

Review Article

A Brief Review of Natural Products with Urate Transporter 1 Inhibition for the Treatment of Hyperuricemia

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Hyperuricemia is a common disease caused by a high level of uric acid. Urate transporter 1 (URAT1) is an important protein and mediates approximately 90% of uric acid reabsorption. Therefore, the URAT1 inhibitor is a class of uricosuric medicines widely used in the clinic for the treatment of hyperuricemia. To find the new medicine with stronger URAT1 inhibition and lower toxicity, researchers have been exploring natural products. This study systematically summarizes the natural products with URAT1 inhibition. The results show that many natural products are potential URAT1 inhibitors, such as flavonoids, terpenoids, alkaloids, coumarins, stilbenes, and steroids, among which flavonoids are the most promising source of URAT1 inhibitors. It is worth noting that most studies have focused on finding natural products with inhibition of URAT1 and have not explored their activities and mechanisms toward URAT1. By reviewing the few existing studies of the structure-activity relationship and analyzing common features of natural products with URAT1 inhibition, we speculate that the rigid ring structure and negative charge may be the keys for natural products to produce URAT1 inhibition. In conclusion, natural products are potential URAT1 inhibitors, and exploring the mechanism of action and structure-activity relationship will be an important research direction in the future.

1. Introduction

Uric acid is the end metabolite derived from the oxidation of purine compounds [1]. Hyperuricemia is a chronic metabolic disease caused by a high level of uric acid. Excessive intake of purine-containing foods and insufficient uric acid excretion are the keys to causing hyperuricemia [2]. In recent years, the incidence of hyperuricemia has continued to increase worldwide, which may be related to changes in lifestyle, such as the prevalence of a high-purine diet, fructose beverages, and alcohol consumption [3, 4]. In China, the overall prevalence of hyperuricemia is 13.3%, and the prevalence in men is higher than in women [5]. In the United States, the prevalence of hyperuricemia is 21.2% in men and 21.6% in women [6]. Hyperuricemia is related to the occurrence of many diseases, such as cardiovascular

disease, metabolic syndrome, and acute kidney injury [7]. Therefore, patients have an urgent need for efficient and safe therapeutic methods or drugs [8].

Reducing purine intake, inhibiting uric acid production, and promoting uric acid excretion are effective ways to treat or improve hyperuricemia [9]. URAT1 inhibitors are a widely used class of uricosuric drugs by inhibiting the reabsorption of uric acid, such as probenecid, sulfapyrazone, and benzbromarone [10]. Although these drugs have good uric acid lowering effects, they all have varying degrees of side effects [11]. Currently, sulfapyrazone has been withdrawn from most countries due to its severe gastrointestinal toxicity [12]. Benzbromarone has severe hepatotoxicity and is currently approved for use in only a few countries [13]. Even the newly approved lesinurad has renal toxicity and cardiovascular toxicity [14]. Therefore, scholars

have been exploring new URAT1 inhibitors with low toxicity [15].

Natural products refer to components or metabolites from animals, plants, insects, and microorganisms, such as proteins, peptides, polysaccharides, and alkaloids [16–18]. Natural products have been used as medicines for thousands of years. Moreover, the importance of natural products is increasing day by day and has become an important source of drug development [19]. At present, long-term clinical practice has demonstrated that traditional Chinese medicine (one of the important sources of natural products) has exact efficacy in lowering serum uric acid without serious adverse effects [20]. With the deepening of research, scholars have found that natural products are expected to be the source of new URAT1 inhibitors. This study systematically summarizes natural products with URAT1 inhibition. The results showed that flavonoids, terpenoids, alkaloids, coumarins, stilbenes, steroids, organic acids, and polysaccharides show inhibitory effects of URAT1, which can inhibit URAT1 activity and promote uric acid excretion. However, most studies have focused on finding natural products with inhibition of URAT1 and have not explored their activities and mechanisms towards URAT1. By reviewing the few existing studies on the structure-activity relationship studies and analyzing common features of natural products with URAT1 inhibition, we speculate that the rigid ring structure and negative charge may be the keys for natural products to produce URAT1 inhibition. In conclusion, natural products are valuable sources of URAT1 inhibitor, and exploring the mechanism of action and structure-activity relationship will be an important research direction in the future.

2. Pathological Processes of Hyperuricemia and the Role of URAT1

Uric acid, also known as 2,6,8-trihydroxypurine, is a heterocyclic carbonyl compound with a relative molecular weight of 168 [21]. Uric acid is mainly produced by the metabolism of endogenous and dietary purine compounds under the action of xanthine oxidase in the liver (Figure 1) [22]. Hyperuricemia refers to an excessively high concentration of uric acid in the blood. That is, uric acid concentration <7.0 mg/dl in men or <6.0 mg/dl in women [23]. As a metabolic disease, hyperuricemia is closely related to the occurrence and development of many diseases, such as gout, hypertension, heart disease, and diabetes [24]. The appearance of gout is most closely related to hyperuricemia. This is because an excessively high concentration of uric acid is easily deposited in the articular cavity in body tissue, causing pain, edema, and inflammation in the joints, finally inducing gout [25].

The metabolic disorder of uric acid includes excessive uric acid production and decreased uric acid excretion [26]. Causes of excess uric acid production include the intake of purine-rich foods, such as seafood and meat, and the increased concentrations or activities of intermediate metabolic enzymes of uric acid, such as xanthine oxidase [27]. Since more than 70% of uric acid in the human body is produced by metabolism, inhibiting the activities of

metabolic enzymes can effectively inhibit the production of uric acid [28]. Therefore, xanthine oxidase inhibitors such as allopurinol, febuxostat, and topiroxostat are the drugs of choice for the clinical treatment of hyperuricemia and gout [29]. In addition, the main reason for the decrease in uric acid excretion is closely related to the insufficient renal excretion capacity. This is because the kidney is the main excretory organ of uric acid, and more than 2/3 of uric acid is excreted from the kidney [30]. Therefore, promoting the excretion of uric acid by the kidney by regulating the activities of uric acid transporters is an effective method to treat hyperuricemia and gout. Current studies have found that uric acid transport-related proteins mainly include uric acid reabsorption-related proteins and uric acid secretion-related proteins (Figure 2). Proteins related to uric acid reabsorption include URAT1, glucose transporter 9 (GLUT9), organic anion transporter 4 (OAT4), and organic anion transporter 10 [31]. Proteins related to uric acid secretion include organic anion transporter 1, organic anion transporter 2, organic anion transporter 3, sodium-dependent phosphate transport protein 1 (NPT1), sodium-dependent phosphate transport protein 4, ATP-binding cassette superfamily G2 (ABCG2), multidrug resistance protein 4 (MRP4), and urate transporter (UAT) [32]. Among these proteins, URAT1 is a highly valuable potential therapeutic target.

URAT1 is encoded by the SLC22A12 gene, which is located on chromosome 11q13, contains 10 exons and 9 introns, encodes 555 amino acids, and has 12 transmembrane domains [33]. URAT1, originally called the renal-specific transporter, is a member of the organic anion transporter family and the first protein to be involved in renal uric acid transport [34]. Figure 2 shows that URAT1 is located in the renal tubule epithelial cell apical membrane and mediates the exchange of uric acid in the lumen with inorganic and organic anions in the proximal tubular epithelial cells, thus reabsorbing uric acid from the lumen into epithelial cells [35]. Although URAT1 is not the only protein that mediates uric acid re-absorption, the importance of URAT1 is reflected in its strong transport capacity: approximately 90% of uric acid re-absorption is mediated by URAT1 [36]. Therefore, considering the important role of URAT1 in uric acid re-absorption, URAT1 inhibitors are considered highly effective and promising drugs for the treatment of hyperuricemia. As early as 2002, related studies explored the possibility and value of URAT1 as a target for reducing uric acid and first proposed the development of URAT1 inhibitors [37]. So far, researchers have developed a variety of URAT1 inhibitors, such as probenecid, benzbromarone, lesinurad, and dotinurad [12]. These drugs can effectively inhibit the reabsorption of uric acid by URAT1 and promote the excretion of uric acid, thus exerting a uric acid-lowering effect.

3. Natural Products with URAT1 Inhibitory Effects

Due to the great potential of URAT1 inhibitors in the treatment of hyperuricemia and gout, researchers have been exploring new URAT1 inhibitors [38]. As an important

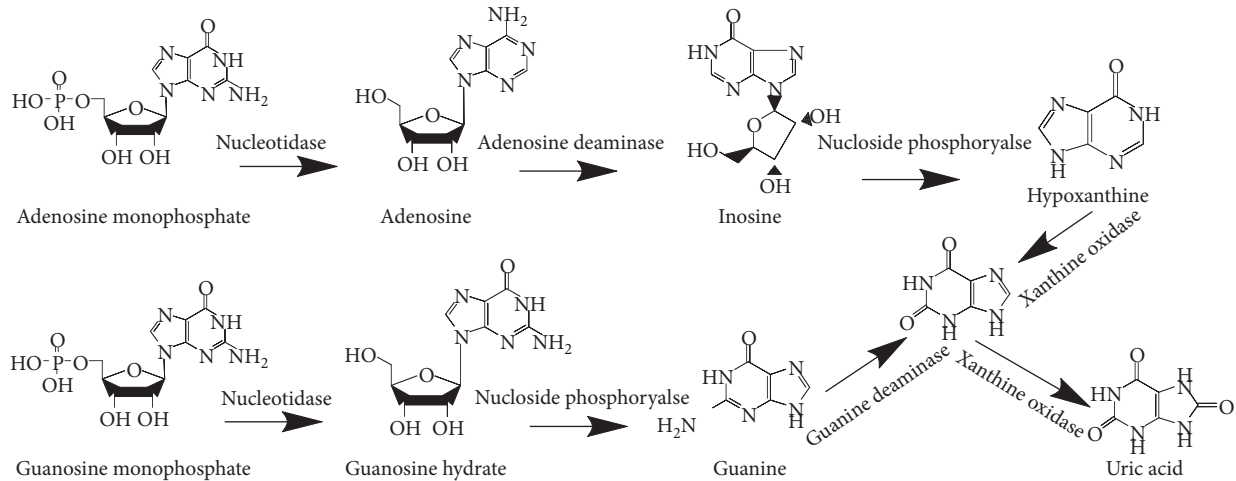


FIGURE 1: The production pathway of uric acid.

