

Research Article

TP73-AS1 rs3737589 Polymorphism is Associated With the Clinical Stage of Colorectal Cancer

Yichang Gao,¹ Shulong Zhang,² and Xueren Gao³ 

¹School of Pharmacy, Nanjing University of Chinese Medicine, Nanjing 210023, China

²Department of General Surgery, Shanghai Xuhui Central Hospital, Zhongshan-Xuhui Hospital, Fudan University, Shanghai 200030, China

³School of Pharmacy, Yancheng Teachers' University, Yancheng 224007, China

Correspondence should be addressed to Xueren Gao; gaoxr@yctu.edu.cn

Received 27 December 2022; Revised 29 January 2023; Accepted 12 February 2023; Published 22 February 2023

Academic Editor: Vijaya Anand

Copyright © 2023 Yichang Gao et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Objective. TP73-AS1 can promote the occurrence and development of a variety of tumors, including colorectal cancer (CRC). The current study aimed to investigate the association between a potentially functional genetic polymorphism (rs3737589 T > C) on the TP73-AS1 gene and the susceptibility and clinical stage of CRC in a Chinese Han population. **Methods.** The polymorphic genotyping was performed by the SNaPshot method. The real-time quantitative PCR method and the luciferase assay were used separately to explore genotype-tissue expression and the function of the genetic polymorphism. **Results.** A total of 576 CRC patients and 896 healthy controls were included in the current study. The rs3737589 polymorphism was not associated with CRC susceptibility but was associated with the CRC stage (CC vs. TT: OR = 0.25, 95% CI = 0.12–0.54, $P = 0.0003$; C vs. T: OR = 0.69, 95% CI = 0.53–0.89, $P = 0.006$; and CC vs. (TC + TT): OR = 0.26, 95% CI = 0.12–0.56, $P = 0.0004$). CRC patients carrying the rs3737589 CC genotype or C allele were less likely to have stage III/IV tumors than those carrying the rs3737589 TT genotype or T allele. The expression of TP73-AS1 was lower in CRC tissues with the rs3737589 CC genotype compared to those with the TT genotype. Bioinformatics analysis and the luciferase assay revealed that the C allele could promote the binding of miR-3166 and miR-4771 to TP73-AS1. **Conclusion.** The TP73-AS1 gene rs3737589 polymorphism affecting miRNAs binding is associated with the CRC stage and may serve as a biomarker for predicting CRC progression.

1. Introduction

Colorectal cancer (CRC) is the third most common malignant tumor and the main cause of cancer death in the world [1]. The occurrence and development of CRC is a complicated and multidimensional process, and epidemiological studies have found multiple risk factors associated with the occurrence of CRC, including various unhealthy diets and lifestyles [2]. In addition, previous genomics studies have revealed that single nucleotide polymorphisms (SNPs) in several CRC-related genes are linked to the risk and progression of CRC [3–8]. For instance, the *interleukins-17A* (*IL-17A*) rs2275913 G > A polymorphism was associated with the occurrence and severity of CRC in the Bulgarian population [6]. Three SNPs (rs2094258 C > T, rs751402 C > T, and rs873601 G > A) on

the xeroderma pigmentosum group G (XPG) gene were found to associate with CRC susceptibility in a Southern Chinese population [8].

Long noncoding RNA (lncRNA) is a class of RNA that is more than 200 nucleotides in length and has no obvious protein-coding capacity. lncRNA can be involved in the occurrence and development of many diseases, including tumors [9–12]. For instance, lncRNA TP73-AS1 transcribed from chromosome 1p36 has been identified as a novel oncogenic molecule in many tumors, such as ovarian cancer, cervical cancer, hepatoma, retinoblastoma, breast cancer, gastric cancer, and CRC [13–19]. Given the important role of TP73-AS1 in tumors, several studies have recently begun to investigate the relationship between SNPs on the TP73-AS1 gene and tumor susceptibility. Fan et al. found that the TP73-AS1 gene rs9800 polymorphism was significantly

related to CRC risk [20]. Chen et al. found that the *TP73-AS1* gene rs3737589 polymorphism might be associated with the risk of gastric cancer [21]. In addition, the rs3737589 polymorphism was a potential biomarker to predict the prognosis of patients with gastric cancer [21]. Considering that the role of the rs3737589 polymorphism in CRC is still unknown, we analyzed the association of the *TP73-AS1* gene rs3737589 polymorphism with the susceptibility and clinical stage of CRC in the current study.

