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Review Article

Pharmacological Effects of Genistein on Cardiovascular Diseases

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Cardiovascular diseases (CVDs) are a group of disorders that involve the heart or blood vessels and are the leading cause of mortality worldwide. Natural products have several pharmacological activities, such as anti-inflammatory, antioxidant, and immunoregulatory properties. This review summarizes the possible therapeutic effects of Genistein on CVD. The information from the current review study was obtained by searching for the keywords such as "Genistein", "Cardiac dysfunction", "hypertrophy", and "Ischemia" "lipid profile" in different online database such as PubMed, Scopus, and Google Scholar, until February 2022. The results of the studies showed that genistein intake has a promising effect on improving cardiac dysfunction, ischemia, and reperfusion of the heart, decreasing cardiac toxicity, modulating lipid profile, and lowering blood pressure. The preventive effects of genistein on experimental models of studies were shown through mechanisms such as anti-inflammatory, antioxidant, and immunomodulatory effects. Pharmacological effects of genistein on cardiac dysfunction, cardiac toxicity, lipid profile, and hypertension indicate the possible remedy effect of this agent in the treatment of CVD.

1. Introduction

Cardiovascular diseases (CVDs) are a group of disorders of the heart and blood vessels that lead to the cause of mortality worldwide, approximately 31% of all global deaths [1]. CVDs consist of many types of diseases such as coronary heart disease (CHD), acute coronary syndrome (ACS), heart failure, stroke, cardiomyopathies, peripheral vascular diseases, hypertension, and dyslipidemias [2].

Diabetes, dyslipidemias, hypertension, smoking, lack of physical activity, and obesity are modifiable cardiovascular-related risk factors [3]. Prevalence of CVDs can be decreased by recognition and prevention of these risk factors. Besides lifestyle modifications, antihypertensive drugs, lipid lowering, and antithrombotic therapies are the main regimes for the prevention and treatment of CVDs [4].

For thousands of years, plants have been used for medicinal purposes and as an origin of the new drug discoveries which help the management of disorders. For example, aspirin, digoxin, and lovastatin are extracted from *salix alba* L. tree, *digitalis purpurea*, and *Monascus purpureus*, respectively [5, 6].

Phytoestrogens are a class of plant which has similar structures to estrogen, which allows them binding to estrogen receptor. Coumestans, isoflavones, and lignans are the main classes of phytoestrogens [7, 8].

Genistein is a phytoestrogen that belongs to the isoflavones family of Leguminosae plants. Genistein is a good choice for the management of some diseases because of its pharmacological and physiological properties [9]. Also, due to the pharmacological and biological properties such as antioxidant, anti-inflammatory, and anti-infective effects, isoflavones are considered for therapy/prevention purposes in medicine [10–12].

Genistein selectivity to estrogen receptor β is 20 times more than α , which is an acceptable substitute for estrogen replacement in postmenopausal women [13]. Genistein has both estrogenic and antiestrogenic activities by binding to estrogen receptor and sex hormone-binding proteins [14]. Genistein consumption reduces the incidence of prostate and breast malignancy by decreasing the testosterone level and competition with natural estrogen and, finally, inhibition of cancer cell growth and tumor development [15, 16].

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The notable effect of genistein on bone density and bone turnover makes it as a principal option for the prevention of osteoporosis [17]. Genistein has safety and tolerability and has no significant change on liver function and hematology test result, and it could be a good reason to use genistein as medicine [8].

Previous studies revealed that dietary intake of genistein was related to lesser risk of cardiovascular diseases; therefore, it is time to assess the evidence of various effects of genistein on cardiovascular diseases. So, evaluating the effects of genistein on cardiovascular disease is the purpose of this systematic review.

2. Method

The information of this study was obtained by searching the keywords such as "Genistein", "Cardiac dysfunction", "hypertrophy", and "Ischemia" "lipid profile" in different online database such as PubMed, Scopus, and Google Scholar until February 2022. Articles published in languages other than English were excluded. A total of 140 articles were selected from the above databases. 22 articles were duplicates, 65 including; reviews, letter to editor, and some articles with poorly and unsuitable information were removed, and at least 53 original articles related to the topic of this review were included in the current study as shown in Figure 1.

3. Results

3.1. Genistein Effects on Cardiac Dysfunction and Ischemia/ Reperfusion

3.1.1. In Vitro Studies. The cardioprotective effect of genistein against cobalt chloride (CoCl2) induced hypoxia in H9c2, up-regulated the expression of Notch-1, reduced cell death, and suppressed HIF-1 α expression. Furthermore, genistein could antagonize CoCl2-induced apoptosis through inhibiting caspase-3 expression and up-regulating Bcl-2/Bax ratio [18].

Genistein treatment (10^{-5} to $10^{-10} \mu mol$) in an acute myocardial ischemia model, increased the endothelial colony-forming cell (ECFC) migration and proliferation, and parallel increased the expression of ILK, α -parvin, Factin, and phosphorylation of ERK 1/2 signaling. ECFCs induction into myocardial ischemic sites, cellular expansion stimulation, and angiogenic modulators secretion at the damaged tissues are the result of genistein treatment. Genistein also improved cardiac function, vascularization, and decreased myocardial fibrosis. The genistein administration to ECFCs before transplantation improved the regenerative potential in ischemic tissues; therefore, this is a new approach for stem cell therapy in ischemic diseases [19].

Treatment of isoproterenol (ISO)-induced H9c2 cardiomyoblast cells with genistein suppressed the mitochondrial pro-apoptotic proteins expression (Bad, caspase-3, caspase-8, and caspase-9). Also in treated cells, genistein expressed many survival proteins such as phosphor p-Bad, (p)-Akt, and p-Erk1/2. Furthermore, through the expression

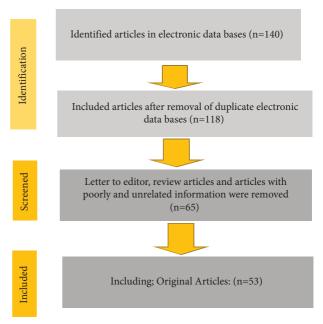


FIGURE 1: Flowchart of the process for selecting studies for the review.

of NF κ B, Akt, and Erk1/2 proteins, genistein has a protective effect [20]. These studies showed that genistein is a natural and safe selective estrogen receptor modulator (SERM) substitute to hormone replacement in cardioprotection.

