

Review Article

A Comprehensive Insight into the Phytochemical, Pharmacological Potential, and Traditional Medicinal Uses of *Albizia lebbeck* (L.) Benth.

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Background. Albizia lebbeck is a deciduous tree having tremendous medicinal utilities, for example, respiratory, skin, gastrointestinal, oral disorders, eye, urinary, genital, anorectal, inflammatory, and neurological disorders, and venereal diseases. Several studies have been undertaken on the medicinal and traditional values of *A. lebbeck. Objective.* The detailed information about its medicinal uses and pharmacological implications is highly scattered and distributed in different data sources. Hence, the study was conducted to supply an inclusive review of its ethnomedicinal uses, phytochemicals, and the available pharmacological attributes supporting its efficiency in traditional medicine. *Method.* Literature surveys were conducted on this medicinal plant *via* search engines like Google Scholar, PubMed, and Science Direct, and obtained information up to December 2020 has been assessed and analyzed for this study. *Results.* Systematic investigation revealed that *A. lebbeck* consists of various phytochemicals, including major alkaloids, flavonoids, saponins, and terpenoids. Its crude extract, fraction, and bioactive compounds exhibited potent adulticidal, antialergic, anticancer, anticonvulsant, antidiabetic, antidiarrheal, anti-inflammatory, antimicrobial, antinociceptive, antioxidant, antiparasitic, antipyretic, antivenom, estrogenic, neuroprotective, nootropic, ovicidal, and wound healing activities. *Conclusions.* This study proposes that *A. lebbeck* remains a rich source of phytochemicals with various biological activities which possess outstanding therapeutic benefits to humanity across the world. However, studies are required to estimate the potential side effects. Moreover, mechanistic physiognomies of the isolated compounds with known bioactivities are quite limited; thus, forthcoming research needs to focus on the mechanisms of these active phytochemicals to facilitate their potential enrolling for drug discovery.

1. Introduction

Medicinal plants and their derived natural products have long served as the primary healthcare requirements of millions of populations for centuries. Among these medicinal plants, many plants have been scientifically documented and validated for their exceptional medicinal efficacy. The genus *Albizia* comprises 150 taxonomically accepted species, which are widely distributed in Asia, Africa, and Australia, as well as tropical and subtropical America [1]. *Albizia lebbeck* mainly grows in the Indian subcontinent and Myanmar (Burma) and is also widely distributed in Western and Southeast Asia, Australia, Northern and West Africa, throughout the Caribbean, Central America, and the northern and eastern regions of South America (Figure 1) [2]. This species is reported to have incredible therapeutic properties, and it is utilized in several countries throughout the world to treat a variety of diseases and disabilities. The plant has been traditionally used against various diseases such as ulcers, night blindness, respiratory disorders, skin disorders, snake, bite, piles, and leprosy [3–5]. It is also used against gonorrhea, scorpion bite, gum problems, cough, pharyngitis, and so on [6–8]. In Sanskrit nomenclature, it is known as Sirisha, Bhandi, and Sirisa, while it is also entitled in many other languages throughout the world, for example, Acacia amarilla, cabellos de ángel, and lengua de mujer in Spanish; Bois noir and Viellefille in Franz; Darash in Urdu; Karuvagei and Vagei in Tamil; Khago and Ka se in Thai. In Burmese, it is spelled Kokko; Lebbek, siris tree, and woman's tongue tree in English; Mara in Sinhalese; Sarin and Shrin in Punjabi; Siris, SIrish, and Sirisha in Bengali; Siris and Sirisha in Hindi; Sultanaulasjar in Arabic; and Tekik in Javanese [2, 9].

It is a deciduous tree that is mostly found in the garden or along the roadside and grows from sea level to 1500 m elevation, attaining height up to 18 m. A. lebbeck contains numerous phytochemicals related to alkaloids, anthraquinones, essential oils, flavonoids, glycosides, phenolics, phytosterol, saponins, steroids, and triterpenoids [9-13]. According to various pharmacological studies, this species exhibited excellent antinociceptive, anti-inflammatory [11], anticancer [9], antimalarial [14], antiallergic [15], antihyperglycemic [16], antidiabetic [17, 18], wound healing [19], nootropic [20], and neuroprotective activities such as anti-Parkinson's and anti-Alzheimer activities [12, 21]. Furthermore, zinc oxide nanoparticles synthesized from Albizia lebbeck stem bark extract caused concentrationdependent organoprotective effect by changing mean body weight, alanine aminotransferase, serum alkaline phosphatase, urea, creatinine, bilirubin, protein, globulin, albumin, total cholesterol, triacylglycerol, and low- and high-density lipoprotein [22]. Other than its medicinal applicability, it is also used for reforestation of degraded sites, fuelwood plantations, and agroforestry systems in Asia [2].

This species contains a huge number of phytochemicals, out of which several phytochemicals have excellent medicinal properties and also showed tremendous pharmacological activities. There are a couple of compounds that have been exposed to pharmacological examinations and deficiently summed up with dispersed and scant data accessible on traditional uses. Additionally, there has been a lack of information that relates the pharmacological attributes of this plant to its ethnomedicinal applications. Likewise, patented formulations and safety profiles have been inadequately explored.

Even though many studies have been published on the biological activity of *A. lebbeck* extracts and their phytoconstituents [23–25], none of the reviews has been published with comprehensive information on pharmacological activities and elaborative insights of countrywise medicinal uses as well as different medicinal systemwise therapeutic potential. This prompted us to write this study, which covers botanical description, taxonomy, geographic distribution, medicinal usage, phytochemistry, and pharmacological qualities of *A. lebbeck*. The obtained information on phytochemicals, therapeutic uses, and pharmacological credits would optimistically assist the scientific community in planning safe tests that incorporate bioactive mixtures.

2. Materials and Methods

For this paper, an inclusive literature search was conducted up to January 2021. To identify appropriate statistics on the botanical description, traditional medicinal uses, phytochemistry, and pharmacological activities of A. lebbeck, information was retrieved from various resources, including Google Scholar, Science Direct, PubMed, and literature books. The keywords used for the database were "Albizia lebbeck," "Medicinal Uses," "Traditional Uses," "Botany," "Chemical Constituents," "Pharmacology," and "Biological Activities" with Boolean operators. Database that was unsuccessful in meeting the inclusion and quality criteria required in traditional uses, phytochemistry, and pharmacological attributes was excluded. The scientific name of the plant was authenticated by different databases like "the plant list" and "plants of the world online" (http:// www.plantsoftheworldonline.org/; http://www.theplantlist. org/).

3. Botanical Description

Albizia lebbeck grows as a deciduous tree with a length up to 18 m and a straight bole. Its bark is brownish-gray in color. The leaves of the plant are bipinnate, which are alternately arranged on the smooth, green twigs. The leaves turn a deep yellow color before falling during the dry season. The inflorescence is of corymb type with 30–40 flowers. Flowers are dimorphic, puberulent, and fragrant white to greenish-yellow in color. Calyx and Corolla are funnel-shaped; their pod is pale, flat, and straw-colored and remains on trees after a long-time of ripening. Seeds are brown, ellipsoidal (4–12) ca. $10 \times 6-7$ mm, and their pleurogram is parallel to margins of the seed [2]. The picture of *A. lebbeck* plant and its different parts is shown in Figure 2.

4. Traditional Medicinal Uses

A. lebbeck has been used in various countries of Africa, Asia, and Australia for the prevention of scabies, lung ailments, piles, bronchitis, abdominal tumors, cough, eye disorders, and so on. It is recommended in several medicinal systems, for example, Ayurveda, Sidha, and Unani medicine (Table 1) [11, 14, 31]. It has been used in numerous traditional uses; among them, it is mostly used in the treatment of respiratory disorders with 16%, skin disorders with 11%, and gastrointestinal disorders and oral disorders with 7% (Figure 3). In all these ethnomedicinal and traditional entities, the plant is ordinarily used to treat asthma, bronchitis, diarrhea, and gum inflammation with 4.88%, piles with 4.27%, parasitic infestation and snakebite with 3.66%, ulcer, scorpion sting, leprosy, and boils with 3.05%, and abdominal tumor, arthritis, cough, dysentery, night blindness, and poisoning with 2.44% in various countries. All plant parts, including root, leaves, flowers, bark, and seed, are useful in Indian traditional medicine in the treatment of several health ailments, for example, allergies, asthma, bronchitis, arthritis, fractures, gingivitis, gum inflammation, toothache, hemorrhage, leprosy, leukoderma, malaria, night blindness, scorpion sting, snakebite, and syphilis [10, 15, 26]. The bark is the most used plant part with 33.33% usage, followed by leaves, flower, seed (16.67%), root (9.52%), root bark, stem, and pods (2.38%) (Figure 4). A. lebbeck has many

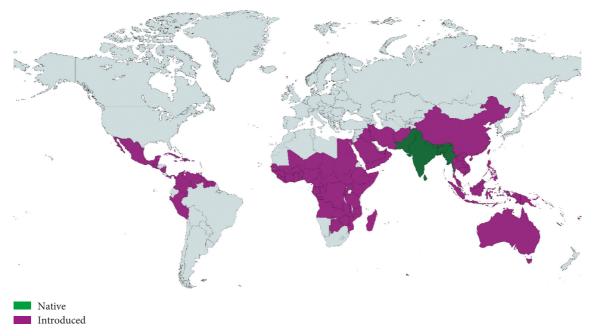


FIGURE 1: Global distribution of medicinal species Albizia lebbeck (L.) Benth. (created with mapchart.net).

therapeutic values such as astringent, pectoral, rejuvenation, and tonic [31].

