraction remain unknown, making it difficult to rationalize its bioactivity or evaluate the safety of this material as a therapeutic agent. Therefore, there is an urgent need to develop an analytical method capable of determining the chemical compositions in the tannin fraction.

The therapeutic effects of traditional Chinese medicines (TCM) are based on the complex interactions of complicated chemical constituents as a whole system. It is obviously unreasonable to use only a few ingredients for quality control. It is also necessary to associate ingredients with activity. Thus, choosing the right ingredients to reflect the quality of traditional Chinese medicine is the key issue. We researched the relevant literature on the quality control of the tannin fraction of *Phyllanthus emblica* L. Some scholars used HPLC to determine the content of a few compounds in *Phyllanthus emblica* L. [21, 22], but there was no correlation between ingredients and efficacy.

This research established a new method to find biomarkers for the quality control of traditional Chinese medicine. We firstly used the UPLC-MSⁿ method to obtain high-resolution mass spectra of the different components. A total of 110 compounds including 45 hydrolysable tannins, 22 mucic acids, 15 phenolic acids, 15 flavonoids, 11 organic acids, and 2 other compounds were tentatively identified by comparing their retention times and mass spectrometry data with those of the reference compounds and reviewing the literature. Then, a network pharmacology method was used to find biomarkers for quality control based on the 110 identified compounds and anti-hepatocellular carcinoma effect. Network pharmacology results showed that gallic acid, punicalagin A, punicalagin B, methyl gallate, geraniin, corilagin, chebulinic acid, chebulagic acid, and ellagic acid might be the biomarkers of the tannin fraction, and these 9 components were detected in the serum, which also proves that they could be biomarkers, because we generally believe that the ingredients those are absorbed into the blood are effective. In the end, a simple method for simultaneously determining the contents of the 9 compounds was constructed using HPLC-DAD. To the best of our knowledge, this is the first report using UPLC-MSⁿ and network pharmacology approaches to find the boimarkers for the quality control of the tannin fraction of Phyllanthus emblica L. The method developed in our study also provides a scientific foundation for the study of anticancer effective substances of the tannin fraction of Phyllanthus emblica L.

2. Materials and Methods

2.1. Samples and Reagents. Methanol (HPLC grade and MS grade) was purchased from Fisher Scientific (Waltham, MA, USA). Distilled water was purchased from Watson's Food & Beverage Co., Ltd. (Guangzhou, China). Acetic acid (MS grade) was purchased from Fisher Scientific (Waltham, MA, USA). Reference standards of gallic acid (98%, CAS No: 149-91-7), punicalagin A (63%, CAS No: 65995-63-3), punicalagin B (37%, CAS No: 65995-63-3), methyl gallate (98%, CAS No: 99-24-1), geraniin (98%, CAS No: 2360976-49-0), corilagin (98%, CAS No: 23094-69-1), chebulinic acid (98%, CAS No: 18942-26-2), chebulagic acid (98%, CAS No:

23049-71-5), and ellagic acid (98%, CAS No: 476-66-4) were purchased from Chengdu-PUSH Bio-Technology Co., Ltd. (Chengdu, Sichuan, China).

2.2. Plant Materials and Sample Preparation. Phyllanthus emblica L. was purchased from Tibet and authenticated by Professor Chun-Sheng Liu (School of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing, China). Voucher specimens (PE001) of the plant were deposited at the authors' laboratory. The crude drug was extracted with ethanol and separated by HPD-400 macroporous resin column chromatography. The sample was dried and powdered, before being sieved through a 40 mesh sieve. A sample of the powder (approximately 25 mg) was suspended in 50 mL of methanol, and the resulting mixture was filtered through a $0.22 \,\mu$ m PTFE syringe filter. The filtrate was collected and subjected to centrifugation (13,000 rpm, 10 min). The supernatant was then transferred to an autosampler vial for analysis by UPLC-MS/MS and HPLC-DAD.

2.3. Apparatus and Parameters. The LTQ-Orbitrap XL UPLC-MS/MS instrument (Thermo Fisher, USA) was equipped with an ESI source used in negative ionization mode. The interface and MS parameters were as follows: nebulizer pressure, 100 kPa; dry gas, N₂ (1.5 L/min); drying gas temperature, 200°C; spray capillary voltage, 4000 V; scan range, m/z 100–1500. Mobile phase: A (methanol); B (H₂O : CH₃COOH, 100:0.2, ν/ν). Column: ACQUITY UPLC BEH C18 1.7 μ m (2.1 × 100 mm, Column; Part No: 1860023452; Serial No: 0246325825758), maintained at 30°C with flow rate of 0.3 mL·min⁻¹. The injection volume was 5μ L. Gradient elution procedure: 0 min (5% A) \rightarrow 5 min (15% A) \rightarrow 3 min (25% A) \rightarrow 10 min (30% A) \rightarrow 18 min (60% A) \rightarrow 26 min (90% A).

A Waters Alliance HPLC 2695 series instrument (Waters, Manchester, UK) was used to perform the high-performance liquid chromatography (HPLC) analysis. Mobile phase: A (methanol); B (H₂O:CH₃COOH, 100:0.2, ν/ν). Column: DiamansilTM C18 (250 × 4.6 mm, 5 μ m), maintained at 30°C with flow rate of 1.0 mL·min⁻¹. The detection wavelength was set at 270 nm for acquiring chromatograms. The injection volume was 20 μ L. Gradient elution procedure: 0 min (5% A) \rightarrow 10 min (15% A) \rightarrow 15 min (25% A) \rightarrow 30 min (30% A) \rightarrow 50 min (60% A) \rightarrow 55 min (90% A) \rightarrow 62 min (90% A).

2.4. Optimization of Analytical Conditions. To obtain better chromatographic separation and mass spectrometric detection, we evaluated three different mobile phase systems, including aqueous methanol, aqueous acetonitrile, and aqueous acetonitrile-formic acid solutions. The aqueous methanol solution resulted in the best separation of the major components of the tannin fraction of *Phyllanthus emblica* L. Furthermore, the addition of 0.2% acetic acid to this mobile phase resulted in a considerable improvement in the symmetry properties of the most chromatographic peaks. We also varied the flow rate (0.8, 1.0, and 1.2 mL/min) for HPLC analysis and (0.25, 0.3, and 0.35 mL/min) UPLC analysis, column temperature (25, 30, and 35°C) for HPLC and UPLC analysis, and injection volume (3, 5, and 10 μ L) for UPLC analysis during method development. The results of these optimization experiments established the following conditions for the chromatographic separation of the different components of the tannin fraction of *Phyllanthus emblica* L.

2.5. Structure Analysis Procedure. In the negative scan mode, based on the high-accuracy precursor ions and product ions obtained from UPLC-MS/MS, the elemental compositions were calculated when the maximum tolerance of mass error for the precursor ions and product ions was set at 1.5 ppm, which can satisfy the requirements for positive identification. Based on the elemental compositions of the precursors, the most rational molecular formula was sought in different chemical databases such as the Spectral Database for Organic Compounds SDBS, m/z cloud, and ChemSpider. Meanwhile, by searching literature sources, such as PubMed of the U.S. National Library of Medicine and the National Institutes of Health, Scifinder Scholar of the American Chemical Society, Science Direct of Elsevier, and Chinese National Knowledge Infrastructure (CNKI) of Tsinghua University, all components reported in the literature on Phyllanthus emblica L. and plants from the same family were summarized in a Microsoft Office Excel table to establish an in-house library [5, 7–13, 23] for searching the most rational molecular formula. When several matching compounds with the same formula were found, the fragmentation patterns and pathways of the compounds were analyzed and then validated by Mass Frontier 7.0 (Thermo Scientific) for positive identification.