Genistein outcome on lipopolysaccharide (LPS)-induced myocarditis in H9c2 cells enhanced the potential of cells growth and prevented cells apoptosis and inflammatory reaction. The results also showed that MAPK as the signaling pathway and Myc (a well-target gene of genistein) as the potential target of genistein in myocarditis [21].

Genistein protective effects against oxidative stress on cardiomyoblasts (H9C2) were studied. Treatment of cells with genistein (10 pM and 100 nM; for 24 h) modulated NO release, induced endothelial NOS-dependent NO production through modulation of intracellular signaling related to Akt, p38MAPK, and ERK1/2, increased the content of glutathione, and decreased reactive oxygen species production [22].

3.1.2. In Vivo Studies. Intraperitoneal administration of genistein (5 mg/kg) showed cardioprotective effects in both sexes of rats. It decreased postischemic contractile recovery (PICR) and pressure/total heat rate (P/Ht) during low-flow ischemia/reperfusion (LFI)/R. Genistein also improved the heart dysfunction by Na⁺/Ca²⁺-exchanger (NCX) blockade with mitochondrial permeability transition pore (mPTP) opening [23].

Genistein administration (250 mg/kg) in ovariectomized (OVX) rats increased contractility (259 mm Hg/sec) and cardiac output (7 mL/min) above baseline. Genistein-treated hearts (n = 5) demonstrated ischemic tolerance, greater cardiac output, and markedly improvements of contractility after reperfusion. Also, genistein decreased mean glucose transporter protein 4 content [24].

Once daily injection of genistein (0.1 and 0.2 mg·kg) in male, Wistar rats showed that genistein prevents increase in heart weight to body weight ratio induced by isoproterenol, fibrosis, left ventricular mass, myocardial oxidative stress, myocyte size, and myocardial 1-OH proline. Inducible nitric oxide synthase (iNOS) inhibition and improvement of endothelial nitric oxide synthase (Enos) activities are the important effects of genistein on isoproterenol-induced cardiac hypertrophy [25].

Intraperitoneal administration of genistein improved pressure and left ventricular pressure/total heat rate in aged female and young male rats but not in young female rats in stunning induced by ischemia/reperfusion (I/R) in isolated hearts. In young male rats, genistein and estradiol have synergistic cardioprotective effect. After ischemia, SR Ca leak causing diastolic contracture was increased due to genistein. Results suggested that genistein was more cardioprotective and synergistic with estradiol, which depends on the mKATP and protein-kinase C channel pathway activation [26].

Administration of genistein (10, 50, and 100 mg/kg/day) on pressure overload-induced cardiac to mice markedly attenuated fibrosis hypertrophy and fibrosis 8 weeks later [27].

TGF β 1-induced proliferation, myofibroblast transformation, and production of collagen on cardiac fibroblasts (CFs) were inhibited by genistein (20, 50, and 100 μ M) administration. Genistein also decreased the expression of TGF β -activated kinase 1 (TAK1) and induced antifibrotic effects. It also increased estrogen-dependent expression of metastasis-associated gene 3 (MTA3), which was a negative modulator of TAK1 [27].

The effects of genistein (as tyrosine kinase inhibitor) on hypertrophy induced by phenylephrine in ventricular myocytes cultured from neonatal rats prevented activation of three promoters, such as atrial natriuretic factor (ANF), Fos, and the myosin light chain 2 (MLC-2) in induced cells. Inhibition of the GTP loading of the Ras protein and activation of the mitogen-activated protein (MAP) kinases Erk1 and Erk2 in phenylephrine-induced cells were another effect of genistein [28].

The results demonstrated that for the activation of the Ras-MAP kinase pathway, which is a critical pathway in hypertrophic response regulation, genistein is important.

Genistein effects on pathological cardiac hypertrophy in mice model showed 7 weeks administration of genistein (40 mg/kg/day) significantly increased collagen volume fraction, cross-sectional area of cardiomyocytes, upregulated expression of beta-myosin heavy chain (β -MHC) promoter, atrial natriuretic peptide (ANP), collagen I α , fibronectin, collagen III, TGF- β 1, and connective tissue growth factor (CTGF) [29].

In pulmonary hypertension (PH) rats, genistein (1 mg/kg/day) administration showed decreased right ventricular ejection fraction to $41.99 \pm 1.27\%$ and increased peak systolic right ventricular pressure to 66.35 ± 1.03 mm Hg. Similar to the control group, right ventricular pressure was reduced, and right ventricular ejection fraction was completely restored to $65.67 \pm 1.08\%$. Proliferation of the smooth muscle

cell of the human pulmonary artery through estrogen receptor- β was inhibited by genistein [30].

Genistein (0.075 mg/min) infusion in pigs increased coronary blood flow by about 16.3%. Also, genistein induced NO production through p38 mitogen-activated protein kinase (MAPK), ERK 1/2, and Akt pathways. In addition, genistein caused coronary vasodilation via release of NO by vasodilatory beta (2)-adrenoreceptor [31].

In hypersensitive necrotic area, genistein IV administration attenuated the necrosis of myocardia and myocardial myeloperoxidase activity (MPO). It also decreased serum levels of TNF- α , creatinine phosphokinase activity (CPK), and ventricular arrhythmias and slowed intercellular adhesion molecule-1 (ICAM-1) expression in the injured myocardium [32]. These data suggested that myocardial ischemia-reperfusion injury protection and inflammatory response inhibition are some effects of genistein.

Genistein administration as dietary supplement (54 mg/ day) in postmenopausal women for one year showed a significant development of left ventricular ejection fraction (LVEF) and left atrial (LA) area fractional change that followed 1 year of treatment. LA longitudinal strain peak and body surface area indexed LA systolic volume were changed by genistein. In women with metabolic syndrome (MetS), genistein intake (54 mg/day/year) improved both LA remodeling and function and LV ejection fraction [33]. Administration of dietary intake of the genistein in humans showed that genistein mitigates ischemic stroke-induced damage by alterations in molecular pathways. Genistein treatment blocked targeting estrogen, nuclear factor (NF)kappa B, direct antioxidant action, androgen-mediated molecular, and Akt signaling pathways which are important for stroke damages decreasing and cell survival increasing [34]. Subcutaneous genistein administration of 0.2 mg/kg/day in ovariectomized New Zealand white female rabbits did not have a protection effect on ischemic myocardium in both nonovariectomized or ovariectomized animals for 4 weeks [35]. Compared to control groups, genistein treatment (600 mg/kg, diet) for 4 weeks in both sexes of C57BL/6J mice made echocardiographic changes in function, increased left ventricle internal dimension (LVIDs), cardiac GLUT4 protein expression, and significantly decreased fractional shortening and whole heart surface area [36].