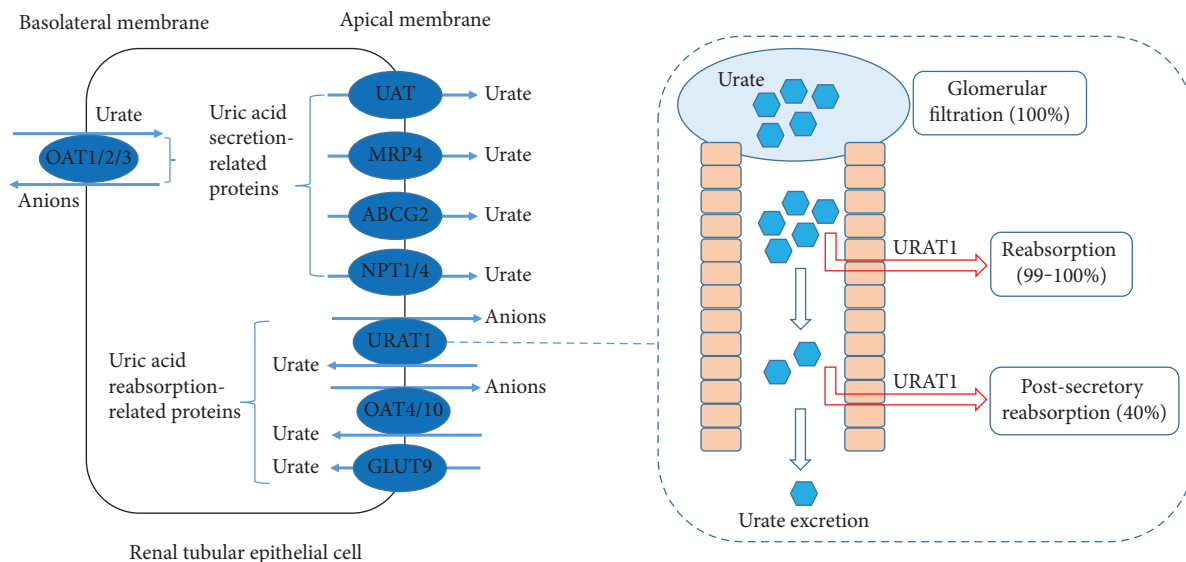


FIGURE 2: The distribution of uric acid transporters and the effects of URAT1 on uric acid metabolism.

source of new drugs, natural products have received more and more attention for their inhibitory effects on URAT1 [39]. Table 1 summarizes the species, main sources, and inhibitory effects of URAT1 of these natural products. It can be seen that many of the natural products with URAT1 inhibition are flavonoids [45, 52, 53]. In addition, some terpenoids, alkaloids, coumarins, stilbenes, and steroids also show a URAT1 inhibitory effect [60, 66, 68, 74, 77]. However, most studies have focused on finding natural products with inhibition of URAT1 and have not explored their mechanisms toward URAT1 [63, 64]. Therefore, exploring the mechanism of action will be an important research direction in the future.

3.1. Flavonoids. Flavonoids are a class of secondary plant metabolites widely present in a variety of plants and are the active components of many Chinese herbal medicines. Chemical structure generally refers to the connection of two

benzene rings (ring A and ring B) through three carbon atoms to form the structure $C_6-C_3-C_6$ [78]. Flavonoids contain many subclasses based on the connection position of the B and C rings as well as the degree of saturation, oxidation, and hydroxylation of the C ring [79]. Currently, studies have shown that many natural products with URAT1 inhibitory effects belong to flavonoids, and the subclasses include flavones, flavonols, flavanols, flavonones, flavanols, isoflavones, and xanthenes. Figure 3 further summarizes the structural formulas of these flavonoids.

3.1.1. Flavones. It can be seen in Figure 3 that flavones are characterized by containing a double bond between positions 2 and 3 and a ketone in position 4 of the C ring [80]. Currently, flavones with the inhibitory effect of URAT1 include chrysin, apigenin, baicalein, nobiletin, and luteolin. The structures of these flavones are very similar, except for nobiletin (the substituents are all methoxy). They have hydroxyl groups at

TABLE 1: Natural products with an URAT1 inhibitory effect.

Category	Name	Common or primary source	Cell lines/model	Dosage	Ref.
Flavones	Nobiletin	Citrus fruits	URAT1-expressing 293A cells	IC ₅₀ = 17.6 μM/l	[40]
	Baicalein	<i>Scutellaria baicalensis</i>	URAT1 and potassium oxonate-induced hyperuricemia mice	IC ₅₀ = 31.56 μM/l and 200 mg/kg, respectively	[41]
	Apigenin	The leafy herbs parsley and dried chamomile flowers	URAT1, URAT1-expressing HK-2 cells and hyperuricemia nephropathy mice	IC ₅₀ = 0.64 μM/l, 3.125–100 μM/l and 100 mg/kg	[42, 43]
	Chrysin	Propolis, blue passion flower, and honey	High fructose corn syrup-induced hyperuricemia rats	50–100 mg/kg	[44]
	Luteolin	Fruits and vegetable	Potassium oxonate-induced hyperuricemia mice	3–10 mg/kg	[45]
	Luteolin-4'-O-glucoside	Fruits and vegetable	Potassium oxonate-induced hyperuricemia mice	20–100 mg/kg	[45]
	Fisetin	Vegetables and fruits	Potassium oxonate-induced hyperuricemia mice	50–100 mg/kg	[46]
	Morin	Plants and fruits of the Moraceae family	Potassium oxonate-induced hyperuricemia mice	10–40 mg/kg	[47, 48]
Flavonols	Rutin	Vegetables and fruits	Potassium oxonate-induced hyperuricemia mice	25–100 mg/kg	[49]
	Gossypetin	Flowers of <i>Hibiscus sabdariffa</i>	URAT1-expressing 293A cells	IC ₅₀ = 31.3 μM/l	[50]
	Quercetagenin	Tagetes flowers	URAT1-expressing 293A cells	IC ₅₀ = 18.4 μM/l	[50]
	Quercetin	Vegetables and fruits	URAT1-expressing 293A cells	IC ₅₀ = 12.6 μM/l	[50]
Flavonones	Naringenin	Citrus fruits	URAT1-expressing 293A cells	IC ₅₀ = 16.1 μM/l	[51]
	Hesperetin	Citrus fruits	URAT1-expressing 293A cells	IC ₅₀ = 17.6 μM/l	[51]
	Isobavachin	<i>Psoralea corylifolia</i> L.	URAT1-expressing HEK293 cells and hyperuricemia mice	IC ₅₀ = 0.39 μM/l and 10 mg/kg, respectively	[52]
Flavanols	Epigallocatechin-3-gallate	Green tea	Potassium oxonate-induced hyperuricemia mice	10–50 mg/kg	[53]
	Astilbin	<i>Smilax glabra</i>	Potassium oxonate-induced hyperuricemia mice	5–20 mg/kg	[54, 55]
Isoflavones	Genistein	Leguminosae plants	Potassium oxonate-induced hyperuricemia mice	10–20 mg/kg	[56]
	Mangiferin	<i>Mangifera indica</i> L.	Potassium oxonate-induced hyperuricemia mice	1.5–24.0 mg/kg	[57]
Xanthones	Mangiferin aglycon	<i>Mangifera indica</i> L.	Potassium oxonate-induced hyperuricemia mice	10–30 mg/kg	[58]
	Psoralen	<i>Cullen corylifolium</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[59]
Coumarins	Isopsoralen	<i>Cullen corylifolium</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[59]
	Imperatorin	<i>Angelica dahurica</i> and <i>Angelica sinensis</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[59]
	Isoimperatorin	<i>Angelica dahurica</i> and <i>Angelica sinensis</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[59]
	Xanthoxin	<i>Zanthoxylum bungeanum</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[59]
	Fraxetin	<i>Fraxinus chinensis</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[60]
	Fraxin	<i>Fraxinus chinensis</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[60]
Stilbenes	Osthol	<i>Clinopodium megalanthum</i>	URAT1 and potassium oxonate-induced hyperuricemia mice	IC ₅₀ = 78.8 μM/l and 20–40 mg/kg	[61]
	Resveratrol	Grapes, soybeans, berries, pomegranate, and peanuts	Potassium oxonate-induced hyperuricemia mice	10–40 mg/kg	[62]
	Polydatin	<i>Polygonum cuspidatum</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[63]
	Mulberroside A	<i>Morus alba</i> L.	Potassium oxonate-induced hyperuricemia mice	10–40 mg/kg	[64]

TABLE 1: Continued.

Category	Name	Common or primary source	Cell lines/model	Dosage	Ref.
Terpenes	Loganin	<i>Cornus officinalis</i>	Potassium oxonate-induced hyperuricemia mice	20–40 mg/kg	[63]
	Geniposide	<i>Gardenia jasminoides</i>	Potassium oxonate-induced hyperuricemia mice	100–200 mg/kg	[65]
	13 β , 18-dihydroxyeurycomanol	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	$\Delta^{4,5}$,14-hydroxyglauucarubol	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	13 β ,	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	21-dihydroxyeurycomanol	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	Eurycomanol	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	13 β ,	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	21-dihydroxyeurycomanone	<i>Eurycoma longifolia</i>	URAT1-expressing HEK293T cells	50 μ M/l	[66]
	13 α 21-epoxyeurycomanone	<i>Elaeagnus pungens</i>	Potassium oxonate-induced hyperuricemia mice	25–100 mg/kg	[67]
Alkaloids	Emodinol	Beet	Potassium oxonate-induced hyperuricemia mice	5–40 mg/kg	[68]
	Betaine	<i>Nelumbo nucifera</i>	Potassium oxonate-induced hyperuricemia mice	5–40 mg/kg	[69]
	Nuciferine	<i>Coptis chinensis</i> and <i>Phellodendron chinense</i>	Potassium oxonate-induced hyperuricemia mice	6.25–25.0 mg/kg	[70]
	Berberine	<i>Coptis chinensis</i> and <i>Phellodendron chinense</i>	Potassium oxonate-induced hyperuricemia mice	25–50 mg/kg	[71]
	Dihydroberberine	<i>Coptis chinensis</i> and <i>Phellodendron chinense</i>	Potassium oxonate-induced hyperuricemia mice	319.22–1276.86 mg/kg	[72]
Steroids	Dioscin	Fenugreek plant	URAT1-expressing HCT116 cells	10–100 μ M/l	[73]
	Tigogenin	<i>Agave sisalana</i>	Potassium oxonate-induced hyperuricemia mice	3–10 mg/kg	[74]
Phenolic acids	Withaferin A	<i>Withania somnifera</i>	Potassium oxonate-induced hyperuricemia mice	0.75 mmol/l	[75]
	Chlorogenic acid	Honeysuckle	Potassium oxonate-induced hyperuricemia mice	20–80 mg/kg	[76]
Acetophenone	2,5-Dihydroxyacetophenone	<i>Ganoderma applanatum</i>	Potassium oxonate-induced hyperuricemia mice		