2.6. Biomarkers Selected by Network Pharmacology and Ingredients Absorbed into the Blood. We followed the methods of Luo et al. 2020 [24]. Firstly, a network pharmacology method was used to find biomarkers for quality control based on the compounds identified by UPLC-MSⁿ and antihepatocellular carcinoma effect. Then to confirm that these compounds were proper quality control markers, animal experiments were conducted, with rats as test animals. We check whether these active ingredients are absorbed into the blood, because we generally believe that the ingredients those are absorbed into the blood are effective. The use of animals in the present study was permitted by the Ethics Committee of Beijing University of Chinese Medicine, and all animal studies were carried out according to the Guide for Care and Use of Laboratory Animals.

3. Results

3.1. Identification of the Compounds Present. UPLC-MS/MS method was employed to identify the components in the tannin fraction of *Phyllanthus emblica* L. The total ion chromatogram profile of the tannin fraction of *Phyllanthus emblica* L. was presented in negative mode, as shown in Figure 1(a). Molecular weights and fragmentation information (Table 1) were obtained. The possible structures of all peaks were deduced as shown in Figure 2. Under the optimized MS conditions, the negative mode was used to identify the peaks. 110 compounds including 45 hydrolysable tannins, 22 mucic acids, 15 phenolic acids, 15 flavonoids, 11 organic acids, and 2 other compounds have been tentatively identified by comparing their retention times and mass spectrometry data with that of reference compounds and reviewing the literature. Data for all of these compounds are summarized in Table 1.

3.1.1. Identification of Hydrolysable Tannins. 45 hydrolysable tannins have been identified in the tannin fraction of Phyllanthus emblica L., accounting for more than 41% (45/ 110). As shown in Table 2, in the negative mode ESI-MS^1 spectra, the [M–H]⁻ ion was observed for all compounds. In the negative mode ESI-MS² spectra, the [M–galloyl–H]⁻ ion was observed for 36 compounds, such as compounds 10, 17, 19, 21, 25, 26, 27, 31, 38, 42, 47, 49, 50, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 71, 74, 75, 76, 77, 78, 79, 86, 88, and 102. The [M-2 galloyl-H]⁻ ion was observed for 18 compounds, such as compounds 17, 19, 23, 25, 26, 38, 48, 56, 57, 62, 64, 66, 71, 74, 75, 76, 78, and 88. The [2M-H]⁻ ion was observed for 9 compounds, such as compounds 27, 31, 36, 46, 47, 52, 69, 70, and 76. The [M-galloyl-HHDP-H] ⁻ ion was observed for 6 compounds, such as compounds 47, 49, 55, 60, 61, and 86. The $[M-HHDP-H]^-$ ion was observed for 4 compounds, such as compounds 35, 36, 47, and 101. The [M-Ela-H]⁻ ion was observed for 2 compounds, such as compounds 39 and 41. The [M-3 galloyl-H]⁻ ion was observed for 1 compound, compound 88. The [M-THBDF-H]⁻ ion was observed for 1 compound, compound 65. From this result, it can be seen that [M-H]⁻ and [M-galloyl-H]⁻ are the most fragmented ions, followed by $[M-2 \text{ galloy}]-H]^-$ ions and $[2M-H]^-$ ions.

3.1.2. Identification of Mucic Acids. 22 mucic acids have been identified in the tannin fraction of Phyllanthus emblica L., accounting for more than 20% (22/110). In the negative mode ESI-MS1 spectra, the [M-H]⁻ ion was observed for all the compounds. In the negative mode ESI-MS2 spectra, the [M-galloyl-H]⁻ ion was observed for 8 compounds, such as compounds 7, 12, 14, 16, 24, 28, 34, and 37. The [2M-H]⁻ ion was observed for 5 compounds, such as compounds 4, 24, 28, 34, and 37. The [M-2CH₂-H]⁻ ion was observed for 3 compounds, such as compounds 18, 20, and 22. The [M-H₂O-H]⁻ ion was observed for 3 compounds, such as compounds 3, 5, and 7. The [M-CO₂-H]⁻ ion was observed for 2 compounds, such as compounds 3 and 4. The $[M-2CO_2-H]^-$ ion was observed for 1 compound, compound 4. From this result, it can be seen that [M-H]⁻ and [M-galloyl-H]⁻ are the most fragmented ions, followed by [2M-H]⁻ ions.

3.1.3. Identification of Phenolic Acids and Phenolic Acid Glycosides. 15 phenolic acids and their glycosides have been identified in the tannin fraction of *Phyllanthus* emblica L., accounting for about 14% (15/110). In the negative mode



FIGURE 1: UPLC-MSⁿ chromatogram of the tannin fraction of *Phyllanthus emblica* (L) in the negative mode (a); HPLC chromatogram of the tannin fraction of *Phyllanthus emblica* (L) (b) gallic acid (1), punicalagin A (2), punicalagin B (3), methyl gallate (4), geraniin (5), corilagin (6), chebulinic acid (7), chebulagic acid (8), and ellagic acid (9).

ESI-MS1 spectra, the [M-H]⁻ ion was observed for all compounds. In the ESI-MS2spectra, the [M-galloyl-H]⁻ ion was observed for 4 compounds, such as compounds 6, 9, 13, and 43. The [2M-H]⁻ ion was observed for 4 compounds, such as compounds 6, 9, 13, and 83. The [M-A-H]⁻ ion was observed for 4 compounds, such as compounds 33, 68, 72, and 80. (A represents various sugar groups). The [M-CO₂-H]⁻ ion was observed for 2 compounds, compounds 15 and 33. The $[M-2CO_2-H]^-$ ion was observed for 1 compound, compound 29. The $[M-CH_3-H]^-$ ion was observed for 1 compound, compound 53. The $[M-H_2O-H]^-$, $[M-CO-H]^-$ and $[M-CO-CO_2-H]^-$ ion was observed for 1 compound, compound 85. From this result, it can be seen that [M-H]⁻ and [M-galloyl-H]⁻ are the most fragmented ions, followed by [M-A-H]⁻ ions and $[2M-H]^{-}$ ions.

3.1.4. Identification of Flavonoids. 15 flavonoids have been identified in the tannin fraction of *Phyllanthus* emblica L.,

accounting for about 14% (15/110). In the negative mode ESI-MS1 spectra, the $[M-H]^-$ ion was observed for all compounds. In the negative mode ESI-MS2 spectra, the $[M-A-H]^-$ ion was observed for 6 compounds, such as compounds 81, 84, 90, 91, 98, and 99 (A represents various sugar groups). The $[M-galloyl-H]^-$ ion was observed for 2 compounds, such as compounds 77 and 102. The $[M-CO_2-H]^-$ ion was observed for 1 compound, compound 89. The $[M-CH_2-H]^-$ ion was observed for 1 compound, compound 67. The $[M-CH_3-H]^-$ ion was observed for 1 compound, compound 67. It can be seen that $[M-H]^-$ and $[M-A-H]^-$ are the most fragmented ions, followed by $[2M-H]^-$ ions.