2. Materials and Methods

2.1. Study Population. Peripheral blood samples from 576 CRC patients and 896 healthy controls were recruited from Shanghai Xuhui District Central Hospital. Patients were pathologically diagnosed with CRC. Healthy controls recruited from medical examinations had no history of cancer, no intestinal disease, and no systemic disease. In addition, CRC and normal paracancerous tissues were obtained from 50 CRC surgery patients who had not received radiochemotherapy before surgery. All individuals were genetically unrelated to ethnic Han Chinese.

2.2. Genotyping. Genomic DNA was extracted from all peripheral blood samples using the TIANamp Genomic DNA Kit according to the manufacturer's instructions. The genotyping of the rs3737589 polymorphism was performed by the SNaPshot method. To verify the accuracy of the genotyping results, 10% of the DNA samples were randomly selected for direct sequencing. The concordance of sequencing results was 100%.

2.3. Real-Time Quantitative PCR and Bioinformatics Analysis. Total RNA was isolated from CRC and normal paracancerous tissues using the RNAsimple total RNA kit (Tiangen) according to the manufacturer's instructions. ReverTra Ace qPCR RT Master Mix Kit (Toyobo) was used to synthesize cDNA. FastStart Universal SYBR Green Master (Roche) was used to conduct real-time quantitative PCR. The expression of *TP73-AS1* was normalized to the internal control *GAPDH*. The $2^{-\Delta\Delta C_t}$ method was used to calculate gene relative expression. The specific primer sequences are presented in Table 1. In addition, the GTEx database (<https://www.gtexportal.org/home/>) was also used to analyze the expression of *TP73-AS1* in colon tissue of different rs3737589 genotypes [22]. The lncRNASNP v3 database (https://gong_lab.hzau.edu.cn/lncRNASNP3/) was used to analyze the effect of the rs3737589 polymorphism on miRNA binding [23]. To assess the influence of the rs3737589 polymorphism on the secondary structure of *TP73-AS1*, the RNAfold web server (<https://rna.tbi.univie.ac.at/cgi-bin/RNAWeb%20Suite/RNAfold.cgi>) was used to draw a plain structure and mountain plot of *TP73-AS1* (NR_033708.1) based on the minimum free energy [24].

2.4. Luciferase Assay. A 200 bp sequence carrying rs3737589 C or T allele was inserted into the psiCHECK2 vector and then cotransfected with miR-3166, miR-4771, or

miRNA-NC, respectively, into 293T cells using Liposome 2000. Transfected cells were collected 48 hours after transfection and their luciferase activity was evaluated using a dual-luciferase reporter assay kit and a luminometer. Each test was performed in triplicate and at least three times.

2.5. Statistical Analysis. Hardy-Weinberg equilibrium (HWE) for the control group was tested by a goodness-of-fit χ^2 test. The association of the rs3737589 polymorphism with CRC susceptibility was evaluated using adjusted odds ratios (ORs) with their 95% confidence intervals (CIs). The Student's *t*-test was used to check the difference in age variable between CRC patients and healthy controls, and compare the relative luciferase activity of different alleles. χ^2 test was used to assess the difference in gender variables between CRC patients and healthy controls. In the real-time quantitative PCR experiment, *TP73-AS1* expression among different genotypes was compared using one-way ANOVA. All statistical analyses were performed by SAS 9.4 (SAS Institute, Cary, USA). $P < 0.05$ was defined as the level of significance.