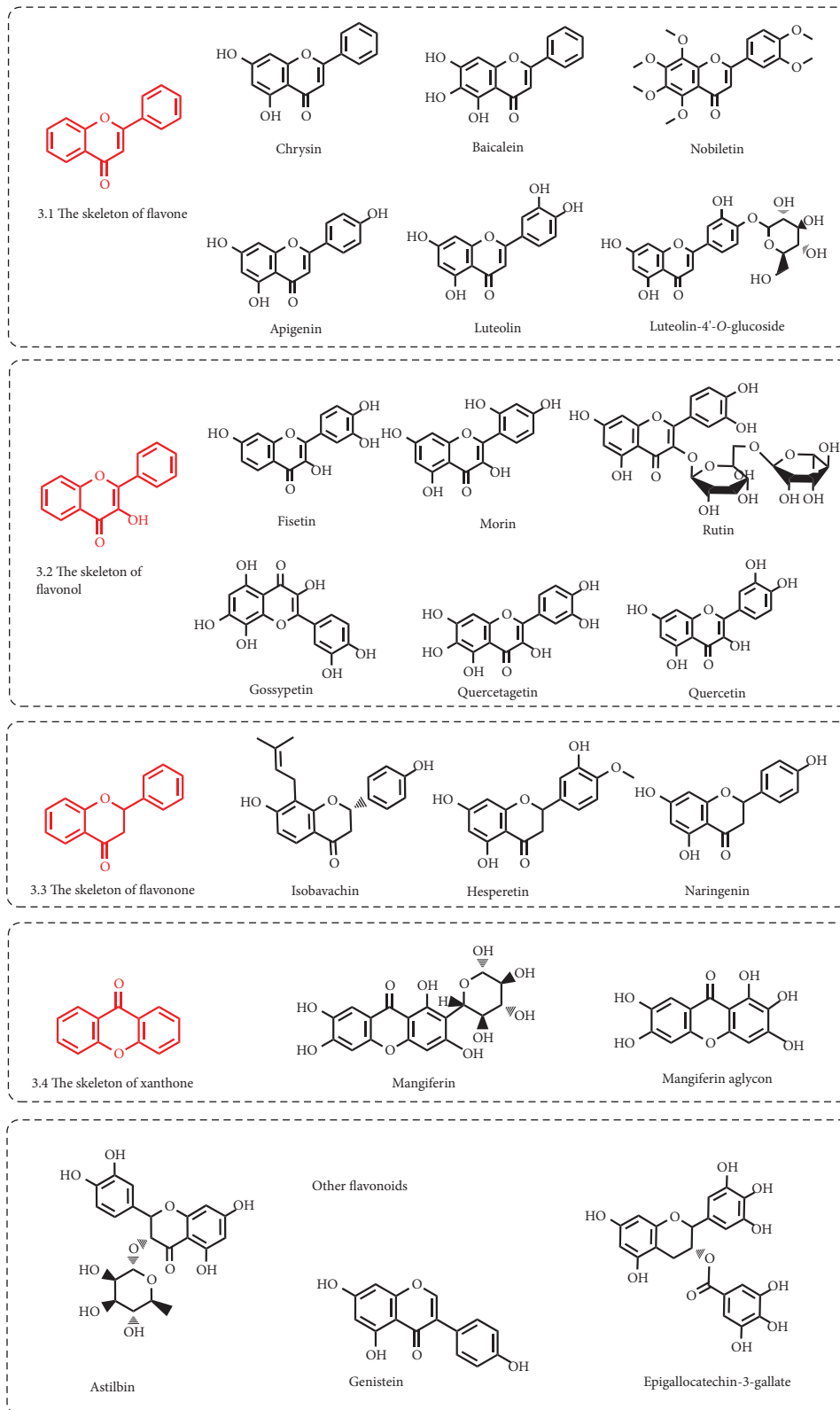


FIGURE 3: Structural formula of flavonoids with URAT1 inhibitory effect.

positions 5 and 7 of the A ring, and the differences are reflected in the number of hydroxyl groups at positions 3, 4, and 5 of the B ring. Chrysin is mainly derived from propolis, blue passion flower, and honey [81]. In rats induced by high fructose corn

symp syrup hyperuricemia, chrysin (50–150 mg/kg) could inhibit the expression of URAT1 and promote uric acid excretion [44]. The main sources of apigenin are the leafy herbs parsley and dried chamomile flowers [82]. Cellular experiments showed

that apigenin (3.125–100 $\mu\text{M/l}$) could inhibit cellular uptake of uric acid in HK-2 cells treated with uric acid by inhibiting URAT1 expression [42]. Li et al. found that apigenin ($\text{IC}_{50} = 0.64 \mu\text{M/l}$) not only competitively inhibited URAT1 activity in vitro, but also (100 mg/kg) promoted uric acid excretion by inhibiting URAT1 activity in potassium oxonate-induced hyperuricemic nephropathy mice [43]. The main source of baicalin is the root of *Scutellaria baicalensis*. Baicalin ($\text{IC}_{50} = 31.56 \mu\text{M/l}$) could non-competitively inhibit URAT1 activity in vitro and (200 mg/kg) improved renal urate excretion by inhibiting URAT1 expression in potassium oxonate-induced hyperuricemia mice. Protein docking analysis revealed that baicalin interacted with Ser35 and Phe241 of URAT1 [41]. Nobiletin is a highly methoxylated flavone compound, especially abundant in citrus [40]. Cell experiments showed that nobiletin ($\text{IC}_{50} = 17.6 \mu\text{M/l}$) could inhibit URAT1 expression and uric acid uptake in 293A cells expressing URAT1 treated with uric acid [51]. Luteolin is widely found in fruits and vegetables [83]. The animal experiment showed that both luteolin (3–10 mg/kg) and luteolin-4'-*O*-glucoside (20–100 mg/kg) could inhibit URAT1 expression and promote uric acid excretion in potassium oxonate-induced hyperuricemia mice [45].

3.1.2. Flavonols. It can be seen in Figure 3 that flavonols are characterized by containing a hydroxyl group at position 3 of the C ring [84]. Current studies show that six flavonols are promising as URAT1 inhibitors, which are gossypetin, quercetagenin, quercetin, fisetin, morin, and rutin. Cellular experiments showed that gossypetin (isolated from *Hibiscus sabdariffa* flowers), quercetagenin (isolated from tagetes flowers), and quercetin (widespread in vegetables and fruits) could inhibit URAT1 expression and uric acid uptake in 293A cells expressing URAT1, and the IC_{50} values were 31.3 $\mu\text{M/l}$, 18.4 $\mu\text{M/l}$, and 12.6 $\mu\text{M/l}$, respectively [50, 85–87]. Rutin, also called rutoside, quercetin-3-rutinoside, and sophorin, is abundant in vegetables and fruits, such as passion flower, tea, apple, asparagus, blackberry, quince, cherry, and red plum [88]. Fisetin is also widely found in vegetables and fruits, such as strawberry, blueberry, apple, grape, persimmon, kiwi, and cucumber [89]. The source of morin is mainly Moraceae plants [90]. Animal studies have shown that fisetin (50–100 mg/kg), morin (10–40 mg/kg), and rutin (25–100 mg/kg) could inhibit URAT1 activity and promote uric acid excretion in potassium oxonate-induced hyperuricemia mice [46–49].

3.1.3. Flavanols. Compared to flavonol, the structural characteristic of flavanol is that the C ring has no carbonyl group and the double bond at positions 2 and 3 is hydrogenated [91]. Flavanols are divided into flavan-3-ols and flavan-3,4-diols according to the position of the hydroxyl group in the C-ring. Current research has shown that only epigallocatechin-3-gallate, the main component of green tea polyphenols, has a URAT1 inhibitory effect [92]. As can be seen from the structure, epigallocatechin-3-gallate is an ester formed by epigallocatechin and gallic acid and belongs to the flavan-3-ols. The animal study showed that epigallocatechin-

3-gallate (10–50 mg/kg) inhibited the expression of URAT1 and promoted uric acid excretion in hyperuricemia mice induced by potassium oxonate [53].

3.1.4. Flavonones. It can be seen in Figure 3 that the structural characteristic of flavonone is that the double bond at positions 2 and 3 of the C ring is hydrogenated [93]. Current research shows that flavonones with the inhibitory effect of URAT1 include hesperetin, naringenin, and isobavachin [94]. Both hesperetin and naringenin derive mainly from citrus fruits such as oranges and lemons [95]. Isobavachins are derived from the seeds of *Psoralea corylifolia* L. [96]. Hesperetin ($\text{IC}_{50} = 17.6 \mu\text{M/l}$) and naringenin ($\text{IC}_{50} = 16.1 \mu\text{M/l}$) could inhibit URAT1 expression and uric acid uptake in URAT1-expressing 293A cells [51]. Isobavachin could also inhibit URAT1 expression and uric acid uptake in URAT1-expressing HEK293 cells ($\text{IC}_{50} = 0.39 \mu\text{M/l}$) and promote uric acid excretion in potassium oxonate-induced hyperuricemia mice (10 mg/kg) [52].

3.1.5. Flavanonols. It can be seen in Figure 3 that flavanonol is produced by hydrogenation of the double bond at positions 2 and 3 of the C ring of flavonol [97]. The current study showed that only astilbin, a flavanonol glucoside of *Smilax glabra*, has the inhibitory effect of URAT1 [98]. In potassium oxonate-induced hyperuricemia mice, astilbin (5–20 mg/kg) inhibited URAT1 expression and promoted the excretion of uric acid [54, 55].

3.1.6. Isoflavones. Compared to flavone, the structural characteristic of isoflavone is that the B ring is attached to the 3-position of the C ring [99]. Current research has shown that only genistein derived from plants of Leguminosae has the inhibitory effect of URAT1 [100]. In potassium oxonate-induced hyperuricemia mice, genistein (10–20 mg/kg) inhibited URAT1 expression and promoted uric acid excretion [56].

3.1.7. Xanthonnes. Xanthonnes (dibenzo- γ -pyrones) constitute an important class of oxygenated heterocycles and occur as secondary metabolites in plants and microorganisms. Xanthonnes do not conform to the basic skeleton of $\text{C}_6\text{-C}_3\text{-C}_6$, but are also classified as flavonoids due to their benzo γ -pyranone structure [101]. Current research has shown that xanthonnes with the inhibitory effect of URAT1 include mangiferin and mangiferin aglycon, and they mainly derive from *Mangifera indica* L. [102]. The animal experiment showed that mangiferin (1.5–24.0 mg/kg) and mangiferin aglycon (10–30 mg/kg) inhibited URAT1 expression and promoted uric acid excretion in potassium oxonate-induced hyperuricemia mice [57, 58].

3.2. Terpenoids. Terpenoids consist of isoprene units and can be divided into hemiterpenes, monoterpenes, sesquiterpenes, diterpenes, disesquiterpenes, triterpenes, and polyterpenes according to the number of units containing

