

Research Article

Preclinical Trial of *Ocotea puberula* (Rich.) Nees ("Canela-Guaicá") in Wound Healing: Validation of a Traditional Medicine Practice Used by Indigenous Groups in Southern Brazil

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Background. "Canela-guaicá," "guaicá," or "canela-sebo" [*Ocotea puberula* (Rich.) Nees] is a native species that is traditionally used by Kaingang indigenous groups for wound healing in southern Brazil. The aim of this study was to extract the mucilage from *O. puberula* barks, perform its phytochemical and physicochemical characterization, and investigate its healing potential. *Methods.* A murine wound model was used as a preclinical trial for authentication of the traditional knowledge from Kaingang indigenous communities. *Results.* Alkaloids and polysaccharides were identified by usual qualitative reactions and Fourier-transform infrared spectroscopy. This natural product showed thermal stability and pseudoplastic properties that were considered suitable for the intended use. A higher initial exacerbation of inflammatory response after 7 days, an improved angiogenesis after 14 days, and an increased wound shrinkage after 21 days were statistically significant for the "canela-guaicá" bark extract in the preclinical trial when compared to the silver calcium alginate dressing (positive control). *Conclusion.* The healing potential of the "canela-guaicá" bark extract, traditionally used by the Kaingang indigenous community from southern Brazil, was preclinically validated. This study paves the way for designing novel wound dressings containing this natural product in order to treat acute and chronic wounds.

1. Introduction

Chronic wounds are a major health problem with high costs and impacts on care programs, primarily affecting adults and elderly individuals. In the UK's National Health Service (NHS), the cost of wound care is conservatively estimated at \$54 billion, including various outpatient facilities. In developed countries, it has been estimated that 1–2% of the whole population may suffer from chronic wounds at some point in their lifetime [1–3]. Wound healing is a complex process that involves an orderly sequence of events, divided into four steps: hemostasis, inflammation, proliferative, and remodeling phase [4]. There are commercial dressings for treating acute and chronic wounds that include silver, nanocrystalline silver, alginate, and hydrogel [5–7].

However, this dressing might not be completely effective due to their high cost and prolonged time for treating chronic wounds, such as ulcers and scars, which remain as unresolved health problems [1]. In that sense, innovative Ocotea puberula (Rich.) Nees is an evergreen tree that belongs to the Lauraceae. Its barks present the well-known aporphine alkaloids as dicentrine, which demonstrated antituberculosis activity and antinociceptive effects [12, 13]. In addition, there are studies that the mucilaginous gel can be used for wound healing, as reported by taxonomic and ethnopharmacological studies [6, 14]. However, no previous report was devoted to investigate the wound healing effect of the mucilaginous "canela-guaicá" bark extract (CGBE) in order to provide an ethnopharmacological-based strategy for wound repair.

The aim of this paper was to characterize the mucilaginous CGBE and to treat standardized lesions in a wound healing preclinical model by comparing it with a commercially available ionic silver wound dressing.

2. Methods

2.1. Plant Material and Processing. Samples of barks of Ocotea puberula (Rich.) Ness, Lauraceae, were collected from six tree specimens at the State University of Ponta Grossa, Ponta Grossa, Parana, Brazil (latitude $25^{\circ}05'39''S$ and longitude $50^{\circ}06'05''W$) in October 2018. The plant material was identified, and a voucher specimen was stored under the number HUPG 19107 at the Herbarium of the State University of Ponta Grossa. Ocotea puberula barks were cut into 4 cm²-sized fragments, dried at $25 \pm 2^{\circ}C$, and then ground into a Wiley-type knife mill (Solab, SL31 model, Piracicaba, Brazil) to obtain a 500 μ m sized powder.

This standardized powder was stored in sealed containers in dark conditions at room temperature $(20 \pm 2^{\circ}C)$.

2.2. Preparation and Characterization of "Canela-Guaicá" Bark Extract. The "canela-guaicá" bark extract (CGBE) was prepared in triplicate using 20 g of the previously standardized powder of Ocotea puberula barks and 800 mL of freshly obtained distilled water (Milli-Q EQ 7000 ultrapure water system, Millipore, Burlington, MA, USA) by stirring (Fisaton stirrer, 713 model, São Paulo, Brazil) at $60 \pm 2^{\circ}$ C and 300 rpm for 1 h. The extract was filtered through cotton wool to remove solid particles and then centrifuged twice at $20 \pm 2^{\circ}$ C and $3600 \times g$ for 15 min. Finally, ultracentrifugation at $8 \pm 1^{\circ}$ C and $15000 \times g$ for 15 min. Finally, ultracentrifugation at $8 \pm 1^{\circ}$ C and $15000 \times g$ for 10 min was carried out in order to obtain a soluble fraction. The mucilaginous CGBE was obtained after precipitation at $4 \pm 2^{\circ}$ C for 24 h using ethanol 95% (Vetec Quimica Fina, Duque de Caxias, Brazil) at 1:3 volume ratio (soluble fraction/ethyl alcohol).

