

### Research Article

## Investigating the Molecular Mechanism of Quercetin Protecting against Podocyte Injury to Attenuate Diabetic Nephropathy through Network Pharmacology, Microarray Data Analysis, and Molecular Docking

# Xiaoqin Ma<sup>(b)</sup>,<sup>1,2</sup> Chenxia Hao<sup>(b)</sup>,<sup>1,3</sup> Meixiang Yu<sup>(b)</sup>,<sup>1</sup> Zhaokang Zhang<sup>(b)</sup>,<sup>1</sup> Jingjing Huang<sup>(b)</sup>,<sup>1</sup> and Wanhua Yang<sup>(b)</sup>

<sup>1</sup>Department of Pharmacy, Ruijin Hospital, Shanghai Jiaotong University School of Medicine, Shanghai 200025, China

<sup>2</sup>Department of Pharmacy, Xi'an Children's Hospital, Xi'an, China

<sup>3</sup>Department of Pharmacy, Shanghai Children's Medical Center, Shanghai Jiao Tong University School of Medicine, Shanghai, China

Correspondence should be addressed to Jingjing Huang; huangjingjing0112@163.com and Wanhua Yang; yangwanhuaxy@163.com

Received 20 July 2021; Revised 3 March 2022; Accepted 29 April 2022; Published 16 May 2022

Academic Editor: Valeria Sülsen

Copyright © 2022 Xiaoqin Ma et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Quercetin (QUE), a health supplement, can improve renal function in diabetic nephropathy (DN) rats by ameliorating podocyte injury. Its clinical trial for renal insufficiency in advanced diabetes (NCT02848131) is currently underway. This study aimed to investigate the mechanism of QUE protecting against podocyte injury to attenuate DN through network pharmacology, microarray data analysis, and molecular docking. QUE-associated targets, genes related to both DN, and podocyte injury were obtained from different comprehensive databases and were intersected and analyzed to obtain mapping targets. Candidate targets were identified by constructing network of protein-protein interaction (PPI) of mapping targets and ranked to obtain key targets. The major pathways were obtained from Kyoto Encyclopedia of Genes and Genomes (KEGG) and Gene Ontology (GO) term enrichment analysis of candidate targets via ClueGO plug-in and R project software, respectively. Potential receptor-ligand interactions between QUE and key targets were evaluated via Autodocktools-1.5.6. 41. Candidate targets, of which three key targets (TNF, VEGFA, and AKT1), and the major AGE-RAGE signaling pathway in diabetic complications were ascertained and associated with QUE against podocyte injury in DN. Molecular docking models showed that QUE could closely bind to the key targets. This study revealed that QUE could protect against podocyte injury in DN through the following mechanisms: downregulating inflammatory cytokine of TNF, reducing VEGF-induced vascular permeability, inhibiting apoptosis by stimulating AKT1 phosphorylation, and suppressing the AGE-induced oxidative stress via the AGE-RAGE signaling pathway.

#### 1. Introduction

Podocyte injury is a critical event resulting in the eventual podocyte loss in the development and progression of diabetic nephropathy (DN), accounting for 40–45% of patients with diabetes mellitus [1–3]. Moreover, podocytes and podocyte-specific proteins are potential urinary markers to detect the early diagnosis of DN, and low podocyte density, correlating directly with the magnitude of proteinuria, is a

strongest predictor for progression of DN [4]. Furthermore, podocyte injury results in permanent alterations in the glomerular filtration barrier in DN [5, 6]. Podocytes are core cells of the glomerular filtration barrier and terminally differentiated parietal epithelial cells with a very limited proliferation ability [7].

Recently, podocyte injury has been regarded as a novel early mechanism involved in DN [8]. In the progress of DN, hyperglycemia (HG) induces the excessive accumulation of advanced glycation end products (AGEs) with reactive oxygen species (ROS), initiating podocyte injury accompanied with proteinuria and ultimately accelerating the development of DN [9, 10]. Podocytes are also targets of AGEs in diabetes by increasing AGE receptor (RAGE) expression [11]. The activated AGE-RAGE signaling pathway (AGEs binding to RAGE) increasing expressions of proinflammatory cytokine and oxidative stress is closely associated with podocyte injury and has been confirmed exactly one of the mechanisms of DN occurrence [12, 13]. However, the alleviation of podocyte injury in DN is mainly to control HG or proteinuria for the management of renal damage [3, 14], but lacks targeted agents. So, drugs targeting podocyte injury are urgently needed to treat DN and will be one of the most promising field of inquiry [13].

Quercetin (3,3',4',5,7-pentahydroxyflavone, QUE) belongs to natural flavonoids that are commonly defined as dietary antioxidants [15]. It has significant therapeutic effects on DN by reducing proteinuria which is a typical clinical manifestation mostly resulted from the podocyte injury [16, 17]. Moreover, it can reduce the oxidative stress, inflammatory responses and apoptosis involved in the progression of DN [18, 19]. Many clinical trials of QUE, including clinical research for renal insufficiency in advanced diabetes (clinicaltrials.gov ID NCT02848131) are (https://www.clinicaltrials.gov/) currently underway [20, 21]. In vitro experiments have confirmed that QUE reverses diabetes-induced podocyte injury by increasing the expression level of nephron and podocin in podocytes [20, 22, 23].

QUE is a phytochemical contained in many Chinese herbs such as *Astragalus membranaceus* (ASM) and *Salvia miltiorrhiza bunge* (SMB). ASM has been reported to have protective effects on podocyte injury and SMB can ameliorate diabetic vascular injury in streptozotocin-induced diabetic rats [24, 25]. However, the molecular mechanism of protects against podocyte injury in DN is lacking. Fortunately, network pharmacology can decipher the mechanism of drugs action with a holistic perspective, which breaks through the "one drug, one target" in the traditional drug discovery model and realizes the synergy of multiple targets [26]. Hence, this study aimed to investigate the mechanism of QUE protecting against podocyte injury to treat DN through network pharmacology [26], microarray data analysis, and molecular docking.

#### 2. Materials and Methods

The flowchart of this study design about the network pharmacology method used to clarify the key targets and the major pathway of QUE protecting against podocyte injury is shown in Figure 1, including six parts: searching QUE-associated targets, screening genes related to DN and podocyte injury, retrieving of mapping target interaction proteins, constructing protein-protein interaction (PPI) network, enrichment analysis, and molecular docking.