3.1.5. Identification of Fatty Acid. 11 fatty acids have been identified in the tannin fraction of *Phyllanthus emblica* L., accounting for about 10% (11/110). In the negative mode $ESI-MS^1$ spectra, the $[M-H]^-$ ion was observed for all compounds. In the ESI-MS² spectra, the $[M-H_2O-H]^-$ ion

TABLE 1: The UPLC- MS^n data and compound names of the 110 peaks.

Peak no.	t _R (min)	Molecular formula	[M–H] ⁻	ppm	Negative mode	Identification
1^d	0.75	C ₇ H ₈ O ₇	203.0186	-0.18	MS ¹ :203.0186 [M–H] ⁻ , MS ² :159.0324 [M–CO ₂ –H] ⁻	2-oxo-3-carboxyadipic acid
2 ^d	0.86	$C_6H_{12}O_6$	179.0562	0.25	$MS^{1}:179.0562 \ [M-H]^{-}, \ MS^{2}:101.0242 \ [C_{4}H_{5}O_{3}]^{-}, \\ 89.0242 \ [C_{3}H_{5}O_{3}]^{-}, \ 71.01382 \ [C_{3}H_{2}O_{2}]^{-}$	Glucose
3 ^a	0.89	$C_6 H_{10} O_8$	209.0378	0.33	$MS^{4}: 209.0378 \ [M-H]^{-}, \ MS^{2}: 191.0267 \ [M-H_{2}O-H]^{-}, \\ 147.0304 \ [M-H_{2}O-CO_{2}-H]^{-}$	Mucic acid
4 ^a	0.93	$C_6H_8O_7$	191.0264	-0.14	MS ¹ :191.0264 [M–H] ⁻ , 383.0234 [2M–H] ⁻ MS ² : 147.0304 [M–CO ₂ –H] ⁻ , 119.0221 [M–CO ₂ –CO–H] ⁻	Mucic acid lactone
5 ^a	0.97	$C_4H_6O_5$	133.0131	0.23	$MS^{1}:133.0131 [M-H]^{-}, MS^{2}:115.0321 [M-H_{2}O-H]^{-}$	Malic acid
6 ^e	1.01	$C_{13}H_{16}O_{10}$	331.0659	0.34	MS ¹ : 331.0659 [M–H] ⁻ , 663.1325 [2M–H] ⁻ , MS ² : 169.0145 [galloyl]	6-O-galloylglucose
7 ^a	1.05	$C_{13}H_{14}O_{12}$	361.0406	0.22	MS ¹ : 361.0406 [M–H] ⁻ , MS ² : 209.0378 [M–galloyl–H] ⁻ , 191.0264 [M–galloyl–H ₂ O–H] ⁻	Mucic acid 2-O-gallate
$8^{\rm b}$	1.13	$C_{14}H_{12}O_{11}$	355.0345	-0.55	MS ¹ : 355.0345 [M–H] ⁻ , MS ² : 331.0666	Chebulic acid
9 ^e	1.17	$C_{13}H_{16}O_{10}$	331.0659	0.44	MS ¹ :331.0659 [M–H] [–] , 663.1325 [2M–H] [–] , MS ² : 169.0145 [galloyl]	2-O-galloylglucose
10 ^b	1.20	$C_{20}H_{18}O_{16}$	513.0589	0.33	MS ¹ : 513.0589 [M–H] ⁻ , MS ² : 361.0444 [M–H–galloyl] ⁻ , 209.0344 [M–H–2 galloyl] ⁻	Mucic acid digallate
11 ^d	2.57	$C_6H_8O_7$	191.0198	0.93	$MS^{-}:191.0198 [M-H]$, 383.0234 [2M-H], $MS^{-}:$ 147.1211 [M-CO ₂ -H] ⁻ 129.0392 [M-CO ₂ -H ₂ O-H] ⁻	Citric acid
12 ^a	1.25	C ₁₃ H ₁₂ O ₁₁	343.0305	-0.29	$MS^{1}: 343.0305 [M-H]^{-}, MS^{2}: 191.0198 [M-galloyl-H]^{-}$	Mucic acid lactone gallate
13 ^e	1.97	$C_{13}H_{16}O_{10}$	331.0659	0.27	MS ¹ : 331.0659 [M–H] ⁻ , 663.1325 [2M–H] ⁻ , MS ² : 169.0145 [gallovl]	1-O-galloylglucose
14^{a}	2.16	$C_{13}H_{12}O_{11}$	343.0305	0.77	MS ¹ : 343.0305 [M-H] ⁻ , MS ² : 191.0198 [M-galloyl-H] ⁻	Mucic acid lactone gallate
15 ^e	2.32	$C_7H_6O_5$	169.0122	-0.38	$MS^{1}:169.0122 [M-H]^{-}, MS^{2}:125.0338 [M-CO_{2}-H]^{-}$	Gallic acid