Genistein effects on cardiac dysfunction and ischemia/ reperfusion are shown in Table 1.

3.2. Genistein Effects on Induced Cardiac Toxicity

3.2.1. In Vitro Studies. Dietary intake of genistein effect (20 microM) on ventricular myocytes in high glucose medium induced diabetic cardiomyopathy in adult rats reduced cardiac mechanical dysfunction, so it concluded that genistein had a significant effect against cardiac dysfunction induced by diabetes [38].

3.2.2. In Vivo Studies. Administration of genistein in endotoxic shock guinea pig model in the chronic and acute

Table 1: The effects of genistein on cardiac dysfunction and ischemia/reperfusion.

H9c2 embryonic rat cardiac cells (0, 50, 100, 150, 200) μ mol/L Human umbilical cord blood (HUCB) 10^{10} – 10^{-5} μ mol	00) μmol/L Expose nool Expose nool Expose	leath n tosis	
		BCl-2/Bax ratio	[18]
		ECFCs' proliferation and migration Expression of α-parvin, F-actin, ILK Phosphorylation of the signaling of ERK 1/2 ECFCs (GS-ECFCs) Proliferation of the Cells Angiogenic cytokines Neovascularization Myocardial fibrosis Cardiac function Cardiac function Regenerative potential in ischemic tissues	[19]
H9c2 cardiomyoblast cells 10 ⁻¹⁰ -10 ⁻⁵ M	M Expose		[20]
H9c2 cells $10\mu\mathrm{M}$	Expose	↓Damage of H9c2 cells ↑Cells growth ability ↓Apoptosis ↓Inflammatory response ↓Myc expression ↓P-P38 MAPK and p-JNK expression	[21]
Cardiomyoblasts (H9C2) 10 Pm-100 nM	nM Expose	†Endothelial NOS-dependent NO by the modulation of intracellular signaling related to p38MAPK, ERK1/2, and Akt	[22]
Human Heart cells	Expose	↓PICR and P/Ht ↑Dysfunction ↓MNCX ↑MKATP channels	[23]
Cultured cardiac fibroblasts (CFs) $10,50,100\mu\mathrm{M}$	M Expose	<pre>LTGFβ1-induced proliferation</pre>	[27]
Culture of ventricular myocytes from neonatal rats	Expose	$ \begin{array}{c} \downarrow \text{ANF} \\ \downarrow \text{Fos} \\ \downarrow \text{MLC-2} \\ \downarrow \text{(MAP) kinases} \\ \downarrow \text{GTP} \end{array} $	[28]
Rats 250 mg/kg	Injected once daily for 2 days	Contractility Cardiac output Tischemic tolerance Mean (SEM) recovery of contractility and cardiac output Mean glucose transporter protein 4	[24]

TABLE 1: Continued.

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Study Design	Doses	Route of administration	Effects	Ref
Rats	0.1, 0.2 mg/kg	Subcutaneous injection	↓Heart weight to body weight ratio ↓Left ventricular mass ↓1-OH proline and oxidative stress ↓ Myocardial fibrosis	[25]
Rats	5 mg/kg or 3 mg/kg	Intraperitoneal	↑SR Ca leak ↑Diastolic contracture	[26]
C57/BL6 mice	40 mg/kg/day for 7 weeks	Gavage	f Cross-sectional area of cardiomyocytes LAKT/GSK-3β signaling and MAPK (INK1/2, P38, and ERK1/2) f Collagen volume fraction f ANP J BNP fβ-MHC J CTGF LFibronectin, collagen III TGF-β1 LTGF-β1 LMAPK (JNK1/2, P38, and ERK1/2)	[37]
Rats	1 mg/kg/day	Injection	↓Right ventricular ↓PH-induced pulmonary vascular	[30]
Pigs	0.075 mg/min	Infused	†Coronary blood flow †Phosphorylation of NO synthase †NO production through p38 MAPK, ERK 1/2, and Akt pathways †Coronary vasodilation †Release of NO	[31]
Rats	(0.25, 0.5, 1.0, 1.5, 3, and 5 mg kg)	I.V., 5 min after coronary artery occlusion	↓Necrosis of myocardia ↓MPO activity ↓Serum CPK activity ↑Myocardial contractility ↓Ventricular arrhythmias ↓TNF-alpha and blunted ICAM-1 ↓Inflammatory response	[32]
In 22 postmenopausal patients	54 mg/day	One-year genistein dietary supplementation	†LA area fractional change †LV ejection fraction †LA remodeling	[33]
Human	I	Dietary intake	↓NF-kappa B ↓Akt signaling ↓Direct antioxidant action ↓Estrogen and androgen-mediated molecular pathways	[34]
Rabbits	0.2 mg/kg/day	Subcutaneously	A single dose of genistein had no protective effect	[35]
Mice	600 mg/kg	Were fed	fEchocardiographic changes in function fLVID fWhole heart surface area fCardiac GLUT4 protein expression fGLUT4 protein expression	[36]

TABLE 2: The effects of genistein on cardio toxicity.