2.1. Searching QUE-Associated Targets. Targets of QUE were searched from the following three databases with the keyword

"quercetin." One is the Traditional Chinese Medicine Systems Pharmacology database [27] (TCMSP, https://lsp.nwu.edu. cn/) which focuses on the exploration of the targets from the HIT database, SysDT model, and targets validated by experiments [27]. Another is the SwissTargetPrediction database [28] (https://www.swisstargetprediction.ch) which estimates the most probable targets of QUE in view of 2D and 3D similarity between QUE and known activities in this database [29]. The third is the SymMap database [30] (https:// www.symmap.org/) which builds a large heterogeneous network by combining 19595 herbal ingredients and 4302 target genes related to symptoms [30]. After deleting repeated targets, all the unique targets obtained were considered to be regulated by QUE.

2.2. Obtaining Genes Related to DN and Podocyte Injury. Genes associated with DN were retrieved from five comprehensive databases, including the Online Mendelian Inheritance in Man database [31] (OMIM, https://www.omim. org/), DrugBank database [32] (https://www.drugbank.ca/), the Kyoto Encyclopedia of Genes and Genomes Pathway Database [33] (KEGG, https://www.kegg.jp/), and Therapeutic Target Database [34], (TTD, https://db.idrblab.net/ ttd/) with the keyword "diabetic nephropathy," as well as GeneCards database [35] (https://www.genecards.org/), with the keyword "[all] (diabetic nephropathy) and [all] (Homo sapiens)."

Genes related to podocyte injury were searched from four databases: OMIM and DigSee database [36] with the "Podocyte injury" keyword (https://210.107.182.61/ digseeOld/), GeneCards database with the keyword "[all] (podocyte injury) and [all] (Homo sapiens)," and Gene Expression Omnibus (GEO) database [37] with the keyword "(Podocyte injury) AND "Homo sapiens" [porgn: txid9606]." From the GEO database, a human gene expression data series (GSE51834) [38] titled "Indoxyl sulfate, a uremic toxin and aryl-hydrocarbon receptor ligand, mediates progressive glomerular disease by damaging podocytes" published in 2014, was selected to explore differential genes (DEGs) of podocyte injury. There was a series matrix file that included three podocyte injury samples and three control samples in this series. The DEGs were gathered by comparing these two types of samples with fold change (FC) of genes expression ( $|\log FC| \ge 1$ ) and false discovery rate (P < 0.05) using the Limma package [39] of the R project software.

2.3. Constructing Protein-Protein Interaction (PPI) of Mapping Targets and Identifying Candidate Targets. QUE-associated targets, podocyte injury-related genes, and DN-associated genes were subjected to intersection analysis to identify the mapping targets that were considered to be highly relevant to QUE protecting against podocyte injury in DN. The "protein-protein interaction (PPI)" topological network of the mapping targets was constructed using the STRING database [40] (https://string-db.org) using Cytoscape 3.71 [41]. The nodes of this network represent proteins, and the edges represent the interactions between the two



FIGURE 1: Flow chart of network pharmacology method used in this study.

proteins. The targets having interactions with a probabilistic association confidence a score  $\geq 0.4$  were identified candidate targets. The network topology parameters, including the "degree" of targets in the PPI network were analyzed using the Network Analyzer plug-in of Cytoscape. The top three targets with the highest "degree" values were defined as three key targets for QUE protecting against podocyte injury in DN.

2.4. Enrichment Analysis of GO Term and KEGG for Candidate Targets using ClueGO and R Project, Respectively. The candidate targets were imported into the ClueGO plug-in [42] of Cytoscape and R project for Gene Ontology (GO) term and KEGG enrichment analysis, respectively, to decipher the molecular mechanisms of QUE protecting against podocyte injury. The results gathered from above two enrichment software were further analyzed and compared, and the most reliable signaling pathway (the largest percentage or the lowest *P* value) was considered to be the major pathway for QUE protecting against podocyte injury in DN.

GO terms describe the biological function of genes through three semantic terms, namely, biological process (BP), cellular component (CC), and molecular function (MF) [43]. KEGG consists of artificially annotated metabolic pathways and defines the complex interrelationship between genes and metabolites [33]. R project software has been widely used in network pharmacology studies for screening of DEGs, enrichment, and annotation analysis [37, 44].

2.5. Docking QUE with Key Targets. The interaction between QUE (ligand) and key targets (receptors) were evaluated using Autodocktools-1.5.6 [45] and visualized through

PyMOL [46], being able to calculate and analyze the binding affinity and binding energy. The structure of QUE (as a mol2 file) and the targets (as a PDB file) were downloaded from PubChem (https://pubchem.ncbi.nlm.nih.gov/) and the Protein Data Bank database (PDB) (https://www.rcsb.org/ pages/contactus), respectively.

#### 3. Result

3.1. QUE-Associated Targets. A total of 355 targets of QUE (Supplementary Table 1) were obtained from TCMSP (148 targets), SwissTargetPrediction (99 targets), and SymMap (108 targets) databases, and 247 unique targets of QUE were gathered after deleting 108 repeated targets. And, there were six targets (AKT1, TOP1, PARP1, MMP9, MMP3, and MMP2) recorded in those three databases.

3.2. 3387 Genes Associated with DN and 816 Genes Related to Podocyte Injury. A total of 4895 human genes (Supplementary Table 2) associated with DN were identified from those five databases, and 3387 unique genes of DN were obtained after deleting 1508 duplications.

848 human genes (Supplementary Table 3) related to podocyte injury were identified, of which 316 were DEGs (130 upregulated genes, including TNFAIP6, TNFAIP3, and VEGFA and 186 downregulated genes, including TNFRSF19 and COL1A1) (Figure 2(a)), 532 of which were obtained from the OMIM (191 genes), GeneCards (339 genes), and DigSee (2 genes) databases. Totally 816 unique genes of podocyte injury were gathered after deleting 32 duplications.

3.3. 41 Candidate Targets from PPI Network Analysis of 42 Mapping Targets Related to QUE, DN, and Podocyte Injury.



FIGURE 2: (a) Volcano map of 316 differential genes of podocyte injury. The 130 upregulated genes are presented in red, whereas 186 downregulated genes are presented in green. (b) Venn diagram and PPI network showed the 42 mapping targets of QUE protects against podocytes injury in DN.

Through intersection analysis QUE-associated targets, DNassociated genes and podocyte injury-related genes, 42 mapping targets (Figure 2(b), Supplementary Table 4) were identified closely related to QUE protecting against podocyte injury in DN.

