

Research Article

Protective Effects of a Polyphenolic Phytochemical Quercetin against Oxidative Dysfunctions in Rats

Ahmed I. Foudah,¹ Mohammad A. Salkini,¹ Hasan Soliman Yusufoglu,²
Huda Mohammed Alkreathy,³ and Rahmat Ali Khan ⁴

¹Department of Pharmacognosy, College of Pharmacy, Prince Sattam Bin Abdulaziz University, Al-Kharj, Saudi Arabia

²Department of Pharmacognosy & Pharmaceutical Chemistry, College of Dentistry & Pharmacy, Buraydah Private Colleges, Buraydah 51418, Saudi Arabia

³Department of Pharmacology, Faculty of Medicine, King Abdulaziz University, Jeddah, Saudi Arabia

⁴Department of Biotechnology, University of Science and Technology Bannu, Bannu, KPK, Pakistan

Correspondence should be addressed to Rahmat Ali Khan; rahmatgenetics@gmail.com

Received 6 June 2022; Revised 28 August 2022; Accepted 31 August 2022; Published 20 April 2023

Academic Editor: Alamgeer Yuchi

Copyright © 2023 Ahmed I. Foudah et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Quercetin has traditionally been used in various oxidative and urinary tract dysfunctions. The current project is consequently set to evaluate the defensive efficacy of Quercetin against potassium bromate (KBrO₃) induced testicular tissue oxidative dysfunctions through biochemical, hormonal, and genotoxic markers. **Methods.** To observe the protective efficacy of Quercetin against urinogenital oxidative dysfunction in rats, thirty six albino male rats were divided into six groups. Protective efficacies of Quercetin were checked on reproductive hormonal levels, antioxidant enzyme activities, lipids peroxidation (LP), and DNA damages. **Results.** Potassium bromate exposure in experimental animals caused a reduction in the activities of antioxidant enzymes and disturbed hormonal secretions while enhancing the peroxidation of lipids and fragmentations of DNA. Cotreatment of Quercetin considerably ($P < 0.01$) reversed these abnormalities with admiration to levels of hormones, antioxidant enzymes activities, and peroxidations of lipids secure to those seen in untreated rats. ($P < 0.01$) **Conclusion.** The findings of the current project revealed that various doses of Quercetin are able to keep the testicular organ from abnormal free radical dysfunctions. These improvements might be due to the antioxidant ability of polyphenolic bioactive constituent, i.e., Quercetin.

1. Introduction

Oxidative stress and dysfunction inside the cell take place as soon as the meditation of reactive oxygen species production exceeds the system's antioxidant capability. In the aging process, oxidative stress plays an important role, and many pathogenesis are responsible for diseases such as diabetes, cancer, neurodegenerative diseases, and respiratory diseases [1]. Various toxic reports revealed that KBrO₃ an oxidizing agent causes hepatotoxicity, and mesothelioma tumor development in investigational animals causes thyroid, kidney failure, and neurotoxicity [2]. Various toxic reports revealed that KBrO₃, an oxidizing agent, causes hepatotoxicity and mesothelioma tumor development

in investigational animals, as well as causes thyroid, kidney failure, and neurotoxicity [3]. Experimental models have been the subject of numerous studies on oxidative injury and KBrO₃-investigating possible mechanisms of induced carcinogenicity [4]. The KBrO₃ produced reactive species combined with polyunsaturated fatty acids (PUFA) present in the tissue membrane to form DNA fragments [5] and decrease the activities of antioxidant enzymes and nonenzymatic antioxidants [6]. To prevent pathology, it is necessary to supply external antioxidant compounds and maintain a balance between oxidants and antioxidants. However, conventional and synthetic drugs used to treat oxidative stress are sometimes inadequate and can have many side effects [7]. However, most consumers prefer to

use natural, more effective antioxidants for a safer approach. Accordingly, plant extracts and their metabolites such as flavonoids, terpenoids, and phenolic components, provide an opportunity in this regard [8]. The use of natural antioxidants to combat tissue damage has been suggested as a healing agent as well as a coagent of medicine. Quercetin is used in the treatment of different types of cancer [9, 10], inflammation [11], oxidative damage [12], and antitumor effects [13]. Therefore, we designed to explore the protective role of Quercetin against potassium-bromated induced testicular carcinogenesis in rats.

2. Materials and Methods

2.1. Experimental Method. The present project is composed of thirty albino male rats were divided into 06 groups, each group containing 06 rats:

Group I as a control group.