According to the Ayurvedic Pharmacopoeia of India (2016), the stem bark possesses therapeutic uses such as Pama (eczema), Kustha (leprosy), Kandu (pruritus), Visarpa (erysipelas), Kasa (cough), Vrana (ulcer), Sotha (inflammation), Svasa (dyspnea), Musaka Visa, Sita Pitta (urticarial), Raktadusti (hypertension), Pinasa (catarrh), Vismajvara (irregular fever), Pratisyaya (common cold), Sarpdansa (snakebite), Visadusti, Suryavarta (migraine), Ardhavabhedaka (headache in half side of the head), KrmiRoga (worm infestation), and Netrabhiasanda (conjunctivitis). It retains various properties and actions; for example, Rasa is Madhura (sweet), Katu (pungent), Tikta (bitter), and Kasaya (astringent); Guna is Laghu (lightness); Virya and Vipaka are Anusna (lukewarm) and Katu (pungent), respectively; and Karma is Sothahara (alleviate swelling), Tridosahara (pacifies the three doshas), Visghna (neutralizing poison), Tvagdosa (skin disease), and Varnya (skin lightening). A. lebbeck has been widely used as an ingredient in several polyherbal formulations, for example, Vajraka Taila, Dasanga Lepa, Ayakrti, Devadarvarista, and Brhanmaricyadi Taila [32]. Bark and flowers are helpful in arthritis, and they are used in the Siddha system [18]. About 5-6 g of fresh leaves and 4-5 g of misree (refined sugar) in 1 glass of water, ground in a clay pot, can be taken 3 times a day to prevent tuberculosis. Fresh leaves are chewed, and then their extract from the mouth is poured into the eyes after filtration with a clean thin piece of cloth to soothe the reddishness of the eyes. 10-15 g of seeds is ground in a clay pot with water and consumed twice a day after filtration for the cure of boils by Sindh Indigenous people [33]. Moreover, the Bhils tribes used powder of crushed stem bark that can be applied on boils and pimples and paste of leaves and bark to cure insect bite and scorpion sting [8].

The stem bark paste is applied on ulcer and flower decoction and leaves for gargling to cure weak and spongy gums and chronic pharyngitis by the Meena tribe [8]. The Zulu tribes from Africa use bark and roots in the treatment of scabies, inflamed eyes, piles, and bronchitis [11]. In Tibetan traditional medicine, it is recommended in the treatment of kapha, pitta, poisoning, erysipelas, and ulcer [34]. In Taiwan, it is used as an anthelmintic, diuretic, stimulant, and tonic [35]. The people of Tamil Nadu use plants to fix bone fractures. The tribal communities in Himachal Pradesh and Kashmir use plants to relieve inflammation [28]. It is commonly called Shirish, Koroi, and Parrot tree in Bangladesh and has been used by the local people in the treatment of ophthalmia. Additionally, its barks and seeds are used as astringent and are given in piles, diarrhea, toothache, and gum problems. Further, bark and leaf decoctions are recommended against bronchial asthma and other allergic disorders [36]. Moreover, saponins of A. lebbeck have been reported to be used in Alzheimer's and Parkinson's disease treatment [37]. The ethnomedicinal uses, including data from various countries and medicinal practices of A. lebbeck, are given in Table 2.

5. Phytochemistry

Phytochemical studies of *A. lebbeck* have exposed the presence of various chemical constituents, including alkaloids, phenols, flavonoids, saponins, phytosterols, and terpenes [14]. Besides, seeds are good source of protein 2.272%, lipids 0.27%, fatty acid (linolenic acid, oleic acid, palmitic acid, and steric acid), tetradecane, hexadecane, phytol, nonadecane, eicosane, vitamin E, stigmastadiene, and octadecane [21, 45]. Complex triterpenoid saponin, that is, 21-[(2E,6S)-6-[6-deoxy-4-O-[(2E,6S)-6-hydroxy-2-(hydroxymethyl)-6-methyl-1-oxo-2,7-octadienyl]-[(β -D-



FIGURE 2: Leaves, flowers, and pods of Albizia lebbeck (source: Patanjali Herbal Museum).

glucopyranosyl) oxy]-2-(hydroxymethyl)-6-methyl-1-oxo-2,7-octadienyl]-[(β -D-glucopyranosyl)oxy]-2,6-dimethyl-1oxo-2,7-octadienyl]oxy]-16-hydroxy-3-[[O- β -D-xylopyranosyl-(1 \longrightarrow 2)-O- α -L-arabinopyranosyl-(1 \longrightarrow 6)-2-(acetylamino)-2-deoxy- β -D-glucopyranosyl]oxy]-(3 β , 16 α , 21 β)olean-12-en-28-oic acid O- α -L-arabinofuranosyl-(1 \longrightarrow 4)-O-[β -D-glucopyranosyl-(1 \longrightarrow 3)]-O-6-deoxy- α -L-mannopyranosyl-(1 \longrightarrow 2)- β -D-glucopyranosyl ester, is isolated from the bark [46]. Other than that, leaves contain essential oil in which 2-pentylfuran (16.4%), (E)-geranyl acetone (15.46%), (E)- α -ionone (15.45%), and 3-Octanone (11.61%) are abundantly found [11]. The present review suggests that the majority of phytochemicals contained in *A. lebbeck* should be explored and isolated from its bark and seeds, and additionally, other parts should be investigated too in the wake of the maximum utility of this plant to mankind.

The bark contains albiziasaponins (A–E) and lebbeckoside C, which possesses anticancer activity [9, 38]. Lebbeckosides A-B isolated from root showed an inhibitory effect on high-grade human brain tumor cells [31]. However, the seed contains lebbeckalysin (hemolysin), which possesses potent antitumor and antimicrobial effects [47]. Flavonoids (geraldone, luteolin, and isookanin) were

Parts used	Medicinal system	Mode of administration	Ethnomedicinal uses	References
Bark	Indian traditional medicine		Asthma, bronchitis, arthritis, gingivitis, toothache, allergies, leukoderma, leprosy, snakebites, malaria, and fractures	[15, 26]
Leaves	medicine		Night blindness and syphilis	[26]
All parts			Snakebite, scorpion sting, hemorrhage, and gum inflammation	[10]
Bark and flowers	Siddha system		Arthritis	[18]
Flowers	Traditional Chinese medicine		Anxiety, depression, and insomnia	[27]
			Nasya, pittaja, prameha, asthma, arthritis, burns, diarrhea, edema,	
	Ayurveda		poisoning, bronchitis, consumption, night blindness, respiratory disorders, skin disorders, snakebite, and scorpion sting	[3, 4, 27–29]
Root			Wounds	[30]
Bark			Bronchitis, leprosy, paralysis, gum inflammation, and helminthic infection	[3]
Leaves		Poultice	Night blindness and ulcer	[3]
Flower		Juice	Poisoning, hikka (hiccup), shwasa (asthma), and eye disease	[16]
Seed		-	Piles and diarrhea	[5]

TABLE 1: Ethnomedicinal uses of different parts of A. lebbeck in various traditional medicinal systems.

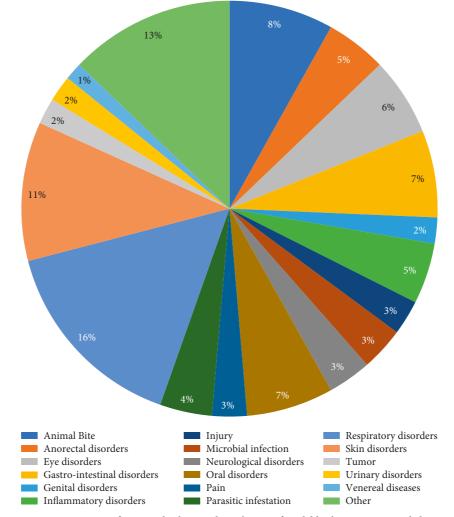


FIGURE 3: Percentage of reported ethnomedicinal uses of A. lebbeck against myriad diseases.

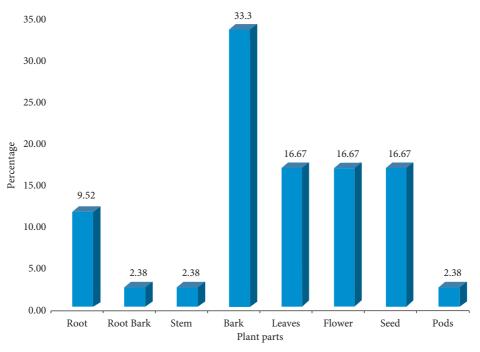


FIGURE 4: Parts usage (%) of A. lebbeck reported for various ethnomedicinal uses.

isolated from the bark having the capability of inhibiting the α -glucosidase and α -amylase activity [17]. Among reported chemical compounds, 45 bioactive molecules have been discussed in the pharmacological section. These studies suggested that most of the phytochemicals have been isolated from bark and seeds, and other parts are still needed to be explored. Plenty of molecular structures of various phytochemicals are procured from PubChem, and their detailed information is given in Table 3 and Figure 5.

6. Pharmacological Activities

Several pharmacological studies showed that extracts/fraction/compounds of leaves, bark, and flower of *Albizia lebbeck* (L.) Benth exhibited significant antiallergic activity, anticancer, anticonvulsant, antidiabetic, anti-inflammatory, antimicrobial, antinociceptive, antioxidant, antiparasitic, antivenom, neuroprotective, nootropic, antipyretic, antidiarrheal, ovicidal, adulticidal activity estrogenic, and wound healing activities. The foremost pharmacological attributes, extract/fraction/compound extracted from different parts of the plant, investigational doses, experimental models, and their results have been given in Figure 6, and pharmacological activities are also described as follows.