3. Results

3.1. Association between the rs3737589 Polymorphism and the Susceptibility and Clinical Stage of CRC. The basic characteristics of CRC patients and healthy controls were shown in Table 2. The age and gender distributions in the case and control groups did not differ statistically. The genotype frequency distribution of the control group was consistent with HWE (Table 3). There was no link found between the rs3737589 polymorphism and CRC susceptibility (Table 3). However, the analysis based on the clinical stage showed that the rs3737589 polymorphism was associated with the CRC stage (Table 4). CRC patients carrying the rs3737589 CC genotype or C allele were less likely to have stage III/IV tumors than those carrying the TT genotype or T allele (CC vs. TT: OR = 0.25, 95% CI = 0.12–0.54, $P = 0.0003$; C vs. T: OR = 0.69, 95% CI = 0.53–0.89, $P = 0.006$). In addition, this significant association was also present under the recessive model (CC vs. (TC+TT): OR = 0.26, 95% CI = 0.12–0.56, $P = 0.0004$).

3.2. Genotype-Tissue Expression. The rs3737589 polymorphism was not associated with *TP73-AS1* expression in normal paracancerous tissues but was significantly associated with *TP73-AS1* expression in CRC tissues (Figure 1). *TP73-AS1* expression was significantly lower in CRC tissues with the CC genotype compared to those of the TT genotype. In addition, *TP73-AS1* expression was significantly higher in colon tissues with the rs3737589 CC genotype (Figure 2).

3.3. Bioinformatics Analysis and the Luciferase Assay. The rs3737589 C allele contributed to the binding of miR-3166 and miR-4771 to *TP73-AS1* (Figure 3). In addition, the rs3737589 polymorphism could affect the secondary structure of *TP73-AS1* (Figure 4).

TABLE 1: The specific primer sequences for real-time quantitative PCR.

Genes	Primer sequences
TP73-AS1	Forward: 5'-ACTCCGGACACTGTGTTTTCTC-3' Reverse: 5'-GCATCTTTTAAGGCGGCCATATC-3'
GAPDH	Forward: 5'-GTCTCCTCTGACTTCAACA-3' Reverse: 5'-TGAGGGTCTCTCTCTCTCT-3'

TABLE 2: Basic characteristics of CRC patients and healthy controls.

Variables	CRC patients (N = 576)	Healthy controls (N = 896)	P value
Age, mean \pm SD	59.4 \pm 7.3	59.2 \pm 6.4	0.54
Gender			
Male	351 (60.9%)	516 (57.6%)	0.20
Female	225 (39.1%)	380 (42.4%)	
Clinical stage			
I + II	300 (52.1%)		
III + IV	276 (47.9%)		

TABLE 3: Association between TP73-AS1 gene rs3737589 polymorphism and CRC susceptibility.

Genotype or allele	CRC patients (N = 576)	Healthy controls (N = 896)	^a OR (95% CI)	^a P value
Genotype				
TT	318 (55.2%)	508 (56.7%)	Reference	0.85
TC	213 (37.0%)	333 (37.2%)	1.03 (0.82–1.28)	
CC	45 (7.8%)	55 (6.1%)	1.33 (0.87–2.03)	
Ptrend				0.36
PHWE				0.97
TT	318 (55.2%)	508 (56.7%)	Reference	0.59
TC + CC	258 (44.8%)	388 (43.3%)	1.07 (0.86–1.32)	
TT + TC	531 (92.2%)	841 (93.9%)	Reference	
CC	45 (7.8%)	55 (6.1%)	1.32 (0.88–1.99)	
Allele				
T	849 (73.7%)	1349 (75.3%)	Reference	0.33
C	303 (26.3%)	443 (24.7%)	1.09 (0.92–1.29)	

^aAdjusted for age and gender.

TABLE 4: The association between TP73-AS1 gene rs3737589 polymorphism and clinical stage of CRC.