Study Design Doses Route of administration Adult rat ventricular myocytes 20 microM Dietary intake Guinea pig model — Chronic: daily subcutaneous injection for 10 day Mice 5 mg/kg Injection Diabetic rats 300 mg/kg/day Administered po for 24 weeks Mice (0.25, 0.5 or 1 mg/kg) intraperitoneal injection					
ea pig model — 5 mg/kg fic rats 300 mg/kg/day (0.25, 0.5 or 1 mg/kg)	tudy Design	Doses	Route of administration	Effects	Ref
ea pig model 5 mg/kg etic rats 300 mg/kg/day (0.25, 0.5 or 1 mg/kg)	dult rat ventricular myocytes	20 microM	Dietary intake	[38] Uclucose toxicity-induced cardiac mechanical dysfunction	[38]
5 mg/kg stic rats 300 mg/kg/day (0.25, 0.5 or 1 mg/kg)	tuinea pig model	I	Chronic: daily subcutaneous injection for 10 days Acute: intraperitoneal injection 1 hour before endotoxic shock	↑plasma nitrate concentration ↑cGMP ↓APD	[39]
etic rats 300 mg/kg/day (0.25, 0.5 or 1 mg/kg)	lice	5 mg/kg	Injection	↓Expressions of IL-6, IL-8, TNF-α ↑Protein expressions of HO-1, Nrf-2, NQO1 ↓Apoptosis	[40]
(0.25, 0.5 or 1 mg/kg)	viabetic rats	300 mg/kg/day	Administered po for 24 weeks		[41]
).25, 0.5 or 1 mg/kg		↓Burn-induced myocardial injury ↑LVEF ↓Lactate dehydrogenase ↓level of creatine kinase ↓Apoptosis ↓Inducible NO synthase ↓NO and superoxide anions production ↑Expression of Hes1 and NICD1 ↓Burn-induced myocardial injury ↓Oxidative/nitrative stress	[42]

treatment part (s.i And i.p, respectively) showed that genistein elevated the myocardial guanosine 3',5'-cyclic monophosphate (cGMP) and concentration of plasma nitrate after 6-hour endotoxic shock. Genistein significantly attenuated the cardiac action potential duration (APD) shortening in endotoxic shock by lowering the plasma nitrate and the cardiac cGMP production in both kinds of administration [39].

The protective effects of genistein (5 mg/kg) against cardio toxicity induced by doxorubicin (DOX) in the mice model significantly reduced cardiac troponin and redox markers such as lipid peroxidation (LPO) and reactive oxygen species (ROS) in serum that increased by DOX. In addition, genistein reduced the expressions of interleukin (IL)-6, IL-8, and tumor necrosis factor (TNF)- α which increased during DOX-induced inflammatory and regulated antioxidant response by increased protein expressions of Nrf-2, HO-1, nitroquinoline oxide (NQO1) [40].

Oral administration of genistein (300 mg/kg/day) in streptozotocin (STZ)-induced diabetic rats showed decreasing in C-reactive protein, HbA1c, blood glucose, and expression of TGF- β 1 and NF- α proteins that were increased in STZ-induced diabetes. In addition, compared to control groups, genistein treatment increased total antioxidant reserves of the hearts and reduced the ultrastructural degenerative changes in the cardiac tissues [41]. These results indicated that genistein had an anti-inflammatory and antioxidant effect.

Intraperitoneal injection of genistein (0.25, 0.5, or 1 mg/kg) in mice revealed cardioprotective and antinitrative/oxidative effects after burning injury in mice. Genistein reduced myocardial injury such as improved left ventricle ejection fraction, reduced creatine kinase levels, and lactate dehydrogenase in the serum. It also reduced expressions of inducible nitric oxide (NO) synthase, NO and superoxide anions production, and reduced peroxynitrite. Treatment with genistein significantly up-regulated the expression of the enhancer of split (Hes-1) and Notch-1 intracellular domain (NICD1) after burning injury [42]. Genistein effects on cardio toxicity are summarized in Table 2.

3.3. Effects of Genistein on Lipids Profile

3.3.1. In Vitro Studies. Treatment of HepG2 cells cultured with genistein (0, 0.01, 1.00, 10.00, and 50.00 μ M) for 24 hr, decreased the secretion of apolipoprotein A1 (Apo-A1) and high-density lipoprotein-cholesterol (HDL-C), and, meanwhile at dose 1.00 μ M, increased total cholesterol (TC). Also, genistein upregulated 3-hydroxy-3-methyl glutaryl coenzyme A reductase (HMGCR), mRNA levels of low-density lipoprotein receptor (LDLR), and the protein and mRNA levels of sterol regulatory element binding proteins-2 (SREBP-2) at dose dependently and decreased peroxisome proliferator-activated receptor- γ (PPAR γ) mRNA level and liver X receptor (LXR α) [43]. These results indicated that genistein could inhibit cellular cholesterol efflux.

Genistein (1000 nM) administration in human umbilical vein endothelial cells (HUVECs) for 30 min showed

inhibition of ox-LDL-induced lowering of the activity of SA- β -gal levels and P16 and P21 proteins. The genistein effect was bound up with elevating autophagic flux [44].

3.3.2. In Vivo Studies. L-carnitine and genistein (50 mg/kg) administration on serum lipid and cytokine profiles in induced nephrotic syndrome demonstrated genistein effect on cardiovascular diseases by inducing alterations in metabolism of lipid and production of cytokine. In addition, after treatment with genistein, low-density lipoproteins (LDL) and interleukin-6 markedly decreased. The effect of genistein on triglyceride and high-density lipoproteins (HDL) levels was less than L-carnitine [45].

Genistein (20 mg/kg) administration for 42 days on polycystic ovary syndrome (PCOS) rats significantly increased insulin level. Genistein also significantly lower the levels of MDA TNF- α and higher SOD activity in the serum. Furthermore, in the histopathological analysis, genistein led to fewer cysts development and increase in luteinization [46]. Compare to control groups, estradiol (E2) and genistein (Gen) administration to ovariectomized (OVX) rats showed that treatment with E2 had significantly increased LDL chol, total cholesterol (Total-C), total oxidant status, and oxidative stress index. Also, in the genistein group, LDL chol and total-C decreased more than in the E2 treatment group. So we could conclude that Gen treatment might be preferred to E2 treatment for alleviating the menopausal symptoms and those who are at risk for cardiovascular diseases [47].

Genistein (600 mg/kg diet) supplement and soy protein (200 g/kg diet) in STZ-induced diabetic rat increased the glucokinase level and plasma insulin level but decreased glucose-6-phosphatase, thiobarbituric acid reactive substances, and HbA(IC) level of the STZ-induced diabetic rats. This combination administration significantly increased hepatic SOD, glutathione peroxidase, and catalase activities compared to the control groups [48].