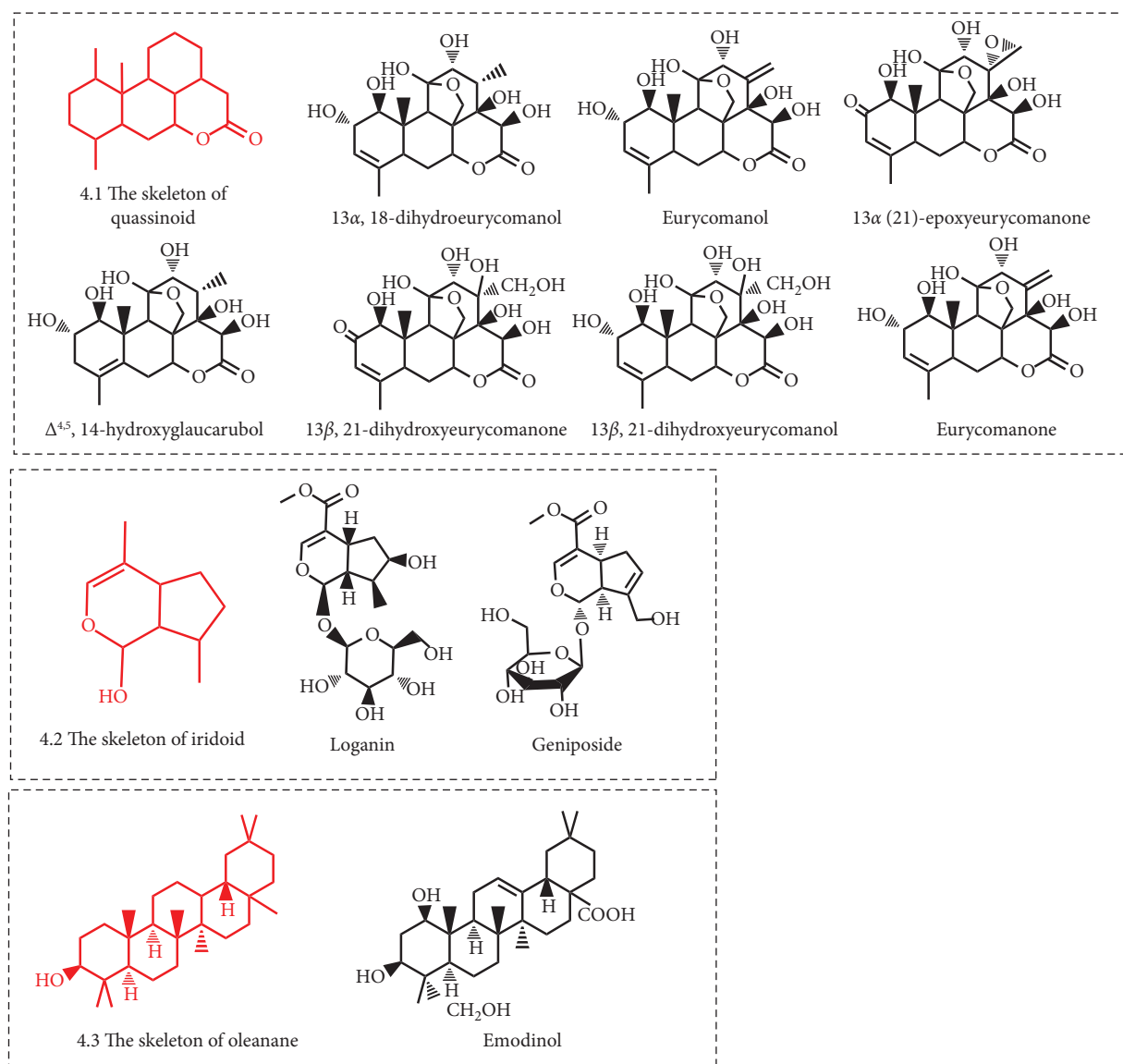


FIGURE 4: Structural formula of terpenes with URAT1 inhibitory effect.

isoprene [103]. The current study shows that terpenoids with an URAT1 inhibitory effect include monoterpenes and triterpenes (Figure 4).

Only two iridoids among monoterpenes show the URAT1 inhibitory effect, including loganin and geniposide. Loganin is a common iridoid glycoside derived from *Cornus officinalis* [104]. Geniposide is also an iridoid glycoside and an important active ingredient of *Gardenia jasminoides* [105]. Studies have shown that both loganin (20–40 mg/kg) and geniposide (100–200 mg/kg) could inhibit URAT1 activity and promote uric acid excretion in potassium oxonate-induced hyperuricemia mice [63, 65].

Triterpenoids with URAT1 inhibitory effect are mainly a series of quassinoids extracted from *Eurycoma longifolia*, including eurycomanol, eurycomanone, 13 β ,18-dihydroeurycomanol, $\Delta^{4,5}$,14-hydroxyglauucarubol, 13 β ,21-dihydroxyeurycomanol, 13 β ,21-dihydroxyeurycomanone, and 13 α (21)-epoxyeurycomanone [66]. The structural

differences of these quassinoids are reflected in the differences of the substituents of the 2 and 21 positions. The cellular experiment showed that these quassinoids (50 μ M/l) decreased urate uptake in HEK293T cells expressing URAT1 by inhibiting URAT1 activity [66]. Furthermore, emodinol, a triterpenoid extracted from *Elaeagnus pungens*, also had the inhibitory effect of URAT1, which (25–100 mg/kg) inhibited the expression of URAT1 and promoted uric acid excretion in potassium oxonate-induced hyperuricemia mice [67, 106].

3.3. Coumarins. Coumarin is a general term for a class of natural compounds with benzo- α -pyrone core, which can be regarded as lactones formed by the dehydration of cis-*o*-hydroxycinnamic acid [107]. Currently, studies have shown that a variety of coumarins have an inhibitory effect on URAT1 (Figure 5), including psoralen and isopsoralen

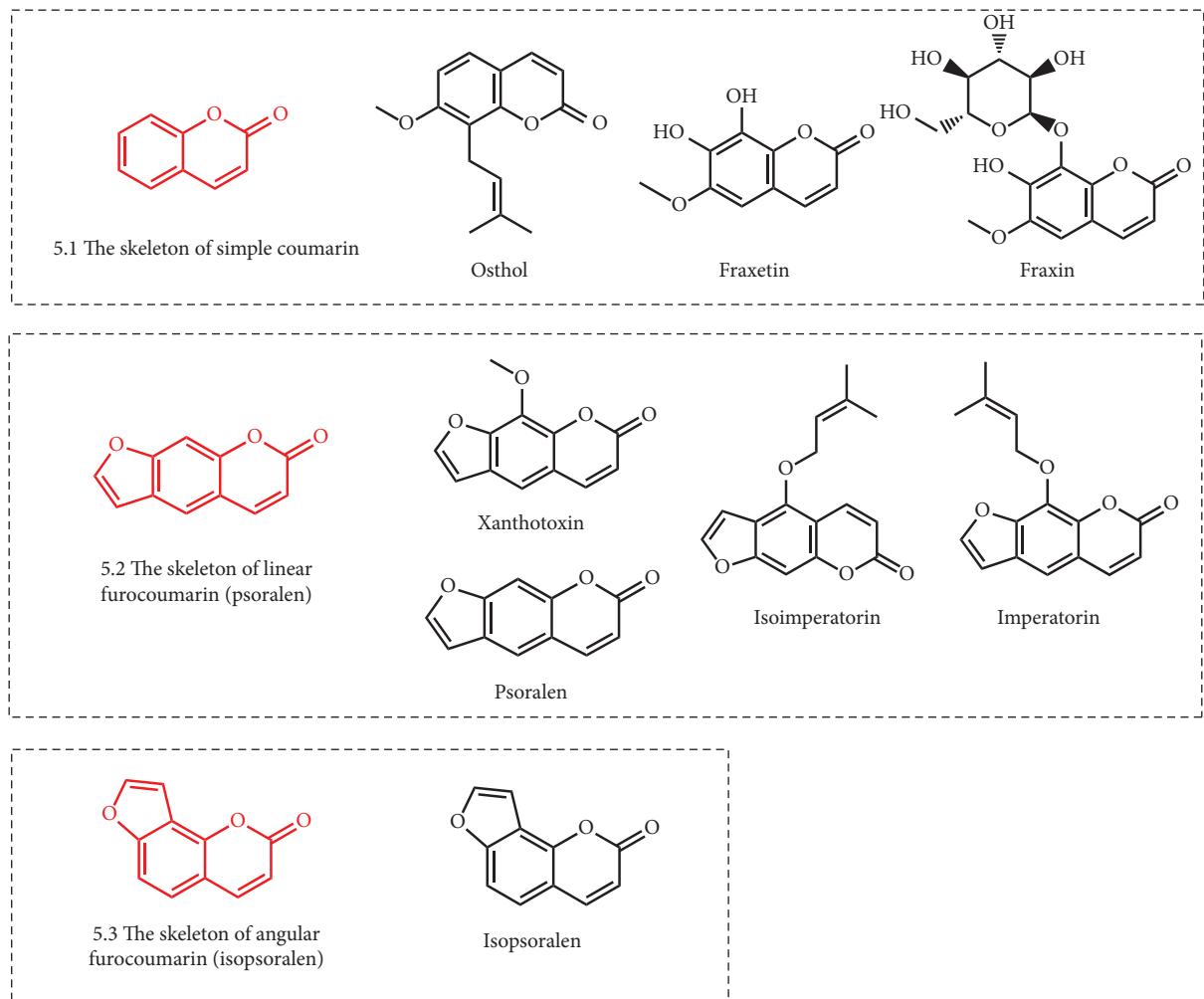


FIGURE 5: Structural formula of coumarins with the URAT1 inhibitory effect.

(extracted from *Cullen corylifolium*) [108], imperatorin and isoimperatorin (extracted from *Angelica dahurica* and *Angelica sinensis*) [109], xanthotoxin (extracted from *Zanthoxylum bungeanum*) [110], fraxetin and fraxin (extracted from *Fraxinus chinensis*) [111], and osthol (extracted from *Clinopodium megalanthum*) [112]. Osthol, fraxetin, and fraxin are simple coumarins characterized by the 7-position hydroxyl group not forming a furan or pyran ring with the 6- or 8-position isopentenyl group. Studies have shown that osthol ($IC_{50} = 78.8 \mu M/l$) could non-competitively inhibit URAT1 activity in vitro, and both fraxetin and fraxin (20–40 mg/kg) could negatively regulate URAT1 expression in potassium oxonate-induced hyperuricemia mice [60, 61]. Xanthotoxin, psoralen, imperatorin, and isoimperatorin are linear furocoumarins formed by the condensation of the 7-position hydroxyl group with the 6-position isopentenyl group. Isopsoralen is an angular furocoumarin formed by condensation of the 7-position hydroxyl group with the 8-position isopentenyl group. Animal studies have shown that these furocoumarins (20–40 mg/kg) inhibited URAT1 activity and promoted uric acid excretion in mice with potassium oxonate-induced hyperuricemia nephropathy [59, 60].

3.4. Stilbenes. Stilbenes refer to the general term of monomers with a 1,2-diphenylethylene skeleton and their polymers [113]. Current studies show that stilbenes with URAT1 inhibitory effects include resveratrol, polydatin, and mulberroside A (Figure 6). Resveratrol is a well-known stilbene compound present in grapes, soybeans, berries, pomegranate, and peanuts [114]. Animal experiments have shown that resveratrol (10–40 mg/kg) could promote uric acid excretion by inhibiting URAT1 activity in potassium oxonate-induced hyperuricemia mice. Furthermore, the researchers believed that this was related to inhibiting the activation of the inflammatory response, namely inhibiting the NLRP3 (NOD-like receptor family, pyrin domain-containing 3) inflammasome and TLR4 (toll-like receptor 4)/MyD88 (myeloid differentiation factor 88)/NF- κ B (nuclear factor- κ B) signaling pathway [62]. In addition to resveratrol, Chen et al. found that the resveratrol tetramer (20–60 mg/kg) could also inhibit URAT1 activity and promote uric acid excretion in mice with potassium oxonate induced hyperuricemia mice [77]. Polydatin and mulberroside A are two stilbene compounds with a very similar structure derived from *Polygonum cuspidatum* and *Morus alba* L., respectively [115, 116]. In potassium oxonate-