Genotype or allele	III + IV (n = 276)	I + II (n = 300)	Comparison	^a OR (95% CI)	^a P value
Genotype					
TT	161 (58.3%)	157 (52.3%)	TC vs. TT	0.98 (0.69–1.38)	0.96
TC	106 (38.4%)	107 (35.7%)	CC vs. TT	0.25 (0.12–0.54)	0.0003
CC	9 (3.3%)	36 (12.0%)	C vs. T	0.69 (0.53–0.89)	0.006
Allele					
T	428 (77.5%)	421 (70.2%)	(TC + CC) vs. TT	0.79 (0.57–1.10)	0.19
C	124 (22.5%)	179 (29.8%)	CC vs. (TC + TT)	0.26 (0.12–0.56)	0.0004

^aAdjusted by age and gender.

4. Discussion

Cai et al. found that TP73-AS1 could sponge miR-194 to promote CRC cell proliferation, migration, and invasion by up-regulating TGF α [19]. Jia et al. observed that TP73-AS1 could enhance CRC proliferation by functioning as a ceRNA for miR-103, which controlled PTEN expression

[25]. Furthermore, Li et al. found that TGF- β 1 could be activated by TP73-AS1 to promote CRC cell migration and invasion [26]. These previous findings suggested that TP73-AS1 could promote the progression of CRC. Since SNPs on cancer-associated lncRNA genes might be involved in the occurrence and development of cancers, we evaluated the association of a potentially functional polymorphism

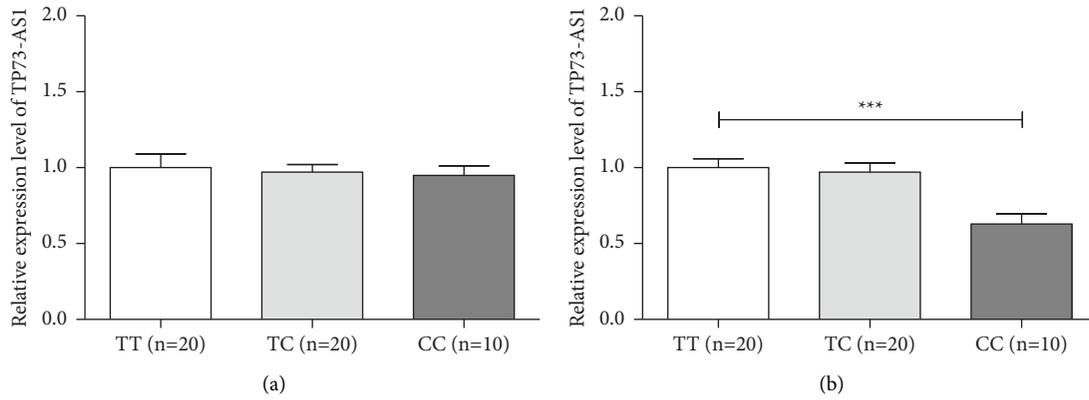


FIGURE 1: Genotype-tissue expression analysis of the rs3737589 polymorphism in colorectal tissues: (a) normal paracancerous tissues; (b) CRC tissues.

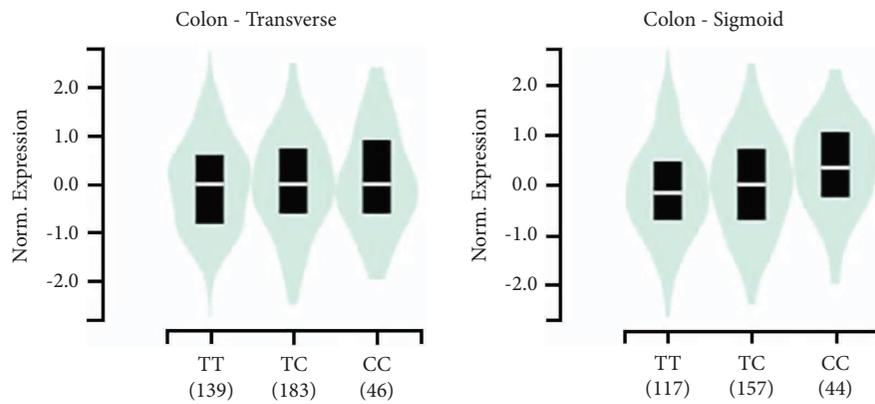


FIGURE 2: Genotype-tissue expression analysis of the rs3737589 polymorphism in colon tissues from the GTEx database.