Group II was given a 3 ml/kg DMSO dose.

Group III was given high grade 20 mg/kg KBrO₃.

Group IV was coadministered 75 mg/kg quercetin after 48 hrs of KBrO₃ treatment.

Group V was treated with 150 mg/kg bw quercetin after 48 hrs of KBrO₃ treatment.

Group VI was given 150 mg/kg quercetin alone.

For four week treatments were given twice a week. Upon the end of the experiment, all animals were kept on a normal diet for 24 hrs without any treatment. The animals were anesthetized, and blood was isolated from the ventral side and collected into a falcon tube, centrifuged, and refrigerated. Then the testicular tissue was removed and dried with blotting papers and weighed. After tissue coagulation, it was divided into 2 parts. For histology, one portion was cut and frozen another part at -70°C after treatment with liquid N₂ for further molecular and biochemical studies.

2.2. Serum Biochemistry. Various parameters of serum including endocrine hormones such as testosterone, estradiol, luteinizing hormones (LH), and follicle-stimulating hormones (FSH) and prolactin were calculated using a kit purchased from 10227-Czech Republic (IM1447-IM3286) IMMUNOTECH Company for serum levels.

2.3. Antioxidant Profile. (γ80 mg of tissue was homogenized in phosphate buffer at 4°C and centrifuged at 10,000 rpm. Then, from the upper clear phase, total tissue protein, glutathione (GSH), thiobarbituric acid reactive substances (TBARS) levels, catalase (CAT), peroxidase (POD), superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), gamma glutamyl transferase (γ-GT), glutathione reductase (GSR), and glutathione-S-transferase (GST) activities were measured as described [14–23].

2.4. Genotoxicity Assays. Quantitative DNA damages were estimated using the protocol of Lee and Jeong [24].

2.5. Histopathological Studies. Cellular changes were observed under a light microscope at 40x.

2.6. Statistical Analysis. To determine treatment effects, the variables were analyzed unilaterally using SPSS13.0, a well-known computer software. The significance levels in different treatments were determined by LSD at 0.05% and 0.01% probability levels.

3. Results

3.1. Effects of Quercetin on Reproductive Hormonal Secretions. Hypothalamus, the pituitary axis (HPA axis) of hormonal secretion were highly affected by ROS. The effect of different doses of Quercetin on serum levels of endocrine hormones such as testosterone, estradiol, luteinizing hormones (LH), and follicle-stimulating hormones (FSH) and prolactin were shown in Figure 1. Potassium bromated increased ($P < 0.01$) hormonal secretions of FSH, prolactin, and estradiol comparatively normal control group. Subsequent treatment with different doses of Quercetin significantly eliminated the toxic effects of ($P < 0.01$) KBrO₃ and improved near-control serum levels of prolactin and estradiol. Serum levels of FSH, testosterone, and LH were significantly increased ($P < 0.05$, $P > 0.01$) at 75 mg/kg and 150 mg/kg by treatment of Quercetin. Administration of Quercetin was more potential as it significantly restored the serum levels of luteinizing hormones and follicle stimulating hormones with 75 mg/kg and 150 mg/kg treatment in the control group ($P < 0.01$).

3.2. Effects of Quercetin on Tissue Homogenate Protein, SOD, POD, and CAT Activity. Administrations of Quercetin in different doses on tissue proteins and antioxidant enzymes such as POD, SOD, and CAT effects were shown in Figure 2. The concentrations of soluble tissue protein and the activity of SOD, POD, and CAT were significantly reduced by the treatment of potassium bromated. Coadministration of various doses of Quercetin recovered these abnormalities and maintained ($P < 0.01$) near the control group ($P < 0.01$).

3.3. Effects of Quercetin on QR, γ-GT, GSH-Px, GST, and GSR Activity. The protective effects of different doses of quercetin against KBrO₃ at different enzyme activities such as QR, γ-GT, GSH-Px, GST, and GSR are shown in Figure 3. In rats, 20 mg/kg BW of KBrO₃ significantly ($P < 0.01$) reduces the activity of phase-II metabolic enzymes such as GST, GSR, and GSH-Px and increases ($P < 0.01$) the activities of γ-GT and QR. After treatment with different doses of quercetin, enzyme activity was significantly restored near the control group ($P < 0.01$).