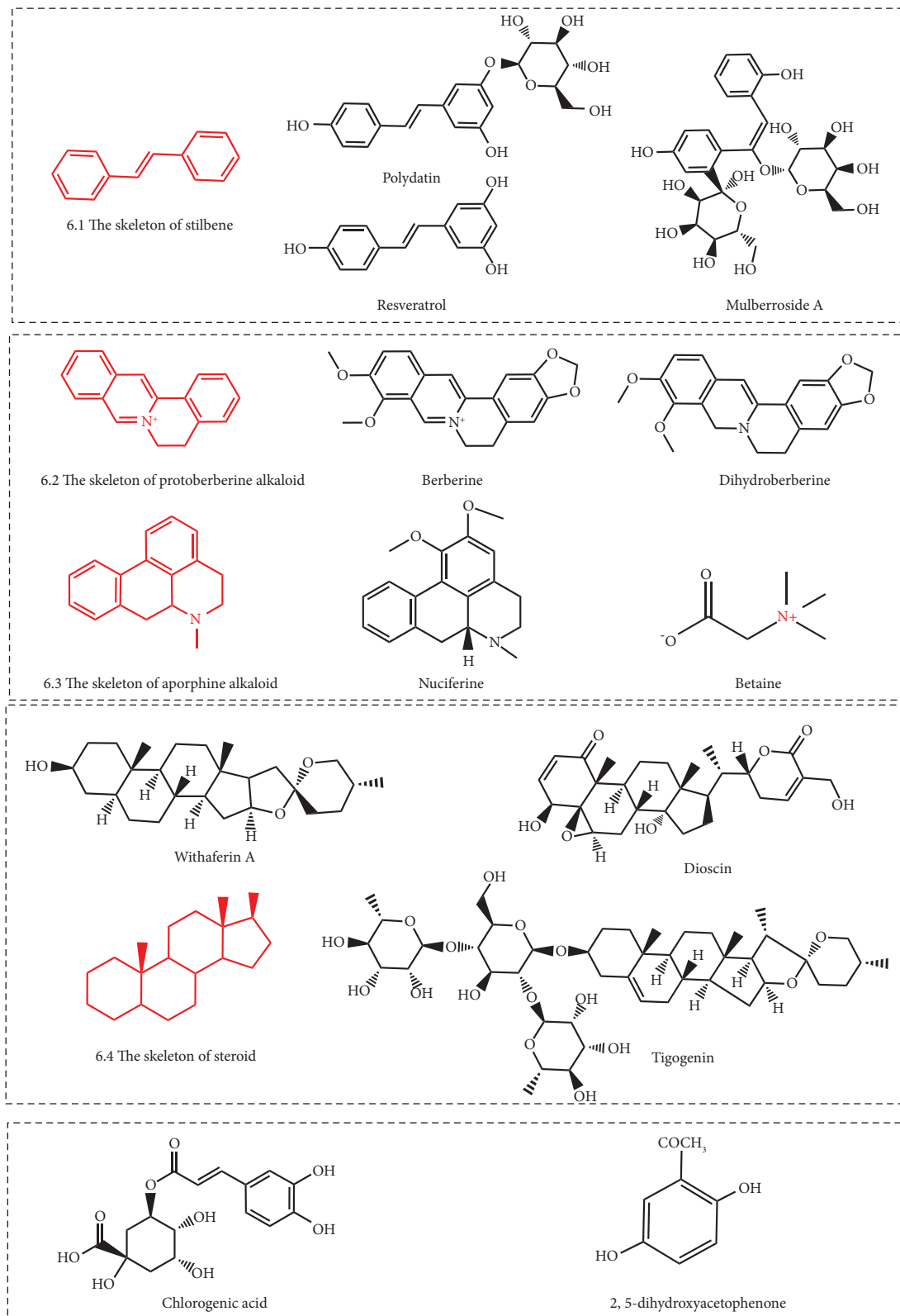


FIGURE 6: Structural formula of stilbenes, alkaloids, steroids, and other natural products with an URAT1 inhibitory effect.

induced hyperuricemia mice, polydatin (20–40 mg/kg) and mulberroside A (10–40 mg/kg) could down-regulate the expression of URAT1 in the kidney and promote uric acid excretion [63, 64].

3.5. Alkaloids. Alkaloids are a class of nitrogen-containing organic compounds derived primarily from plants [117]. Current studies show that alkaloids with URAT1 inhibition include betaine, nuciferine, berberine, and dihydroberberine

(Figure 6). Betaine is a quaternary ammonium-type alkaloid derived from beet [118]. Nuciferine is an aporphine alkaloid extracted from *Nelumbo nucifera* [119]. Berberine and dihydroberberine are two isoquinoline alkaloids derived from *Coptis chinensis* and *Phellodendron chinense* [119, 120]. Animal experiments showed that betaine (5–40 mg/kg), nuciferine (5–40 mg/kg), berberine (6.25–25.0 mg/kg) and dihydroberberine (25–50 mg/kg) could inhibit URAT1 expression and promote the excretion of uric acid in potassium oxonate-induced hyperuricemia mice [68–71].

3.6. Steroids. Steroid is a general term for a large class of compounds with the basic skeleton structure of perhydrocyclopentano-phenanthrene [121]. Current studies have shown that steroids with an inhibitory effect of URAT1 include withaferin A, dioscin, and tigogenin (Figure 6). Dioscin is an isospirostane glycoside mainly extracted from the fenugreek plant [122]. Su et al. found that dioscin (319.22–1276.86 mg/kg) could negatively regulate URAT1 expression in potassium oxonate-induced hyperuricemia mice [72]. Tigogenin is a spirostane glycoside extracted from *Agave sisalana* [123]. Zhang et al. found that tigogenin (10–100 μ M/l) could decrease uric acid uptake in URAT1-expressing HCT116 cells by inhibiting URAT1 activity [73]. Withaferin A extracted from *Withania somnifera*, is a steroidal lactone [124]. In potassium oxonate-induced hyperuricemia mice, withaferin A (3–10 mg/kg) negatively regulated URAT1 expression in the kidney and promoted uric acid excretion [74].

3.7. Other Natural Products. Chlorogenic acid is an organic acid derived from honeysuckle [125]. In vitro research showed that chlorogenic acid (0.75 mmol/l) could inhibit uric acid reuptake in URAT1 expressing-HEK293T cells by inhibiting URAT1 expression [75]. Liang et al. isolated 2,5-dihydroxyacetophenone from *Ganoderma applanatum*, which (20–80 mg/kg) could inhibit URAT1 activity in mice induced by potassium oxonate hyperuricemia [76]. Li et al. isolated a pure polysaccharide ULP from *Ulva lactuca* consisting of rhamnose, glucuronic acid, galactose and xylose at a molar ratio of 32.75:22.83:1.07:6.46 with a molecular weight of 2.24×10^5 Da. In potassium oxonate-induced hyperuricemia mice, ULP (10–50 mg/kg) inhibited the expression of URAT1 and promoted uric acid excretion [126].

4. Review and Speculation of the Structure-Activity Relationship

Currently, most studies have focused on finding natural products with inhibition of URAT1 and have not explored their activities and mechanisms toward URAT1. Therefore, in addition to exploring the mechanism of action, exploring the structure-activity relationship will be another important research direction in the future. By reviewing the few existing studies on the structure-activity relationship studies and analyzing common features of natural products with URAT1 inhibition, we speculate that the rigid ring structure

and negative charge may be the keys for natural products to produce URAT1 inhibition. We hope that the analysis and speculation in this chapter can broaden the current understanding and trigger further interest in exploring the relationship between structure and URAT1 inhibition of natural products.

First, natural products may need to contain rigid structures. It can be found that among these natural products with URAT1 inhibition, almost all of them have a rigid ring structure. For example, one third of natural products with URAT1 inhibition are flavonoids. It can be seen from the structure of the flavonoid that two benzene rings connected to an oxygen-containing pyranol group are a typical rigid plane molecular structure [127]. Polycyclic terpenoids, such as quassinoids, also contain unique rigid ring backbones [128]. In addition, other compounds also contain rigid ring structures, such as the benzene ring, the naphthalene ring, or the pyridine ring. Therefore, the presence of a rigid structure may be one of the elements that natural products must use to inhibit URAT1.

Second, natural products may require anions to act as URAT1 inhibitors. Wempe et al. evaluated the inhibitory effect of a series of (2-ethylbenzofuran-3-yl) (substituted-phenyl) methanone compounds on URAT1 activity in oocytes expressing hURAT1. The experimental data indicated that a potent hURAT1 inhibitor requires an anion (that is, a formal negative charge) to interact with the positively charged URAT1 binding pocket [129]. The C-ring of flavonoids is an electron-rich region with a strong negative charge. This partially explains the inhibitory activities of flavonoids in URAT1 [130]. Furthermore, most natural products with URAT1 inhibition also contain phenolic hydroxyl groups, so that these compounds can show acidity and generate anions [131]. However, alkaloids including betaine, nuciferine, berberine, and dihydroberberine also show inhibition of URAT1, so the presence of anions may not be a determinant of whether a natural product has inhibition of URAT1.

5. Conclusion and Prospects

In summary, current studies have shown that many natural products have a URAT1 inhibitory effect and are expected to be developed as URAT1 inhibitors, including flavonoids, terpenoids, alkaloids, coumarins, stilbenes, steroids, organic acids, and polysaccharides. The number of flavonoids is the largest among them, including many subtypes. Animal experiments have shown that these natural products can inhibit URAT1 activity in hyperuricemia mice and promote uric acid excretion. By reviewing the few existing studies on the structure-activity relationship studies and analyzing common features of natural products with URAT1 inhibition, we speculate that the rigid ring structure and negative charge may be the keys for natural products to produce URAT1 inhibition.

Although studies have confirmed that natural products are promising as URAT1 inhibitors, there are still some issues that need to be addressed in the future. First, the mechanism by which these natural products inhibit URAT1

is unclear. Therefore, more research is needed to explore the mechanism of action. Second, current research is still in the experimental study stage and it is necessary to carry out clinical research to further explore its therapeutic effects. Third, the relationship between structure and URAT1 inhibitory activity requires further investigation. In addition to the rigid ring structure and negative charge, what other structural features are essential for the URAT1 inhibitory effect of natural products? Fourth, structural modification is a common method to improve the therapeutic effect of drugs and reduce side effects. Therefore, structural modification based on clarifying the structure-activity relationship of natural products to improve the inhibitory activity of URAT1 may be a key research direction in the future.

Data Availability

Data sharing are not applicable to this article, as no new data were created or analyzed in this study.

Disclosure

Qianghua yuan, Yuan cheng, Rong sheng and Mei hu should be regarded as the co first author.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Qianghua yuan, Yuan cheng, Rong sheng and Mei hu contributed equally to this work.