Preliminary qualitative assays were carried out in order to confirm the nature of the CGBE extract obtained. The chemical tests included were: Molish's test for carbohydrates, ruthenium red test for mucilage, Benedict's test and Fehling's test for reducing/nonreducing sugar, ninhydrin test for proteins, Folin–Ciocalteu test for total phenolic content, Shinoda test for flavonoid, ferric chloride test for tannins, and Dragendorff's test for alkaloids [15–17]. The freeze-dried CGBE was investigated by thermogravimetric analysis (TGA) and derivative thermogravimetric analysis (DTG). TGA and DTG analyses were both performed using the STA 6000 instrument (Perkin Elmer, Waltham, MA, USA) with a mass of 5 mg CGBE into an alumina crucible. The equipment was previously calibrated using indium (In, M.P. = 156.6°C, ΔH fusion = 28.54 J/g) and was heated at a 10°C/min constant rate, from 20 to 600°C, under constant nitrogen flow (50 mL/min) [18].

The CGBE rheological profile was performed in triplicate using a Discovery DR2TA-Instrumentals hybrid rheometer (New Castle, DE, USA) coupled to the TC81 Peltier temperature controller kept at 36.5°C (similar to human skin temperature) and to the PP40Ti plate-plate sensors. A volume of 20 mL of the mucilaginous CGBE was placed on the Peltier plate in order to obtain the complex viscosity data including storage (G') and loss (G'') after moduli. The rheological analysis was carried out using 30 s immersion time, 0.5 Pa tension, and logarithmic weep with 0.1 to 100 rad/s angular frequency and 5 points per decade.

The fourier-transform infrared (FTIR) spectrum was recorded by using a KBr tablet (4 mg of CGBE and 196 mg of spectroscopic grade KBr, 2% w/w) in the 4000–400 cm⁻¹ range, with a resolution of 4 cm⁻¹ at 32 scan/min in the IR Prestige-21 spectrophotometer (Shimadzu, Kyoto, Japan) [19].

2.3. Preclinical Trial: Wound Healing Assay. All the experiments were approved by the Ethics Committee on Animals Experimentation of the State University of Ponta Grossa (CEUA/UEPG) under process number 016/2018. The wound healing assay was carried out in accordance with the AR-RIVE Guidelines [20]. The sample number was calculated using the Gpower 3.1 program (2020 version, Heinrich Heine University Düsseldorf, Düsseldorf, Germany) at 80% test power based on previous studies [4, 21].

The animals were selected in accordance with age and weight (standardized age approximately 3 months, weighing around 290 g to 380 g). The rats were weighed and separated by the probabilistic random-choice process into 3 groups of 15 animals each, amounting to a 45 Male Wistar *Rattus norvegicus albinus*. These animals were kept in the light-dark cycle of 12 hours, temperature of $21 \pm 2^{\circ}$ C, adequate ventilation system, and free access to food and water.

For the surgical procedure, rats were intraperitoneally (i.p.) anesthetized with 40 mg/kg of 10% ketamine and 10 mg/kg of 2% xylazine [22]. The back of each animal was shaved, and a dorsal lesion $(2 \text{ cm} \times 2 \text{ cm})$ was made with a scalpel up to the level of the aponeurotic tissue, and then the lesions were cleaned with 0.9% sterile saline solution. Each treatment was then applied every other day using an aseptic procedure. The animals were divided into 3 groups: positive control (PC)–4×4 cm foam of alginate containing silver (Megisorb Ag, Mölnlycke Health Care, Mölnlycke, Sweden); negative control (NC)–5 mL of 0.9% sterile saline solution (Fresenius Kabi, Aquiraz, Brazil), and CGBE (5 g of mucilaginous gel). CGBE presented a mean dry matter content of 105.2 mg/g and showed mucilage (92.1 mg/g) and

alkaloids $(43.6 \,\mu g/g)$ as the most representative chemical biomarkers. All lesions were covered with sterile bandages.

Each wound was macroscopically inspected after 7, 14, and 21 days and photographed by standardized apparatus. The wound healing index (WHI) was obtained as a percentage of re-epithelialization using the lesion areas provided by standard photographs assessed by Image J 1.44 software (National Institute of Health, Bethesda, MD, USA) on days 0, 7, 14, and 21. The initial areas represented the lesion areas on the day of the surgical procedure, and the final areas corresponded to the lesion areas on each day of euthanasia. Then, the WHI value was calculated by the following formula WHI (%) = [(initial area – final area/initial area)] * 100.