3.4. Effects of Different Doses of Quercetin on GSH, TBARS, H₂O₂, and Nitrate Content. Figure 4 presented the content of GSH, TBARS, H₂O₂, and nitrates in tests of different experimental groups of rats. Lipid peroxidation is caused by

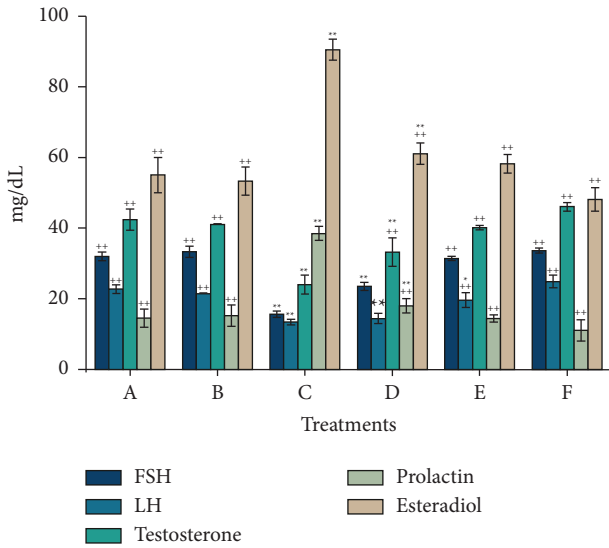


FIGURE 1: The effect of various doses of quercetin on serum male hormones in rat.

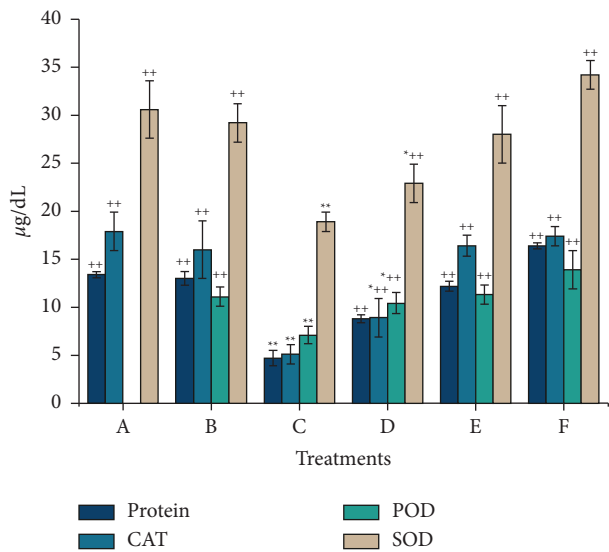


FIGURE 2: The effect of various doses of quercetin on testis protein, CAT, POD, and SOD.

20 mg/kg $KBrO_3$ and significantly reduces GSH content ($P < 0.01$) while increasing nitrate, TBARS, and H_2O_2 content as compared to the control group. The content of GSH, tissue nitrate, TBARS, and H_2O_2 was significantly higher ($P < 0.01$) than in the control group. Cotreatment of various doses of quercetin caused all these contents to improve to a normal level.

3.5. Effects of Quercetin on Testis Weight, Relative Weight, and % DNA Fragments. The effects of Quercetin against $KBrO_3$ toxicity on testis weight, relative testisweight, AgNORs, and DNA damages were shown in Figure 5. Administrations of $KBrO_3$ caused abnormalities in tissue weight, relative tissue

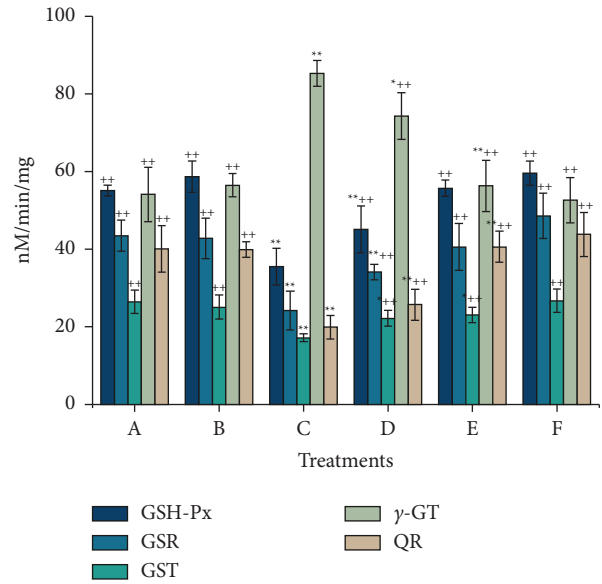


FIGURE 3: The effect of various doses of quercetin on testis GST, GSR, GSH-Px, γ -GT, and QR.