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References

- [1] D. Xie, H. Zhao, J. Lu et al., "High uric acid induces liver fat accumulation via ROS/JNK/AP-1 signaling," *American Journal of Physiology—Endocrinology And Metabolism*, vol. 320, no. 6, pp. E1032–E1043, 2021.
- [2] C. G. Li, M. C. Hsieh, and S. J. Chang, "Metabolic syndrome, diabetes, and hyperuricemia," *Current Opinion in Rheumatology*, vol. 25, no. 2, pp. 210–216, 2013.
- [3] B. Jakše, B. Jakše, M. Pajek, and J. Pajek, "Uric acid and plant-based nutrition," *Nutrients*, vol. 11, p. 1736, 2019.
- [4] C. Yokose, N. McCormick, and H. K. Choi, "Dietary and lifestyle-centered approach in gout care and prevention," *Current Rheumatology Reports*, vol. 23, no. 7, p. 51, 2021.
- [5] Y. F. Guo, H. Lu, J. Gan, D. D. Li, J. D. Gao, and C. M. Zhang, "Efficacy of Chinese herbal medicine jiangniaosuan formula for treatment of hyperuricemia: study protocol for a double-blinded non-inferiority randomized controlled clinical trial," *Trials*, vol. 23, no. 1, p. 1, 2022.
- [6] P. Liu, H. C. Xu, Y. C. Shi, L. Deng, and X. Y. Chen, "Potential molecular mechanisms of plantain in the treatment of gout and hyperuricemia based on network pharmacology," *Evidence-Based Complementary and Alternative Medicine*, vol. 2020, Article ID 3023127, 20 pages, 2020.
- [7] A. M. Abeles and M. H. Pillinger, "Gout and cardiovascular disease: crystallized confusion," *Current Opinion in Rheumatology*, vol. 31, no. 2, pp. 118–124, 2019.
- [8] Z. L. Wang, Y. C. Li, W. H. Liao et al., "Gut microbiota remodeling: a promising therapeutic strategy to confront hyperuricemia and gout," *Frontiers in Cellular and Infection Microbiology*, vol. 12, Article ID 935723, 2022.
- [9] M. Gliozzi, N. Malara, S. Muscoli, and V. Mollace, "The treatment of hyperuricemia," *International Journal of Cardiology*, vol. 213, pp. 23–27, 2016.
- [10] Y. Chen, R. You, K. Wang, and Y. Wang, "Recent updates of natural and synthetic URAT1 inhibitors and novel screening methods," *Evidence-Based Complementary and Alternative Medicine*, vol. 2021, Article ID 5738900, 12 pages, 2021.
- [11] L. Strilchuk, F. Fogacci, and A. F. Cicero, "Safety and tolerability of available urate-lowering drugs: a critical review," *Expert Opinion on Drug Safety*, vol. 18, no. 4, pp. 261–271, 2019.
- [12] D. N. Song, X. Zhao, F. Q. Wang, and G. Wang, "A brief review of urate transporter 1 (URAT1) inhibitors for the treatment of hyperuricemia and gout: current therapeutic options and potential applications," *European Journal of Pharmacology*, vol. 907, Article ID 174291, 2021.
- [13] W. Zhang, M. Doherty, T. Bardin et al., "EULAR evidence based recommendations for gout. Part II: management. Report of a task force of the EULAR standing committee for international clinical studies including therapeutics (ESCI-SIT)," *Annals of the Rheumatic Diseases*, vol. 65, no. 10, pp. 1312–1324, 2006.
- [14] S. M. Hoy, "Lesinurad: first global approval," *Drugs*, vol. 76, no. 4, pp. 509–516, 2016.
- [15] Y. Dong, T. Zhao, W. Ai et al., "Novel urate transporter 1 (URAT1) inhibitors: a review of recent patent literature (2016–2019)," *Expert Opinion on Therapeutic Patents*, vol. 29, no. 11, pp. 871–879, 2019.
- [16] F. Shakeri, V. Bianconi, M. Pirro, and A. Sahebkar, "Effects of plant and animal natural products on mitophagy," *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 6969402, 11 pages, 2020.
- [17] N. A. Ratcliffe, C. B. Mello, E. S. Garcia, T. M. Butt, and P. Azambuja, "Insect natural products and processes: new treatments for human disease," *Insect Biochemistry and Molecular Biology*, vol. 41, no. 10, pp. 747–769, 2011.
- [18] L. Chen, X. Y. Wang, R. Z. Liu, and G. Y. Wang, "Culturable microorganisms associated with sea cucumbers and microbial natural products," *Marine Drugs*, vol. 19, no. 8, p. 461, 2021.
- [19] H. D. Yoo, S. J. Nam, Y. W. Chin, and M. S. Kim, "Misassigned natural products and their revised structures," *Archives of Pharmacal Research*, vol. 39, no. 2, pp. 143–153, 2016.
- [20] J. P. Lin, S. Q. Chen, S. Z. Li, M. L. Lu, Y. A. Li, and Y. X. Su, "Efficacy and safety of Chinese medicinal herbs for the treatment of hyperuricemia: a systematic review and meta-analysis," *Evidence-Based Complementary and Alternative Medicine*, vol. 2016, Article ID 2146204, 12 pages, 2016.

- [21] X. D. Hu, Y. Yang, X. N. Hu et al., "Effects of sodium-glucose cotransporter 2 inhibitors on serum uric acid in patients with type 2 diabetes mellitus: a systematic review and network meta-analysis," *Diabetes, Obesity and Metabolism*, vol. 24, no. 2, pp. 228–238, 2022.
- [22] A. F. G. Cicero, F. Fogacci, R. I. Cincione, G. Tocci, and C. Borghi, "Clinical effects of xanthine oxidase inhibitors in hyperuricemic patients," *Medical Principles and Practice*, vol. 30, no. 2, pp. 122–130, 2021.
- [23] E. Dominguez-Zambrano, J. Pedraza-Chaverri, A. L. López-Santos et al., "Association between serum uric acid levels, nutritional and antioxidant status in patients on hemodialysis," *Nutrients*, vol. 12, no. 9, p. 2600, 2020.
- [24] S. S. Zhang, Y. Wang, J. S. Cheng et al., "Hyperuricemia and cardiovascular disease," *Current Pharmaceutical Design*, vol. 25, no. 6, pp. 700–709, 2019.
- [25] A. Danve, S. T. Sehra, and T. Neogi, "Role of diet in hyperuricemia and gout," *Best Practice & Research Clinical Rheumatology*, vol. 35, no. 4, Article ID 101723, 2021.
- [26] L. J. Li, Y. P. Zhang, and C. C. Zeng, "Update on the epidemiology, genetics, and therapeutic options of hyperuricemia," *American Journal of Tourism Research*, vol. 12, no. 7, pp. 3167–3181, 2020.
- [27] M. Kakutani-Hatayama, M. Kadoya, H. Okazaki et al., "Nonpharmacological management of gout and hyperuricemia: hints for better lifestyle," *American Journal of Lifestyle Medicine*, vol. 11, no. 4, pp. 321–329, 2017.
- [28] F. Liote, "Gout furunculosis," *Joint Bone Spine*, vol. 86, no. 1, p. 103, 2019.
- [29] T. Wakabayashi, S. Ueno, T. Nakatsuji et al., "Safety profiles of new xanthine oxidase inhibitors: a post-marketing study," *International Journal of Clinical Pharmacology & Therapeutics*, vol. 59, no. 05, pp. 372–377, 2021.
- [30] A. Nakayama, H. Nakaoka, K. Yamamoto et al., "GWAS of clinically defined gout and subtypes identifies multiple susceptibility loci that include urate transporter genes," *Annals of the Rheumatic Diseases*, vol. 76, no. 5, pp. 869–877, 2017.
- [31] A. K. Mandal, A. Mercado, A. Foster, K. Zandi-Nejad, and D. B. Mount, "Uricosuric targets of tranilast," *Pharmacology Research & Perspectives*, vol. 5, Article ID e00291, 2017.
- [32] S. K. Nigam and V. Bhatnagar, "The systems biology of uric acid transporters: the role of remote sensing and signaling," *Current Opinion in Nephrology and Hypertension*, vol. 27, no. 4, pp. 305–313, 2018.
- [33] P. K. Tan, S. Liu, E. Gunic, and J. N. Miner, "Discovery and characterization of verinurad, a potent and specific inhibitor of URAT1 for the treatment of hyperuricemia and gout," *Scientific Reports*, vol. 7, no. 1, p. 665, 2017.
- [34] M. Ljubojevic, D. Balen, D. Breljak et al., "Renal expression of organic anion transporter OAT2 in rats and mice is regulated by sex hormones," *American Journal of Physiology—Renal Physiology*, vol. 292, no. 1, pp. F361–F372, 2007.
- [35] A. Enomoto and H. Endou, "Roles of organic anion transporters (OATs) and a urate transporter (URAT1) in the pathophysiology of human disease," *Clinical and Experimental Nephrology*, vol. 9, no. 3, pp. 195–205, 2005.
- [36] W. Q. Cai, W. Liu, C. Y. Liu, J. W. Wang, and G. L. Zhao, "A systematic review of uric acid transporter 1 (URAT1) inhibitors for the treatment of hyperuricemia and gout and an insight into the structure-activity relationship (SAR)," *Chinese Journal of Structural Chemistry*, vol. 36, no. 6, pp. 897–910, 2017.
- [37] A. Enomoto, H. Kimura, A. Chairoungdua et al., "Molecular identification of a renal urate-anion exchanger that regulates blood urate levels," *Nature*, vol. 417, no. 6887, pp. 447–452, 2002.
- [38] F. Sivera, M. Andres, and N. Dalbeth, "A glance into the future of gout," *Therapeutic Advances in Musculoskeletal Disease*, vol. 14, 2022.
- [39] X. Ling and W. Bochu, "A review of phytotherapy of gout: perspective of new pharmacological treatments," *Pharmazie*, vol. 69, no. 4, pp. 243–256, 2014.
- [40] H. Mizuno, H. Yoshikawa, and T. Usuki, "Extraction of nobiletin and tangeretin from peels of shekwasha and ponkan using C(2)mim (MeO)(H)PO2 and centrifugation," *Natural Product Communications*, vol. 14, no. 5, 2019.
- [41] Y. Y. Chen, Z. A. Zhao, Y. M. Li et al., "Baicalein alleviates hyperuricemia by promoting uric acid excretion and inhibiting xanthine oxidase," *Phytomedicine*, vol. 80, Article ID 153374, 2021.
- [42] J. X. Zhu, H. Y. Yang, W. Q. Hu et al., "Active components from *Lagotis brachystachya* maintain uric acid homeostasis by inhibiting renal TLR4-NLRP3 signaling in hyperuricemic mice," *Inflammopharmacology*, vol. 29, no. 4, pp. 1187–1200, 2021.
- [43] Y. M. Li, Z. A. Zhao, J. Luo et al., "Apigenin ameliorates hyperuricemic nephropathy by inhibiting URAT1 and GLUT9 and relieving renal fibrosis via the Wnt/ β -catenin pathway," *Phytomedicine*, vol. 87, Article ID 153585, 2021.
- [44] Y. H. Chang, Y. F. Chiang, H. Y. Chen et al., "Anti-inflammatory and anti-hyperuricemic effects of chrysin on a high fructose corn syrup-induced hyperuricemia rat model via the amelioration of urate transporters and inhibition of NLRP3 inflammasome signaling pathway," *Antioxidants*, vol. 10, no. 4, p. 564, 2021.
- [45] Y. Lin, P. G. Liu, W. Q. Liang et al., "Luteolin-4'-O-glucoside and its aglycone, two major flavones of *gnaphalium affine* D. Don, resist hyperuricemia and acute gouty arthritis activity in animal models," *Phytomedicine*, vol. 41, pp. 54–61, 2018.
- [46] Q. Ren, S. B. Tao, F. Guo et al., "Natural flavonol fisetin attenuated hyperuricemic nephropathy via inhibiting IL-6/JAK2/STAT3 and TGF- β /SMAD3 signaling," *Phytomedicine*, vol. 87, Article ID 153552, 2021.
- [47] Y. W. Shi, C. P. Wang, X. Wang et al., "Uricosuric and nephroprotective properties of ramulus mori ethanol extract in hyperuricemic mice," *Journal of Ethnopharmacology*, vol. 143, no. 3, pp. 896–904, 2012.
- [48] J. J. Zhang, X. Shuai, J. B. Li, N. X. Xiang, T. Gong, and Z. R. Zhang, "Biodistribution, hypouricemic efficacy and therapeutic mechanism of morin phospholipid complex loaded self-nanoemulsifying drug delivery systems in an experimental hyperuricemic model in rats," *Journal of Pharmacy and Pharmacology*, vol. 68, no. 1, pp. 14–25, 2016.
- [49] Y. S. Chen, Q. H. Hu, X. Zhang, Q. Zhu, and L. D. Kong, "Beneficial effect of rutin on oxonate-induced hyperuricemia and renal dysfunction in mice," *Pharmacology*, vol. 92, no. 1-2, pp. 75–83, 2013.
- [50] Y. Toyoda, T. Takada, H. Saito et al., "Identification of inhibitory activities of dietary flavonoids against URAT1, a renal urate Re-absorber: in vitro screening and fractional approach focused on rooibos leaves," *Nutrients*, vol. 14, no. 3, p. 575, 2022.
- [51] Y. Toyoda, T. Takada, H. Saito et al., "Inhibitory effect of Citrus flavonoids on the in vitro transport activity of human