Administration of 200 ppm genistein in rats by a 4-week feeding decreased cholesterol levels and improved calcium absorption efficiency. In addition, in OVX rats receiving an AIN-93M diet supplement with 200 ppm genistein, restored total and trabecular bone mineral density at the distal femur similar to the levels of sham rats [49].

Subcutaneous administration of Gen (10 mg/kg) in orchidectomized (Orx) and intact (IA) middle-aged male rats significantly reduced lipoprotein cholesterol and TC levels. Gen showed significant cholesterol-lowering effect but increased total triglycerides only in the Orx group [50]. Dietary of genistein on induced hyperlipidemia in male hamsters showed genistein (2 g/kg) significantly lowered plasma cholesterol, triglyceride, LDL cholesterol, MDA, and liver cholesterol than those in the high-fat diet (HFD) group. In addition, genistein significantly up-regulated the expression of α and β mRNA estrogen and hepatic LDL receptors compared to the HFD group [51].

Administration of 250 μ mol/L genistein that dissolved in dimethyl sulfoxide in humans showed that by decreasing lipid elements, genistein has beneficial effects on

TABLE 3: Genistein effects on lipids profile.

		7		
Study Design	Doses	Route of administration	Effects	Ref
HepG2 cells cultured	0, 0.01, 1.00, 10.00, and $50.00\mu\mathrm{M}$	Expose	f Total-cholesterol levels JHDL-C JApo-A1 [Sterol regulatory element binding proteins-2)SREBP-2) at protein and mRNA levels [Low-density lipoprotein receptor (LDLR) mRNA levels f3-hydroxy-3-methyl glutaryl coenzyme A reductase (HMGCR) [Intracellular cholesterol synthesis JMRNA levels of (PPARy) JX receptor of liver (LXRα) LXRα and ATP-binding cassette transporter A1 (ABCA1) protein levels fIntracellular cholesterol levels fSREBP-2/LDLR/HMGCR pathway JPPARy/LXRα/ABCA1 pathway JCholesterol plasma levels fCholesterol absorption JCholesterol efflux	[43]
Human umbilical vein endothelial cells (HUVECs)	1000 nM	Pretreated with genistein	<pre></pre>	[44]
Rats	50 mg/kg	Gavaged	$\downarrow Triglyceride$ and HDL level more than LCG or LC $\downarrow TNF-\alpha,$ and IL-6	[45]
Rats	20 mg/kg	Gavage	Levels of malondialdehyde (MDA)	[46]
Rats	10 mg/kg/day	Expose	↓LDL cholesterol ↓Total cholesterol levels	[47]
Rats	600 mg/kg diet	In the diet	↑Plasma insulin level ↓HbA(1C) level ↑Glucokinase level ↓Hyperglycemia and Glucose-6-phosphatase	[48]
Rats	200 ppm	A 4-week feeding study	↓Serum cholesterol ↑Total and trabecular bone mineral density	[49]

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TABLE

		Table 3: Continued.		
Study Design	Doses	Route of administration	Effects	Ref
Rats	10 mg/kg	Subcutaneous administration	↓Total cholesterol levels ↓Lipoprotein cholesterol levels ↑TT levels ↓Serum cholesterol levels ↑Serum triglycerides	[50]
Hamsters	2 g/kg	Diet		[51]
Volunteers human	250 µmol/L	Was dissolved in dimethyl sulfoxide injection	↓Agonist-induced platelet aggregation ↓Collagen- and ADP-dependent platelet activation ↓Coronary atherosclerosis ↓Platelet activity	[52]
Human	0.5–2.5 microM	Subjects consuming	↓Lipid peroxidation of HDL ↑Conjugated dienes ↓Alterations of apoprotein structure	[53]
Women	90 mg/day	Supplement for 12 week	↓ Triglyceride levels ↓Total cholesterol ↓Ldl	[54]
Postmenopausal women	54 mg	PO for 24 month	↓Fasting glucose ↓HOMA-IR and insulin ↓Homocysteine and Fibrinogen	[55]
Postmenopausal women	60 mg/day	Genistein or placebo capsule	↓Total cholesterol, LDL-C, and TG levels ↑HDL-C levels ↑Expression of LDLR, LXRα, and ABCG1 ↓Cholesterol	[56]

atherosclerotic risk and lowering platelet aggregation. Genistein decreased collagen- and ADP-dependent platelet activation and inhibited agonist-induced platelet aggregation by dose-dependent manner [52]. In a human study, administration of Gen (0.5–2.5 microM) inhibited Cu (⁺⁺)-induced lipid peroxidation (LP) of HDL and increased the levels of conjugated dienes in lipoproteins oxidized. In addition, genistein blocked the alterations of the structure and physico-chemical properties of apoprotein, related to Cu (⁺⁺) triggered LP of lipoproteins [53].

Supplementation with genistein (90 mg/day), vitamin D, and calcium in healthy postmenopausal women in clinical double blind study indicated variant allele rs9340799, rs928554, and rs4986938, and estrogen receptors (ERs) polymorphisms were associated with a greater reduction in LDL cholesterol, total cholesterol, and triglyceride levels. These results indicated the relationship between ER genes polymorphisms and improvement in the serum lipid profile after treatment with genistein in postmenopausal women [54].

The results of a placebo-controlled trial on postmenopausal women (n = 389) with low bone mass showed treatment with of genistein aglycone (54 mg) for 24 months significantly declined fasting glucose and insulin, homocysteine, and fibrinogen. Genistein did not affect HDL cholesterol and triglycerides through serum osteoprotegerin. These results showed that a healthy diet and treatment with genistein aglycone plus calcium showed beneficial effects on some CVD risk factors and homocysteine levels in low bone mass postmenopausal women [55].

Treatment of hyperlipidemic postmenopausal women in China with genistein (60 mg/day) for six months significantly lowered TC, TG, and LDL cholesterol levels, while HDL cholesterol levels as well as mRNA expression and protein levels of LDLR, and liver X receptor α (LXR α) were markedly increased [56]. The effects of genistein on lipids profile are shown in Table 3.