16 ^a	2.43	$C_{14}H_{16}O_{12}$	375.0558	-0.72	MS ¹ : 375.0558 [M–H] ⁻ , MS ² : 223.0434 [M–galloyl–H] ⁻	gallate
17 ^b	2.50	$C_{20}H_{18}O_{16}$	513.0589	0.66	MS ¹ : 513.0589 [M–H] ⁻ , MS ² : 361.0444 [M–H–galloyl] ⁻ , 209.0344 [M–H–2 galloyl] ⁻	Mucic acid digallate
18 ^a	2.57	$C_{14}H_{16}O_{12}$	375.0558	0.90	MS ¹ : 375.0558 [M–H] ⁻ , MS ² : 345.0323 [M–H–2CH ₃] ⁻ , 331.0711	Mucic acid methyl ester gallate
19 ^b	2.69	$C_{20}H_{18}O_{16}$	513.0589	-0.33	MS ¹ : 513.0589 [M–H] ⁻ , MS ² : 361.0444 [M–H–galloyl] ⁻ , 209.0344 [M–H–2 galloyl] ⁻	Mucic acid digallate
20 ^a	2.89	$C_{14}H_{16}O_{12}$	375.0558	-0.48	MS ¹ : 375.0558 [M–H] ⁻ , MS ² : 345.0323 [M–H–2CH ₃] ⁻ , 331.0711	Mucic acid methyl ester gallate
21 ^b	3.11	$C_{20}H_{18}O_{16}$	513.0589	0.78	MS ¹ : 513.0589 [M–H] ⁻ , MS ² : 361.0444 [M–H–galloyl] ⁻ , 209.0344 [M–H–2galloyl] ⁻	Mucic acid digallate
22 ^a	3.24	$C_{14}H_{16}O_{12}$	375.0558	0.92	MS ¹ : 375.0558 [M–H] ⁻ , MS ² : 345.0323[M–H–2CH ₃] ⁻ , 331.0711	Mucic acid methyl ester gallate
23 ^b	4.08	$C_{20}H_{16}O_{15}$	495.0405	-0.93	MS ¹ : 495.0405 [M–H] ⁻ , MS ² : 343.0356 [M–H–galloyl] ⁻ , 191.0223 [M–H–2 galloyl] ⁻	Mucic acid lactone digallate
24 ^a	4.75	$C_{15}H_{18}O_{12}$	389.0714	0.22	MS ¹ : 389.0714 [M–H] ⁻ , 779.1529 [2M–H] ⁻ , MS ² : 237.0564 [M–H–gallov]] ⁻	Mucic acid dimethyl ester gallate
25 ^b	5.10	$C_{20}H_{18}O_{16}$	513.0589	0.44	MS ¹ : 513.0589 [M–H] ⁻ , MS ² : 361.0444 [M–H–galloyl] ⁻ , 209.0344 [M–H–2 galloyl] ⁻	Mucic acid digallate
26 ^b	5.19	$C_{21}H_{20}O_{16}$	527.0667	0.73	MS ¹ : 527.0667 [M–H] ⁻ , MS ² : 375.0557 [M–H–galloyl] ⁻ , 223.0448 [M–H–2 galloyl] ⁻	Mucic acid methyl ester digallate
27 ^b	5.41	$C_{20}H_{20}O_{14}$	483.0769	-0.92	MS ¹ :483.0769 [M–H] ⁻ , 967.1610 [2M–H] ⁻ MS ² : 331.0712 [M–H–gallovl] ⁻ , 271.0523, 169.0139	1, 4-di-O-galloylglucose
28 ^a	4.79	$C_{11}H_{10}O_9$	285.0241	0.4	MS ¹ : 285.0250 [M–H] ⁻ , 571.0583 [2M–H] ⁻ , MS ² : 133.0122 [M–H–galloyl] ⁻	Malic acid gallate
29 ^e	5.89	$C_{14}H_{16}O_{10}$	343.0659	0.38	MS ¹ : 343.0659 [M–H] ⁻ , MS ² : 255.0522 [M–H–2CO ₂] ⁻ , 169.0141	3-Galloylquinic acid
30 ^b	6.17	$C_{27}H_{26}O_{20}$	669.0933	-0.65	MS ¹ : 669.0933 [M–H] ⁻ , MS ² : 337.0199 [M–H–galloyl–H ₂ O–Hex] ⁻	Phyllanemblinin D
31 ^b	6.44	$C_{20}H_{20}O_{14}$	483.0769	0.89	MS ⁺ :483.0769 [M–H] ⁻ , 967.1610 [2M–H] ⁻ MS ² : 331.0712 [M–H–galloyl] ⁻ , 271.0523, 169.0139	1, 6-di-O-galloylglucose
32 ^b	6.52	$C_{27}H_{26}O_{20}$	669.0933	-0.19	MS ¹ : 669.0933 [M–H] ⁻ , MS ² : 337.0199 [M–H–galloyl–H ₂ O–Hex] ⁻	Phyllanemblinin E