At each time interval, 5 animals were euthanized per group for histochemical biopsies. Skin fragments were picked up through a skin incision, coating the healthy skin and the preinjured area. A part of the material was promptly fixed in 10% phosphate-buffered formalin (pH 7.3 and 0.1 M) for 24 hours. Subsequently, the specimens were dehydrated in ethyl alcohol in ascending concentrations, diaphanized by xylol, and impregnated with liquid paraffin in an oven set at 59°C and embedded in paraffin. The blocks were cut on a Minot microtome (LEICA®, model M2125RT), adjusted to five micrometers, and processed for hematoxylin (Merck, Darmstadt, Germany) and eosin (Merck, Darmstadt, Germany) (HE) staining to evaluate inflammatory cells and angiogenesis. Masson's trichrome staining (Biognost, Zagreb, Croatia) was used to evaluate collagen fibers. The analyses were accomplished using a light microscope (LEICA® DM500 Trinocular Microscope, Wetzlar, Germany) coupled with an ICC50 HD camera, and the images were digitalized by computer using LEICA® LAS EZ software (Wetzlar, Germany).

All analyses were performed blindly by two evaluators since the histological slides were identified by coding to which the examiner had no knowledge of which group of animals these slides belonged to. In the morphohistological evaluation, inflammatory infiltrate, vascular formation, neocollagenesis and re-epithelialization were observed at all times, according to the standard protocol for histological evaluation [20]. The scores attributed to these histological analyses were absence (0), presence (1), intensely infiltrated (2), and severely infiltrated (3) [4, 21].

The statistical data were obtained using the GraphPad Prism 5 program (San Diego, CA, USA). The results were expressed as mean ± standard deviation. The one-way ANOVA and Tukey's post hoc test were used for the histological assays. Values were considered significant when p < 0.05.

3. Results

3.1. Preparation and Characterization of "Canela-Guaicá" Bark Extract. The dried and powdered barks of O. puberula showed a suitable yield of freeze-dried mucilage (13.43%). The phytochemical results demonstrated the remarkable presence of carbohydrates and mucilage in CGBE using Molish's test and ruthenium red test, respectively. Benedict's test and Fehling's test revealed no change in color,

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which corresponded to the presence of nonreducing sugar. The positive result was also observed using Dragendorff's reagent, which means that CGBE contains alkaloids. Other photochemical assays for proteins, phenolic compounds, flavonoids, and tannins presented negative results.

The presence of phytochemicals in O. puberula barks mainly represented by mucilage corroborates with the indigenous use of CGBE for wound healing. The water retention capacity along with the moisturizing properties makes mucilages ideal for the production of dressings for wounds and skin burns [23, 24]. In particular, mucilage from CGBE can also act on healing due to the fact that it is biocompatible and nontoxic since it is composed of carbohydrates. Furthermore, the CGBE is a herbal traditional practice provided by the culture-nature relations of Kaingang groups in southern Brazil and is an ethnobotanical approach for valuing the biodiversity of the Brazilian flora at the same time. There is an extensive report about the presence of aporphine alkaloids in O. puberula with antinociceptive effects by nullifying or reducing the perception and transmission of pain-causing stimuli [12, 13]. These alkaloids can increase the wound contraction rate and can reduce the time interval for epithelialization [25, 26].

Figure 1(a) depicts the thermal results of TGA and DTG. TGA and DTG curves showed four events of mass loss. The thermal gravimetric data of the CGBE sample recorded a decomposition process starting at 38°C. This event may be due to the thermal desorption of the remaining free water molecules in the sample, causing a small amount of mass loss with a loss rate of 4.4%. A second event of water loss occurred at 174°C and was assigned to the water adsorbed to carbohydrate polymers due to the binding of functional groups. After a period of heating up, the temperature reached 329°C and entered the third stage of decomposition. The sample revealed a large amount of mass loss with a loss rate of 53.8% related to the alkaloid and mucilage compounds. The temperature continued to rise and the sample further decomposed at 445°C, and the remaining sample mass was 19.3%. This event was attributed to amino group degradation. The obtained data by TGA/DTG revealed a regular mass degradation, which indicated that CGBE showed high purity and could be used as a phytotherapeutic material for wound healing. Furthermore, the thermal degradation of the CGBE compounds occurred above 300°C, which makes it suitable for the proposed use as a dressing for wounds, since the temperature of human skin is remarkably lower.