3.4. The Effects of Genistein on Hypertension

3.4.1. In Vitro Studies. The effects of genistein on the vascular calcification in rat monocytes, osteoblasts cultures, and aortic vascular cell (in vitro) were evaluated. The mRNA expression of intercellular adhesion molecule-1 was reduced in the cells exposed to genistein (10 nM or 1 μ M). Also, the cell proliferation and migration on vascular muscle cells were also markedly reduced.

Both osteoblastic markers such as deposition of calcium nodules and alkaline phosphatase activity were markedly reduced after genistein treatment [57].

The acute treatment of thoracic aortae was isolated from adult male SHR rats with genistein ($10\,\mu\text{M}$) restores the impaired endothelial functions in the aorta.

Genistein transactivates epidermal growth factor receptor (EGFR) through membrane $ER\alpha$ via G protein-coupled pathways [58]. This is enhanced eNOS phosphorylation and hence endothelial function in the aorta. Genistein treatment (50 micromol/l) inhibited the expressions of

p22phox NADPH oxidase subunit and angiotensin II type 1 receptor (AT1) in aortic endothelial cells from stroke-prone SHR. Genistein also significantly reduced the angiotensin II induced superoxide by reduction of nitroblue tetrazolium, endothelin-1 production, and inhibition of nitrotyrosine formation [59].

3.4.2. In Vivo Studies. The effects of genistein (0.1-30 micromol/L) on aortic smooth muscle cells (SMC) from hypertensive rats showed inhibitory effects on proliferation and DNA synthesis of SMC in a dose-dependent manner. It also showed inhibitory effects on platelet-derived growth factor (PDGF) induced aortic SMC proliferation [60]. These results indicated that genistein treatment may be useful for reducing cell proliferation that involved in atherosclerotic vascular changes. Administration of genistein for 10 days (1 mg/kg) significantly improved lung and heart function by reduction in right ventricular pressure and restored right ventricular ejection fraction similar to the control group. Genistein also reversed pulmonary hypertension (PH)-induced pulmonary vascular remodeling (in vivo) and inhibited human pulmonary artery SMC proliferation (*in vitro*) likely through treatment with estrogen receptor- β . It also reversed right ventricular (RV) hypertrophy, inhibited neonatal rat ventricular myocyte hypertrophy, and restored PH-induced loss of capillaries in the RV [30]. The results indicated that genistein prevents the progression of severe PH to right heart failure.

Treatment of fructose-fed hypertensive rats with genistein (1 mg/kg) lowered blood pressure (BP), restored angiotensin-converting enzyme (ACE), protein kinase C- β II (PKC- β II), and expression of endothelial NO synthase (eNOS), and preserved renal ultrastructural integrity [61].

Effects of genistein on pulmonary arterial hypertension (PAH) in monocrotaline-induced rat model were also studied. Genistein (20, 80, and $200\,\mu\text{g/kg}$) significantly improved diameter of pulmonary artery, speed of tricuspid regurgitation, mean pulmonary artery pressure, and RV hypertrophy index. Treatment with genistein also improved proliferation of smooth muscle, stenosis of pulmonary artery, RV hypertrophy, and myocardial hypertrophy. In rat lung tissue, phosphorylated protein kinase B (P-Akt), phosphorylated eNOS, and expressions of nitric oxide (NO) were remarkably increased in the genistein treatment group compared to the PAH group [62].

The genistein fed diet (600 mg/kg food) remarkably less weight than male's mice fed the genistein-free diet (0G). The Genistein fed diet also exhibited remarkably elevated serum insulin and decreased serum glucose levels. Basal vascular reactivity of isolated aortic rings (IAR) was increased remarkably by the genistein diet in both males and females mice [63].

Administration of genistein (1 mg/kg) on lipopolysaccharide (LPS)-induced rat cardiovascular abnormalities causes to reduction in contractile response to noradrenaline (NA) in vascular tissue, and inhibition in hyporesponsiveness to NA, increasing in nitrite plasma levels, and iNOS expression production by endotoxin. In addition,

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	TABLE 4: THE EHECIS OF BEHISTEIN ON HYPETTENSION.	ın on nypertension.		
Study Design	Doses	Route of administration	Effects	Ref
Postmenopausal rat monocytes, aortic vascular cell, and osteoblasts cultures	$10\mathrm{nM}$ or $1\mu\mathrm{M}$	Exposed	↓Cell adhesion molecules ↓Expression of intercellular adhesion molecule 1 ↓Osteoblastic ↑Antiosteogenic action ↑Bone cells ↑Alkaline phosphatase activity	[57]
Rats	10 microM	Exposure	↓Contractions ↓Acetylcholine and A23187-induced relaxation	[58]
Rats	50 micromol/l	Exposed	LExpressions of the angiotensin II (Ang II) type 1 (ATI) receptor Lp22phox NADPH oxidase subunit LAngiotensin II-induced superoxide Unitroblue tetrazolium LNitrotyrosine formation Angiotensin II LP22phox and ATI receptor expression	[59]
Rats	0.1–30 micromol/L	SMC were cultured in dishes	↓Proliferation of natural and PDGF-BB-induced SMC ↓Atherosclerotic cardiovascular diseases	[09]
Rats	1 mg/kg/day	Daily injection of genistein	Usight ventricular pressure ↓Pulmonary vascular remodeling ↓50% in human pulmonary artery smooth muscle cell proliferation ↓Hypertrophy of right ventricular ↓Hypertrophy of ventricular myocyte in neonatal rat	[30]
Wistar rats	1 mg/kg/day	Feed a diet	Uslood pressure Restored PKC-βII, eNOS, and ACE expression [61] Preserved ultrastructural integrity of renal	[61]
Rats	20,80,200 μg/kg	Subcutaneous injection for two weeks	LSpeed of tricuspid regurgitation LVentricular hypertrophy index LPulmonary artery stenosis LSmooth muscle proliferation LHypertrophy of right ventricular LHypertrophy of myocardia LPulmonary artery pressure	[62]

TABLE 4: Continued.