- urate transporter 1 (URAT1/SLC22A12), a renal re-absorber of urate,” *Npj Science of Food*, vol. 4, p. 3, 2020.
- [52] X. Chen, Z. Zhao, J. Luo et al., “Novel natural scaffold as hURAT1 inhibitor identified by 3D-shape-based, docking-based virtual screening approach and biological evaluation,” *Bioorganic Chemistry*, vol. 117, Article ID 105444, 2021.
- [53] C. Zhu, Y. Xu, Z. H. Liu, X. C. Wan, D. X. Li, and L. L. Tai, “The anti-hyperuricemic effect of epigallocatechin-3-gallate (EGCG) on hyperuricemic mice,” *Biomedicine & Pharmacotherapy*, vol. 97, pp. 168–173, 2018.
- [54] M. Wang, J. Zhao, N. Zhang, and J. H. Chen, “Astilbin improves potassium oxonate-induced hyperuricemia and kidney injury through regulating oxidative stress and inflammation response in mice,” *Biomedicine & Pharmacotherapy*, vol. 83, pp. 975–988, 2016.
- [55] S. W. Wang, Y. J. Fang, X. F. Yu, L. Guo, X. X. Zhang, and D. Z. Xia, “The flavonoid-rich fraction from rhizomes of *Smilax glabra* Roxb. ameliorates renal oxidative stress and inflammation in uric acid nephropathy rats through promoting uric acid excretion,” *Biomedicine & Pharmacotherapy*, vol. 111, pp. 162–168, 2019.
- [56] W. Y. Bi and C. L. Zhu, “Genistein ameliorates hyperuricemia-associated nephropathy in hyperuricemic mice,” *Food and Agricultural Immunology*, vol. 32, no. 1, pp. 778–797, 2021.
- [57] H. Yang, L. H. Gao, Y. F. Niu et al., “Mangiferin inhibits renal urate reabsorption by modulating urate transporters in experimental hyperuricemia,” *Biological and Pharmaceutical Bulletin*, vol. 38, no. 10, pp. 1591–1598, 2015.
- [58] Z. Z. Qin, S. B. Wang, Y. H. Lin et al., “Antihyperuricemic effect of mangiferin aglycon derivative J99745 by inhibiting xanthine oxidase activity and urate transporter 1 expression in mice,” *Acta Pharmaceutica Sinica B*, vol. 8, no. 2, pp. 306–315, 2018.
- [59] X. Wang, Y. J. Lou, M. X. Wang, Y. W. Shi, H. X. Xu, and L. D. Kong, “Furocoumarins affect hepatic cytochrome P450 and renal organic ion transporters in mice,” *Toxicology Letters*, vol. 209, no. 1, pp. 67–77, 2012.
- [60] J. M. Li, X. Zhang, X. Wang, Y. C. Xie, and L. D. Kong, “Protective effects of cortex *Fraxini* coumarines against oxonate-induced hyperuricemia and renal dysfunction in mice,” *European Journal of Pharmacology*, vol. 666, no. 1–3, pp. 196–204, 2011.
- [61] Y. Tashiro, R. Sakai, T. Hirose-Sugiura et al., “Effects of osthol isolated from *Cnidium monnieri* fruit on urate transporter 1,” *Molecules*, vol. 23, no. 11, p. 2837, 2018.
- [62] X. M. Zhang, Q. Nie, Z. M. Zhang et al., “Resveratrol affects the expression of uric acid transporter by improving inflammation,” *Molecular Medicine Reports*, vol. 24, no. 2, p. 564, 2021.
- [63] O.-K. Kim, J.-M. Yun, M. Lee, D. Kim, and J. Lee, “Hypouricemic effects of *Chrysanthemum indicum* L. and *Cornus officinalis* on hyperuricemia-induced HepG2 cells, renal cells, and mice,” *Plants*, vol. 10, no. 8, p. 1668, 2021.
- [64] C. P. Wang, Y. M. Wang, X. Wang et al., “Mulberroside A possesses potent uricosuric and nephroprotective effects in hyperuricemic mice,” *Planta Medica*, vol. 77, no. 8, pp. 786–794, 2011.
- [65] C. Liu, H. N. Zhou, R. R. Zhang et al., “Anti-hyperuricemic and nephroprotective effect of geniposide in chronic hyperuricemic mice,” *Journal of Functional Foods*, vol. 61, Article ID 103355, 2019.
- [66] R. X. Bao, M. Y. Liu, D. Wang et al., “Effect of *Eurycoma longifolia* stem extract on uric acid excretion in hyperuricemic mice,” *Frontiers in Pharmacology*, vol. 10, p. 1464, 2019.
- [67] H. Wu, Y. L. Yuan, Y. D. Chen et al., “Hypouricemic and nephroprotective effects of emodinol in oxonate-induced hyperuricemic mice are mediated by organic ion transporters and OIT3,” *Planta Medica*, vol. 82, no. 4, pp. 289–297, 2016.
- [68] Y. L. Liu, Y. Pan, X. Wang et al., “Betaine reduces serum uric acid levels and improves kidney function in hyperuricemic mice,” *Planta Medica*, vol. 80, no. 1, pp. 39–47, 2014.
- [69] M. X. Wang, Y. L. Liu, Y. Yang, D. M. Zhang, and L. D. Kong, “Nuciferine restores potassium oxonate-induced hyperuricemia and kidney inflammation in mice,” *European Journal of Pharmacology*, vol. 747, pp. 59–70, 2015.
- [70] G. S. Lin, Q. X. Yu, L. Q. Xu et al., “Berberine attenuates potassium oxonate- and hypoxanthine-induced hyperuricemia by regulating urate transporters and JAK2/STAT3 signaling pathway,” *European Journal of Pharmacology*, vol. 912, Article ID 174592, 2021.
- [71] L. Q. Xu, G. S. Lin, Q. X. Yu et al., “Anti-hyperuricemic and nephroprotective effects of dihydroberberine in potassium oxonate- and hypoxanthine-induced hyperuricemic mice,” *Frontiers in Pharmacology*, vol. 12, Article ID 645879, 2021.
- [72] J. X. Su, Y. H. Wei, M. L. Liu et al., “Anti-hyperuricemic and nephroprotective effects of *Rhizoma Dioscoreae septemlobae* extracts and its main component dioscin via regulation of mOAT1, mURAT1 and mOCT2 in hypertensive mice,” *Archives of Pharmacological Research*, vol. 37, no. 10, pp. 1336–1344, 2014.
- [73] Y. Zhang, L. J. Jin, J. Liu et al., “Effect and mechanism of dioscin from *Dioscorea spongiosa* on uric acid excretion in animal model of hyperuricemia,” *Journal of Ethnopharmacology*, vol. 214, pp. 29–36, 2018.
- [74] X. Zhao, J. Wang, L. Y. Tang, P. Li, J. Ru, and Y. Z. Bai, “Withaferin A protects against hyperuricemia induced kidney injury and its possible mechanisms,” *Bioengineered*, vol. 12, no. 1, pp. 589–600, 2021.
- [75] H. N. Dai, S. Lv, X. Q. Fu, and W. N. Li, “Identification of scopoletin and chlorogenic acid as potential active components in sunflower calathide enzymatically hydrolyzed extract towards hyperuricemia,” *Applied Sciences*, vol. 11, no. 21, Article ID 10306, 2021.
- [76] D. L. Liang, T. Q. Yong, S. D. Chen et al., “Hypouricemic effect of 2, 5-dihydroxyacetophenone, a computational screened bioactive compound from *Ganoderma applanatum*, on hyperuricemic mice,” *International Journal of Molecular Sciences*, vol. 19, no. 5, p. 1394, 2018.
- [77] Y. S. Chen, C. J. Chen, W. Yan, H. M. Ge, and L. D. Kong, “Anti-hyperuricemic and anti-inflammatory actions of vaticaffinol isolated from *Dipterocarpus alatus* in hyperuricemic mice,” *Chinese Journal of Natural Medicines*, vol. 15, no. 5, pp. 330–340, 2017.
- [78] M. Sisa, S. L. Bonnet, D. Ferreira, and J. H. Van der Westhuizen, “Photochemistry of flavonoids,” *Molecules*, vol. 15, no. 8, pp. 5196–5245, 2010.
- [79] D. M. Kopustinskiene, V. Jakstas, A. Savickas, and J. Bernatoniene, “Flavonoids as anticancer agents,” *Nutrients*, vol. 12, no. 2, p. 457, 2020.
- [80] A. Berim and D. R. Gang, “Methoxylated flavones: occurrence, importance, biosynthesis,” *Phytochemistry Reviews*, vol. 15, no. 3, pp. 363–390, 2016.
- [81] R. Mani and V. Natesan, “Chrysin: sources, beneficial pharmacological activities, and molecular mechanism of action,” *Phytochemistry*, vol. 145, pp. 187–196, 2018.