6.1. Antiallergic Activity. Ethanolic extract (200 mg/200–250 gm b. w., p.o.) of *A. lebbeck* stem bark exhibited excellent antiallergic activity in toluene-2,4-diisocyanate- (TDI-) sensitized allergy model Brown Norway rats and HeLa cells expressing endogenous H1R with a significant decrease in the numbers of sneezing, nasal rubbing, and mRNA expression which have been found to elevate TDI-induced H1R and HDC, although the least doses of extract (0.1 to

 $10 \,\mu$ g/ml) also reduced PMA- or histamine-induced upregulation of H1R mRNA in HeLa cells [48]. Besides, catechin present in the ethanolic extract from *A. lebbeck* bark showed potent activity by modulating histamine release and cytokine expression. *In vitro*, chloroform, methanol, and water extracts of leaf and bark showed a significant mast cell stabilizing effect with 19.71–59.69% against compound 48/80 [15, 51].

6.2. Anticancer Activity. Bark and leaves of A. lebbeck showed a potent anticancer effect from diverse cell lines. A saponin-rich fraction from the bark of A. lebbeck exerted antiproliferative activity via MTT assay in human breast cancer cell line MCF-7 by inhibiting the growth with IC_{50} 1 µg/ml and inducing apoptosis at 10 µg/ml by promoting activation of caspases 3 and 8. Furthermore, in shell-less chick embryo culture assay, there was a significant (p < 0.05) reduction in the number of extremities, nodes, junctions, and total branches length between 0 and 3 hr and 0-6 hr of drug exposure (0.1, 0.5, and $1 \mu g/ml$) and elevation of chromosomal aberration observed [40]. In another study, lebbeckosides A and B isolated from the root showed significant cytotoxic activity against U-87 MG, TG1 high-grade human brain tumors cells with IC₅₀ 3.46, 1.36, and 2.10, 2.24 µM, respectively [31]. The isolated compounds lebbeckosides A and B are responsible for initiating apoptosis in the cancerous cell by the activation of caspase 8 (Figure 7). Apart, crude methanol extract from leaves exerted a cytotoxic effect on hepatocarcinoma (HepG2) cancer cell line with IC₅₀ 24.03 µg/ml [52]. In another study, gold nanoparticles isolated from aqueous leaf extract of A. lebbeck showed cytotoxicity against HCT-116 colon cancer cells with IC₅₀ 48 mg/ml and also induced apoptosis by increased

S. no.	Country	Parts used	Mode of administration	Medicinal uses	References
	Africa	Leaves, stem bark, pods, and seeds	_	Dysentery, diarrhea, bronchial asthma, eczema, insect bite, allergy, piles, hernia, malaria, gonorrhea, scrofulous swellings, earache, antiprotozoal, and anthelmintic	[7]
1	Zulu of Southern Africa	Bark and roots	_	Scabies, inflamed eyes, piles, and bronchitis	[11]
	West Africa		_	Diarrhea, dysentery, hemorrhoids, bronchitis, asthma, eczema, and leprosy	[31, 38]
2	Asia	Stem Bark	_	Abdominal tumors, boils, cough, eye disorders, and lung ailments	[31]
3	Australia	Seed, = stem bark, and root bark	_	Diarrhea, gastroenteritis, hemorrhoids, bronchitis, leprosy, paralysis, parasitic infestation, ulcer, snakebite, gum ailments, abdominal tumors, boils, cough, eye disorders, and lung ailments	[31, 39]
4	Bangladesh	Bark, seed, and leaves	Decoction	Piles, diarrhea, toothache, gum ailments, bronchial asthma, allergic disorder, and ophthalmia	[36]
5	China India	Flowers Bark and seed	Powder and juice	Anxiety, depression, and insomnia Astringent, tonic, restorative, and anus pain Arthritis, bone fracture, edema, poisoning, asthma, bronchitis, skin disease, cold and	[27] [40, 41]
6		Bark, flowers, seeds, and roots		asuma, bronchins, skin disease, cold and cough, itching, pruritus, wounds healing, leprosy, malaria, gonorrhea, abscesses, boils and abdominal tumors, snakebite, scorpion sting, hemorrhage, and gum inflammation Spermatorrhea	[9, 10] [42]
	India (Bhils and Meena tribes)	Stem bark, flowers, and leaves	Powder, paste, and decoction	Stone, boil, pimples, ulcer, gums ailments, pharyngitis, insect bite, and scorpion sting	[8]
	India (tribes of Himachal Pradesh and Kashmir)			Inflammation	[28]
7	India (Tamil Nadu) Myanmar (Burma)			Bone fractures Abdominal tumors Snakebite, scorpion sting, hemicrania,	[9]
8	Nepal	Root, leaves, flowers, bark, and seed	Bark aqueous extract (leaf), decoction (seed), ointment, and powder	strengthen gum, ophthalmia, cough, bronchitis, asthma, prevent conception in women, anus pain, night blindness, astringent, piles, diarrhea, dysentery, gums ailment (spongy and ulcerated gums), emollient for boils, eruption, carbuncle, swelling, eye disease, and scrofulous enlargement of glands	[6, 40, 43]
9 10	Nigeria Philippines	Bark and leaves	Aqueous extract Decoction	Fever, pain, epilepsy, and inflammation Dysentery, diarrhea, and ulcer	[11] [44]
10	Taiwan	Bark		Anthelmintic, diuretic, stimulant, tonic, and	[35]
12	Tibet			vermifuge Kapha, pitta, poisoning, erysipelas, and ulcer	[34]

TABLE 2: Medicinal uses of A. lebbeck in different countries of the world.

ROS production, decreased $\Delta \Psi m$, apoptotic morphological changes by AO/EtBr, and altering pro- and antiapoptotic protein expressions [53].

6.3. Anticonvulsant Activity. The methanolic fraction of chloroform soluble part of the ethanolic extract of *A. lebbeck* (20, 40, or 100 mg/kg i.p.) exhibited remarkable anticonvulsant activity against pentylenetetrazole-induced convulsions and maximum electroshock in mice by delaying the

onset of spasms and clonic convulsions. Fraction also delayed the latency to stage 4 significantly in lithium-pilocarpine-induced seizures. Moreover, in electrical kindling, fractions decreased the behavioral score. However, the fraction showed no protective effect against strychnine-induced convulsions [54]. Furthermore, 200 and 400 mg/kg (p.o.) ethanolic extract of *A. lebbeck* leaves demonstrated a considerable anticonvulsant effect by reducing the duration of hind limb extensor in the MES model and delaying the onset of convulsions in the PTZ mode [55].

TABLE 3: Chen	nicals constituen	ts of A. lebbeck.
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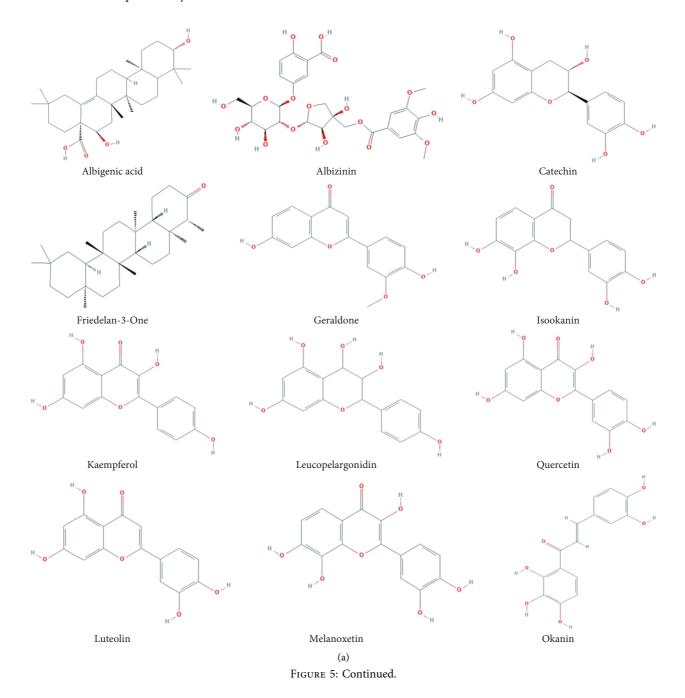
Chemical compounds	Plant part	References
Alkaloids (budmunchiamines L1–L6), α -amyrine, catechins, echinocystic acid or acacic acid, flavonoids (kaempferol, quercetin, and quercetin 3-O-alpha-rhamnopyranosyl (1 \rightarrow 6)-beta-		
glucopyranosyl $(1 \rightarrow 6)$ -beta-galactopyranosides), friedelan-3-one, (-)-leucopelargonidin, lupeol, melanoxetin, okanin, oleanolic acid, (+) pinitol, polyphenols, saponins (lebbekanin A-H) g-	Plant	[15, 28, 48]
sitosterol, and triterpenoids Oleanane-type saponins (lebbeckosides A and B)	Roots	[31]
Alkaloids, flavonoid (geraldone, luteolin, isookanin, epicatechin, and procyanidins B-2, B-5, and C-3), glycoside (albizinin), hemolysin (lebbeckalysin), oleanane triterpene (albiziasaponins A–E), phenols, phytosterols, saponins, and triterpenoid saponin (lebbeckoside C, 21-[[(2e,6S)-6-[6-deoxy-4-O-[(2e,6S)-6-hydroxy-2-(hydroxymethyl)-6-methyl-1-oxo-2,7-octadienyl]-[(β -D-		
glucopyranosyl) oxy]-2-(hydroxymethyl)-6-methyl-1-oxo-2,7-octadienyl]-[(β -D-gluco- pyranosyl) oxy]-2,6-dimethyl-1-oxo-2,7-octadienyl]oxy]-16-hydroxy-3-[[O- β -D-xylopyranosyl-(1 \longrightarrow 2)-O- α -L-arabinopyranosyl-(1 \longrightarrow 6)-2-(acet-ylamino)-2-deoxy- β -D-glucopyranosyl]oxy]-(3 β ,16 α ,21 β)-olean-12-en-28-oic acid O- α -L-arabinofuranosyl-(1 \longrightarrow 4)-O-[β -D-glucopyranosyl-(1 \longrightarrow 3)]-O-6-deoxy- α -L-mannopyranosyl-(1 \longrightarrow 2)- β -D-glucopyranosyl ester)	Bark	[4, 12, 14, 17, 38, 46, 47]
Alkaloids, glycosides, saponin (albiziahexoside) steroids, tannins, terpenoids, flavonoids (kaempferol 3-O-a rhamnopyranosyl(1/6)-b-glucopyranosyl(1/6)-o-galactopyranoside, quercetin 3- O-a rhamnopyranosyl(1/6)-b-glucopyranosyl(1/6)-b-galactopyranoside, kaempferol, and 3- rhamnosyl (1–6) glycosyl (1–6) galactoside)	Leaves	[4,49,50]
Alkaloids, anthraquinones, eicosane, fatty acid (linolenic acid, oleic acid, palmitic acid, and steric acid), flavonoids, glycosides, nonadecane, octadecane, phenolics, phytol, saponins (glycosaponins), steroids, stigmastadiene, tetradecane, and vitamin E	Seed	[10,21,45]
3',5-Dihydroxy-4',7 dimethoxy flavone and N-benzoyl-L-phenyl alaninol Albigenic acid	Pod Bean	[19]