Considering the rheological data, the storage modulus (G') and the loss modulus (G'') versus angular frequency (rad/s) plots for the CGBE sample are described in Figure 1(b). CGBE presented a higher elastic modulus (G')than viscous modulus (G''), and there was no crossing point between G' and G''' for the range of angular frequencies applied. In that sense, CGBE demonstrated a pseudoplastic behavior that enabled it to be deformable as the share force increased. This material showed resistance to mechanical stresses such as tearing, shearing, and elongation, which is an important feature for healing applications [27, 28].

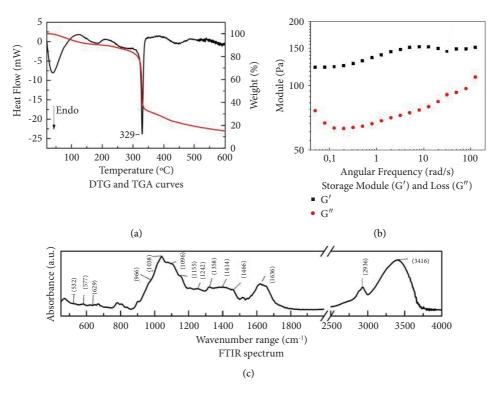


FIGURE 1: Characterization of "canela-guaicá" bark extract (CGBE) by thermogravimetric analysis (TGA) ((a), redline), derivative thermogravimetric analysis (DTG) ((a), blackline), rheological analysis (b), and fourier-transform infrared (FTIR) spectrum (c).

The FTIR spectrum of CGBE is summarized in Figure 1(c). A broad O-H band at $3,416 \text{ cm}^{-1}$ was assigned to the mucilage polysaccharides. The band at $2,936 \text{ cm}^{-1}$ was attributed to the alkyl C-H stretching. The aporphine alkaloids presented bands at $1,466 \text{ and } 1,414 \text{ cm}^{-1}$ due to the aromatic ring. The FTIR data confirmed the results obtained in the phytochemical qualitative analysis and demonstrated the potential of CGBE for wound [24] and pain [12] treatment since mucilage and alkaloids have been reported as healing compounds [29] and angiogenic and anti-inflammatory substances [30], respectively. However, these findings lack in vivo biological evidence to validate the CGBE traditional use by indigenous Kaingang groups in southern Brazil.

3.2. Preclinical Trial: Wound Healing Assay. The results of the macroscopic evaluation for experimental groups are illustrated in Figure 2. The CGBE group showed an increased lesion area after seven days of injury as well as the positive control. These findings are expected in the inflammatory phase of the murine lesion since the inflammation proceeds as usual and the dominant macrophage population shifts from an inflammatory to a tissue repair phenotype characterized by secretion of various paracrine factors. These factors promote angiogenesis as well as further proliferation of fibroblastic cells, mesenchymal stem/stromal cells, and other local stem and progenitor cells [31, 32].

After 14 days of injury, the proliferative phase of wound healing is typically occurring, and the new connective tissue or granulation tissue is formed concurrently with other healing processes, including re-epithelialization, neovascularization, and immune modulation [33]. However, the macroscopic images of the lesions do not allow to examine these processes, and further histological data are required to confirm the healing progression.

After 21 days of injury, CGBE demonstrated a substantial healing potential with a suitable regression of murine skin lesions. This performance was distinctly superior to the positive control, as shown in Figure 2. In particular, the negative control presented the lesion regression behavior in an intermediate degree between the CGBE group and the positive control due to the natural course of the healing process, as previously reported [34, 35].

These data can also be demonstrated by the wound healing index (WHI) as represented in Table 1. The WHI values were consistent with the observational data provided by the macroscopic evaluation. In particular, CGBE was the only experimental group that provided lesion regressions of above 90%, which denotes its positive effect on wound healing.

Considering the promising macroscopic and WHI findings, histological analysis was carried out in order to elucidate some probable processes involved in the wound healing progression such as the inflammatory response, the angiogenesis, and the neocollagenesis after 7, 14, and 21 days of injury, respectively. These results are described in Figure 3 and Table 2.

CGBE demonstrated a statistically higher histological score than the respective controls for the inflammatory response. In addition, an intense concentration of poly and mononuclear inflammatory cells (purple nuclei) was

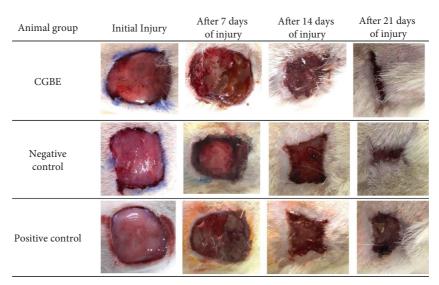


FIGURE 2: Macroscopic aspect of wounds for experimental groups at different time intervals. Images are representative of wounds in Wistar rats using a standard photography apparatus. Note: CGBE – "canela-guaicá" bark extract.