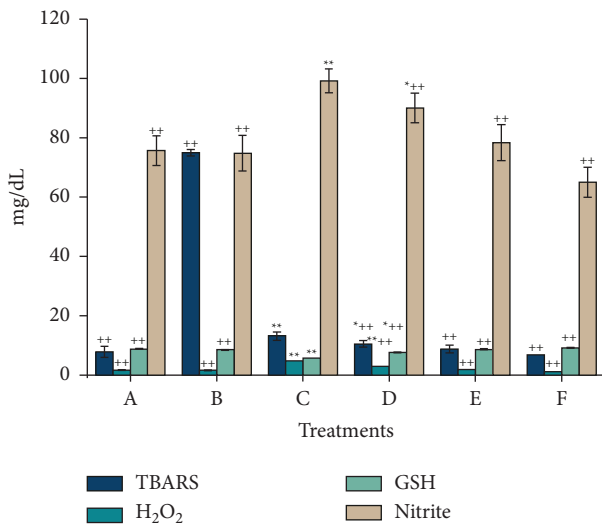


FIGURE 4: The effect of various doses of quercetin on testis GSH, TBARS, H_2O_2 , and nitrite contents.

weight, % DNA damage, and AgNORs. Cotreatment of rats with different doses of quercetin significantly improved ($P < 0.01$) tissue and relative tissue weight, DNAfragments, and number of NORS per cell near the control group.

3.6. Effect of Quercetin on the Histopathology. Microscopic examinations of the malereproductive system of the control group showed normal shape seminiferoustubules and sperm concentrations. Sertoli cells were not clear. Stromaappearance and histological appearance of fibrous muscle surrounding theprostate gland were found to be normal. Administration of $KBrO_3$ caused degeneration ofseminiferous tubules, aberration of epithelium, obstruction of meiosis, andabnormal shape and concentration of semen.

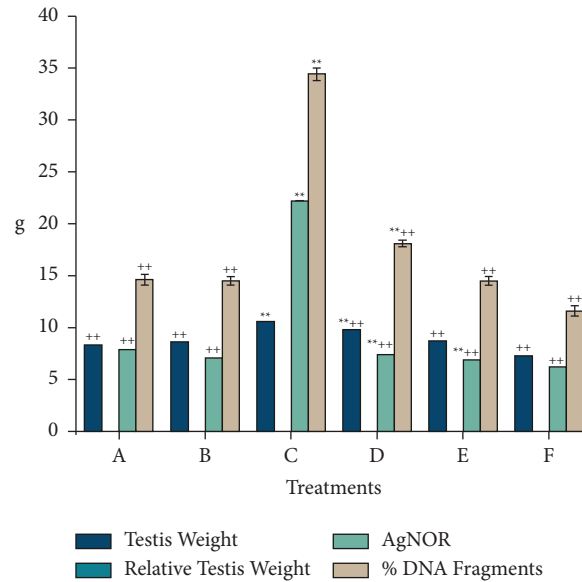


FIGURE 5: The effect of various doses of quercetin on testis weight, relative testis weight, AgNORs, and % DNA fragmentation.

TABLE 1: Effect of various doses of quercetin on histopathology.

Treatment	Tubules blockage	Meiotic interruption	Somniferous tubules	Germinative epithelium
Control	—	—	—	—
DMSO	—	—	—	—
20 mg/kg KBrO ₃	++	++	++	++
75 mg/kg quercetin + KBrO ₃	-/+	—	—	-/+
150 mg/kg quercetin + KBrO ₃	—	—	-/+	—
150 mg/kg quercetin alone	—	—	—	—

—, normal; -/+, mild; ++, medium.

Administration of different doses of Quercetin revealed clear repair of testicular abnormalities induced by KBrO₃ near the control group as shown in Table 1.

4. Discussions

Medicinal plants play an important role in the detoxification of free radicals due to the presence of bioactive ingredients. In the present study, it was reported that various doses of Quercetin have significantly reversed the KBrO₃-induced pancreatic stress. Induction of KBrO₃ generated free radicals that caused the production of highly reactive trichloroethylene and peroxy radical by the system of cytochrome P₄₅₀ oxygenase trigger the initiation of lipid peroxidation [24]. Lipid peroxidation further causes strand breakage and DNA mutation [24]. In the present study, KBrO₃ caused DNA damages in the testis, which were significantly improved by various doses of Quercetin. Related to these findings, other reports revealed that plant extract and its various components comprehensively improved injuries caused by KBrO₃ intoxication and DNA strand breakage [1].