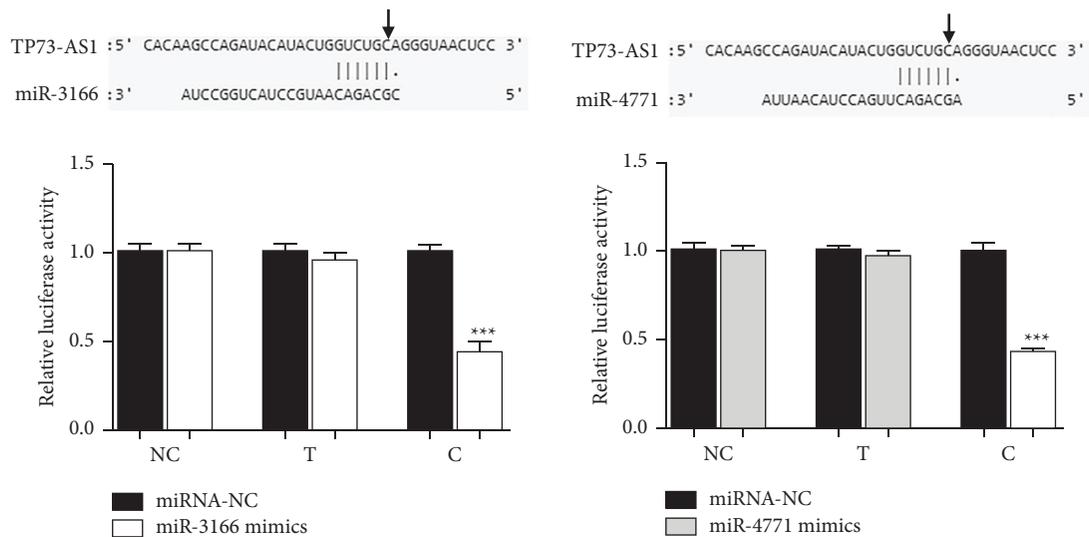


FIGURE 3: The effect of the rs3737589 polymorphism on miRNA binding.

(rs3737589) on the *TP73-AS1* gene with CRC susceptibility and clinical stage and found no significant association between this genetic polymorphism and CRC

susceptibility. However, the rs3737589 polymorphism was associated with the CRC stage. CRC patients carrying the rs3737589 CC genotype or C allele were less likely to have