TABLE 1: Wound healing index (WHI) for the experimental groups at different time intervals.

Animal group	WHI (%)		
	After	After	After
	7 days of injury	14 days of injury	21 days of injury
	Mean ± SD	Mean ± SD	Mean ± SD
CGBE	-7.8 ± 1.7^{a}	$+64.1 \pm 3.2^{a}$	$+95.4 \pm 3.3^{a}$
Negative control	$+1.8 \pm 1.1^{b}$	$+63.6 \pm 4.7^{a}$	$+87.8 \pm 5.9^{a,b}$
Positive control	-5.5 ± 2.2^{a}	$+68.9 \pm 4.5^{a}$	$+78.8 \pm 4.9^{b}$

*one-way nonparametric ANOVA (Kruskal–Wallis test); different letters on the columns indicate the statistical difference (p < 0.05). Note. CGBE, "canela-guaicá" bark extract; SD, standard deviation.

observed in the extracellular matrix and inflammatory infiltrate in the CGBE-treated group after 7 days, when compared to the other groups (Figure 3, photomicrograph I.1). This enhanced inflammatory process, but not excessive, was essential for the inflammatory phase of wound healing since it allowed a suitable activation of cellular and humoral elements fundamental to healing. In addition, the process of wound inflammation is crucial for the optimal completion of hemostasis, as well as the detection and elimination of pathogenic microorganisms, the removal of damaged tissues, and wound cleaning. Furthermore, leukocytes (mainly macrophages) are responsible for the timely and adequate preparation of wound tissues for proliferation [36, 37].

Significant angiogenesis was reported for the CGBE group after 14 days of injury when compared to control groups. The formation of new vessels (arrows) is observed in higher concentration in the CGBE group when compared to the other groups (Figure 3, photomicrograph A.1). Remarkably, these findings could not be verified by the macroscopic analysis and were only histologically observed. This experimental evidence may be related to the presence of aporphine alkaloids in the CGBE composition. Alkaloids, according to previous studies, can stimulate new vessels in lesions and can promote fibroblasts proliferation and/or collagen production [24–26, 29, 38]. As a result of this neovascularization, with suitable supply of micronutrients

and tissue oxygenation [32], a better lesion regression was macroscopically achieved after 21 days of injury.

Considering the neocollagenesis, no statistically significant difference was obtained. The acute wound in a healthy rat model has a rapid course and is usually closured in 21 days, which provides an expanded window of observation and identification of critical events involved in wound repair [39]. Thus, the end of the acute process for all groups did not allow for establishing differences between them.

The results highlighted in this paper revealed the promising healing effect of CGBE; however, a study limitation must be reported. Some additional experiments such as enzyme-linked immunosorbent assay (ELISA), western blotting or immunohistochemistry for inflammatory cytokines and vascular endothelial growth factor (VEGF) expression, and alpha-smooth muscle actin (α -SMA) for angiogenesis could be further performed in order to elucidate the possible mechanism of action.

In brief, CGBE was essential in the inflammatory and proliferative stages of wound healing due to the improvement of cell migration and angiogenesis [40–42]. In that sense, the healing potential of CGBE, traditionally used by the Kaingang indigenous community from southern Brazil, was preclinically confirmed. This study can support the design of novel wound dressings containing this ethnopharmacological-based strategy for wound repair.

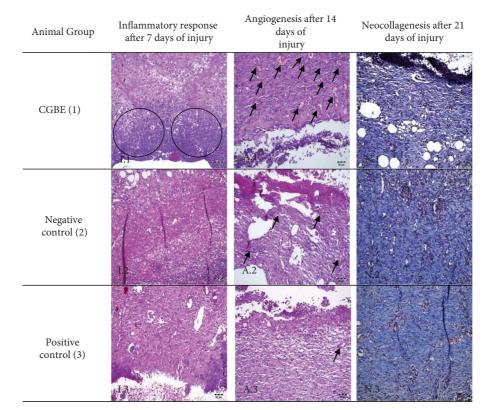


FIGURE 3: Microscopic aspect of wounds throughout the experiment according to such criteria: inflammatory response, angiogenesis, and neocollagenesis for experimental groups at different time intervals. Circles areas reveals a increased presence of inflammatory cells (poly and mononuclear). Arrows indicate new vessels. Several microphotographs were taken using a $\times 200$ magnification and the images are representative of the experimental results. Note: CGBE – "canela-guaicá" bark extract.

TABLE 2: Histological scores of wounds throughout the experiment according to such criteria: inflammatory response, angiogenesis, and neocollagenesis for experimental groups at different time intervals.