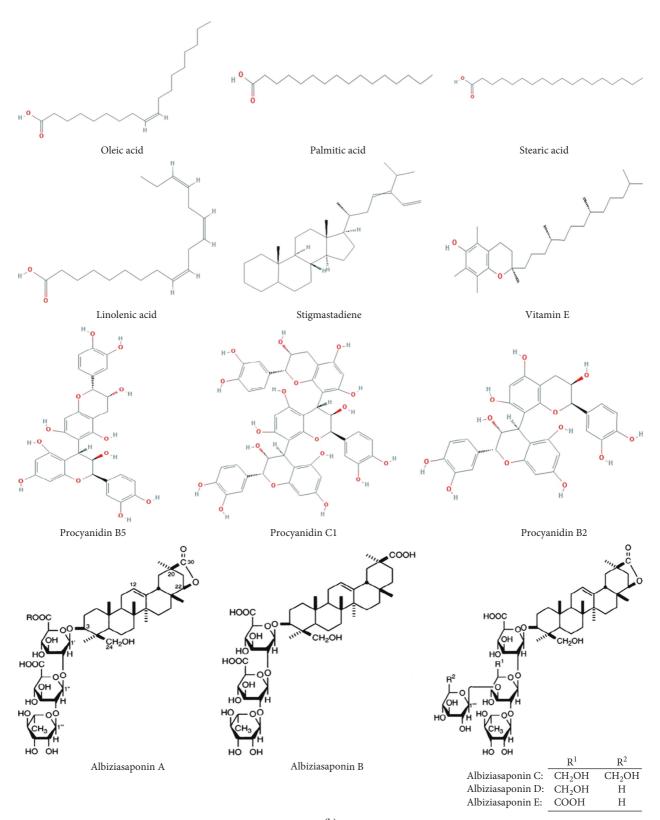
6.4. Antidiabetic Activity. The bark of A. lebbeck demonstrated noteworthy antidiabetic activity. The methanol extract (200, 350, and 620 mg/kg) exhibited antihyperglycemic activity against streptozotocin-nicotinamide stimulated type II diabetes mellitus rats by significantly decreasing the level of serum glucose, creatinine, urea, cholesterol, triglycerides, LDL-cholesterol, and VLDL-cholesterol and increasing HDL levels as compared to diabetic control [16]. A study was conducted to evaluate in vitro antidiabetic activity of geraldone, isookanin, and luteolin isolated from methanolic extract of A. lebbeck bark, which showed potent inhibition against α -glucosidase and α -amylase (73.14 to 93.98%). The mechanistic approach of geraldone, isookanin, and luteolin has been graphically represented in Figure 7. In another study, it was demonstrated that methanol/dichloromethane extract of A. lebbeck bark possesses antidiabetic activity in streptozotocin-induced diabetic rats via significant reduction of blood glucose, BUN, SCr, GSP, TC, TG, LDL-c, and VLDL-c and increases plasma insulin level, hepatic enzymes, SOD, CAT, GSH, and HDL-c levels [17,18].

6.5. Anti-Inflammatory Activity. Administration of leaf essential oil (100, 200, and 400 mg/kg) caused significant inhibition of carrageenan-induced edema [11]. Leaves aqueous and ethanolic extract showed anti-inflammatory effect at 200 mg/kg with percentage inhibition of 39.36% and 42.55% in carrageenan-induced paw edema and also reduced granuloma formation with 38.55% and 42.33%, respectively [49]. In another study, petroleum ether and ethanol extracts (400 mg/kg) exhibited maximum inhibition of carrageenaninduced inflammation with percentage inhibition of 48.6% and 59.57%; dextran-induced group 45.99% and 52.93%; cotton pellet-induced models 34.46% and 53.57%, and Freund's adjuvant-induced animal group 64.97% and 68.57%, respectively [28], while bark petroleum ether: ethyl acetate: methanol extract (1:1:1) significantly (p < 0.001) reduces carrageenan-induced rat hind paw edema at 400 mg/kg with 36.68% [56]. Moreover, n-hexane, dichloromethane, ethyl acetate, and n-butanol fraction from flowers reduce inflammation in carrageenan-induced paw edema. Among tested fractions, the most potent activity was shown at 1 g/kg by dichloromethane (71.6%) followed by ethyl acetate (60.3%) [37].

6.6. Antimicrobial Activity. The zinc nanoparticle from the stem bark of A. lebbeck demonstrated activity against B. cereus, S. aureus, E. coli, K. pneumoniae, and S. typhi, with inhibition zones ranging from 1 to 10.57 mm, with S. typhi showing maximum inhibition at 0.1 M, which was comparable to ciprofloxacin (12.53 mm) [57]. In another study, ethanolic extract of root exerted antibacterial activity against E. coli, S. flexneri, P. aeruginosa, S. typhi, K. pneumonia, S. boydii, S. aureus, and E. faecalis with 9.05-15.77 mm inhibition range, where S. typhi showed maximum inhibition followed by S. flexneri (15.50 mm) at 200 mg/ml with MIC 0.20 and 0.39 mg/ml, respectively [30]. Similarly, petroleum ether, ethyl acetate, and methanol extracts from the stem bark and leaves exhibited antimicrobial activity against selective microbes among Gram-positive bacteria, that is, B. polymyxa, B. subtilis, B. megaterium, S. lutea, and S. aureus; Gram-negative bacteria such as V. mimicus, V. cholera, S. typhi, S. boydii, S. flexneri type-1, S. dysenteriae, P. aeruginosa, K. pneumoniae, E. coli, and P. vulgaris; fungal

Evidence-Based Complementary and Alternative Medicine





(b) Figure 5: Continued.

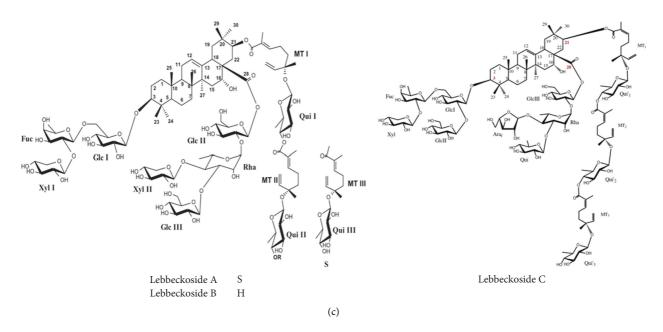


FIGURE 5: Molecular structure of various phytochemicals extracted from different parts of Albizia lebbeck.

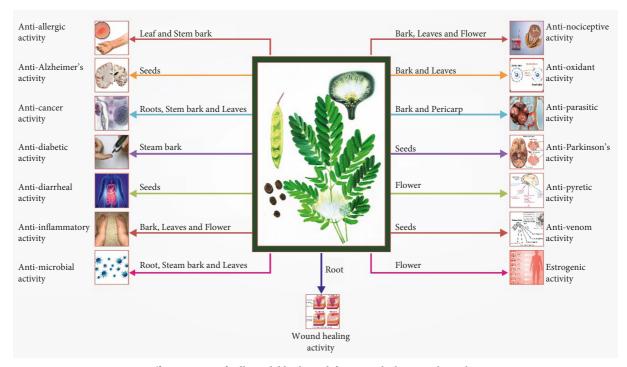


FIGURE 6: Different parts of Albizia lebbeck used for several pharmacological investigations.

strains as *C. arrizae*, *A. fumigatus*, *A. Niger*, *R. oryzae*, *C. albicans*, *C. krusei*, and *Saccharomyces cerevisiae*. Stem bark extract was shown to have action with a zone of inhibition of 6–14 mm, with ethyl acetate extract having the best activity against *B. subtilis*, *S. typhi* (14 mm), and *C. arrizae* (10 mm). However, leaves extract had an antimicrobial activity with a zone of inhibition of 3–23 mm, whereas methanolic extract demonstrated the highest effective action against *S. typhi* at 500 mg [19, 58]. Moreover, leaves crude ethanolic extract at

10 mg/ml exerted activity against *S. aureus* (6 mm) and *E. coli* (7.5 mm), with IC_{50} 7.97, 5.62 mg/ml [52].