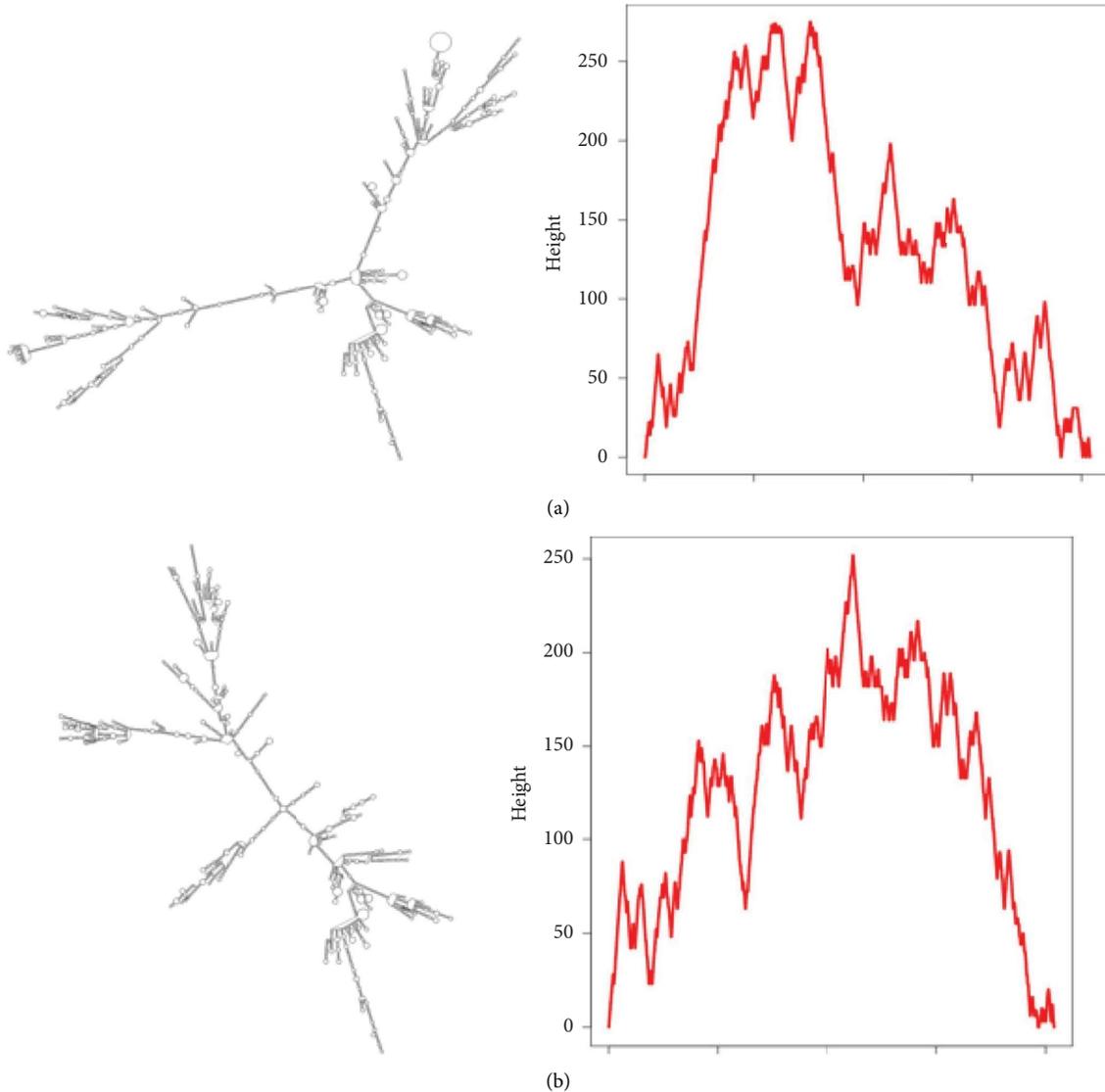


FIGURE 4: The influence of the rs3737589 polymorphism on the secondary structure of TP73-AS1: (a) TP73-AS1 with the rs3737589 A allele; (b) TP73-AS1 with the rs3737589 G allele.

stage III/IV tumors than those carrying the TT genotype or T allele. Further analysis showed that TP73-AS1 expression was significantly lower in CRC tissues with the rs3737589 CC genotype compared to those of the TT genotype. The C allele for this genetic polymorphism located on the TP73-AS1 transcript could promote the binding of miR-3166 and miR-4771. Therefore, we speculated that the rs3737589 polymorphism might influence the regulation of TP73-AS1 expression by miR-3166 and miR-4771 and thus associated CRC progression. In addition, the rs3737589 polymorphism might also affect TP73-AS1 stability by altering the structure of TP73-AS1 and thus associating CRC progression.

Although the current study has made some interesting findings, there are still some issues that need improvement. GTEx data showed that the rs3737589 polymorphism was associated with TP73-AS1 expression in colon tissues, which was not confirmed by the current study due to the

limited sample size. Since information on the lifestyle and diet of the studied individuals and clinical data were not fully collected, the study did not adjust for other confounding factors such as smoking, alcohol consumption, and red meat intake. In addition, the current study did not elucidate the exact function of the rs3737589 polymorphism in CRC. Thus, more studies are needed to confirm our findings.