	Inflammatory response after	Angiogenesis after 14 days	Neocollagenesis after 21 days
Animal group	7 days of injury	of injury	of injury
	Mean ± SD	Mean ± SD	Mean \pm SD
CGBE	2.20 ± 0.33^{a}	2.40 ± 0.06^{a}	2.00 ± 0.71^{a}
Negative control	$1.00\pm0.00^{\rm b}$	1.60 ± 0.33^{b}	2.40 ± 0.55^{a}
Positive control	$1.00\pm0.00^{\rm b}$	$1.20 \pm 0.25^{\rm b}$	2.60 ± 0.55^{a}

*one-way nonparametric ANOVA (Kruskal–Wallis test); different letters on the columns indicate the statistical difference (p < 0.05). Note. CGBE, "canela-guaicá" bark extract; SD, standard deviation.

4. Conclusion

The mucilaginous "canela-guaicá" bark extract (CGBE) was effective as an indigenous traditional practice for treating wounds. CGBE presented alkaloids and polysaccharides as the main chemical constituents. This natural product showed thermal stability and pseudoplastic rheological behavior. CGBE provided a remarkable regression of skin lesions in the macroscopic evaluation of tissue repair after 21 days. The histological analysis demonstrated an enhanced inflammatory response and angiogenesis. CGBE proved to be an expressive ethnopharmacological knowledge of the Kaingang community from southern Brazil.

Data Availability

The data used to support the findings of the study are available from the corresponding author upon request.

Additional Points

"Canela-guaicá" [*Ocotea puberula* (Rich.) Nees] bark extract (CGBE) presented mucilage and alkaloids. This natural product showed thermal stability and pseudoplastic properties. A statistically higher angiogenesis was observed for CGBE after 14 days of treatment. An improved macroscopic wound repair was observed for CGBE after 21 days.

Ethical Approval

This research was approved by the institutional animal care and use committee (CEUA Process 014/2016 and UEPG protocol 6378/2016).

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

All authors had significant contributions to the evaluation and framing of the manuscript. G.A., P.V.F., M.D.R.G., and L.A.P. were responsible for the broad construction of the paper. P.V.F. addressed ethical and botanical considerations. G.M.B., J.M.N., and A.N. collected characterization data. L.C.L., A.Y.K., and B.C. performed the preclinical trial. L.A.P. carried out the statistical analyses. G.A. and G.M.B. wrote the first draft of the manuscript, and G.A., P.V.F., and L.A.P. contributed to writing the final versions of the manuscript.

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References

- S. R. Nussbaum, M. J. Carter, C. E. Fife et al., "An economic evaluation of the impact, cost, and medic are policy implications of chronic non healing wounds," *Value in Health*, vol. 21, no. 1, pp. 27–32, 2018.
- [2] J. F. Guest, G. W. Fuller, and P. Vowden, "Cohort study evaluating the burden of wounds to the UK's National Health Service in 2017/2018: update from 2012/2013," *BMJ Open*, vol. 10, Article ID e045253, 2020.
- [3] S. Gupta, S. Sagar, G. Maheshwari, T. Kisaka, and S. Tripathiet, "Chronic wounds: magnitude, socioeconomic burden and consequences," *Wounds Asia*, vol. 14, no. 1, 2021.
- [4] B. Carletto, A. Y. Koga, A. Novatski, R. M. Mainardes, L. C. Lipinski, and P. V. Farago, "Ursolic acid-loadedlipidcore nanocapsules reduce damage caused by estrogen deficiency in wound healing," *Colloids and Surfaces B: Biointerfaces*, vol. 203, Article ID 111720, 2021.
- [5] P. Basu, U. Narendra Kumar, and I. Manjubala, "Wound healing materials - a perspective for skin tissue engineering," *Current Science*, vol. 112, no. 12, p. 2392, 2017.
- [6] T. G. Silva, A. P. L. Vasconcelos, E. V. C. Ramos, and J. P. Farias Neto, "Evaluation of the quality of life of patients with chronic wounds treated at the Healing Clinic of the University Hospital of Sergipe," *Revista Brasileira de Qualidade de Vida*, vol. 9, no. 3, pp. 234–246, 2017.
- [7] M. Mir, M. N. Ali, A. Barakullah et al., "Synthetic polymeric biomaterials for wound healing: a review," *Progress in Biomaterials*, vol. 7, pp. 1–21, 2018.

vol. 16, no. 1, pp. 53–60, 2006.
[9] P. d. A. Sarmento, T. d. R. Ataíde, A. P. F. Barbosa, J. X. d. Araújo-Júnior, I. M. L. Lúcio, and M. L. d. A. Bastos, "Evaluation of *Zeyheria tuberculosa* extract in the perspective of a wound-healing product," *Revista Latino-Americana de Enfermagem*, vol. 22, no. 1, pp. 165–172, 2014.