In total, 41 candidate targets (Figure 3(a), Supplementary Table 5), including 13 DEGs (2 downregulated and 11 upregulated genes (Figure 3(b)) were identified through the PPI network of 42 mapping targets. The PPI network contained 41 nodes, 340 edges with an average "degree" value (the mean number of connections per node) of 16.585. There were 21 candidate targets with a "degree" value  $\geq$ average "degree" (Table 1), and the top three targets TNF, VEGFA, and AKT1 ranked by degree were identified as the key targets of protecting against podocyte injury in DN.

3.4. Results of Enrichment Analysis about Candidate Targets. A total of five signaling pathways (Figure 4(a)) and 10 biological functions (P < 0.05, Figure 4(b)) involving in QUE protecting against podocyte injury were obtained via ClueGO, respectively. Detailed information of ClueGO enrichment results is listed in Table 2. 118 signaling pathways (P < 0.05, Supplementary Table 6) and 52 biological



FIGURE 3: (a) 41 candidate targets obtained from network of "protein-protein" interaction (PPI), including three key targets TNF (degree = 34), VEGFA (degree = 33), and AKT1 (degree = 31). The color of the nodes is shown in a gradient from to red to transparent according to the degree value. (b) 13 DEGs contained in 41 candidate targets, the red bars represents11 upregulated genes (log C 1), and the green bars represents 2 upregulated genes (log FC < -1).

Target	Uniprot ID	Description	Degree
TNF	P01375	Tumor necrosis factor	34
VEGFA	P15692	Vascular endothelial growth factor	33
AKT1	P31749	AKT serine/threonine kinase 1	31
TP53	P04637	Tumor protein p53	29
PTGS2	P35354	Prostaglandin G/H synthase 2	28
CXCL8	Q9UI36	C-X-C motif chemokine ligand 8	28
JUN	P05412	Jun proto-oncogene,	27
IL10	P22301	Interleukin-10	25
IL1B	P01584	Interleukin-1 beta	24
MAPK1	P10911	Mitogen-activated protein kinase 1	24
CCND1	P24864	Cyclin D1	22
HMOX1	P09601	Heme oxygenase 1	21
STAT1	P42224	Signal transducer and activator of transcription 1-alpha/beta	21
APP	P05067	Amyloid beta precursor protein	21
VEGF2	P35968	Vascular endothelial growth factor receptor 2	20
PTEN	P60484	Phosphatase and tensin homolog	20
CRP	P02741	C-reactive protein	20
SELE	Q5TI75	Selectin E	18
F2	P16930	Coagulation factor II, thrombin	17
HIF1A	P01892	Hypoxia inducible factor 1 subunit alpha	17
SOD1	P00441	Superoxide dismutase 1	17

TABLE 1: 21 candidate targets with a degree greater than average.

functions (P < 0.05, Supplementary Table 7) were obtained via R project, respectively. The top 20 signaling pathways with low P values are shown in Figure 5.

The top pathway obtained in ClueGO (accounting for 94.44%) and R project ( $P = 3.655 \times 10^{-19}$ ; count = 15) both was the AGE-RAGE signaling pathway in diabetic complications that was also identified as the major signaling pathway of QUE protecting against podocyte injury in DN. Oxidoreductase activity, antioxidant activity, and peroxidase activity from R project were consistent with the regulation of reactive oxygen metabolism (36.92%) from ClueGO [47],

and growth factors, cytokine receptors, and protein phosphatase 2A from R project were consistent with endothelial cell proliferation (26.92%) from ClueGO [48].

3.5. *Results of Molecular Docking*. The molecular docking analysis showed that QUE (ZINC3869685) could easily enter and bind to the key target TNF (2JG9), VEGFA (1MKK), and AKT1 (1UNR) with several interactions, hydrogen bonds, and amino acid residues, shown in Figure 6. QUE can form five H-bonds with GLY-201 (2.1), HIS-101 (2.5), SER-207



FIGURE 4: The enrichment results of KEGG (a) and GO terms analysis (b) for the 41 candidate targets via ClueGO. The AGE-RAGE signaling pathway is the most reliable pathway in ClueGO (94.44%), regulation of reactive oxygen metabolism (36.92%) and endothelial cell proliferation (26.92%) are top two reliable biological functions.

(2.8), and HIS-101 (2.5 and 1.8) of TNF, eight H-bonds with LEU-66 (2.8), CYS-26 (2.6 and 2.4), GLU-64 (2.4), PHE-47 (2.4), SER-50 (2.5), and SER-24 (2.2 and 3.5) of VEGFA. And, the binding energy of QUE and TNF, VEGFA and AKT1 were -6.35 kJ/mol, -6.75 kJ/mol, and -5.36 kJ/mol, respectively.

#### 4. Discussion

The present study shows that QUE would protect against podocyte injury in DN mainly by regulating the major AGE-RAGE signaling pathway and three key targets: TNF mediating the proinflammatory, VEGF promoting vascular permeability and proliferation, and AKT1 participating in apoptosis. Furthermore, QUE, having the five hydroxy groups (placed at the 3-, 3'-, 4'-, 5- and 7-positions), should have suitable binding sites with three key targets and interacts with amino acid residues of targets through multiple hydrogen bonding and Van der Waals using molecular docking analysis.

QUE regulates the oxidative stress-associated AGE-RAGE signaling pathway to protect against podocyte injury in DN. It is known that the binding of AGEs to the receptor RAGE can induce oxidative stress and inflammation, eliciting podocyte injuries [49, 50]. Encouragingly, Li et al. validated that QUE can reduce the production of AGEs by

ID	Description	Percentage	Count
А			
KEGG: 04933	AGE-RAGE signaling pathway in diabetic complications	94.44%	36
KEGG: 05418	Fluid shear stress and atherosclerosis	1.39%	15
KEGG: 04610	Complement and coagulation cascades	1.39%	4
KEGG: 05014	Amyotrophic lateral sclerosis (ALS)	1.39%	3
KEGG: 04923	Regulation of lipolysis in adipocytes	1.39%	3
В			
GO: 1903409	Reactive oxygen species biosynthetic process	36.92%	18
GO: 0001937	Positive regulation of endothelial cell proliferation	26.92%	24
GO: 0048662	Positive regulation of smooth muscle cell proliferation	12.31%	20
GO: 0001936	Regulation of endothelial cell proliferation	8.84%	17
GO: 2000377	Regulation of reactive oxygen species metabolic process	5.34%	31
GO: 0014074	Response to purine-containing compound	5.34%	17
GO: 0045766	Positive regulation of angiogenesis	2%	17
GO: 0007589	Body fluid secretion	2%	6
GO: 0042730	Fibrinolysis	2%	5
GO: 0001937	Negative regulation of endothelial cell proliferation	2%	3