In conclusion, our study confirmed that the TP73-AS1 rs3737589 polymorphism was not associated with CRC susceptibility in the Chinese Han population, but was associated with the CRC stage. This polymorphism may serve as a biomarker for predicting CRC progression.

Data Availability

The data used to support this study are available from the corresponding author upon request.

Ethical Approval

This study was approved by the Ethical Committee of Shanghai Xuhui District Central Hospital (No. 047-001). All participants have given their informed consent for participation in the study.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

This work was supported by the Opening Project of Jiangsu Province Engineering Research Center of Tumor Targeted Nano Diagnostic and Therapeutic Materials (no. JETNM202201).

References

- [1] Y. Xi and P. Xu, "Global colorectal cancer burden in 2020 and projections to 2040," *Translational Oncology*, vol. 14, no. 10, Article ID 101174, 2021.
- [2] N. Keum and E. Giovannucci, "Global burden of colorectal cancer: emerging trends, risk factors and prevention strategies," *Nature Reviews Gastroenterology & Hepatology*, vol. 16, no. 12, pp. 713–732, 2019.
- [3] X. Gao and X. Wang, "HMGA2 rs968697 T > C polymorphism is associated with the risk of colorectal cancer," *Nucleosides, Nucleotides & Nucleic Acids*, vol. 40, no. 8, pp. 821–828, 2021.
- [4] E. Bakhshian-Dehkordi, M. Safaei, S. Fattahi, M. Faghani, F. Deris, and M. H. Chaleshtori, "The association of VEGF rs833061 and rs2010963 polymorphisms with susceptibility to colorectal cancer in an Iranian population," *Cancer epidemiology*, vol. 75, Article ID 102041, 2021.
- [5] C. K. Choi, M. H. Shin, S. H. Cho et al., "Association between ALDH2 and ADH1B polymorphisms and the risk for colorectal cancer in Koreans," *Cancer Research and Treatment*, vol. 53, no. 3, pp. 754–762, 2021.
- [6] E. Aleksandrova, T. Vlaykova, J. Ananiev, and M. Gulubova, "Protective role of IL7A-197 A/G heterozygosity in the development and severity of colorectal cancer in the Bulgarian population," *Medicina (Kaunas)*, vol. 58, no. 11, p. 1632, 2022.
- [7] R. X. Hua, J. Zhu, D. H. Jiang et al., "Association of XPC gene polymorphisms with colorectal cancer risk in a Southern Chinese population: a case-control study and meta-analysis," *Genes*, vol. 7, no. 10, p. 73, 2016.
- [8] R. X. Hua, Z. J. Zhuo, J. Zhu et al., "XPG gene polymorphisms contribute to colorectal cancer susceptibility: a two-stage case-control study," *Journal of Cancer*, vol. 7, no. 12, pp. 1731–1739, 2016.
- [9] H. Huang, D. Xing, Q. Zhang et al., "LncRNAs as a new regulator of chronic musculoskeletal disorder," *Cell Proliferation*, vol. 54, no. 10, Article ID e13113, 2021.
- [10] H. Zhang, B. Liu, X. Shi, and X. Sun, "Long noncoding RNAs: potential therapeutic targets in cardiocerebrovascular diseases," *Pharmacology & Therapeutics*, vol. 221, Article ID 107744, 2021.
- [11] F. Xu, L. Jin, Y. Jin, Z. Nie, and H. Zheng, "Long noncoding RNAs in autoimmune diseases," *Journal of Biomedical Materials Research, Part A*, vol. 107, no. 2, pp. 468–475, 2019.