TABLE 1: Continued.

Peak no.	t _R (min)	Molecular formula	[M–H] [–]	ppm	Negative mode	Identification
33 ^e	6.77	$C_{15}H_{18}O_9$	341.0867	-0.28	MS ¹ : 341.0867 [M–H] ⁻ , MS ² : 297.0989 [M–H–CO ₂] ⁻ , 179.0234 [M–H-glc] ⁻	Caffeic acid-3-glucoside
34 ^b	6.88	$C_{14}H_{14}O_{11}$	357.0452	-0.23	MS ¹ : 357.0452 [M–H] ⁻ , 715.5998 [2M–H] ⁻ , MS ² : 205.0332 [M–H–galloyl] ⁻	Mucic acid lactone methyl ester digallate
35 ^b	3.84	$C_{48}H_{28}O_{30}$	1083.0581	0.99	MS ¹ :1083.0581 [M–H] ⁻ , MS ² :541.0836 [M–2H] ^{2–} , 781.3025 [M–H–HHDP] ⁻ , 300.9921	Punicalagin A
36 ^b	5.90	$C_{48}H_{28}O_{30}$	1083.0581	0.34	MS ¹ :1083.0581 [M–H] ⁻ , MS ² :541.0836 [M–2H] ^{2–} , 781.3025 [M–H–HHDP] ⁻ , 300.9921	Punicalagin B
37 ^b	7.36	$C_{14}H_{14}O_{11}$	357.0452	0.92	MS ¹ : 357.0452 [M–H] ⁻ , 715.5998 [2M–H] ⁻ , MS ² : 205.0433 [M–H–galloyl] ⁻	Mucic acid lactone methyl ester digallate
38 ^b	7.46	$C_{21}H_{20}O_{16}$	527.0667	0.66	MS : 527.0667 [M-H] , MS : 375.0557 [M-H-galloy1] , 223.0448 [M-H-2 galloy1]	digallate
39 ^b	7.51	$C_{33}H_{28}O_{24}$	807.0896	0.71	MS ¹ : 807.0896 [M–H] ⁻ , MS ² : 483.0789 [M–H–Ela] ⁻ , 331.0189 [M–H–Ela–galloyl] ⁻ , 169.0143	Mallonin
40 ^c	7.91	$C_{15}H_{14}O_7$	305.0655	0.59	MS ¹ : 305.0655 [M–H] ⁻ , MS ² : 215.0098	Gallocatechin
41^{b}	8.00	$C_{33}H_{28}O_{24}$	807.0896	0.49	$MS^{+}: 807.0896 [M-H]^{-}, MS^{2}: 483.0789 [M-H-Ela]^{-},$ 331.0189 [M-H-Ela-gallovl] ⁻ 169.0143	Mallonin
42 ^b	8.03	$C_{20}H_{16}O_{15}$	495.0405	0.88	MS ¹ : 495.0405 [M–H] ⁻ , MS ² : 343.0356 [M–H–galloyl] ⁻ , 191.0223 [M–H–2 galloyl] ⁻	Mucic acid lactone digallate
43 ^e	8.34	$C_{14}H_{10}O_9$	321.0251	-0.46	MS ¹ : 321.0251 [M–H] ⁻ , MS ² : 169.1144 [galloyl] ⁻	Digallate
44 ^b	8.37	C ₂₇ H ₂₆ O ₂₀	669.0933	0.81	$MS^{1}: 669.0933 [M-H]^{-}, MS^{2}: 337.0199$	Phyllanemblinin F
45 ^e	8.60	$C_{10}H_{12}O_7$	243.1600	0.24	$[M-H-galloyI-H_2O-Hex]$ MS ¹ :243.1600 $[M-H]^-$, MS ² :169.0144 [galloyl] ⁻ , 125.0246	1-O-galloyl-glycerol
46 ^b	8.78	$C_{20}H_{20}O_{14}$	483.0769	0.34	MS ¹ :483.0769 [M–H] ⁻ , 967.1610 [2M–H] ⁻ MS ² : 331.0712 [M–H–galloyl] ⁻ , 271.0523, 169.0139	3, 6-di-O-galloylglucose
47 ^b	8.84	$C_{46}H_{36}O_{31}$	1083.1156	0.66	MS ¹ :1083.1156 [M–H] ⁻ , MS ² :541.0836 [M–2H] ²⁻ , 781.0341 [M–H–HHDP] ⁻ , 629.0609 [M–H–HHDP–galloy] ⁻ , 301.0115	Putranjivain A
48 ^b	9.03	$C_{21}H_{20}O_{16}$	527.0667	0.71	MS ¹ : 527.0667 [M–H] ⁻ , MS ² : 375.0557 [M–H–galloyl] ⁻ , 223.0448 [M–H–2 galloyl] ⁻	Mucic acid methyl ester digallate
49 ^b	9.46	$C_{41}H_{28}O_{27}$	951.0734	0.26	MS ¹ :951.0734 [M–H] ⁻ , MS ² :799.0563 [M–H–galloyl] ⁻ , 497.0219 [M–H–galloyl–HHDP] ⁻ , 301.0113	Geraniin
50 ^b	9.53	$C_{41}H_{30}O_{27}$	969.0839	0.22	$MS^{1}:969.0839 [M-H]^{-}, MS^{2}:817.0783 [M-H-galloyl]^{-}$ $MS^{1}:971.0996 [M-H]^{-}, MS^{2}:953.0906 [M-H-H_2O]^{-}$	Phyllanemblinin C
51 ^b	9.72	$C_{41}H_{32}O_{28}$	971.0996	0.30	935.0800 $[M-H-2H_2O]^-$, 467.0361 $[M-2H-H_2O]^{2-}$, 300.9911	Neochebulagic acid
52 ^b	9.39	$C_{41}H_{30}O_{27}$	953.0890	0.50	MS ¹ :953.0890 [M–H] ⁻ , MS ² :476.0412 [M-2H] ²⁻ , 300.9983	Terchebin
53 ^e 54 ^b	10.14 10.25	$\begin{array}{c} C_8 H_8 O_5 \\ C_{27} H_{24} O_{19} \end{array}$	183.0259 651.0828	0.61 0.11	MS ¹ :183.0259 [M–H] ⁻ , MS ² :169.0064 [M–H–CH ₃] ⁻ MS ¹ :651.0828 [M–H] ⁻ , MS ² :499.0782[M–H–galloyl] ⁻	Methyl gallate Chebulanin
55 ^b	10.33	$C_{47}H_{34}O_{32}$	1109.0949	0.03	MS ⁻ :1109.0949 [M–H], MS ⁻ :957.0466 [M–galloyl–H] ⁻ , 655.0497 [M–galloyl–HHDP–H] ⁻ , 300.0001	Elaeocarpusin
56 ^b	10.48	$C_{27}H_{24}O_{18}$	635.0878	0.06	MS ¹ :635.0878 [M–H] ⁻ , MS ² :465.0679 [M–galloyl–H ₂ O–H] ⁻ , 313.0560 [M–H–galloyl–galloyl–H ₂ O] ⁻ , 169.1045	Trigalloylglucose
57 ^b	10.64	$C_{41}H_{30}O_{26}$	937.0941	0.09	MS ¹ :937.0941 [M–H] ⁻ , MS ² :785.0648 [M–galloyl–H] ⁻ , 633.0566 [M–2 galloyl–H] ⁻ , 331.0516 [M–H–2 galloyl–HHDP–H] ⁻ 300 9416	Punicafolin
58 ^b	10.56	$C_{41}H_{28}O_{26}$	935.0785	0.31	MS ¹ :935.0785 [M–H] ⁻ , MS ² :783.0648 [M–galloyl–H] ⁻	Casuarinin
59 ^b	10.73	$C_{27}H_{22}O_{18}$	633.0728	0.92	MS ¹ : 633.0728 [M–H] ⁻ , MS ² : 463.0511 [M–galloyl–H] ⁻ , 301.0221	Corilagin
60 ^b	10.73	$C_{27}H_{22}O_{18}$	633.0738	0.39	MS ¹ : 633.0738 [M–H] ⁻ , MS ² : 481.0511 [M–galloyl–H] ⁻ , 331.0598 [M–HHDP–H] ⁻ , 300.9601	Phyllanemblinin B
61 ^b	10.73	$C_{27}H_{22}O_{18}$	633.0722	0.34	MS ¹ :633.0722 [M–H] ⁻ , MS ² :481.0511 [M–galloyl–H] ⁻ , 331.0598 [M–HHDP–H] ⁻ , 300.9601	Isostrictinin
62 ^b	11.14	$C_{20}H_{16}O_{15}$	495.0405	0.66	MS ¹ : 495.0405 [M–H] ⁻ , MS ² : 343.0356 [M–H–galloyl] ⁻ , 191.0223 [M–H–2 galloyl] ⁻	Mucic acid lactone digallate