TABLE 2: KEGG (A) and GO (B) term enrichment results from ClueGO.

trapping 50.5% of glyoxal and 80.1% of methylglyoxal which are the crucial reactive dicarbonyl precursors of AGEs [51]. Moreover, QUE decreases the expression of RAGE [52] and increases the expression of superoxide dismutase (SOD) to suppress oxidative stress accelerated by the activated AGE-RAGE pathway, protecting the cell from injury [53]. Additionally, QUE is an antioxidant, it can not only increase the expression of podocyte slit diaphragms and sensitive markers of podocyte nephrin and podocin to the maintenance of the skeletal structure and function of podocytes [23, 54] but also lower the kidney hypertrophy index (KI), blood urea nitrogen (BUN), and blood creatinine (Scr) to improve kidney function in diabetic rats [52]. Therefore, it can be inferred that QUE can prevent podocytes from the stimulation of oxidative stress by inhibiting HG which induced excessive accumulation of AGEs, lowering ROS synthesis. Furthermore, accumulating evidence has also shown there is a close relation between the AGE-RAGE signaling pathway and other complications of diabetes such as diabetic peripheral neuropathy and diabetic retinopathy [55-57]. Therefore, QUE might also have preventive effects on complications of diabetes.

QUE inhibits key targets of TNF mediating the proinflammatory and regulates VEGF promoting vascular permeability and AKT mediating apoptosis to protect podocytes from injury in DN. Experiments in the streptozocin-induced diabetic rat have demonstrated that the inflammatory cytokine of TNF- $\alpha$ , a member of the TNF receptors superfamily, is an intermediate factor for excessive ROS-induced podocyte injury and apoptosis [58-60]. Moreover, TNF- $\alpha$ plays a predictive role in DN, attributing to its involvement in the onset and progression of DN [61]. Encouragingly, QUE has been proven to decrease the renal TNF- $\alpha$  and ROS synthesis [62] induced by high homocysteine (Hcy) [63] which is an independent risk factor for DN [64]. High Hcy can also directly cause podocyte injury, with subsequent progression of glomerular permeability induced by oxidative stress [64, 65], and can affect the function of renal endothelium and mesangial cells during the progression of DN

[66, 67]. While, as DN progresses further, abnormal elevation of Hcy directly damages vascular endothelial cells and aggravates microalbuminuria and ultimately forms a vicious circle between DN and Hcy [65, 68]. Interestingly, QUE can also reduce the level of Hcy and increase the level of the Hcy's metabolite, taurine, an antioxidant that has been demonstrated to improve glomerular sclerosis and attenuate the progression of DN in mice [69, 70]. Metabolomic studies have consistently shown that QUE increases the level of taurine in mice serum and urine [70, 71]. Treatment with taurine significantly downregulates the protein levels of podocyte homeostasis regulator and consequently the reduction of glucose-induced podocytes injuries in DN mice model [72]. Furthermore, high Hcy-induced endothelial cell apoptosis is commonly associated with increased VEGF [73]. VEGF is the important mediator in endothelial cell proliferation and glomerular mesangial proliferation at the endstage of DN [74]. It is regulated by candidate target ERK1/2 (also known as MAPK1, degree = 24) and key target AKT (degree = 31), and the excessive production of VEGF subtype A (VEGFA), resulting from the interaction of AGEs and RAGE, is a novel risk factor in the pathogenesis of the endstage renal disease [55, 75]. But excessive inhibition of VEGF causes glomerular injury with prominent podocyte injury [76, 77]. So, it would be speculated that the therapeutic index of VEGF for podocyte injury is narrow, which is in line with the statements of Oe et al. [78]. Interestingly, QUE can moderately regulate the expressions of VEGFA and alleviate podocyte injury and kidney function in diabetic rats [23]. Hence, QUE is a new appropriate product that targeted VEGFA to ameliorate podocyte injury. The phosphorylation of another key target AKT can significantly prevent from podocyte apoptosis, foot process shrinkage, and renin loss [78]. However, the levels of phospho-Akt are downregulated by long-term HG, causing the increased activation of p38 MAPK and renal proximal tubule cell apoptosis [79]. Noteworthily, QUE can increase the phosphorylation of AKT to promote the synthesis of liver glycogen with lowering blood sugar and regulate the downstream proteins of



FIGURE 5: Enrichment results of KEGG (a) and ClueGO (b) from R project. The AGE-RAGE signaling pathway ( $P = 3.655 \times 10^{-19}$ ; count = 15) is also the most reliable pathway R project.



FIGURE 6: 3D molecular binding model of QUE to key targets TNF, VEGFA, and AKT1. Three key targets are represented as light blue flat strips, and amino acid residues of key targets are represented as colored sticks and QUE is represented as the yellow stick. The yellow dashed lines demarcate hydrogen bonds, and the interaction distances are indicated next to the bonds.



FIGURE 7: Effects of QUE protecting against podocytes injury in DN via key targets TNF, VEGFA, and AKT1 in AGE-RAGE signaling pathways.

AKT to facilitate lipid metabolism [80]. Thus, it can be assumed that QUE promotes glycogen synthesis via AKT phosphorylation to prevent podocytes from injury. In addition, QUE inhibits the expression of candidate target TP53 being a key regulator of p53 apoptotic signaling pathways which are involved in podocyte senescence and apoptosis [80, 81], and the downstream signaling pathways of TP53, such as NF-kB signaling pathways, that participate in HGinduced podocyte injury [82, 83].

Moreover, this study also indicates that QUE protects against podocytes injury having relation with autophagy from enrichment analysis (P = 0.0048 from the R project). Autophagy, which can accelerate the metabolism of ROS induced by HG, significantly accelerates the metabolism of ROS and inhibits the activation of VEGF, showing its importance to maintain the postmitotic podocytes cells [84, 85]. More and more researches prove that QUE can suppress ROS synthesis through induction of autophagy to cure liver fibrosis and CVD [86–88]. Additionally, QUE can significantly upregulate autophagy by suppressing oxidative stress and downregulating TNF- $\alpha$  and AKT and ameliorating doxorubicin-induced podocyte injury in rats [89, 90]. Thus, it can be hypothesized that QUE may act on autophagy associated with the reduction of ROS to participate in the protection of podocytes injury [91, 92].