Less., Asteraceae," Brazilian Journal of Pharmacognosy,

- [10] T. H. L. Bernardo, R. C. Sales Santos Veríssimo, V. Alvino et al., "Antimicrobial analysis of an antiseptic made from ethanol crude extracts of *P. granatum* and *E. uniflora* in wistar rats against *Staphylococcus aureus* and *Staphylococcus epidermidis*," *The Scientific World Journal*, vol. 2015, Article ID 751791, 7 pages, 2015.
- [11] V. P. Santos and L. M. P. Trindade, "Nursing in the use of medicinal plants and phytotherapy with emphasis in public health," *Revista Científica Fac Mais*, vol. 8, no. 1, pp. 17–34, 2017.
- [12] D. P. Montrucchio, M. M. Córdova, and A. R. Soares Santos, "Plant derived aporphinic alkaloid S(+)-Dicentrine induces antinociceptive effect in both acute and chronic inflammatory pain models: evidence for a role of TRPA1 channels," *PLoS One*, vol. 8, p. e67730, 2013.
- [13] M. C. Avila Murillo, L. T. Albarracin Melo, L. E. Cuca Suárez et al., "Aporphine alkaloids with antitubercular activity isolated from Ocotea discolor Kunth (Lauraceae)," Revista Colombiana de Química, vol. 46, no. 3, pp. 22–27, 2017.
- [14] M. L. Brotto, "Taxonomic Study of the Ocotea aubl. Genus (Lauraceae) in the dense ombrophylous Forest of paraná state, Brazil," Dissertation presented as a partial requirement for obtaining the master's degree, Federal University of Paraná, Curitiba, Brazil, 2013.
- [15] P. D. Choudhary and H. A. Pawar, "Recently investigated natural gums and mucilages as pharmaceutical excipients: an overview," *Journal of Pharmaceutics*, vol. 2014, Article ID 204849, 9 pages, 2014.
- [16] A. Singhal, R. Mazumder, A. Mazumder, and G. S. Chakraborthy, "Phytochemical screening and Antioxidant property of methanolic extract of *Bauhinia variegate* leaves," *Advance Pharmaceutical Journal*, vol. 1, no. 4, pp. 90–94, 2016.
- [17] C. D. Machado, V. Raman, J. U. Rehman et al., "Schinus molle: anatomy of leaves and stems, chemical composition and insecticidal activities of volatile oil against bed bug (Cimex lectularius)," *Revista Brasileira de Farmacognosia*, vol. 29, no. 1, pp. 1–10, 2019.
- [18] J. M. Chassot, D. Ribas, E. F. Silveira et al., "Beclomethasone Dipropionate-Loaded Polymeric Nanocapsules: Development, In Vitro Cytotoxicity, and In Vivo Evaluation of Acute Lung Injury><>><>," Journal of Nanoscience and Nanotechnology, vol. 15, no. 1, pp. 855–864, 2015.
- [19] M. L. S. Gomes, N. da Silva Nascimento, D. M. Borsato et al., "Long-lastinganti-platelet activity of cilostazol from poly(εcaprolactone)-poly(ethylene glycol) blend nanocapsules," *Materials Science and Engineering: C*, vol. 94, pp. 694–702, 2019.
- [20] N. Percie du Sert, V. Hurst, A. Ahluwalia et al., "The ARRIVE guidelines 2.0: updated guidelines for reporting animal research," *PLoS Biology*, vol. 18, no. 7, Article ID e3000410, 2020.
- [21] T. S. Santos, D. Santos, R. N. Pereira-Filho et al., "Histological evidence of wound healing improvement in rats treated with oral administration of hydroalcoholic extract of *Vitis*

8

labrusca," Current Issues in Molecular Biology, vol. 43, no. 1, pp. 335–352, 2021.