6.7. Antinociceptive Activity. Essential oil isolated from leaves significantly inhibited nociceptive mediators at both neurogenic and inflammatory phases in the formalin hind paw with an average of 44% and 100% at 200 and 400 mg/kg, respectively [11]. Leaves aqueous and ethanolic extract was

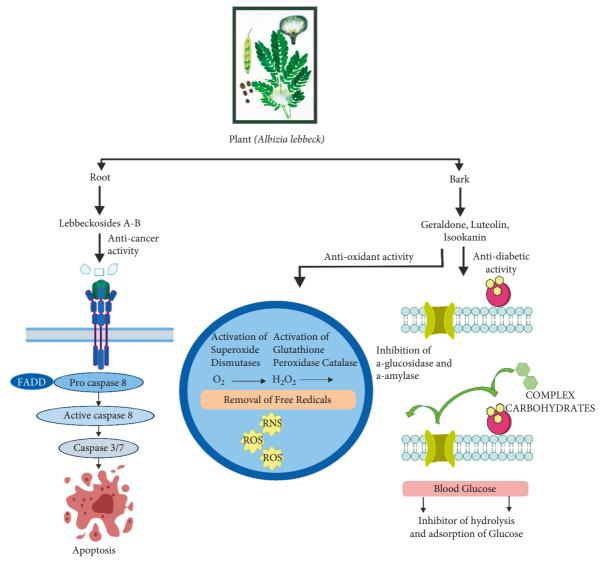


FIGURE 7: Mechanistic representation of different phytochemicals extracted from A. lebbeck.

administered orally to evaluate analgesic activity by eddy's hot plate and tail-flick test. In the hot plate method, a significant elevation was observed in the mean basal reaction time, and an elevation in the latency time was found in the tail-flick method [26]. In another study, among n-hexane, dichloromethane, ethyl acetate, and n-butanol fraction from flower, only dichloromethane fraction (1 g/kg) significantly increases in pain threshold in the hot plate test [37]. Bark petroleum ether: ethyl acetate: methanol extract (1:1:1) showed a significant reduction in the number of writhes by 52.4% and significant elongation of tail flicking time with 61.48% at 400 mg/kg [56].

6.8. Antioxidant Activity. Increased production of reactive oxygen species is a cause of most human diseases, including cardiovascular disease and cancer. Cells enable upregulation of antioxidant defenses and other protective systems against mild oxidative stress, although severe stress can harm the

integrity of DNA, proteins, and lipids and lead to cell death by apoptotic or necrotic mechanisms [59]. Therefore, the antioxidant effect of A. lebbeck is evaluated. Geraldone, isookanin, and luteolin isolated from the bark of the plant are tested for DPPH-free radical scavenging assay, where geraldone showed the best activity (IC₅₀ 21.5 μ M) [17]. These isolated compounds are able to neutralize the free radicals, including RNS and ROS, by activating antioxidant enzymes (Figure 7). Zinc oxide nanoparticles from the stem bark exhibited the most potent antioxidant effect against hydrogen peroxide-free radical with IC_{50} 48.5 µg/ml [57]. Petroleum ether, ethyl acetate, and methanol barks extracts of A. lebbeck were evaluated for DPPH-free radical scavenging activity, where ethyl acetate (81.13%) and methanol extract (78.23%) showed high radical scavenging activity, followed by petroleum ether (74.82%) at $100 \,\mu\text{g/ml}$ [58]. Additionally, leaves crude methanol extract showed DPPH and ABTS radical scavenging activity with IC₅₀ 34.22 and 108.7 µg/ml, respectively [52].

TABLE 4: Pharmacological activities of various parts of A. lebbeck.

Ref.	[15]	[51]	[21]		[6]			[57]	[31]	[52]	[16]	[17]	[18]	[63]
Result	Dose-dependent mast cell stabilization activity at 200 and 300 mg/kg dose extract protected the degranulation (53 and 61%, resp.). There was significant protection from degranulation (compound 48/80 induced) of mast cells, dose- dependent, that is, 61 and 74% of inhibition of	histamine release at 200 and 300 mg/kg, respectively All the extracts showed significant mast cell stabilization activity. However, methanolic and water extracts of the bark showed the maximum activity along with the leaf methanolic extract	Extracts significantly improved the memory and cognitive impairments, ↑GSH, SOD, CAT, and ↓ AChE	Fraction inhibits the growth of MCF-7 with IC_{50} $$1\mu g/ml$$	Fraction increases apoptosis and promotes activation of caspases 3 and 8 Reduction in number of extremities nodes	junctions, and total branches length between 0 and	5 hr and 0 and6 hr of drug exposure 1 Total chromosomal aberrations	Extract significantly inhibited the viability	Lebbeckoside A and B showed cytotoxicity against TG1 and U-87 MG, with IC ₅₀ 2.10, 2.24, 3.46, and $1.36\mu\text{M}$, respectively	Extract significantly decreased the cell viability with ${\rm IC_{50}}~24.03\mu {\rm g/ml}$	Extract significantly decreased the level of serum GLU, creatinine, urea, triglycerides, cholesterol, low- density lipoprotein-cholesterol, and very low-density lipoprotein-cholesterol and increased high-density linomrotein levels	All three compounds significantly inhibit the a-glucosidase and a-amylase enzymes	Significant reduction of blood glucose, BUN, SGr, GSP, TC, TG, LDL-c, and VLDL-c and increasing plasma insulin level, hepatic enzymes, SOD, CAT, GSH, and HDL-c,	Extract significantly inhibited the cathartic effect of castor oil in a dose-dependent manner
f Study model/parameter	Mast cell stabilization, compound 48/80-induced systemic anaphylaxis	<i>In vitro</i> mesenteric mast cell stabilization against compound 48/80	In vivo aluminum chloride (100 mg/kg, p.o.)- induced Alzheimer's disease in Wistar albino rats Morris water maze, open field, hole board, Y-maze, and T-maze test	In vitro MTT assay in human breast cancer MCF-7	Apoptosis assay	Shell-less chick embryo culture assay	Chromosomal aberration (CA) assay	In vitro MTT assay in human breast cancer MCF-7 and MDA-MB 231	<i>In vitro</i> cytotoxicity against the glioblastoma stem- like TG1 cells and human glioblastoma U- 87 MG cell lines	In vitro MTT assay against human hepatocarcinoma (HepG2) cancer cell line	Streptozotocin-nicotinamide-induced type II diabetes mellitus using female Sprague-Dawley rats	In vitro a-glucosidase and a-amylase inhibitory assay	<i>In vivo</i> streptozotocin-induced diabetic rats using male albino Wistar rats	In vivo castor oil-induced diarrhea using albino rats and mice
Standard	DSCG (50 mg/kg, i.p.)	1% DMSO	Galantamine 0.5 mg/kg	Doxorubicin 500 nM	Staurosporine 1 ug/ml				Tamoxifen		Metformin 45 mg/kg	Acarbose 10 mg/ml	Glibenclamide 1 mg/kg	Loperamide 1 mg/kg i.p.
Dose/mode of administration	50 to 300 mg/kg, p.o.	50 µg/ml	100–300 mg/kg p.o.	0.001, 0.01, 0.1, 1, and 10 μg/ml	$10\mu{ m g/ml}$	0.1, 0.5, and $1 \mu \mathrm{g/ml}$		5, 25, 50, and 100 μg/ ml of 0.1 M, 0.05 M, and 0.01 M ZnO NPs		1, 10, 25, 50, 75, 100, 125, and 150 μg/ml	200, 350, and 620 mg/ kg/day, p.o.		100-400 mg/kg	2.5–5 mg/kg i.p.
Parts used	Stem bark	Leaf and stem bark	Seed		Bark			Stem bark	Root	Leaves	Bark	Bark	Stem bark	Seed
Extract, fraction, and isolate	Ethanolic extract	Chloroform, methanol, and water extracts	Hydromethanolic extract		Saponin-rich fraction			Zinc oxide nanoparticles	Lebbeckosides A-B	Crude methanol extract	Methanolic extract	Geraldone, isookanin, and luteolin	Methanol/ dichloromethane extract	Aqueous methanol extract
Pharmacological activity	Antiallergic activity	(Anti-Alzheimer's activity				:	Anticancer activity				Antidiabetic activity		Antidiarrheal activity
S. no.	1		2					ŝ				4		5