TABLE 1: Continued.

Peak no.	t _R (min)	Molecular formula	[M–H] [–]	ppm	Negative mode	Identification
63 ^b	11.25	$C_{33}H_{26}O_{24}$	805.0564	0.66	$MS^{1}: 805.0564 [M-H]^{-}, MS^{2}: 653.0432 [M-galloyl-H]^{-}$	Mallonin
64 ^b	11.65	$C_{27}H_{24}O_{18}$	635.0878	-0.39	$[M-H-galloy]-H_2O]^-$, 313.0560 $[M-H-2]$ galloyl-H ₂ O] ⁻ , 169.1045	Trigalloylglucose
65 ^b	11.88	$C_{27}H_{20}O_{17}$	615.0616	0.09	MS ¹ : 615.0616 [M–H] ⁻ , MS ² : 463.0506 [M–H–galloyl] ⁻ , 177.0534 [M–H–galloyl–THBDF] ⁻	Phyllanemblinin A
66 ^b	11.92	$C_{34}H_{26}O_{22}$	785.0831	0.01	MS ¹ :785.0831 [M–H] ⁻ , MS ² :633.0726 [M–H–galloyl] ⁻ , 463.0685 [M–H–2 galloyl–H ₂ O] ⁻ , 300.9482	Digalloyl-HHDP-glucose
67 ^c	12.31	$C_{15}H_{14}O_{6}$	289.0708	0.22	MS ¹ :289.0708 [M–H] ⁻ , MS ² :275.0192 [M–H–CH ₂] ⁻ , 215.0094	Epicatechin
68 ^e	12.35	$C_{20}H_{16}O_{13}$	463.0507	0.11	MS ¹ :463.0507 [M–H] ⁻ , 927.1101 [2M–H] ⁻ , MS ² : 300.9982 [M–H–Hex] ⁻	Ellagic acid hexose
69 ^b	12.47	$C_{41}H_{30}O_{27}$	953.0890	0.51	$MS^{1}:953.0890 [M-H]^{-}, MS^{2}:476.0412 [M-2H]^{2-},$ 300.9983	Chebulinic acid
70 ^b	12.47	$C_{41}H_{30}O_{27}$	953.0890	0.62	MS ¹ :953.0890 [M–H] ⁻ , MS ² :476.0412 [M–2H] ^{2–} , 300.9983	Chebulagic acid
71 ^b	12.88	$C_{34}H_{26}O_{22}$	785.0831	-0.71	MS ¹ :785.0831 [M–H] ⁻ , MS ² :633.0726 [M–H–galloyl] ⁻ , 463.0685 [M–H–2 galloyl–H ₂ O] ⁻ , 300.9922	Digalloyl-HHDP-glucose
72 ^e	13.00	$C_{19}H_{14}O_{12}$	433.0401	0.66	$MS^{1}:433.0401 [M-H]^{-}, 867.0895 [2M-H]^{-}, MS^{2}:$ 300.9991 [M-H-pent]^	Ellagic acid pentose
73 ^c	13.23	$C_{15}H_{14}O_{6}$	289.0716	-0.11	MS ¹ : 289.0716 [M–H] ⁻ , MS ² : 241.0354 [M–H–2CH ₃ –H ₂ O] ⁻ , 215.0094	Catechin
74 ^b	13.73	$C_{34}H_{26}O_{22}$	785.0831	0.19	MS ¹ :785.0831 [M–H] ⁻ , MS ² :633.0726 [M–H–galloyl] ⁻ , 463.0685 [M–H–2 gallovl–H ₂ O] ⁻ , 300.9911	Digalloyl-HHDP-glucose
75 ^b	13.80	$C_{34}H_{28}O_{22}$	787.0988	0.29	MS ¹ :787.0988 [M–H] ⁻ , MS ² :483.0785 [M–H–2 galloyl] ⁻	1, 2, 3, 6-tetra-O- galloylglucose
76 ^b	14.03	$C_{41}H_{30}O_{26}$	937.0941	-0.13	MS ¹ :937.0941 [M–H] ⁻ , MS ² :785.0848 [M–H–galloyl] ⁻ , 633.0737 [M–H–2 galloyl] ⁻ , 481.0411[M–H–3 galloyl] ⁻ , 468.0442 [M–2H] ^{2–} 300.9991	Trigalloyl-HHDP-glucose
77 ^c	14.06	$C_{28}H_{24}O_{16}$	615.0980	0.51	MS ¹ : 615.0980 [M–H] ⁻ , MS ² : 463.0893 [M–H–galloy] ⁻	2'-O-galloylhyperin
78 ^b	14.12	$C_{34}H_{22}O_{22}$	781.0518	0.11	$MS^{-}:781.0518 [M-H], MS^{-}:629.0521 [M-H-galloy1], 477.0443 [M-H-2 galloy1]^{-}, 175.0551 [M-H-2 galloy1]^{-}, 200.0550 [M-10 gall0]^{-}, 200.0550 [M-1$	Emblicanin A
79 ^b	10.40	C ₄₈ H ₃₂ O ₃₂	1119.0792	0.19	MS ¹ :1119.0792 [M–H] ⁻ , MS ² :967.0893 [M–H–galloyl] ⁻	Mallotusinic acid
80 ^e	14.53	$C_{19}H_{14}O_{12}$	433.0401	0.17	$MS^{1}:433.0401 [M-H]^{-}, 867.0895 [2M-H]^{-}, MS^{2}:$ 300.9991 $[M-H-pent]^{-}$	Ellagic acid pentose
81 ^c	17.20	$C_{28}H_{24}O_{15}$	599.1046	-0.22	MS ¹ :599.1046 [M–H] ⁻ , MS ² :285.0393 [M–galloylgalactoside–H] ⁻ , 153.01813	Kaempferol-3-(6"- galloylgalactoside)
82 ^c	14.86	$C_{15}H_{12}O_5$	271.0611	0.92	$MS^{-}: 2/1.0611 [M-H], MS^{-}: 1/7.0197 [M-C_{6}H_{7}O-H], $ $151.0035 [M-C_{6}H_{7}O-2CH_{3}-H]^{-}, 119.0012 $ $[M-C_{7}H_{7}O-2CH_{3}-2CH_{4}-H]^{-}$	Naringenin
83 ^e	15.02	$C_{20}H_{16}O_{12}$	447.0558	0.62	$MS^{-1}:447.0558 [M-H]^{-1}, MS^{-2}:300.9991$	Ellagic acid deoxyhexose
84° 85°	15.10 15.22	$C_{21}H_{20}O_{12}$ $C_{14}H_6O_8$	463.0871 300.9978	-0.15 -0.71	$MS^{2}: 463.0871 [M-H]$, $MS^{2}: 301.0312 [M-H-hexose]$ $MS^{1}: 300.9978 [M-H]^{-}$, $MS^{2}: 283.2637 [M-H-H_{2}O]^{-}$,	Quercetin hexose Ellagic acid
86 ^b	15 49	CuHacOas	917 0679	0.66	$M^{-1}=CO^{-$	Mallotusinin
87 ^c	15.97	$C_{41}H_{26}O_{25}$ $C_{15}H_{14}O_{5}$	273.0769	0.92	463.0335 [M–H–galloyl–HHDP] ⁻ , 300.9091 MS ¹ : 273.0769 [M–H] ⁻ , MS ² : 215.0100	Epiafzelechin
88 ^b	16.17	$C_{41}H_{30}O_{26}$	937.0941	-0.39	MS ¹ :937.0941 [M–H] ⁻ , MS ² :785.0848 [M–H–galloyl] ⁻ , 633.0737 [M–H–2 galloyl] ⁻ , 481.0411 [M–H–3 galloyl] ⁻ ,	Trigalloyl-HHDP-glucose
89 ^e	16.21	C ₉ H ₁₀ O ₅	197.0444	0.09	468.0442 $[M-2H]^{-7}$, 300.9922 MS ¹ :197.0444 $[M-H]^{-7}$, MS ² :153.322 $[M-CO_2-H]^{-7}$	Vanillylmandelic acid
90 ^c	16.24	C ₂₁ H ₂₀ O ₁₁	447.0921	0.01	$MS^{1}:447.0921 [M-H]^{-}, MS^{2}:300.9988 [M-H-decyrbay]^{-}$	Quercetin deoxyhexose
91 ^c	16.24	$C_{21}H_{20}O_{11}$	447.0927	0.44	MS ¹ :447.0921[M–H] ⁻ , MS ² :287.0545 [M–H–Glc] ⁻	Luteolin-7-galactoside
92 ^e	16.68	$\mathrm{C_{15}H_8O_8}$	315.0147	-0.72	MS ⁺ : 315.0147 [M–H] ⁻ , MS ² : 300.0991 [M–H–CH ₃] ⁻ , 212.9015 [M–H–CH ₃ –2CO ₂] ⁻	3-O-Methylellagic acid