- [12] R. Sun, X. Y. He, C. Mei, and C. L. Ou, "Role of exosomal long non-coding RNAs in colorectal cancer," *World Journal of Gastrointestinal Oncology*, vol. 13, no. 8, pp. 867–878, 2021.
- [13] X. Wang, B. Yang, Y. She, and Y. Ye, "The lncRNA TP73-AS1 promotes ovarian cancer cell proliferation and metastasis via modulation of MMP2 and MMP9," *Journal of Cellular Biochemistry*, vol. 119, no. 9, pp. 7790–7799, 2018.
- [14] J. Xu and J. Zhang, "LncRNA TP73-AS1 is a novel regulator in cervical cancer via miR-329-3p/ARF1 axis," *Journal of Cellular Biochemistry*, vol. 121, no. 1, pp. 344–352, 2020.
- [15] C. X. Ma, W. C. Gao, and L. Tian, "LncRNA TP73-AS1 promotes malignant progression of hepatoma by regulating microRNA-103," *European Review for Medical and Pharmacological Sciences*, vol. 23, no. 11, pp. 4713–4722, 2019.
- [16] Z. Xia, X. Yang, S. Wu et al., "LncRNA TP73-AS1 down-regulates miR-139-3p to promote retinoblastoma cell proliferation," *Bioscience Reports*, vol. 39, no. 5, Article ID BSR20190475, 2019.
- [17] J. Yao, F. Xu, D. Zhang et al., "TP73-AS1 promotes breast cancer cell proliferation through miR-200a-mediated TFAM inhibition," *Journal of Cellular Biochemistry*, vol. 119, no. 1, pp. 680–690, 2018.
- [18] Z. Ding, H. Lan, R. Xu, X. Zhou, and Y. Pan, "LncRNA TP73-AS1 accelerates tumor progression in gastric cancer through regulating miR-194-5p/SDAD1 axis," *Pathology, Research & Practice*, vol. 214, no. 12, pp. 1993–1999, 2018.
- [19] Y. Cai, P. Yan, G. Zhang, W. Yang, H. Wang, and X. Cheng, "Long non-coding RNA TP73-AS1 sponges miR-194 to promote colorectal cancer cell proliferation, migration and invasion via up-regulating TGF α ," *Cancer Biomarkers*, vol. 23, no. 1, pp. 145–156, 2018.
- [20] J. Fan, H. Xu, B. Liu et al., "Association of a novel antisense lncRNA TP73-AS1 polymorphisms and expression with colorectal cancer susceptibility and prognosis," *Genes & Genomics*, vol. 44, no. 7, pp. 889–897, 2022.
- [21] W. Chen, J. Xiao, L. Shi et al., "Association of TP73-AS1 gene polymorphisms with the risk and survival of gastric cancer in a Chinese Han Population," *Artificial Cells, Nanomedicine, and Biotechnology*, vol. 47, no. 1, pp. 3814–3822, 2019.
- [22] Gtex Consortium, "The genotype-tissue expression (GTEx) project," *Nature Genetics*, vol. 45, no. 6, pp. 580–585, 2013.
- [23] Y. Yang, D. Wang, Y. R. Miao et al., "lncRNASNP v3: an updated database for functional variants in long non-coding RNAs," *Nucleic Acids Research*, vol. 51, no. 1, pp. D192–D198, 2023.
- [24] D. H. Mathews, M. D. Disney, J. L. Childs, S. J. Schroeder, M. Zuker, and D. H. Turner, "Incorporating chemical modification constraints into a dynamic programming algorithm for prediction of RNA secondary structure," *Proceedings of the National Academy of Sciences*, vol. 101, no. 19, pp. 7287–7292, 2004.
- [25] Z. Jia, J. Peng, Z. Yang et al., "Long non-coding RNA TP73-AS1 promotes colorectal cancer proliferation by acting as a ceRNA for miR-103 to regulate PTEN expression," *Gene*, vol. 685, pp. 222–229, 2019.
- [26] M. Li, Y. Jin, and Y. Li, "LncRNA TP73-AS1 α - β 1 to promote the migration and invasion of colorectal cancer cell," *Cancer Management and Research*, vol. 11, pp. 10523–10529, 2019.