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100. 00, ond 400 mg/ kg p.n. Indomethani 10 mg/kg In visce antititis units granted witstar rass induced antititis units granted witstar rass Des-dependent and significant inhibition of induced antititis units granted witstar rass Des-dependent and significant inhibition of induced antititis units granted witstar rass 200 md 400 mg/s Pensyhuazane 100 mg/s In viso arragerana-induced at huld pave defan Des-dependent and significant inhibition of units and a domong/s All ractions aboved approximation in vitro age diffusion method using <i>Evants</i> . Des-dependent and significant inhibition of units and <i>Evants</i> . 200 md 005M, and 0.01M, and 0.01M, and 0.01M, and 0.0200 mg/ml Cprofloxacin 10.gg/dis. In vitro age diffusion method using <i>Evants</i> . Des-dependent and significant inhibition of units and <i>Evants</i> . 300 ug/dis Cprofloxacin 10.gg/dis. Misrata diatoral diffusion method using <i>Evants</i> . Des-dependent and significant inhibition of units and <i>Evants</i> . 300 ug/dis Cprofloxacin 10.gg/dis Misrata diatoral diffusion method using <i>Evants</i> . Des-dependent and significant inhibition of units and distro- section and distro- section and dimension. Des-dependent and dignificant inhibition of units and dimension. 300 ug/dis Cprofloxacin 10.gg/dis Misrata dimension. Des-dependent inhibition of units and dimension. 300 ug/dis Significant inhibition of units anterus Des-dependent inhibition of units	olic		L	Leaves	50-200 mg/kg, p.o.	Diclofenac 20 mg/kg and indomethacin 10 mg/kg	<i>In vivo</i> carrageenan-induced paw edema and cotton pellet-induced granuloma models using Wistar rats	Dose-dependent and significant inhibition of inflammation	[49]
0.2.5 and 1 g/kg, ip. Didofenae solum 20 mg/kg In vire carrageome induced pave deam aung Wistau ratas All fractions showed significant inhibition of unimmation 200 and 400 mg/kg Provis/burazone 100 mg/kg Daviso carrageome induced pave deam aung ponound Daviso carrageome induced pave deam Dose-dependent and agnificant inhibition of unimmation 200 and 400 mg/kg Provis/burazone 100 mg/kg Daviso carrageome induced pave deam aung gang and interview Daviso carrageome induced pave deam aung gang and interview Daviso carrageome induced pave deam aung gang and interview Daviso carrageome induced pave deam aung gang gan and supply provide sinterview Daviso carrageome induced pave deam aung gang gan and supply gang and gan	Petroleum ether, Anti- chloroform, and ethanol F inflammatoryHevene		щ	Bark	100, 200, and 400 mg/ kg p.o.	Indomethacin 10 mg/kg	In vivo carrageenan - and dextran-induced rat paw edema; cotton pellet-induced granuloma; adjuvant- induced arthritis using female Wistar rats	Dose-dependent and significant inhibition of inflammation	[28]
200 and 400 mg/kg Panylburazone 100 mg/kg In vitro carrageman-induced rat hind pow edem Dose-dependent and significant inhibition zone using long-Frant rate of a control of millammation of the control o	, ethyl ıtanol		Flc	Flower	0.25 and 1 g/kg, i.p.	Diclofenac sodium 20 mg/kg	<i>In vivo</i> carrageenan-induced paw edema using Wistar rats	All fractions showed significant inhibition	[37]
001.M. 0.05M, and 0.1M Ciprofloxacin 10 rgdis Invito age disc diffusion method sup g Backins, intro alse cliftision method using Backins, Singelia Presenvisa: and Simonella Nepti Neptode Statistican Presenvisa: Singelia Poyliti, and Entrencess Jaccasis Neptode Neptode Statistican Presenvisa: Singelia Poyliti, and Entrencess Jaccasis Provide Statistican Presenvisa: Singelia Poyliti, Silverer Type-1, against sleater Statistican Presenvisa: Approprise Schericitati osli, Silverer Type-1, against sleater Statistican Presenvisa: Approprise Schericitati osli, Schericitata osli, Schericitata osli Presenvisa: Approprise Schericitati osli, Schericitata osli Presenvisa: Approprise Schericitati osli, Schericitata osli Presenvisa: Approprise Schericitati osli, Schericitata osli Presenvisa: Antipo Schericitata osli Provest Barters, Presenvisae Schericitata P	r: ethyl I extract		Bé	Bark	200 and 400 mg/kg p.o.	Phenylbutazone 100 mg/kg	<i>In vivo</i> carrageenan-induced rat hind paw edema using long-Evans rats	Dose-dependent and significant inhibition of inflammation	[56]
100-200 mg/ml Ciprofloxacin Staggla / plearent, seata four clinical bacterial statistics <i>sharmonus aerugiona</i> , <i>statistics sharmonus statistis</i> , harmonuch statisti <i>sharbitistis</i> , harmonuch statistis, harmonuch statistis <i>sharbitistis</i> , harmonuch statistis, harmonuch statistis <i>sharbitistis</i> , harmonuch statistis, <i>sharbitistis</i> , harmonuch statistis, <i>sharbitis</i> , harbitis, harbitistis <i>sharbitis</i> , harbit	Zinc oxide nanoparticles Sten		Sten	Stem bark	0.01 M, 0.05 M, and 0.1 M	Ciprofloxacin 10µg/disc	In vitro agar disc diffusion method using Bacillus cereus, Staphylococcus aureus, Escherichia coli, Klebsiella pneumoniae, and Salmonella typhi In vitro disc diffusion method using Escherichia coli,	Extract showed strong activity with inhibition zone ranging from 1 to 10.57 mm	[57]
Sol polymyca, B. subfils, B. megaterium, Sarrian late, 300 pgdits Defension B. minusary. Cholera, Error type-1, for fung Defension B. minusary. Cholera, Submendia bydits, S. fleareri type-1, abereting servitiva 25 pgdits Defension B. minusary. Cholera, Submendia bydits, S. fleareri type-1, abereting fungtuats, S. fleareri type-1, abereting fungtuats, S. fleareri type-1, aperation strentom, streptomycin, strentomycin, for fung Defension B. minusary. Cholera, S. deserteria, Estericina of the Approach in physican attraction and strentomycin. Defension B. minusary. S. ceresistar (8 mm) 300 µgdul Tetracycline, streptomycin, ritampcin, northoasem, streptomycin Tetracycline, streptomycin, triampcin, northoa agains S. ceresistar (8 mm) 300 µgdul Tetracycline, streptomycin, streptomycin, triampcin, northoasem, streptomycin, triampcin, northoa agains S. ceresistar (8 mm) 300 µgdul Tetracycline, streptomycin, streptomycin, triampcin, northoa agains S. ceresistar (8 mm) 300 µgdul Tetracycline, streptomycin, streptomycin, triampcin, northoa agains S. ceresistar (8 mm) 300 µgdul Tetracycline, streptomycin, streptomycin, triampcin, northoa agains S. ceresistar (8 mm) 300 µgdul Tetracycline, streptomycin, streptomycin, tetracycline 20 mg/ms Monog extract showed activity agains 300 µgdul Province 1, mycles are the and thy actate extract showed a significant and dose- tion with a zone of inhithinon rangitors a toh merogene.	Ethanolic extract Ro		Ro	Root	100–200 mg/ml	Ciprofloxacin	Shigella flexneri, Pseudomonas aeruginosa, Staphylococcus aureus, and four clinical bacterial isolates Salmonella typhi, Klebsiella pneumoniae, Shigella boydii, and Enterococcus faecalis In vitro disc cliftusion method using Bacillus	Extract showed activity against all tested bacteria with a zone of inhibition ranging from 9.05 to 15.77 mm and MIC 0.20–1.56 mg/ml	[30]