Peak no.	$t_{\rm R}$ (min)	Molecular formula	[M–H] ⁻	ppm	Negative mode	Identification
93 ^d	16.91	$C_{12}H_{20}O_5$	243.1227	0.41	MS ¹ :243.1227 [M–H] ⁻ , MS ² :225.1125 [M–H–H ₂ O] ⁻ , 207.5586 [M–H–2H ₂ O] ⁻ , 133.2019 [M–H–2H ₂ O–2CH ₃ –CO ₂] ⁻	Methyl 5, 10-dihydroxy-10- methoxydeca-6, 8-dienoate
94 ^b	17.36	$C_{29}H_{36}O_8$	511.2326	0.38	MS ¹ :511.2326 [M–H] ⁻ , MS ² :468.2119 [M–CH ₃ –CO–H] ⁻ , 425.1767 [M–2CH ₃ –2CO–H] ⁻	Mallotojaponin C
95 ^c	17.56	$C_{21}H_{20}O_{10}$	431.0983	-0.15	MS ¹ :431.0983 [M–H] ⁻ , MS ² :416.0093 [M–H–CH ₃] ⁻	Vitexin
96 ^e	18.50	$C_{16}H_{10}O_8$	329.0302	-0.19	MS ¹ : 329.0302 [M–H] ⁻ , MS ² : 255.2328 [M–H–CO ₂ –2CH ₃] ⁻	3, 4-di-O-methylellagic acid
97 ^d	18.56	$C_{10}H_{18}O_4$	201.1132	-0.18	$MS^1:201.1132 [M-H]^-, MS^2:183.1023 [M-H-H_2O]^-$	2-hydroxy-4-oxo-decanoic acid
98 ^c	18.80	$C_{30}H_{28}O_{13}$	595.1446	-0.13	MS ¹ : 595.1446 [M–H] ⁻ , MS ² : 287.0382 [M–H–coumaroylhexose] ⁻	Eriodictyol coumaroylhexose
99 ^c	19.06	$C_{30}H_{28}O_{13}$	595.1446	0.19	MS ¹ : 595.1446 [M–H] ⁻ , MS ² : 287.0382 [M–H–coumaroylhexose] ⁻	Eriodictyol coumaroylhexose
100 ^d	19.20	$C_{12}H_{20}O_4$	227.1288	0.14	MS ¹ :227.1288 [M–H] ⁻ , MS ² :183.0382 [M–H–CO ₂] ⁻ , 157.1079 [M–H–CO ₂ –CH = CH] ⁻	Traumatic acid
101 ^b	20.25	$C_{34}H_{20}O_{22}$	779.0362	0.01	MS ¹ :779.0362 [M–H] ⁻ , MS ² :477.0342 [M–H– HHDP] ⁻ , 300.9809	Emblicanin B
102 ^c	20.56	$C_{28}H_{24}O_{14}$	583.1082	0.91	MS ¹ :583.1082 [M–H] ⁻ MS ² :431.0984 [M–H–galloyl] ⁻ , 331.0561	2"-O-galloylisovitexin
103 ^d	22.31	$C_{11}H_{20}O_4$	215.1277	0.49	MS ¹ :215.1277 [M–H] ⁻ , MS ² :197.1181 [M–H–H ₂ O] ⁻ , 153.1286 [M–H–H ₂ O–CO ₂] ⁻	5-hydroxy-3-methoxydec-2- enoic acid
104 ^b	22.54	$C_{25}H_{30}O_8$	457.1856	0.14	MS ¹ :457.1856 [M–H] ⁻ , MS ² :414.1181 [M–H– CH ₃ CO] ⁻ , 371.1286 [M–H–2CH ₃ CO] ⁻	Mallotojaponin B
105 ^d	25.56	$C_{18}H_{28}O_3$	291.1954	0.66	MS ¹ :291.1954 [M–H] ⁻ , MS ² :247.0332 [M–H–CO ₂] ⁻	Licanic acid
106 ^d	26.07	$C_{18}H_{36}O_4$	315.2535	-0.24	MS ¹ :315.2535 [M–H] ⁻ , MS ² :297.1526 [M–H–H ₂ O] ⁻ , 253.1221 [M–H–H ₂ O–CO ₂] ⁻	Dihydroxystearic acid
107 ^d	26.24	$C_{18}H_{30}O_3$	293.2111	-0.29	MS ¹ :293.2111 [M–H] ⁻ , MS ² :275.2159 [M–H–H ₂ O] ⁻ , 231.1441 [M–H–H ₂ O–CO ₂] ⁻	9-hydroxyoctadeca-5, 10, 12- trienoic acid
108 ^d	27.01	$C_{18}H_{36}O_2$	283.2641	0.33	MS ¹ :283.2641 [M–H] ⁻ , MS ² :265.1576 [M–H–H ₂ O] ⁻ , 237.0132 [M–H–H ₂ O–CO] ⁻	3-Hydroxyoctadecanal
109 ^d	28.73	$C_{30}H_{48}O_3$	455.3519	0.29	MS ¹ :455.3519 [M–H] ⁻ , MS ² :401.0874 [M–H–CO ₂] ⁻	Ursolic acid
110 ^d	30.71	$C_{16}H_{32}O_2$	255.2318	0.33	MS ¹ :255.2318 [M–H] ⁻ , MS ² :211.0521 [M–H–CO ₂] ⁻	Palmitic acid

a: mucic acid, b: hydrolysable tannin, c: flavonoids, d: fatty acid, and e: phenolic acids.

was observed for 6 compounds, such as compounds 93, 97, 103, 106, 107, and 108. The $[M-CO_2-H]^-$ ion was observed for 6 compounds, such as compounds 1, 11, 100, 105, 109, and 110. The $[M-CO_2-H_2O-H]^-$ ion was observed for 4 compounds, such as compounds 11, 103, 106, and 107. The $[M-CH_3-2CO_2-H]^-$ ion and $[M-CH_3-H]^-$ ion were observed for compound 92. From this result, it can be seen that $[M-H]^-$ and $[M-H_2O-H]^-$ are the most fragmented ions, followed by $[M-CO_2-H]^-$ ions.

3.2. Biomarkers Selected by Network Pharmacology and Ingredients Absorbed into the Blood. 228 potential targets related to the 110 compounds were obtained by using Swiss Target Prediction and TCMSP databases. And 7392 potential targets related to hepatocellular carcinoma were obtained according to OncoDB.HCC and Liverome databases. Through protein-protein interaction analysis, 120 targets with higher correlation were obtained, as shown in Figure 3. The DAVID database was used to conduct GO enrichment analysis on 120 targets with *p*-value less than 0.01, as shown in Figure 4. Finally, the Cytoscape 3.7.1 software was used to visualize the "component-targetfunction" network, as shown in Figure 5. 9 compounds, 72 proteins, and 20 pathways were obtained. Of the 20 pathways, PI3K-Akt signaling pathway, HIF-1 signaling pathway, Ras signaling pathway, ErbB signaling pathway, FoxO signaling pathway, and VEGF signaling pathway are related to anticancer effect [25–31], these pathways may be related to the anti-cancer effect of the tannin fraction of *Phyllanthus emblica* L. And the 9 compounds including gallic acid, punicalagin A, punicalagin B, methyl gallate, geraniin, corilagin, chebulinic acid, chebulagic acid, and ellagic acid were all detected in the rat serum by using UPLC-MS/MS, which further verified that these compounds were proper biomarkers. Detailed information about the analysis of chemical components in rat serum can be found in the supplementary materials.

3.3. Validation of the HPLC Method. The method was validated in terms of linearity, precision, stability, repeatability, and recovery test.

The concentrations of gallic acid, punicalagin A, punicalagin B, methyl gallate, geraniin, corilagin, chebulinic acid, chebulagic acid, and ellagic acid in the stock solution were



27

óн



óн

35



(b) FIGURE 2: Continued.







53

ОН







56







59

(c) FIGURE 2: Continued.



FIGURE 2: Continued.



















85









93 (e) FIGURE 2: Continued.



FIGURE 2: Structures of chemical constituents in the tannin fraction of Phyllanthus emblica L.

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		-		
Analytes	Calibration curve	Linear range (ng/mL)	R^2	LOQ (ng/mL)
Gallic acid	Y = 2E + 06X - 30393	4.92~93.60	0.9991	2.56
Punicalagin A	Y = 3E + 06X - 66826	2.88~69.70	0.9992	1.44
Punicalagin B	Y = 2E + 06X - 77857	5.27~125.50	0.9993	1.03
Methyl gallate	Y = 3E + 06X - 55968	2.33~74.55	0.9991	0.46
Geraniin	Y = 741189X - 29791	6.00~192.00	0.9994	1.20
Corilagin	Y = 844347X - 75313	13.75~440.00	0.9997	0.28
Chebulinic acid	Y = 4E + 06X - 87496	2.74~87.65	0.9991	1.35
Chebulagic acid	Y = 886311X - 15698	2.10~67.20	0.9993	1.05
Ellagic acid	Y = 1E + 06X - 65693	0 45~144 00	0 9993	0.22

TABLE 2: Calibration curves of the analytes.