	IABLE 4. COILIILICA	מני		
Study Design	Doses	Route of administration	Effects	Ref
C57BL/6] mice	600 mg/kg food	Genistein-containing diet 1 month	↓Elevated serum insulin ↓Serum glucose ↑Insulin to glucose ratio ↑Isolated aortic rings in both sex ↑Vascular function	[63]
In vivo rats	1 mg/kg	Injection	↓Contractile response ↓Hyporesponsiveness to NA ↑Nitrite plasma levels ↑Nitrite plasma levels ↑INOS expression ↓Long-term hypotension ↓Hypotension and vascular alterations ↓NO overproduction ↑Tyrosine kinase inhibitor effect	[64]
Rats	10 nM	Exposed	↑Antiaggregatory action ↓Platelet aggregation ↑Antiplatelet activity	[65]
Rats	Low-dose $(2/5 \mu g/kg)$ and high-dose $(25 \mu g/kg)$	Subcutaneous treatments	↓Angiotensin II (46%)	[99]
Rats	Low-dose (20 $\mu g/kg$) and high-dose (80 $\mu g/kg$)	I	JPhosphorylation of tyrosine ↓Renal arteries contraction ↓Elevated mean pulmonary arterial pressure ↑HO-1 expression ↓Pulmonary arterial hypertension ↑HO-1 in the lung tissues	[67]
Rats	Normal amounts of genistein in diet (0.06% (wt/wt))	Soy-based diet	↓Hypertension sensitive to NaCl in young ↓Stroke-prone SHRs in male ↓Insulin Plasma ↓Insulin resistance in SHR-SP in NaCl rich [68] diet ↓Cholesterol Plasma ↓Triglycerides	[89]
Rats	(Wistar-Kyoto 10 mg/kg/day); SHR-genistein-faslodex (2.5 mg/kg/day)	Vehicle containing	Lystolic blood pressure ↑Aortic calmodulin-1 protein ↑NOS activity LO2 (-) production LElevated blood pressure ↑ENOS activity LO2 (-) generation	[69]

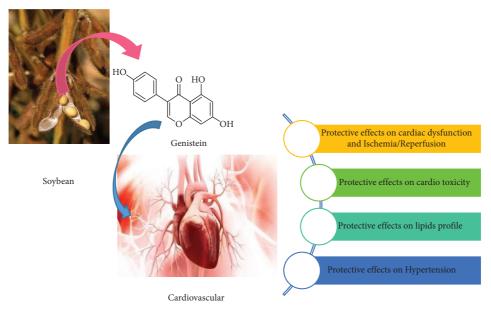


FIGURE 2: Possible therapeutic effects of genistein on cardiovascular diseases.

genistein restored lipid peroxidation, impaired aortic relaxation to acetylcholine, and suppressed long-term hypotension [64]. This result indicated that genistein prevented vascular alterations and hypotension induced by LPS that was mediated by its anti-inflammatory and antioxidant effects as well as inhibitory effect on NO overproduction. The effects of 10 nM genistein in rat aortic strips significantly increased nitric oxide synthesis. It also exhibited an antiaggregatory action, dependent on the NO release from vascular tissue, and suppressed the inhibition of platelet aggregation. Moreover, genistein inhibited platelet aggregation in aortic strips from ovariectomized rats [65].

The vascular effects of genistein in ovariectomized spontaneously hypertensive rats (SHR) were evaluated. Animals treated with genistein (2.5 and 25 mg/kg), for 2 days or 2 weeks, and contractility of the renal arterial rings were studied. The tissue contractions and tyrosine phosphorylation were reduced by the genistein (2.5 mg/kg) for 2 days [66].

Genistein treatment (20 and $80\,\mu\text{g/kg}$) on heme oxygenase-1 (HO-1) expression in PAH induced by monocrotaline (MCT) in rats as dose dependently improved the RV hypertrophy index, increased the expression of HO-1, and decreased the elevated mean PA pressure [67].

Dietary genistein (0.06%, wt/wt) decreased NaCl-sensitive hypertension in young, male stroke-prone spontaneously hypertensive rats (SHRs). Genistein also declined plasma insulin and insulin resistance in SHR on a high NaCl diet and reduced plasma triglycerides and cholesterol in SHR. The antihypertensive effect of genistein was influenced by the autonomic nervous system [68].

Treatment of 23-week-old rats with genistein (10 mg kg) reduced NADPH-induced O_2 ($^-$) production and systolic blood pressure (SBP) and enhanced endothelium-dependent aortic relaxation to acetylcholine

and increased aortic calmodulin-1 protein abundance and eNOS activity [69]. The beneficial effects of genistein on blood pressure and endothelial dysfunction may be due to increased eNOS activity associated with increased calmodulin-1 expression and declined O_2 ($\bar{}$) generation. The effects of genistein on hypertension are shown in Table 4. The possible therapeutic effects of genistein on CVDs are shown in Figure 2.

4. Conclusion

In vitro and in vivo studies reported potential therapeutic effects of genistein on cardiovascular disorders including improving cardiac dysfunction, ischemia and reperfusion of heart, decreasing cardiac toxicity, modulating lipid profile, and lowering the blood pressure which are reviewed in the current article. The preventive or prophylactic effects of genistein on cardiovascular disorders may be due to anti-inflammatory, antioxidant, and immunomodulatory effects. Although the effect of genistein on CVD is well shown in the reviewed studies, while further standard clinical trials and meta-analyzing the previous results are needed to be performed.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

Authors' Contributions

SJ and MS searched in literature and prepared the original draft. MRK contributed to the study design and critically

revised the manuscript. All authors read and approved the final version of the manuscript.