Although it has been confirmed in different experimental models that QUE regulates key targets at TNF, VEGFA, and AKT1, as well as the AGE-RAGE signaling pathway to protect against podocyte injury. In this article, the findings suggest that the multipronged therapeutic effect of QUE on podocyte injury, attributing to its synergistic effects on these multiple targets. However, further experimental studies will be needed to verify it.

#### 5. Conclusion

This study reveals that QUE can reduce the inflammatory response (TNF and IL6), inhibit endothelial cell proliferation (VEGFA) and apoptosis of podocytes (AKT1 and TP53), and suppress the AGE-induced oxidative stress by regulating the AGE-RAGE signaling pathway activated by HG to protect against podocytes injury in DN (Figure 7). This study provides a scientific basis for developing QUE as a potential natural medicine for the treatment of DN.

#### **Data Availability**

The data of this research is obtained through authoritative online databases and software analysis and can be acquired from the Supplementary Materials uploaded with this article.

#### **Conflicts of Interest**

The authors declare that they have no conflicts of interest.

#### **Authors' Contributions**

Chenxia Hao contributed to this work.

#### Acknowledgments

This project was supported by the key scientific research projects of the Science and Technology Commission of Shanghai (no. 17401901100) and Science and Technology Innovation Project of Traditional Chinese Medicine of Shanghai Municipal Commission of Health (no. ZYKC201701007).

#### **Supplementary Materials**

Supplementary Table 1: 355 targets of QUE. Supplementary Table 2: 4895 human genes related to DN. Supplementary Table 3: 848 human genes associated with podocyte injury. Supplementary Table 4: 42 mapping targets. Supplementary Table 5: specific analytical parameters of 41 candidate targets. Supplementary Table 6: 118 signaling pathways from R project. Supplementary Table 7: 52 biological functions from R project. (*Supplementary Materials*)

#### References

- Y. Li, X. Sui, X. Hu, and Z. Hu, "Overexpression of KLF5 inhibits puromycin induced apoptosis of podocytes," *Molecular Medicine Reports*, vol. 18, pp. 3843–3849, 2018.
- [2] D. Zhou, M. Zhou, Z. Wang et al., "PGRN acts as a novel regulator of mitochondrial homeostasis by facilitating mitophagy and mitochondrial biogenesis to prevent podocyte injury in diabetic nephropathy," *Cell Death & Disease*, vol. 10, no. 7, p. 524, 2019.
- [3] L. Gnudi, R. J. M. Coward, and D. A. Long, "Diabetic nephropathy: perspective on novel molecular mechanisms," *Trends in Endocrinology and Metabolism*, vol. 27, no. 11, pp. 820–830, 2016.
- [4] I. Kravets and S. K. Mallipattu, "The role of podocytes and podocyte-associated biomarkers in diagnosis and treatment of diabetic kidney disease," *Journal of the Endocrine Society*, vol. 4, Article ID bvaa029, 2020.
- [5] H. Yang, T. Xie, D. Li et al., "Tim-3 aggravates podocyte injury in diabetic nephropathy by promoting macrophage activation via the NF- $\kappa$ B/TNF- $\alpha$  pathway," *Molecular Metabolism*, vol. 23, pp. 24–36, 2019.
- [6] C. Hu, L. Sun, L. Xiao et al., "Insights into the mechanisms involved in the expression and regulation of extracellular

matrix proteins in diabetic nephropathy," *Current Medicinal Chemistry*, vol. 22, no. 24, pp. 2858–2870, 2015.

- [7] J. B. Kopp, H. J. Anders, K. Susztak et al., "Podocytopathies," *Nature Reviews Disease Primers*, vol. 6, no. 1, p. 68, 2020.
- [8] Y. Cao, Y. Hao, H. Li et al., "Role of endoplasmic reticulum stress in apoptosis of differentiated mouse podocytes induced by high glucose," *International Journal of Molecular Medicine*, vol. 33, no. 4, pp. 809–816, 2014.
- [9] X. Y. Zhang, Y. Mi, and C. l. Wang, "Podocyte injury and diabetic nephropathy," *Chinese Journal of Nephrology, Dial*ysis & Transplantation, vol. 28, pp. 161–165, 2019.
- [10] X. Zhao, F. Li, W. Sun et al., "Extracts of magnolia speciesinduced prevention of diabetic complications: a brief review," *International Journal of Molecular Sciences*, vol. 17, no. 10, p. 1629, 2016.
- [11] T. Bondeva and G. Wolf, "Role of neuropilin-1 in diabetic nephropathy," *Journal of Clinical Medicine*, vol. 4, no. 6, pp. 1293–1311, 2015.
- [12] S. Zhang, D. J. Wang, N. Xue et al., "Nicousamide protects kidney podocyte by inhibiting the TGF $\beta$  receptor II phosphorylation and AGE-RAGE signaling," *American Journal of Translational Research*, vol. 9, pp. 115–125, 2017.
- [13] L. Chen, "Study on the mechanism of diabetic podocyte injury and the treatment of Chinese and Western medicines," *Diabetes New World*, vol. 34, pp. 22-23, 2014.
- [14] H. Y. Chen, J. Q. Chen, J. Y. Li et al., "Deep learning and random forest approach for finding the optimal traditional Chinese medicine formula for treatment of alzheimer's disease," *Journal of Chemical Information and Modeling*, vol. 59, no. 4, pp. 1605–1623, 2019.
- [15] B. K. Ghimire, J. W. Seo, C. Y. Yu, S. H. Kim, and I. M. Chung, "Comparative study on seed characteristics, antioxidant activity, and total phenolic and flavonoid contents in accessions of sorghum bicolor (L.) moench," *Molecules*, vol. 26, no. 13, p. 3964, 2021.
- [16] I. B. S. Gomes, M. L. Porto, M. C. L. F. S. Santos et al., "Renoprotective, anti-oxidative and anti-apoptotic effects of oral low-dose quercetin in the C57BL/6J model of diabetic nephropathy," *Lipids in Health and Disease*, vol. 13, p. 184, 2014.
- [17] Q. Zhang, Q. Ye, X. Huang et al., "Revealing active components, action targets and molecular mechanism of Gandi capsule for treating diabetic nephropathy based on network pharmacology strategy," *BMC Complementary Medicine and Therapies*, vol. 20, no. 1, p. 362, 2020.
- [18] J. Granados-Pineda, N. Uribe-Uribe, P. Garcia-Lopez, M. Ramos-Godinez, J. Rivero-Cruz, and J. Perez-Rojas, "Effect of pinocembrin isolated from Mexican Brown propolis on diabetic nephropathy," *Molecules*, vol. 23, no. 4, p. 852, 2018.
- [19] T. Ding, S. Wang, X. Zhang et al., "Kidney protection effects of dihydroquercetin on diabetic nephropathy through suppressing ROS and NLRP3 inflammasome," *Phytomedicine*, vol. 41, pp. 45–53, 2018.
- [20] A. K. Palmer, M. Xu, Y. Zhu et al., "Targeting senescent cells alleviates obesity-induced metabolic dysfunction," *Aging Cell*, vol. 18, Article ID e12950, 2019.
- [21] L. J. Hickson, L. G. P. Langhi Prata, S. A. Bobart et al., "Senolytics decrease senescent cells in humans: preliminary report from a clinical trial of Dasatinib plus Quercetin in individuals with diabetic kidney disease," *EBioMedicine*, vol. 47, pp. 446–456, 2019.
- [22] C. Gupta and D. Prakash, "Nutraceuticals for geriatrics," *Journal of Traditional and Complementary Medicine*, vol. 5, no. 1, pp. 5–14, 2015.