Free radicals are thought to cause cellular injuries through lipid peroxidation [25]. In the current study, co-administration of various doses of Quercetin considerably

reversed the serum markers such as insulin, lipase, and amylase as well as blood glucose levels. Our results were in agreement with other findings [26], which reported that the same effect may be due to antioxidant activity.

SOD, POD, and CAT are highly effective antioxidant enzymes responsible for the catalytic distribution of highly reactive toxic radicals viz; superoxide and peroxide radicals [27–29]. In the present study, induction of KBrO₃ caused depletion of this enzymatic level which was significantly modulated by various doses of Quercetin. The glutathione system includes GSH, GSH-px and GSH, hydrogen peroxide, and hydroperoxide causes deficiency [30–32]. In the present study, various doses of Quercetin significantly reversed the reduction in the enzymatic level of GST, GR, GPx, and quinone reductase, which were depleted by induction of KBrO₃. Similar observations were reported during the administration of the chemical stimulant *Coriandrum sativum*, against oxidative stress [33, 34].

Lipid peroxidation (LPO) is an automated process and can result in peroxidative tissue damage, inflammation, cancer, and aging as a common result of cell death [35, 36]. In the present study, administration of KBrO₃ resulted in a significant increase in tissue MDA concentration indicating lipid peroxidation. Interestingly, the combined administration of different doses of Quercetin has significantly reduced

theLPO threshold by significantly reducing the MDA concentration, which is the effect of the extract against the lipid peroxidation of the tissue induced by KBrO_3 . Similar reports have been documented in various studies [37, 38].

The administration of KBrO_3 revealed abnormal cellular changes in testicular tissue. Coadministration of different doses of Quercetin showed protective effects and reduced cell degeneration. Our study revealed a similar investigation that agrees with previous findings [39] while examining the protective effect of medicinal plants against KBrO_3 -induced toxicity in rats.

5. Conclusion

The finding of the current study showed that various doses of Quercetin are strong antioxidant and is capable to protect testicular damage from KBrO_3 -induced toxicity. However, further studies are needed on the subject to study the mechanism of action.

Data Availability

All the data and material relevant to the paper are available in the paper.

Ethical Approval

The study was conducted according to the protocol approved by the ethical committee of the university.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

Acknowledgments

The authors extend their appreciation to the Deputyship for Research and Innovation, Ministry of Education in Saudi Arabia for funding this research work through the project number (IF-PSAU-2021/03/17778).