50, 100, 200, and 500 µg/mlTetracycline, streptomycin, iriampicin, northoxacin, and gentamycin <i>In vitro</i> agar dis diffusion method using <i>Bacillus</i> abrilis. <i>Escherichia coli, Klebsiella pneumonia</i> , sentanycinAnnogertaets, methanolic extract showed strong activity with a zone of inhibition ranging from 11 to 23 mm at 500 µg/ml50, 100, 200, and iriampicin, northoxacin, and gentamycin <i>In vitro</i> agar well diffusion method using method admined <i>typhi</i> , and <i>Staphylococcus aureus</i> , <i>Seudomonus aeruginosa</i> , 23 mm at 500 µg/mlAnno extract showed strong activity with a zone of inhibition ranging from 11 to 23 mm at 500 µg/ml10m-400 mg/ml tetracycline 20 mg/ml <i>In vitro</i> agar well diffusion method against <i>Salmonella ablicans</i> , and <i>Escherichia coli</i> Extract showed potent antibacterial activity against <i>S. aureus and E. coli</i> with ZOI 6 and 7.5 mm, <i>La viro formalin hind paw</i> in Wistar rats <i>Both extracts showed a significant and dos- response</i> 50–200 mg/kg, po.Pentazocine 15 mg/kg <i>In viro formalin hind paw</i> in Wistar rats <i>Both extracts showed a significant and dos- exponse</i> 0.25 and 1 g/kg, ip.Aspirin 200 mg/kg <i>In vivo hot plate method using male albino white in the ot plate test and latency of the fift, kull rats200 and 400 mg/kgAminopyrine 50 mg/kg<i>In vivo hot plate method using Swiss albino mice</i>Extract showed significant and dos- response200 and 400 mg/kgAminopyrine 50 mg/kgAretic acid induced writhing test using Swiss albino increases in pain threhold increases in pain threhold mice200 and 400 mg/kgAminopyrine 20 mg/kgAminopyrine 20 mg/kgExtract showed significant and dos- response</i>	Antimicrobial Petroleum ether, ethyl activity acetate, and methanol Stem bark extracts		Stem	bark	300 µg/disc	Ciprofloxacin 10 µg/disc for bacteria: griseofulvin 25 µg/disc for fungi	polymyxa, B. subtilis, B. megaterium, Sarcina lutea, Staphylococcus aureus, Vibrio mimicus, V. Cholera, Salmonella typhi, Shigella boydii, S. flexneri type-1, S. dysenteriae, Pseudomonas aeruginosa, Klebsiella pneumoniae, Escherichia coli, Candida arrizae, Aspergillus fumigatus, A. niger, Rhizopus oryzae, Candida albicans, C. Krusei, and Saccharomyces	Pet. ether and ethyl acetate extract showed activity against selective microbes with ZOI ranging from 6 to 14 mm. Methanol extract is only active against <i>S. cerevisiae</i> (8 mm)	[58]
Ampöcillin 10 mg/ml, 10 mg/mlIn vitro agar well diffusion method against streptomycin 10 mg/ml, and streptomycin 10 mg/ml, and Snaphylococcus aureus, Pseudomonas aeruginosa, tetracycline 20 mg/mlExtract showed potent antibacterial activity against streptomycin streptomycin streptomycin 10 mg/mlIn vitro agar well diffusion method against streptomonas aeruginosa, streptomycin 20 mg/mlExtract showed potent antibacterial activity against streptomycin streptomycin100-400 mg/kg p.o.Piroxicam 10 mg/kg p.o.In vivo formalin hind paw in Wistar rats In vivo Eddy's hot plate and tail-flick test in Wistar ratsExtract inhibited nociceptive mediators at both neurogenic and inflammatory phases Both extracts showed a significant and dose- frast50-200 mg/kg, p.o.Pentazocine 15 mg/kgIn vivo Eddy's hot plate and tail-flick test in Wistar ratsExtract inhibited nociceptive mediators at both neurogenic and inflammatory phases Both extracts showed a significant and dose- dependent increase in the mean basal reaction time in the hot plate test and latency of the flick tail response0.25 and 1 g/kg, i.p.Aspirin 200 mg/kgIn vivo hot plate method using male albino white in the hot plate test and latency of mice200 and 400 mg/kgAminopyrine 50 mg/kgAcetic acid induced writhing test using Swiss albino witeExtract showed a significant and dose- dependent increases in pain threshold mice200 and 400 mg/kgAminopyrine 50 mg/kgAcetic acid induced writhing test using Swiss albino bExtract showed a significant and dose- dependent increases in pain	Petroleum ether, ethyl acetate, and methanol Leaves extract		Leav	res	50, 100, 200, and 500 μg/ml	Tetracycline, streptomycin, erythromycin, lincomycin, rifampicin, norfloxacin, and gentamycin	In vitro agar disc diffusion method using Bacillus subtilis, Escherichia coli, Klebsiella pneumonia, Proteus vulgaris, Pseudomonas aeruginosa, Salmonella typhi, and Staphylococcus aureus	Among extracts, methanolic extract showed strong activity with a zone of inhibition ranging from 11 to $23 \mathrm{mm}$ at $500 \mathrm{\mu g}/\mathrm{ml}$	[19]
100-400 mg/kg p.o.Piroxicam 10 mg/kg p.o.In vivo formalin hind paw in Wistar ratsExtract inhibited nociceptive mediators at both neurogenic and inflammatory phases50-200 mg/kg, p.o.Pentazocine 15 mg/kgIn vivo Eddy's hot plate and tail-flick test in WistarExtract inhibited nociceptive mediators at both neurogenic and inflammatory phases50-200 mg/kg, p.o.Pentazocine 15 mg/kgIn vivo Eddy's hot plate and tail-flick test in WistarExtract inhibited nociceptive mediators at both neurogenic and inflammatory phases50-200 mg/kg, p.o.Pentazocine 15 mg/kgIn vivo Eddy's hot plate and tail-flick test in WistarRependent increase in the mean basal reaction time in the hot plate test and latency of the flick tail response0.25 and 1 g/kg, i.p.Aspirin 200 mg/kgIn vivo hot plate method using male albino whiteOnly dichloromethane fraction (1 g/kg) significantly increases in pain threshold200 and 400 mg/kgAminopyrine 50 mg/kgAcetic acid induced writhing test using Swiss albinoExtract showed a significant and dose-dependent reduction in the number of writhes micep.o.Morphine 2 mg/kgRadiant heat tail-flick method using Swiss albinoExtract showed a significant elongation of tail flicking tract showed significant elongation of tail flicking	Crude methanol extract Leaves		Leav	/es	10 mg/ml	Ampocillin 10 mg/ml, streptomycin 10 mg/ml, and tetracycline 20 mg/ml	In vitro agar well diffusion method against Staphylococcus aureus, Pseudomonas aeruginosa, Candida albicans, and Escherichia coli	Extract showed potent antibacterial activity against S. <i>aureus</i> and E. <i>coli</i> with ZOI 6 and 7.5 mm, respectively	[52]
50-200 mg/kg, p.o.Pentazocine 15 mg/kgIn vivo Eddy's hot plate and tail-flick test in WistarBoth extracts showed a significant and dose- dependent increase in the meen basal reaction time in the hot plate test and latency of the flick tail response0.25 and 1 g/kg, i.p.Aspirin 200 mg/kgIn vivo hot plate method using male albino whiteOnly dichloromethane fraction (1 g/kg) significantly mice200 and 400 mg/kgAminopyrine 50 mg/kgAcetic acid induced writhing test using Swiss albinoExtract showed a significant and dose-dependent ncreases in pain threshold200 and 400 mg/kgMorphine 2 mg/kgRadiant heat tail-flick method using Swiss albinoExtract showed a significant and dose-dependent reduction in the number of writhes mice200 and 200 mg/kgMorphine 2 mg/kgRadiant heat tail-flick method using Swiss albinoExtract showed a significant end ose-dependent reduction in the number of writhes mice	Essential oil Le		Lea	Leaves	100-400 mg/kg p.o.	Piroxicam 10 mg/kg p.o.	In vivo formalin hind paw in Wistar rats	Extract inhibited nociceptive mediators at both neurogenic and inflammatory phases	[11]
0.25 and 1 g/kg, i.p. Aspirin 200 mg/kg In vivo hot plate method using male albino white Only dichloromethane fraction (1 g/kg) significantly mice 0.25 and 1 g/kg, i.p. Aspirin 200 mg/kg In vivo hot plate method using male albino white Only dichloromethane fraction (1 g/kg) significantly increases in pain threshold 200 and 400 mg/kg Aminopyrine 50 mg/kg Acetic acid induced writhing test using Swiss albino Extract showed a significant and dose-dependent mice 200 and 400 mg/kg Morphine 2 mg/kg Radiant heat tail-flick method using Swiss albino Extract showed significant elongation of tail flicking mice	Aqueous and ethanolic Lea extract		Lea	Leaves	50–200 mg/kg, p.o.	Pentazocine 15 mg/kg	<i>In vivo</i> Eddy's hot plate and tail-flick test in Wistar rats	Both extracts showed a significant and dose- dependent increase in the mean basal reaction time in the hot plate test and latency of the flick tail response	[26]
200 and 400 mg/kg Aminopyrine 50 mg/kg Acetic acid induced writhing test using Swiss albino Extract showed a significant and dose-dependent 200 and 400 mg/kg Aminopyrine 50 mg/kg Acetic acid induced writhing test using Swiss albino Extract showed a significant elongation of tail flicking p.o. Morphine 2 mg/kg Radiant heat tail-flick method using Swiss albino Extract showed significant elongation of tail flicking	Antinociceptive n-Hexane, activity dichloromethane, ethyl Fl acetate, and n-butanol fraction		H	Flower	0.25 and 1 g/kg, i.p.	Aspirin 200 mg/kg	<i>In vivo</i> hot plate method using male albino white mice	Only dichloromethane fraction (1 g/kg) significantly increases in pain threshold	[37]
	Petroleum ether: ethyl acetate: methanol extract (1:1:1)			Bark	200 and 400 mg/kg p.o.	Aminopyrine 50 mg/kg Morphine 2 mg/kg	Acetic acid induced writhing test using Swiss albino mice Radiant heat tail-flick method using Swiss albino mice	Extract showed a significant and dose-dependent reduction in the number of writhes Extract showed significant elongation of tail flicking time	[56]