- [22] L. R. de Moura Estevão, P. Cassini-Vieira, A. G. B. Leite, A. A. V. de Carvalho Bulhões, L. da Silva Barcelos, and J. Evencio-Neto, "Morphological evaluation of wound healing events in the excisional wound healing model in rats," *Bio Protocol*, vol. 9, no. 13, Article ID e3285, 2019.
- [23] A. Martelli, T. A. M. Andrade, and G. M. T. Santos, "Perspectives on the use of herbal medicines in tissue healing: a systematic review," *Arch Health Invest*, vol. 7, no. 8, pp. 344–350, 2018.
- [24] K. Dybka-Stępień, A. Otlewska, P. Gózdz, and M. Piotrowska, "The renaissance of plant mucilage in health promotion and industrial applications: A review," *Nutrients*, vol. 13, no. 10, p. 3354, 2021.
- [25] J. P. Fetse, J. Kyekyeku, E. Dueve, and K. Mensah, "Wound healing activity of total alkaloid extract of the root bark of *Alstonia boonei* (Apocynacea)," *British Journal of Pharmaceutical Research*, vol. 4, no. 23, pp. 2642–2652, 2014.
- [26] Y. P. Sukmawan, I. Alifiar, L. Nurdianti, and W. R. Ningsih, "Wound healing effectivity of the ethanolic extracts of Ageratum conyzoides L. Leaf (white and purple flower type) and *Centella asiatica* and astaxanthin combination gel preparation in animal model," *Turkish Journal of Pharmaceutical Sciences*, vol. 18, no. 5, pp. 609–615, 2021.
- [27] G. Stojkov, Z. Niyazov, F. Picchioni, and R. K. Bose, "Relationship between structure and rheology of hydrogels for various applications," *Gels*, vol. 7, no. 4, p. 255, 2021.
- [28] E. P. Milan, M. R. V. Bertolo, V. C. A. Martins, S. Bogusz Junior, and A. M. G. Plepis, "Chitosan and collagen-based materials enriched with curcumin (*Curcuma longa*): rheological and morphological characterization," *Polysaccharides*, vol. 3, no. 1, pp. 236–249, 2022.
- [29] M. M. Tosif, A. Najda, A. Bains et al., "A comprehensive review on plant-derived mucilage: characterization, functional properties, applications, and its utilization for nanocarrier fabrication," *Polymers*, vol. 13, no. 7, p. 1066, 2021.
- [30] K. Lu, M. Bhat, and S. Basu, "Plants and their active compounds: natural molecules to target angiogenesis," *Angio*genesis, vol. 19, no. 3, pp. 287–295, 2016.
- [31] A. L. Mescher, "Macrophages and fibroblasts during inflammation and tissue repair in models of organ regeneration," *Regeneration*, vol. 4, no. 2, pp. 39–53, 2017.
- [32] L. F. S. Gushiken, F. P. Beserra, J. K. Bastos, C. J. Jackson, and C. H. Pellizzon, "Cutaneous wound healing: an update from physiopathology to current therapies," *Life*, vol. 11, no. 7, p. 665, 2021.
- [33] M. Rodrigues, N. Kosaric, C. A. Bonham, and G. C. Gurtner, "Wound healing: a cellular perspective," *Physiological Reviews*, vol. 99, no. 1, pp. 665–706, 2018.
- [34] J. M. Baron, M. Glatz, and E. Proksch, "Optimal support of wound healing: new insights," *Dermatology*, vol. 236, no. 6, pp. 593–600, 2020.
- [35] L. P. F. Abbade, M. A. C. Frade, J. R. P. Pegas et al., "Consensus on the diagnosis and management of chronic leg ulcers Brazilian Society of Dermatology," *Anais Brasileiros de Dermatologia*, vol. 95, no. 1, pp. 1–18, 2020.
- [36] A. C. d. O. Gonzalez, T. F. Costa, Z. d. A. Andrade, and A. R. A. P. Medrado, "Wound healing-a literature review," *Anais Brasileiros de Dermatologia*, vol. 91, no. 5, pp. 614–620, 2016.
- [37] V. Stupin, N. Manturova, E. Silina et al., "The effect of inflammation on the healing process of acute skin wounds under the treatment of wounds with injections in rats,"

Journal of Experimental Pharmacology, vol. 12, pp. 12409–12422, 2020.

- [38] S. Vitale, S. Colanero, M. Placidi et al., "Phytochemistry and biological activity of medicinal plants in wound healing: an overview of current research," *Molecules*, vol. 27, no. 11, p. 3566, 2022.
- [39] A. Grada, J. Mervis, and V. Falanga, "Research techniques made simple - animal models of wound healing," *Journal of Investigative Dermatology*, vol. 138, no. 10, pp. 2095–2105, 2018.
- [40] D. R. Childs and A. S. Murthy, "Overview of wound healing and management," *Surgical Clinics of North America*, vol. 97, no. 1, pp. 189–207, 2017.
- [41] X.-N. Zhang, Z.-J. Ma, Y. Wang, B. Sun, X. Guo, and C.-Q. M. Pan, "Angelica Dahurica ethanolic extract improves impaired wound healing by activating angiogenesis in diabetes," PLoS One, vol. 12, no. 5, Article ID e0177862, 2017.
- [42] H. N. Wilkinson and M. J. Hardman, "Wound healing: cellular mechanisms and pathological outcomes," *Open Biology*, vol. 10, no. 9, pp. 2002–2023, 2020.