- [23] Y. I. Jin, Z. H. Qu, P. P. Yang, B. Y. Yuan, and L. B. Chen, "Effects of quercetin on the expression of nephrin and podocin in kidney pods of diabetic rats," *Chinese Journal of Laboratory Diagnosis JST*, vol. 23, pp. 519–522, 2019.
- [24] W. Zhao, Y. Yuan, H. Zhao, Y. Han, and X. Chen, "Aqueous extract of Salvia miltiorrhiza Bunge-Radix Puerariae herb pair ameliorates diabetic vascular injury by inhibiting oxidative stress in streptozotocin-induced diabetic rats," *Food and Chemical Toxicology*, vol. 129, pp. 97–107, 2019.
- [25] R. Zhai, G. Jian, T. Chen et al., "Astragalus membranaceus and panax notoginseng, the novel renoprotective compound, synergistically protect against podocyte injury in streptozotocin-induced diabetic rats," *Journal of Diabetes Research*, vol. 2019, Article ID 1602892, 14 pages, 2019.
- [26] R. Zhang, X. Zhu, H. Bai, and K. Ning, "Network pharmacology databases for traditional Chinese medicine: review and assessment," *Frontiers in Pharmacology*, vol. 10, p. 123, 2019.
- [27] J. Ru, P. Li, J. Wang et al., "TCMSP: a database of systems pharmacology for drug discovery from herbal medicines," *Journal of Cheminformatics*, vol. 6, no. 1, p. 13, 2014.
- [28] A. Daina, O. Michielin, and V. Zoete, "Swiss target prediction: updated data and new features for efficient prediction of protein targets of small molecules," *Nucleic Acids Research*, vol. 47, no. W1, pp. W357–W364, 2019.
- [29] D. Gfeller, O. Michielin, and V. Zoete, "Shaping the interaction landscape of bioactive molecules," *Bioinformatics*, vol. 29, no. 23, pp. 3073–3079, 2013.
- [30] Y. Wu, F. Zhang, K. Yang et al., "SymMap: an integrative database of traditional Chinese medicine enhanced by symptom mapping," *Nucleic Acids Research*, vol. 47, no. D1, pp. D1110–D1117, 2019.
- [31] J. S. Amberger and A. Hamosh, "Searching online mendelian inheritance in man (OMIM): a knowledgebase of human genes and genetic phenotypes," *Current Protocols in Bioinformatics*, vol. 58, pp. 1 2 1-2 12, 2017.
- [32] D. S. Wishart, Y. D. Feunang, A. C. Guo et al., "DrugBank 5.0: a major update to the DrugBank database for 2018," *Nucleic Acids Research*, vol. 46, no. D1, pp. D1074–D1082, 2018.
- [33] M. Kanehisa, M. Furumichi, M. Tanabe, Y. Sato, and K. Morishima, "KEGG: new perspectives on genomes, pathways, diseases and drugs," *Nucleic Acids Research*, vol. 45, no. D1, pp. D353–D361, 2017.
- [34] Y. H. Li, C. Y. Yu, X. X. Li et al., "Therapeutic target database update 2018: enriched resource for facilitating bench-to-clinic research of targeted therapeutics," *Nucleic Acids Research*, vol. 46, no. D1, pp. D1121–D1127, 2018.
- [35] G. Stelzer, N. Rosen, I. Plaschkes et al., "The GeneCards suite: from gene data mining to disease genome sequence analyses," *Current Protocols in Bioinformatics*, vol. 54, pp. 1–13, 2016.
- [36] J. Kim, S. So, H. J. Lee, and J. C. Park, "DigSee: disease gene search engine with evidence sentences (version cancer)," *Nucleic Acids Research*, vol. 41, no. W1, pp. W510–W517, 2013.
- [37] E. Clough and T. Barrett, "The gene expression Omnibus database," *Methods in Molecular Biology*, vol. 1418, pp. 93– 110, 2016.
- [38] O. Ichii, S. Otsuka-Kanazawa, T. Nakamura et al., "Podocyte injury caused by indoxyl sulfate, a uremic toxin and arylhydrocarbon receptor ligand," *PLoS One*, vol. 9, no. 9, Article ID e108448, 2014.
- [39] M. E. Ritchie, B. Phipson, D. Wu et al., "Limma powers differential expression analyses for RNA-sequencing and microarray studies," *Nucleic Acids Research*, vol. 43, no. 7, p. e47, 2015.