References

- [1] R. A. Khan, M. R. Khan, and S. Sahreen, "Evaluation of *Launaea procumbens* use in renal disorders: a rat model," *Journal of Ethnopharmacology*, vol. 128, no. 2, pp. 452–461, 2010.
- [2] A. B. DeAngelo, M. H. George, S. R. Kilburn, T. M. Moore, and D. C. Wolf, "Carcinogenicity of potassium bromate administered in the drinking water to male B6C3F1 mice and F344/N rats," *Toxicologic Pathology*, vol. 26, no. 5, pp. 587–594, 1998.
- [3] Y. Kurokawa, N. Takamura, C. Matsuoka et al., "Comparative studies on lipid peroxidation in the kidney of rats, mice, and hamsters and on the effect of cysteine, glutathione, and diethyl maleate treatment on mortality and nephrotoxicity after administration of potassium bromate," *Journal of the American College of Toxicology*, vol. 6, no. 4, pp. 489–501, 1987.
- [4] J. Chipman, J. Davies, J. Parsons, J. Nair, G. O'Neill, and J. Fawell, "DNA oxidation by potassium bromate; a direct mechanism or linked to lipid peroxidation?" *Toxicology*, vol. 126, no. 2, pp. 93–102, 1998.
- [5] R. A. Khan, M. R. Khan, S. Sahreen, S. Jan, J. Bokhari, and U. Rashid, "Protective effects of *Launaea procumbens* against KBrO_3 -induced hepatic serum marker enzymes," *African Journal of Pharmacy and Pharmacology*, vol. 5, no. 23, pp. 2639–2641, 2011.
- [6] S. Sahreen, M. R. Khan, and R. A. Khan, "Hepatoprotective effects of methanol extract of *Carissa opaca* leaves on CCl_4 -induced damage in rat," *BMC Complementary and Alternative Medicine*, vol. 11, no. 1, p. 48, 2011.
- [7] K. Wolfe, X. Wu, and R. H. Liu, "Antioxidant activity of apple peels," *Journal of Agricultural and Food Chemistry*, vol. 51, no. 3, pp. 609–614, 2003.
- [8] R. J. Wallace, "Antimicrobial properties of plant secondary metabolites," *Proceedings of the Nutrition Society*, vol. 63, no. 4, pp. 621–629, 2004.
- [9] A. F. Brito, M. Ribeiro, A. M. Abrantes et al., "Quercetin in cancer treatment, alone or in combination with conventional therapeutics?" *Current Medicinal Chemistry*, vol. 22, no. 26, pp. 3025–3039, 2015.
- [10] A. Demiroglu-Zergeroglu, E. Ergene, N. Ayvali, V. Kuete, and H. Sivas, "Quercetin and Cisplatin combined treatment altered cell cycle and mitogen activated protein kinase expressions in malignant mesotelioma cells," *BMC Complementary and Alternative Medicine*, vol. 16, no. 1, p. 281, 2016.
- [11] K. Men, X. Duan, X. W. Wei et al., "Nanoparticle-delivered quercetin for cancer therapy," *Anti-Cancer Agents in Medicinal Chemistry*, vol. 14, no. 6, pp. 826–832, 2014.
- [12] F. Ge, E. Tian, L. Wang et al., "Taxifolin suppresses rat and human testicular androgen biosynthetic enzymes," *Fitoterapia*, vol. 125, pp. 258–265, 2018.
- [13] K. Hu, L. Miao, T. J. Goodwin, J. Li, Q. Liu, and L. Huang, "Quercetin remodels the tumor microenvironment to improve the permeation, retention, and antitumor effects of nanoparticles," *ACS Nano*, vol. 11, no. 5, pp. 4916–4925, 2017.
- [14] O. H. Lowry, N. J. Rosebrough, A. L. Farr, and R. J. Randall, "Protein measurement with Folin phenol reagent," *Journal of Biological Chemistry*, vol. 193, no. 1, pp. 265–275, 1951.
- [15] B. Chance and A. C. Maehly, "Assay of catalase and peroxidases," *Methods in Enzymology*, vol. 11, pp. 764–775, 1955.
- [16] P. Kakkar, B. Das, and P. N. Viswanathan, "A modified spectrophotometric assay of superoxide dismutase," *Indian Journal of Biochemistry & Biophysics*, vol. 21, no. 2, pp. 130–132, 1984.
- [17] W. H. Habig, M. J. Pabst, and W. B. Jakoby, "Glutathione-S-transferases: the first enzymatic step in mercapturic acid formation," *Journal of Biological Chemistry*, vol. 249, no. 22, pp. 7130–7139, 1974.
- [18] I. Carlberg and E. B. Mannervik, "Glutathione level in rat brain," *Journal of Biological Chemistry*, vol. 250, pp. 4475–4480, 1975.
- [19] J. Mohandas, J. J. Marshall, G. G. Duggin, J. S. Horvath, and D. J. Tiller, "Differential distribution of glutathione and glutathione-related enzymes in rabbit kidney. Possible implications in analgesic nephropathy," *Biochemical Pharmacology*, vol. 33, no. 11, pp. 1801–1807, 1984.
- [20] A. M. Benson, M. J. Hunkeler, and P. Talalay, "Increase of NAD(P)H:quinone reductase by dietary antioxidants: possible role in protection against carcinogenesis and toxicity," *Proceedings of the National Academy of Sciences*, vol. 77, no. 9, pp. 5216–5220, 1980.