FIGURE 3: PPI network of "compound-target" in the tannin fraction of *Phyllanthus emblica* L. The bigger the graph, the bigger the degree. PPI: protein-protein interaction.

0.09360, 0.1152, 0.2107, 0.0932, 0.3200, 0.5500, 0.1096, 0.08400, and 0.1800 $\mu g \mu L$ -1, respectively. The stock solution was then diluted to appropriate concentration range for the establishment of calibration curves. The calibration curves were constructed by plotting the peak area (*Y*) versus the concentration (*X*, μg) of each standard solution. Detailed information regarding the calibration curves and linear ranges is listed in Table 2. All calibration curves showed good linear regression within the test ranges.

The precision was determined by replicate injection with the same sample solution six consecutive times. The RSDs of peak area of gallic acid, punicalagin A, punicalagin B, methyl gallate, geraniin, corilagin, chebulinic acid, chebulagic acid, and ellagic acid were all below 3.05%, which showed high precision.

Stability testing was performed with one sample over 24 h. The RSDs of peak area of the 9 constituents were all below 2.71%, which indicated that the samples remained stable during the testing period and the conditions for the analysis were satisfactory.

The repeatability was evaluated by the analysis of six prepared samples. The RSDs of for the contents of 9 constituents were all below 3.61%, which showed high repeatability.



FIGURE 4: GO analysis of potential target genes of the tannin fraction of Phyllanthus emblica L. GO: gene ontology.

The recovery was determined by the standard addition method. Certain amounts of the 9 constituents were spiked into the known sample and then processed and quantified in accordance with the established procedures as shown in Sections 2.2 and 2.3. The average recoveries were between 98.11% and 103.16%, with RSD values of less than 3.01% for the 9 compounds. Therefore, the developed method was precise and sensitive enough for simultaneously quantitative analysis of 0 compounds in the tannin fraction of *Phyllanthus emblica* L.

3.4. Quality Evaluation of the 9 Compounds. The developed quantitative analysis method was subsequently applied to 6 batches of the tannin fraction of *Phyllanthus emblica* L. sample habitat in Nepal. The results demonstrated a successful application of this HPLC-DAD assay for the quantification of 9 major constituents in different samples. The 9 compounds have been eluted within 62 min, giving good separation and acceptable tailings factors. Representative HPLC-DAD chromatograms of standard solutions and sample solutions for quantitative analysis are shown in Figure 1. The contents, summarized in Table 3, were calculated with the external standard methods.

In this experiment, UPLC-MSⁿ was employed to analyze the tannin fraction of *Phyllanthus emblica* L. The total ion chromatograms under both positive and negative modes were investigated at first, but the response intensity in the negative mode was significantly increased, and the number of detected chromatographic peaks increased significantly. Therefore, the negative mode was selected for the detection of Phyllanthus emblica L. We tentatively identified a total of 110 compounds including 45 hydrolysable tannins, 22 mucic acids, 15 phenolic acids, 15 flavonoids, 11 organic acids, and 2 other compounds. It can be seen from this result that most of the compounds in the tannin fraction are hydrolysable tannins, and the number of compounds accounted for more than 41% (45/110). The total tannins in the tannin fraction were determined before, and the content reached more than 60%. It is consistent with the results detected by UPLC-MSⁿ. There are also some mucic acids, phenolic acids, and flavonoids. Next, we will pay attention to these chemical components; the total content of flavonoids, mucic acids, phenolic acids, and organic acids accounts for about 40%; these ingredients may work synergistically with the hydrolysable tannins.

From the results of content determination by HPLC-DAD, the contents of gallic acid (content: 3.42%) and ellagic acid (content: 3.21%) are significantly higher than some hydrolysable tannins (punicalagin A: 0.26%, punicalagin B: 0.42%, chebulinic acid: 0.44%). Analyzing the reasons, we speculate that gallic acid and ellagic acid may be produced by the decomposition of other hydrolysable tannins. As we all know, gallic acid and ellagic acid are the basic structural units of hydrolysable tannins. Hydrolysable tannins are



FIGURE 5: "Component-target-function" network. Note: yellow triangle: compounds; pink ellipse: target; blue triangle: biological process.

TABLE 3: Contents of 9 compounds (n = 6).

Analytas	Contents (%)							
Allalytes	S1	S2	S3	S4	S5	S6	$Mean \pm SD$	
Gallic acid	3.42	3.46	3.39	3.44	3.43	3.38	3.42 ± 0.062	
Punicalagin A	0.26	0.23	0.23	0.26	0.28	0.29	0.26 ± 0.023	
Punicalagin B	0.43	0.44	0.42	0.43	0.4	0.41	0.42 ± 0.013	
Methyl gallate	0.45	0.43	0.45	0.46	0.44	0.45	0.45 ± 0.009	
Geraniin	1.15	1.19	1.18	1.15	1.2	1.18	1.15 ± 0.019	
Corilagin	2.70	2.72	2.74	2.7	2.71	2.72	2.70 ± 0.014	
Chebulinic acid	0.44	0.43	0.45	0.42	0.43	0.44	0.44 ± 0.010	
Chebulagic acid	1.14	1.16	1.11	1.18	1.15	1.15	1.14 ± 0.021	
Ellagic acid	3.21	3.24	3.22	3.26	3.26	3.24	3.21 ± 0.019	

unstable; they are easily decomposed under acids, alkali, enzyme, and high temperatures and used to produce gallic acid, ellagic acid, and polyols. In the process of preparing tannin fraction, the extraction temperature is 60° C and some hydrolysable tannins may decomposed, and these need to be further confirmed.

4. Conclusions

This research established a new method to find biomarkers for quality control of the tannin fraction of Phyllanthus emblica L. by using the UPLC-MSⁿ and network pharmacology methods. 110 compounds were obtained from UPLC-MSⁿ and the characteristic fragmentations were summarized. We found that hydrolysable tannins were the main components of the tannin fraction of Phyllanthus emblica L. Then, a network pharmacology method was used to explore the biomarkers for quality control of the tannin fraction of Phyllanthus emblica L., gallic acid, punicalagin A, punicalagin B, methyl gallate, geraniin, corilagin, chebulinic acid, chebulagic acid, and ellagic acid were filter as the biomarkers. Animal experiments proved these 9 compounds were proper biomarkers, because we generally believe that the ingredients those are absorbed into the blood are effective. Finally, a simple method for simultaneously measuring the contents of 9 biomarkers was established using HPLC-DAD. This method does not require high equipment, and it is suitable for promotion. The method developed in our study also provides a scientific foundation for the study of anticancer effective substances of the tannin fraction of *Phyllanthus emblica* L.

Data Availability

The data used to support the findings of this study are included within the Supplementary Materials.

Ethical Approval

The use of animals in the present study was permitted by the Ethics Committee of Beijing University of Chinese Medicine, and all animal studies were carried out according to the Guide for Care and Use of Laboratory Animals.

Conflicts of Interest

The authors declare no conflicts of interest.

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Supplementary Materials

 Table 1: 128 Blood Absorbed Components. (Supplementary Materials)

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