- [40] D. Szklarczyk, J. H. Morris, H. Cook et al., "The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible," *Nucleic Acids Research*, vol. 45, no. D1, pp. D362–D368, 2017.
- [41] T. Van Parys, I. Melckenbeeck, M. Houbraken et al., "A Cytoscape app for motif enumeration with ISMAGS," *Bioinformatics*, vol. 33, pp. 461–463, 2017.
- [42] G. Bindea, B. Mlecnik, H. Hackl et al., "ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks," *Bioinformatics*, vol. 25, no. 8, pp. 1091–1093, 2009.
- [43] M. Ashburner, C. A. Ball, J. A. Blake et al., "Gene ontology: tool for the unification of biology. The Gene Ontology Consortium," *Nature Genetics*, vol. 25, pp. 25–29, 2000.
- [44] X. Ma, M. Yu, C. Hao, and W. Yang, "Identifying synergistic mechanisms of multiple ingredients in shuangbai tablets against proteinuria by virtual screening and a network pharmacology approach," *Evidence-based Complementary* and Alternative Medicine, vol. 2020, Article ID 1027271, 15 pages, 2020.
- [45] Z. Bikadi and E. Hazai, "Application of the PM6 semi-empirical method to modeling proteins enhances docking accuracy of AutoDock," *Journal of Cheminformatics*, vol. 1, p. 15, 2009.
- [46] D. Seeliger and B. L. de Groot, "Ligand docking and binding site analysis with PyMOL and Autodock/Vina," *Journal of Computer-Aided Molecular Design*, vol. 24, no. 5, pp. 417–422, 2010.
- [47] H. Galehdari, S. Z. Azarshin, M. Bijanzadeh, and M. Shafiei, "Polymorphism studies on microRNA targetome of thalassemia," *Bioinformation*, vol. 14, no. 5, pp. 252–258, 2018.
- [48] L. Janet, J. K. G. Douglas, A. V. Moses, B. J. Dezube, and L. Pantanowitz, "Kaposi sarcoma pathogenesis: a triad of viral infection, oncogenesis and chronic inflammation," *Translational Biomedicine*, vol. 1, p. 172, 2012.
- [49] R. Frédérick, L. Pochet, P. De Tullio, and F. Dufrasne, "31ièmes journées franco-belges de pharmacochimie: meeting report," *Pharmaceuticals*, vol. 10, no. 4, 2017.
- [50] Y. Yamamoto and H. Yamamoto, "Interaction of receptor for advanced glycation end products with advanced oxidation protein products induces podocyte injury," *Kidney International*, vol. 82, no. 7, pp. 733–735, 2012.
- [51] X. Li, T. Zheng, S. Sang, and L. Lv, "Quercetin inhibits advanced glycation end product formation by trapping methylglyoxal and glyoxal," *Journal of Agricultural and Food Chemistry*, vol. 62, no. 50, pp. 12152–12158, 2014.
- [52] L. X. Tang, K. M. Zhu, D. P. Li, and S. M. Gu, "Effects of quercetin linosomes on the formation of advanced glycation end products (AGEs) and receptor for advanced glycation end products (RAGE) in kidney of diabetic rats," *Tianjin Medical Journal*, vol. 44, pp. 71–74+132, 2016.
- [53] J. I. Wang, Z. C. Yang, C. Wang, F. Sun, and X. H. Xu, "The hypoglycemic effect and mechanism of quercetin for diabetic rats," *Jining Medical University*, vol. 42, pp. 135–138, 2018.
- [54] W. Qin, Z. Xu, Y. Lu et al., "Mixed organic solvents induce renal injury in rats," *PLoS One*, vol. 7, no. 9, Article ID e45873, 2012.
- [55] S. I Yamagishi, T. Matsui, K. Nakamura et al., "Olmesartan blocks advanced glycation end products (AGEs)-induced angiogenesis in vitro by suppressing receptor for AGEs (RAGE) expression," *Microvascular Research*, vol. 75, no. 1, pp. 130–134, 2008.
- [56] R. Ramasamy, S. F. Yan, and A. M. Schmidt, "Receptor for AGE (RAGE): signaling mechanisms in the pathogenesis of

diabetes and its complications," *Annals of the New York Academy of Sciences*, vol. 1243, no. 1, pp. 88–102, 2011.

- [57] G. J. Shi, Y. Li, Q. H. Cao et al., "In vitro and in vivo evidence that quercetin protects against diabetes and its complications: a systematic review of the literature," *Biomedicine & Pharmacotherapy*, vol. 109, pp. 1085–1099, 2019.
- [58] M. X. Xu, Y. T. Qin, C. X. Ge et al., "Activated iRhom2 drives prolonged PM2.5 exposure-triggered renal injury in Nrf2defective mice," *Nanotoxicology*, vol. 12, no. 9, pp. 1045–1067, 2018.
- [59] T. A. Mudyanadzo, "Endothelial progenitor cells and cardiovascular correlates," *Cureus*, vol. 10, Article ID e3342, 2018.
- [60] J. Li, B. Liu, H. Xue, Q. Q. Zhou, and L. Peng, "miR-217 is a useful diagnostic biomarker and regulates human podocyte cells apoptosis via targeting TNFSF11 in membranous nephropathy," *BioMed Research International*, vol. 2017, Article ID 2168767, 9 pages, 2017.
- [61] S. N. Uwaezuoke, "The role of novel biomarkers in predicting diabetic nephropathy: a review," *International Journal of Nephrology and Renovascular Disease*, vol. 10, pp. 221–231, 2017.
- [62] H. Y. Wang, J. G. Zhao, Z. G. Wei, and Y.-Q. Zhang, "The renal protection of flavonoid-rich ethanolic extract from silkworm green cocoon involves in inhibiting TNF-α-p38 MAP kinase signalling pathway in type 2 diabetic mice," *Biomedicine & Pharmacotherapy*, vol. 118, Article ID 109379, 2019.
- [63] X. Lin, X. Meng, and Z. Song, "Homocysteine and psoriasis," *Bioscience Reports*, vol. 39, no. 11, 2019.
- [64] R. Zhang, X. Wang, Q. Gao et al., "Taurine supplementation reverses diabetes-induced podocytes injury via modulation of the CSE/TRPC6 axis and improvement of mitochondrial function," *Nephron*, vol. 144, no. 2, pp. 84–95, 2020.
- [65] J. Zhang, J. Li, S. Chen et al., "Modification of platelet count on the association between homocysteine and blood pressure: a moderation analysis in Chinese hypertensive patients," *International Journal of Hypertension*, vol. 2020, Article ID 5983574, 8 pages, 2020.
- [66] J. Wu, S. Yue, J. Geng et al., "Relationship between diabetic retinopathy and subclinical hypothyroidism: a meta-analysis," *Scientific Reports*, vol. 5, no. 1, Article ID 12212, 2015.
- [67] E. Jahangir, J. A. Vita, D. Handy et al., "The effect of L-arginine and creatine on vascular function and homocysteine metabolism," *Vascular Medicine*, vol. 14, no. 3, pp. 239–248, 2009.
- [68] H. Guan, M. D. Xia, M. Wang, Y. J. Guan, and X. C. Lyu, "Methylenetetrahydrofolate reductase genetic polymorphism and the risk of diabetic nephropathy in type 2 diabetic patients," *Medicine (Baltimore)*, vol. 99, no. 35, Article ID e21558, 2020.
- [69] A. B. Oyenihi, A. O. Ayeleso, E. Mukwevho, and B. Masola, "Antioxidant strategies in the management of diabetic neuropathy," *BioMed Research International*, vol. 2015, Article ID 515042, 15 pages, 2015.
- [70] L. Zhang, M. Dong, X. Guangyong, T. Yuan, H. Tang, and Y. Wang, "Metabolomics reveals that dietary ferulic acid and quercetin modulate metabolic homeostasis in rats," *Journal of Agricultural and Food Chemistry*, vol. 66, no. 7, pp. 1723–1731, 2018.
- [71] C. Xu, Quercetin and Isorhamnetin from Metabolism Distribution in Mice Plasma and Tissues after Long Term Administration, Nanchang University, Nanchang, China, 2012.