TABLE 4: Continued.

14

Evidence-Based Complementary and Alternative Medicine

Ref.	[57]	[17]	20	[00]	[52]	[14]		[60]	[12]	[37]	[10]	[37]	[30]
Result	IC ₅₀ 48.7, 60.2, and 48.5 μ g/ml, respectively	All compounds showed activity with IC_{50} 21.5, 31.8, and 29.26 μ M, respectively	Extracts showed DPPH- and H ₂ O ₂ -free radical	$60.21, 70.93, 64.69, and 68.99 \mu g/ml, respectively$	Extract exhibited UPPTH and AB15 radical scavenging activity with IC_{50} 34.22 and 108.7 $\mu g/m$ l, respectively	$\rm IC_{50}$ = 8.2 and 5.1 $\mu g/m$ against MRC2 and RKL9 strains	Dose-dependent chemosuppression was observed with significant schizonticidal activity at 1000 mg/kg with ED> 100 mg/kg. Significant curative and repository activities were exhibited by 750 mg/kg concentration of extract on day 7	Extract showed antiparasitic activity with IC ₅₀ 8.7, 8.1, 37.9, and 50.8 μg/ml against T. cruzi, T. brucei, P. falciparum, and L. infantum, respectively	Extract improved the motor functions and showed significant improvement in catalepsy, time latency, no. of exploration, † SOD, CAT, and GSH	All fractions showed a decrease in temperature	Extract inhibited protease and hyaluronidase (IC ₅₀ 36.32 and 91.95 μ g), hemorrhage (ED ₅₀ 26.37 μ g), serum creatinine kinase, and lactate dehydrogenase (ED ₅₀ 37.5 and 31.44 μ g)	Ethyl acetate (200) and total alcohol fraction (500 mg/kg) significantly decrease and increase uterine weight by 25.2 and 109%, respectively	7 Wound breaking strength in incision model, complete wound contraction was observed on the 22nd day in excision model, 7 wet weight of granulation tissue, total protein, SOD, GSH, hydroxyproline, hexuanine, hexuronic acid levels, J lipid peroxidation, and nitric oxide
Study model/parameter	H ₂ O ₂ -free radical scavenging assay	DPPH radical scavenging assay	DDDH and H O free redivel conversion of	DEFIT- and 11202-tice faultal stavenging assay	DPPH and ABTS radical scavenging assays	In vitro antimalarial activity against Plasmodium falciparum chloroquine (CQ) sensitive (MRC2) and CQ resistant (RKL9) strains	<i>In vivo</i> schizonticidal activity, repository, and curative activities using <i>P. berghei</i> -infected white Swiss albino mice	In vitro antiplasmodial, antileishmanial, and antitrypanosomal activities against Plasmodium falciparum, Leishmania infantum, Trypanosoma cruzi, and T. brucei	In vivo haloperidol-induced catalepsy Assessment of catalepsy, hang test, and narrow beam walk test Open field test	<i>In vivo</i> Brewer's yeast-induced pyrexia using albino mice	<i>Echis carinatus</i> venom- (ECV-) induced local toxicity in Swiss albino mice <i>in vivo</i> and proteolytic and hyaluronidase activities <i>in vitro</i>	Uterine weight using female Albino mice	<i>In vivo</i> incision and excision wound models in nulliparous and nonpregnant healthy female rats
Standard	Ascorbic acid	Trolox	According to the second	ASCOLDIC ACIA	Ascorbic acid	Chloroquine 5 mg/kg	Chloroquine 5 mg/kg and pyrimethamine 1.25 mg/kg	Chloroquine, miltefosine, benznidazole, and suramin	Sinemet-levodopa 100 mg+ carbidopa 25 mg/kg per oral	Aspirin (200 mg/kg)		17-β-Estradiol (0.32μg/animal/ day)	Vitamin E 200 mg/kg
Dose/mode of administration	0.01, 0.05, and 0.1 M		20 100 m	1111/B# 001-07	1, 10, 25, 50, 75, 100, 125, and 150 μg/ml	$5-100\mu g/ml$	100, 250, 500, 750, and 1000 mg/kg/day	20 mg/ml	100–300 mg/kg	0.25 and 1 g/kg, i.p.	1: 1-1: 100 w/w	200 and 500 mg/kg i.p.	250, 500, and 750 mg/ kg p.o.
Parts used	Stem bark	Bark	Stom hould	JICIII DAIN	Leaves	Bark	Bark	Pericarp	Seed	Flower	Seed	Flower	Root
Extract, fraction, and isolate	Zinc oxide nanoparticles	Geraldone, isookanin, and luteolin	Petroleum ether, ethyl	extracts	Crude methanol extract	Ethanolic extract	Ethanolic extract	Methanol extract	Aqueous methanolic extract	n-Hexane, dichloromethane, ethyl acetate, and n-butanol fraction	Methanolic extract	n-Hexane, dichloromethane, ethyl acetate, and n-butanol fraction	Ethanolic extract
Pharmacological activity			Antioxidant	activity			Antiparasitic activity		Anti-Parkinson's activity	Antipyretic activity	Antivenom activity	Estrogenic activity	W ound healing activity
S. no.			a	n			10		11	12	13	14	15

Evidence-Based Complementary and Alternative Medicine

TABLE 4: Continued.

6.9. Antiparasitic Activity. Ethanolic extract from the bark of A. lebbeck showed antimalarial activity against P. falciparum chloroquine-resistant (RKL9) and CQ sensitive (MRC2) strains with IC₅₀ 5.1 and 8.2 μ g/ml, respectively. Furthermore, the extract showed significant (p < 0.001) schizonticidal activity, repository, and curative activities against P. berghei. Moreover, plant extracts at different doses, 100, 250, 500, 750, and 1000 mg/kg/day, exhibited chemosuppression of 69.4, 71.4, 71.9, 79.8, and 84.7%, respectively, on the seventh day of postexposure [14]. In another study, pericarp ethanolic extract exhibited antiparasitic activity against P. falciparum, L. infantum, T. cruzi, and T. brucei with IC₅₀ 37.9, 50.8, 8.7, and 8.1 μ g/ml, respectively [60].

6.10. Antivenom Activity. Albizia lebbeck is used traditionally as medicine in the treatment of snakebite, and several researchers have experimentally evaluated the medicinal use of *A. lebbeck* against snakebite [9, 10, 31]. One of the studies revealed that seed methanolic extract exhibited significant (p < 0.0001) antivenom activity with inhibition of ECV protease and hyaluronidase with IC₅₀ 36.32 µg, 91.95 µg at 1 : 100 w/w, respectively. Moreover, extract neutralizes (p < 0.0001) ECV-induced hemorrhage with ED₅₀ 26.37 µg, myotoxicity by reducing serum creatinine kinase with ED₅₀ 37.5 µg (p < 0.0001), and lactate dehydrogenase 31.44 µg (p = 0.0021) levels at 1 : 50 w/w [10].

6.11. Neuroprotective Activity. The symptoms of Alzheimer's disease include deterioration of memory, judgment, and decision-making power which reduces impairment in the orientation of physical surroundings and language [61]. It was observed that seed hydromethanolic extract (100-300 mg/kg orally) reduced biochemical oxidative stress and improved functional outcomes of behavioral studies by improving memory and cognition functions via inhibiting anticholinesterase, thereby preserving acetylcholine concentration [21]. The second most common neurodegenerative disease is Parkinson's disease which causes parkinsonism that occurs due to the loss of neurons in the substantia nigra and elsewhere in association with the presence of ubiquitinated protein deposits in the cytoplasm of neurons and thread-like proteinaceous inclusions within neurites [61]. The anti-Parkinson activity was evaluated by performing behavioral and biochemical oxidative stress assay in Wistar albino rats. It was observed that the plant extract can be able to ameliorate motor function and prevent biochemical damage in brain cells [12].

6.12. Nootropic Activity. The n-butanol fraction (10 and 25 mg/kg) from dried leaves of *A. lebbeck* exhibited excellent nootropic activity in mice by using the elevated plus maze and passive shock avoidance paradigm. On both doses, the inflexion ratio (IR) was increased significantly, while IR was found to decrease at the utmost dose (50 mg/kg) after 24 h after exposure as well as on day 9 in the passive avoidance test. Moreover, the fraction (10, 25, and 50 mg/kg) dose-dependently reduced the lithium-induced head twitches and

at 50 mg/kg significantly potentiated and prolonged the haloperidol-induced catalepsy [20].

6.13. Miscellaneous Activity. Ovicidal and adulticidal activities were studied against Culex quinquefasciatus, Aedes aegypti, and Anopheles stephensi from hexane, benzene, chloroform, ethyl acetate, and methanol extracts; among tested extracts, methanolic extract obtained from the leaf and seed showed absolute mortality at 200, 250, 150, and 300, 375, and 225 ppm against Ae. aegypti, C. quinquefasciatus, and An. stephensi, respectively. Methanol leaf extract showed the highest adulticidal activity against An. stephensi with LC₅₀ 65.12 ppm [62]. n-Hexane, dichloromethane, ethyl acetate, and n-butanol fraction from flower were evaluated for antipyretic activity. The most potent effect was shown by dichloromethane followed by ethyl acetate at 1 g/kg with a reduction of 8°C and 5°C, respectively [37]. Aqueous methanol extract from seed (5 mg/kg *i.p.*) almost entirely inhibits the castor oil-induced diarrhea [63]. The pharmacological profile of various parts of A. lebbeck is shown in Table 4.

7. Conclusion

Albizia lebbeck is an Ayurvedic plant and has been widely utilized in the treatment of anorectal, eye, gastrointestinal, genital, inflammatory, neurological disorders, oral disorders, respiratory, skin, urinary disorders, and venereal diseases across the world. Different parts of the plant have been used, but bark appears to be the most often used plant part in the employment of traditional medicine. However, in support of its therapeutic uses, more scientific clinical trials extensively are necessary. The phytochemical studies revealed an abundance of saponins with other chemicals, for example, flavonoids, phenols, and glycosides. A. lebbeck has been studied for many pharmacological activities against allergy, cancer, convulsant, diabetes, inflammation, parasitic infestation, snake venom, nootropic, pyrexia, diarrhea, and so on, and there remains still a scarcity of information on the mechanism of action. Additionally, it is worth noting that even though A. lebbeck has been used in the treatment of various ailments, it is an ingredient in several Ayurvedic formulations; nonetheless, studies are required to evaluate the possible toxicities or adverse effects. In forthcoming research, studies should target the discovery of the chemical compounds responsible for the therapeutic action, which comprise the mechanisms of action.

Conflicts of Interest

The authors declare no conflicts of interest with regard to the submitted work.

Authors' Contributions

Acharya Balkrishna performed diconceptualizatiod n, funding acquisition, and provided resources. Ms. Sakshi performed data curation, visualization, formal analysis, and writing the original draft. Mr. Mayur Chauhan performed data curation and formal analysis. Dr. Anurag Dabas performed conceptualization, supervision, investigation, validation, and review and editing of the paper. Dr. Vedpriya Arya did project administration, supervision, and review and editing of the paper.

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