- [82] E. C. Lefort and J. Blay, "Apigenin and its impact on gastrointestinal cancers," *Molecular Nutrition & Food Research*, vol. 57, no. 1, pp. 126–144, 2013.
- [83] J. Cao, W. Chen, Y. Zhang, Y. Q. Zhang, and X. J. Zhao, "Content of selected flavonoids in 100 edible vegetables and fruits," *Food Science and Technology Research*, vol. 16, no. 5, pp. 395–402, 2010.
- [84] S. Pollastri and M. Tattini, "Flavonols: old compounds for old roles," *Annals of Botany*, vol. 108, no. 7, pp. 1225–1233, 2011.
- [85] V. M. Mounnissamy, V. Gopal, R. Gunasegaran, and A. Saraswathy, "Antiinflammatory activity of gossypetin isolated from *Hibiscus sabdariffa*," *Indian Journal of Heterocyclic Chemistry*, vol. 12, no. 1, pp. 85–86, 2002.
- [86] J. J. Alvarado-Sansininea, L. Sánchez-Sánchez, H. López-Muñoz et al., "Quercetagenin and patuletin: antiproliferative, necrotic and apoptotic activity in tumor cell lines," *Molecules*, vol. 23, no. 10, p. 2579, 2018.
- [87] A. T. Jan, M. R. Kamli, I. Murtaza, J. B. Singh, A. Ali, and Q. M. R. Haq, "Dietary flavonoid quercetin and associated health benefits an overview," *Food Reviews International*, vol. 26, no. 3, pp. 302–317, 2010.
- [88] F. Damin, A. Meinhart, L. Caldeirao et al., "Determination of rutin in fruits and vegetables in natura," *Journal of Food and Nutrition Research*, vol. 58, no. 4, pp. 328–338, 2019.
- [89] L. D. Antika and R. M. Dewi, "Pharmacological aspects of fisetin," *Asian Pacific Journal of Tropical Biomedicine*, vol. 11, no. 1, pp. 1–9, 2021.
- [90] C. Park, W. S. Lee, S. I. Go et al., "Morin, a flavonoid from moraceae, induces apoptosis by induction of BAD protein in human leukemic cells," *International Journal of Molecular Sciences*, vol. 16, no. 1, pp. 645–659, 2014.
- [91] M. Á. Martín and S. Ramos, "Impact of cocoa flavanols on human health," *Food and Chemical Toxicology*, vol. 151, Article ID 112121, 2021.
- [92] Y. Hatasa, M. Chikazawa, M. Furuhashi et al., "Oxidative deamination of serum albumins by (-)-epigallocatechin-3-O-gallate: a potential mechanism for the formation of innate antigens by antioxidants," *PLoS One*, vol. 11, Article ID e0153002, 2016.
- [93] L. Zhang, C. X. Wang, J. Wu et al., "Metabolic profiling of mice plasma, bile, urine and feces after oral administration of two licorice flavonones," *Journal of Ethnopharmacology*, vol. 257, Article ID 112892, 2020.
- [94] R. Pingili, S. Vemulapalli, S. S. Mullapudi, S. Nuthakki, S. Pendyala, and N. Kilaru, "Pharmacokinetic interaction study between flavanones (hesperetin, naringenin) and rasagiline mesylate in wistar rats," *Drug Development and Industrial Pharmacy*, vol. 42, no. 7, pp. 1110–1117, 2016.
- [95] A. Khan, M. Ikram, J. R. Hahm, and M. O. Kim, "Antioxidant and anti-inflammatory effects of citrusflavonoid hesperetin: special focus on neurological disorders," *Antioxidants*, vol. 9, no. 7, p. 609, 2020.
- [96] H. Xing, J. Yang, K. D. Ren et al., "Investigation on the metabolic characteristics of isobavachin in *Psoralea corylifolia* L. (Bu-gu-zhi) and its potential inhibition against human cytochrome P450s and UDP-glucuronosyltransferases," *Journal of Pharmacy and Pharmacology*, vol. 72, no. 12, pp. 1865–1878, 2020.
- [97] A. M. Karimov and E. K. Botirov, "Structural diversity and state of knowledge of flavonoids of the *Scutellaria* L. genus," *Russian Journal of Bioorganic Chemistry*, vol. 43, no. 7, pp. 691–711, 2017.
- [98] X. Chen, H. Z. Ge, S. S. Lei et al., "Dendrobium officinale six nostrum ameliorates urate under-excretion and protects renal dysfunction in lipid emulsion-induced hyperuricemic rats," *Biomedicine & Pharmacotherapy*, vol. 132, Article ID 110765, 2020.
- [99] L. Krizova, K. Dadakova, J. Kasparovska, and T. Kasparovsky, "Isoflavones," *Molecules*, vol. 24, p. 1076, 2019.
- [100] A. Sureda, A. Sanches Silva, D. I. Sanchez-Machado et al., "Hypotensive effects of genistein: from chemistry to medicine," *Chemico-Biological Interactions*, vol. 268, pp. 37–46, 2017.
- [101] T. Elsaman, M. S. Mohamed, E. M. Eltayib, A. E. Abdalla, and M. A. Mohamed, "Xanthone: a promising antimycobacterial scaffold," *Medicinal Chemistry*, vol. 17, no. 4, pp. 310–331, 2021.
- [102] A. J. N. Selles, M. Daglia, and L. Rastrelli, "The potential role of mangiferin in cancer treatment through its immunomodulatory, anti-angiogenic, apoptotic, and gene regulatory effects," *BioFactors*, vol. 42, no. 5, pp. 475–491, 2016.
- [103] E. Proshkina, S. Plyusnin, T. Babak et al., "Terpenoids as potential geroprotectors," *Antioxidants*, vol. 9, no. 6, p. 529, 2020.
- [104] L. G. Chen, Y. Y. Jiang, and Z. Yu, "Determining concentrations of loganin in plasma of rat by UPLC-MS/MS method: applications for a pharmacokinetic study," *Latin American Journal of Pharmacy*, vol. 36, no. 12, pp. 2374–2378.
- [105] H. M. Gao, J. Chen, P. Yu et al., "Pharmacokinetic comparisons of five different combinations of Zhi-zi-chi Decoction among rats: competing mechanisms between geniposide and genistein," *Chinese Journal of Analytical Chemistry*, vol. 49, no. 12, pp. 19–25, 2021.
- [106] H. Wu, M. Z. Zhou, G. Lu, Z. L. Yang, H. Ji, and Q. H. Hu, "Emodinol ameliorates urate nephropathy by regulating renal organic ion transporters and inhibiting immune inflammatory responses in rats," *Biomedicine & Pharmacotherapy*, vol. 96, pp. 727–735, 2017.
- [107] M. Loncar, M. Jakovljevic, D. Subaric et al., "Coumarins in food and methods of their determination," *Foods*, vol. 9, no. 5, p. 645, 2020.
- [108] K. Jafarnik, E. Halina, S. Ercisli, and A. Szopa, "Characteristics of bakuchiol-the compound with high biological activity and the main source of its acquisition-cullen *Corylifolium* (L.) Medik," *Natural Product Research*, vol. 35, no. 24, pp. 5828–5842, 2021.
- [109] S. Zschocke, J. H. Liu, H. Stuppner, and R. Bauer, "Comparative study of roots of *Angelica sinensis* and related umbelliferous drugs by thin layer chromatography, high-performance liquid chromatography, and liquid chromatography mass spectrometry," *Phytochemical Analysis*, vol. 9, no. 6, pp. 283–290, 1998.
- [110] M. M. Zhang, J. L. Wang, L. Zhu et al., "Zanthoxylum bungeanum maxim. (Rutaceae): a systematic review of its traditional uses, botany, phytochemistry, pharmacology, pharmacokinetics, and toxicology," *International Journal of Molecular Sciences*, vol. 18, no. 10, p. 2172, 2017.
- [111] I. Kostova and T. Iossifova, "Chemical components of *Fraxinus* species," *Fitoterapia*, vol. 78, no. 2, pp. 85–106, 2007.
- [112] M. N. Sun, M. J. Sun, and J. Y. Zhang, "Osthole: an overview of its sources, biological activities, and modification development," *Medicinal Chemistry Research*, vol. 30, no. 10, pp. 1767–1794, 2021.
- [113] T. Shen, X. N. Wang, and H. X. Lou, "Natural stilbenes: an overview," *Natural Product Reports*, vol. 26, no. 7, pp. 916–935, 2009.

- [114] D. Arora and S. Jaglan, "Therapeutic applications of resveratrol nanoformulations," *Environmental Chemistry Letters*, vol. 16, no. 1, pp. 35–41, 2018.
- [115] J. Komaikul, T. Kitisripanya, H. Tanaka, B. Sritularak, and W. Putalun, "Enhanced mulberroside a production from cell suspension and root cultures of morus alba using elicitation," *Natural Product Communications*, vol. 10, no. 7, 2015.
- [116] L. F. Zhou, S. H. Li, T. Zhang, W. M. Mu, and B. Jiang, "Properties of a novel polydatin-beta-D-glucosidase from aspergillus niger SK34.002 and its application in enzymatic preparation of resveratrol," *Journal of the Science of Food and Agriculture*, vol. 96, no. 7, pp. 2588–2595, 2016.
- [117] I. I. Koleva, T. A. van Beek, A. E. M. F. Soffers, B. Dusemund, and I. M. C. M. Rietjens, "Alkaloids in the human food chain—natural occurrence and possible adverse effects," *Molecular Nutrition & Food Research*, vol. 56, no. 1, pp. 30–52, 2012.
- [118] J. M. Cholewa, L. Guimaraes-Ferreira, and N. E. Zanchi, "Effects of betaine on performance and body composition: a review of recent findings and potential mechanisms," *Amino Acids*, vol. 46, no. 8, pp. 1785–1793, 2014.
- [119] M. Wu, J. Wang, and L. T. Liu, "Advance of studies on anti-atherosclerosis mechanism of berberine," *Chinese Journal of Integrative Medicine*, vol. 16, no. 2, pp. 188–192, 2010.
- [120] Y. Sun, G. B. Lenon, and A. W. H. Yang, "Phellodendri cortex: a phytochemical, pharmacological, and pharmacokinetic review," *Evidence-Based Complementary and Alternative Medicine*, vol. 2019, Article ID 7621929, 45 pages, 2019.
- [121] Y. A. Mostafa and S. D. Taylor, "Steroid derivatives as inhibitors of steroid sulfatase," *Journal of Steroid Biochemistry and Molecular Biology*, vol. 137, pp. 183–198, 2013.
- [122] H. Q. Zeng, L. J. Yang, X. B. Zhang, Y. Chen, and J. H. Cai, "Dioscin prevents LPS-induced acute lung injury through inhibiting the TLR4/MyD88 signaling pathway via upregulation of HSP70," *Molecular Medicine Reports*, vol. 17, no. 5, pp. 6752–6758, 2018.
- [123] Y. C. Wang, X. Li, H. Sun et al., "Biotransformation of steroidal saponins in sisal (agave sisalana perrine) to tigenin by a newly isolated strain from a karst area of guilin, China," *Biotechnology & Biotechnological Equipment*, vol. 28, no. 6, pp. 1024–1033, 2014.
- [124] J. W. Park, K. J. Min, D. E. Kim, and T. K. Kwon, "Withaferin a induces apoptosis through the generation of thiol oxidation in human head and neck cancer cells," *International Journal of Molecular Medicine*, vol. 35, no. 1, pp. 247–252, 2015.
- [125] A. Maalik, S. M. Bukhari, A. Zaidi, K. H. Shah, and F. A. Khan, "Chlorogenic acid: a pharmacologically potent molecule," *Acta Poloniae Pharmaceutica*, vol. 73, no. 4, pp. 851–854, 2016.
- [126] X. Q. Li, Y. H. Chen, X. X. Gao et al., "Antihyperuricemic effect of green alga ulva lactuca ulvan through regulating urate transporters," *Journal of Agricultural and Food Chemistry*, vol. 69, no. 38, pp. 11225–11235, 2021.
- [127] H. Zhang, J. Zhai, L. P. Zhang et al., "In Vitro inhibition of glyoxalase I by flavonoids: new insights from crystallographic analysis," *Current Topics in Medicinal Chemistry*, vol. 16, no. 4, pp. 460–466, 2015.
- [128] Z. K. Duan, Z. J. Zhang, S. H. Dong, Y. X. Wang, S. J. Song, and X. X. Huang, "Quassinoids: phytochemistry and anti-tumor prospect," *Phytochemistry*, vol. 187, Article ID 112769, 2021.
- [129] M. F. Wempe, B. Quade, P. Jutabha et al., "Human uric acid transporter 1 (hurati1): an inhibitor structure-activity relationship (sar) study," *Nucleosides, Nucleotides & Nucleic Acids*, vol. 30, no. 12, pp. 1312–1323, 2011.
- [130] A. G. Veiko, E. A. Lapshina, and I. B. Zavodnik, "Comparative analysis of molecular properties and reactions with oxidants for quercetin, catechin, and naringenin," *Molecular and Cellular Biochemistry*, vol. 476, no. 12, pp. 4287–4299, 2021.
- [131] X. J. Li, X. F. Shang, L. L. Liu, N. K. Xi, J. L. Zhang, and X. F. Xu, "Anion recognition based on phenolic hydroxyl group in competitive media," *Journal of Inclusion Phenomena and Macrocyclic Chemistry*, vol. 73, no. 1–4, pp. 185–192, 2012.