- [72] E. E. Ngowi, M. Sarfraz, A. Afzal et al., "Roles of hydrogen sulfide donors in common kidney diseases," *Frontiers in Pharmacology*, vol. 11, Article ID 564281, 2020.
- [73] A. Tawfik, R. Mohamed, N. M. Elsherbiny, M. DeAngelis, M. Bartoli, and M. Al-Shabrawey, "Homocysteine: a potential biomarker for diabetic retinopathy," *Journal of Clinical Medicine*, vol. 8, no. 1, p. 121, 2019.
- [74] Y. Alam-Faruque, D. P. Hill, E. C. Dimmer et al., "Representing kidney development using the gene ontology," *PLoS One*, vol. 9, no. 6, Article ID e99864, 2014.
- [75] K. Prasad, I. Dhar, Q. Zhou, H. Elmoselhi, M. Shoker, and A. Shoker, "AGEs/sRAGE, a novel risk factor in the pathogenesis of end-stage renal disease," *Molecular and Cellular Biochemistry*, vol. 423, no. 1-2, pp. 105–114, 2016.
- [76] V. Eremina and S. E. Quaggin, "The role of VEGF-A in glomerular development and function," *Current Opinion in Nephrology and Hypertension*, vol. 13, no. 1, pp. 9–15, 2004.
- [77] H. Xiao, W. Shi, S. Liu et al., "1,25-Dihydroxyvitamin D(3) prevents puromycin aminonucleoside-induced apoptosis of glomerular podocytes by activating the phosphatidylinositol 3-kinase/Akt-signaling pathway," *American Journal of Nephrology*, vol. 30, no. 1, pp. 34–43, 2009.
- [78] Y. Oe, T. Fushima, E. Sato et al., "Protease-activated receptor 2 protects against VEGF inhibitor-induced glomerular endothelial and podocyte injury," *Scientific Reports*, vol. 9, no. 1, p. 2986, 2019.
- [79] J. Wang and G. Yu, "A systems biology approach to characterize biomarkers for blood stasis syndrome of unstable Angina patients by integrating MicroRNA and messenger RNA expression profiling," *Evidence-based Complementary* and Alternative Medicine, vol. 2013, Article ID 510208, 21 pages, 2013.
- [80] J. Peng, Q. Li, K. Li et al., "Quercetin improves glucose and lipid metabolism of diabetic rats: involvement of akt signaling and SIRT1," *Journal of Diabetes Research*, vol. 2017, Article ID 3417306, 10 pages, 2017.
- [81] M. D. M. Leiserson, D. Blokh, R. Sharan, and B. J. Raphael, "Simultaneous identification of multiple driver pathways in cancer," *PLoS Computational Biology*, vol. 9, no. 5, Article ID e1003054, 2013.
- [82] Y. Chen, Q. Liu, Z. Shan et al., "The protective effect and mechanism of catalpol on high glucose-induced podocyte injury," *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, p. 244, 2019.
- [83] S. Roy, S. Banerjee, and T. Chakraborty, "Vanadium quercetin complex attenuates mammary cancer by regulating the P53, Akt/mTOR pathway and downregulates cellular proliferation correlated with increased apoptotic events," *Biometals*, vol. 31, no. 4, pp. 647–671, 2018.
- [84] Y. Ding and M. E. Choi, "Autophagy in diabetic nephropathy," *Journal of Endocrinology*, vol. 224, no. 1, pp. R15-R30, 2015.
- [85] W. Miaomiao, L. Chunhua, Z. Xiaochen, C. Xiaoniao, L. Hongli, and Y. Zhuo, "Autophagy is involved in regulating vegf during high-glucose-induced podocyte injury," *Molecular BioSystems*, vol. 12, no. 7, pp. 2202–2212, 2016.
- [86] H. Cao, Q. Jia, L. Yan, C. Chen, S. Xing, and D. Shen, "Quercetin suppresses the progression of atherosclerosis by regulating MST1-mediated autophagy in ox-LDL-induced RAW264.7 macrophage foam cells," *International Journal of Molecular Sciences*, vol. 20, no. 23, p. 6093, 2019.
- [87] X. Li, Q. Jin, Q. Yao et al., "The flavonoid quercetin ameliorates liver inflammation and fibrosis by regulating hepatic

macrophages activation and polarization in mice," *Frontiers in Pharmacology*, vol. 9, p. 72, 2018.

- [88] T. Lagerweij, L. Hiddingh, D. Biesmans et al., "A chemical screen for medulloblastoma identifies quercetin as a putative radiosensitizer," *Oncotarget*, vol. 7, no. 24, pp. 35776–35788, 2016.
- [89] S. Sato, T. Norikura, and Y. Mukai, "Maternal quercetin intake during lactation attenuates renal inflammation and modulates autophagy flux in high-fructose-diet-fed female rat offspring exposed to maternal malnutrition," *Food & Function*, vol. 10, no. 8, pp. 5018–5031, 2019.
- [90] S. R. Khalil, A. T. Mohammed, A. H. Abd El-Fattah, and A. W. Zaglool, "Intermediate filament protein expression pattern and inflammatory response changes in kidneys of rats receiving doxorubicin chemotherapy and quercetin," *Toxicology Letters*, vol. 288, pp. 89–98, 2018.
- [91] E. Tili and J. J. Michaille, "Promiscuous effects of some phenolic natural products on inflammation at least in part arise from their ability to modulate the expression of global regulators, namely microRNAs," *Molecules*, vol. 21, no. 9, p. 1263, 2016.
- [92] L. A. Dutra and T. R. Ferreira de Melo, "The paradigma of the interference in assays for natural products," *Biochemistry & Pharmacology: Open Access*, vol. 5, no. 3, 2016.