- [21] D. J. Jollow, J. R. Mitchell, N. Zampaglione, and J. Gillette, "Bromobenzene-induced liver necrosis. Protective role of glutathione and evidence for 3, 4-bromobenzene oxide as the hepatotoxic metabolite," *Pharmacology*, vol. 11, no. 3, pp. 151–169, 1974.
- [22] M. Iqbal, S. D. Sharma, H. Rezazadeh, N. Hasan, M. Abdulla, and M. Athar, "Glutathione metabolizing enzymes and oxidative stress in ferric nitrilotriacetate mediated hepatic injury," *Redox Report*, vol. 2, no. 6, pp. 385–391, 1996.
- [23] B. Wu, A. Ootani, R. Iwakiri et al., "T cell deficiency leads to liver carcinogenesis in Azoxymethane-treated rats," *Experimental Biology and Medicine*, vol. 231, no. 1, pp. 91–98, 2006.
- [24] K. J. Lee and H. G. Jeong, "Protective effect of Platycodi radix on carbon tetrachloride-induced hepatotoxicity," *Food and Chemical Toxicology*, vol. 40, no. 4, pp. 517–525, 2002.
- [25] L. T. Wang, B. Zhang, and J. J. Chen, "Effect of anti-fibrosis compound on collagen expression of hepatic cells in experimental liver fibrosis of rats," *World Journal of Gastroenterology*, vol. 6, pp. 877–880, 2000.
- [26] J. W. Xu, J. Gong, X. M. Chang et al., "Estrogen reduces CCL4-induced liver fibrosis in rats," *World Journal of Gastroenterology*, vol. 8, no. 5, pp. 883–887, 2002.
- [27] R. J. Reiter, D. X. Tan, C. Osuna, and E. Gitto, "Actions of melatonin in the reduction of oxidative stress," *Journal of Biomedical Science*, vol. 7, no. 6, pp. 444–458, 2000.
- [28] M. B. Kadiiska, B. C. Gladen, D. D. Baird et al., "Biomarkers of oxidative stress study: are plasma antioxidants markers of CCl4 poisoning?" *Free Radical Biology and Medicine*, vol. 28, no. 6, pp. 838–845, 2000.
- [29] A. Srivastava and T. Shivanandappa, "Hepatoprotective effect of the root extract of *Decalepis hamiltonii* against carbon tetrachloride-induced oxidative stress in rats," *Food Chemistry*, vol. 118, no. 2, pp. 411–417, 2010.
- [30] Y. S. Yang, T. H. Ahn, J. C. Lee et al., "Protective effects of Pycnogenol on carbon tetrachloride-induced hepatotoxicity in Sprague-Dawley rats," *Food and Chemical Toxicology*, vol. 46, no. 1, pp. 380–387, 2008.
- [31] S. Sreelatha, P. R. Padma, and M. Umadevi, "Protective effects of *Coriandrum sativum* on carbon tetrachloride-induced hepatotoxicity in rats," *Food and Chemical Toxicology*, vol. 47, no. 4, pp. 702–708, 2009.
- [32] M. Bhadauria and S. K. Nirala, "Reversal of acetaminophen induced subchronic hepatorenal injury by propolis extract in rats," *Environmental Toxicology and Pharmacology*, vol. 27, no. 1, pp. 17–25, 2009.
- [33] L. W. D. Weber, M. Boll, and A. Stampfl, "Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model," *Critical Reviews in Toxicology*, vol. 33, no. 2, pp. 105–136, 2003.
- [34] L. J. Marnett, "Oxyradicals and DNA damage," *Carcinogenesis*, vol. 21, no. 3, pp. 361–370, 2000.
- [35] E. Chavez, K. Reyes-Gordillo, J. Segovia et al., "Resveratrol prevents fibrosis NF- κ B activation and TGF- β increase induced by chronic CCl₄ treatment in rats," *Journal of Applied Toxicology*, vol. 28, no. 1, pp. 35–43, 2008.
- [36] M. Gul, B. Demircan, S. Taysi et al., "Effects of endurance training and acute exhaustive exercise on antioxidant defense mechanisms in rat heart," *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, vol. 143, no. 2, pp. 239–245, 2006.
- [37] S. Taysi, "Oxidant/antioxidant status in liver tissue of vitamin B6 deficient rats," *Clinical Nutrition*, vol. 24, no. 3, pp. 385–389, 2005.
- [38] S. Taysi, F. S. Algburi, Z. Mohammed, O. A. Ali, and M. E. Taysi, "Thymoquinone: a review of pharmacological importance, oxidative stress, COVID-19, and radiotherapy," *Mini Reviews in Medicinal Chemistry*, vol. 22, 2022.
- [39] S. Taysi, A. S. Tascan, M. G. Ugur, and M. Demir, "Radicals, oxidative/nitrosative stress and preeclampsia," *Mini-Reviews in Medicinal Chemistry*, vol. 19, no. 3, pp. 178–193, 2019.