References

- [1] E. J. Benjamin, M. J. Blaha, S. E. Chiuve et al., "Heart disease and stroke statistics—2017 update: a report from the American Heart Association," *Circulation*, vol. 135, no. 10, pp. e146–e603, 2017.
- [2] A. Shaito, D. T. B. Thuan, H. T. Phu et al., "Herbal medicine for cardiovascular diseases: efficacy, mechanisms, and safety," *Frontiers in Pharmacology*, vol. 11, p. 422, 2020.
- [3] S. A. Patel, M. Winkel, M. K. Ali, K. V. Narayan, and N. K. Mehta, "Cardiovascular mortality associated with 5 leading risk factors: national and state preventable fractions estimated from survey data," *Annals of Internal Medicine*, vol. 163, no. 4, pp. 245–253, 2015.
- [4] G. D. Flora and M. K. Nayak, "A brief review of cardiovascular diseases, associated risk factors and current treatment regimes," *Current Pharmaceutical Design*, vol. 25, no. 38, pp. 4063–4084, 2019.
- [5] D. S. Fabricant and N. R. Farnsworth, "The value of plants used in traditional medicine for drug discovery," *Environ*mental Health Perspectives, vol. 109, no. 1, pp. 69–75, 2001.
- [6] G. M. Cragg and D. J. Newman, "Natural products: a continuing source of novel drug leads," *Biochimica et Biophysica Acta (BBA) General Subjects*, vol. 1830, no. 6, pp. 3670–3695, 2013.
- [7] L. Ciumărnean, M. V. Milaciu, O. Runcan et al., "The effects of flavonoids in cardiovascular diseases," *Molecules*, vol. 25, no. 18, p. 4320, 2020.
- [8] D. Altavilla, A. Crisafulli, H. Marini et al., "Cardiovascular effects of the phytoestrogen genistein," *Current Medicinal Chemistry - Cardiovascular and Hematological Agents*, vol. 2, no. 2, pp. 179–186, 2004.
- [9] A. Sureda, A. Sanches Silva, D. I. Sánchez-Machado et al., "Hypotensive effects of genistein: from chemistry to medicine," *Chemico-Biological Interactions*, vol. 268, pp. 37–46, 2017
- [10] M. R. Khazdair, A. Anaeigoudari, and G. A. Agbor, "Anti-viral and anti-inflammatory effects of kaempferol and quercetin and COVID-2019: a scoping review," *Asian Pacific Journal of Tropical Biomedicine*, vol. 11, no. 8, p. 327, 2021.
- Tropical Biomedicine, vol. 11, no. 8, p. 327, 2021.
 [11] M. Kianmehr and M. R. Khazdair, "Possible therapeutic effects of Crocus sativus stigma and its petal flavonoid, kaempferol, on respiratory disorders," *Pharmaceutical Biology*, vol. 58, no. 1, pp. 1149–1158, 2020.
- [12] M. R. Khazdair, M. Kianmehr, and A. Anaeigoudari, "Effects of medicinal plants and flavonoids on Parkinson's disease: a review on basic and clinical evidences," *Advanced Phar-maceutical Bulletin*, vol. 11, no. 2, pp. 224–232, 2021.
- [13] A. Amerizadeh, S. Asgary, G. Vaseghi, and F. Ziba, "Effect of genistein intake on some cardiovascular risk factors: an updated systematic review and meta-analysis," *Current Problems in Cardiology*, vol. 47, no. 9, Article ID 100902, 2022.
- [14] R. A. Dixon and D. Ferreira, "Genistein," *Phytochemistry*, vol. 60, no. 3, pp. 205–211, 2002.
- [15] M. Zhang, K. Wang, L. Chen, B. Yin, and Y. Song, "Is phytoestrogen intake associated with decreased risk of prostate cancer? A systematic review of epidemiological studies based on 17,546 cases," *Andrology*, vol. 4, no. 4, pp. 745–756, 2016.
- [16] J. P. Rigalli, G. N. Tocchetti, M. R. Arana et al., "The phytoestrogen genistein enhances multidrug resistance in breast

- cancer cell lines by translational regulation of ABC transporters," *Cancer Letters*, vol. 376, no. 1, pp. 165–172, 2016.
- [17] K. D. Setchell and E. Lydeking-Olsen, "Dietary phytoestrogens and their effect on bone: evidence from in vitro and in vivo, human observational, and dietary intervention studies," *American Journal of Clinical Nutrition*, vol. 78, no. 3, pp. 593S-609S, 2003.
- [18] Y.-N. Shi, X.-Q. Zhang, Z.-Y. Hu et al., "Genistein protects H9c2 cardiomyocytes against chemical hypoxia-induced injury via inhibition of apoptosis," *Pharmacology*, vol. 103, no. 5-6, pp. 282–290, 2019.
- [19] S. H. Lee, J. H. Lee, T. Asahara et al., "Genistein promotes endothelial colony-forming cell (ECFC) bioactivities and cardiac regeneration in myocardial infarction," *PLoS One*, vol. 9, no. 5, Article ID e96155, 2014.
- [20] W.-S. Hu, Y.-M. Lin, T.-J. Ho et al., "Genistein suppresses the isoproterenol-treated H9c2 cardiomyoblast cell apoptosis associated with P-38, erk1/2, JNK, and NF κ B signaling protein activation," *The American Journal of Chinese Medicine*, vol. 41, no. 05, pp. 1125–1136, 2013.
- [21] C. Huang, Y. Zhang, H. Qi, X. Xu, L. Yang, and J. Wang, "Myc is involved in Genistein protecting against LPS-induced myocarditis in vitro through mediating MAPK/JNK signaling pathway," *Bioscience Reports*, vol. 40, no. 6, 2020.
- [22] S. Farruggio, G. Raina, G. Cocomazzi et al., "Genistein improves viability, proliferation and mitochondrial function of cardiomyoblasts cultured in physiologic and peroxidative conditions," *International Journal of Molecular Medicine*, vol. 44, no. 6, pp. 2298–2310, 2019.
- [23] G. A. Colareda and A. E. Consolini, "Low-flow ischaemia and reperfusion in rat hearts: energetic of stunning and cardioprotection of genistein," *Journal of Pharmacy and Pharmacology*, vol. 70, no. 9, pp. 1174–1187, 2018.
- [24] L. Al-Nakkash, B. Markus, K. Bowden, L. M. Batia, W. C. Prozialeck, and T. L. Broderick, "Effects of acute and 2day genistein treatment on cardiac function and ischemic tolerance in ovariectomized rats," *Gender Medicine*, vol. 6, no. 3, pp. 488–497, 2009.
- [25] S. K. Maulik, P. Prabhakar, A. K. Dinda, and S. Seth, "Genistein prevents isoproterenol-induced cardiac hypertrophy in rats," *Canadian Journal of Physiology and Phar*macology, vol. 90, no. 8, pp. 1117–1125, 2012.
- [26] G. A. Colareda, M. I. Ragone, P. Bonazzola, and A. E. Consolini, "The mKATP channels and protein-kinase C are involved in the cardioprotective effects of genistein on estrogen-deficient rat hearts exposed to ischemia/reperfusion: energetic study," *Journal of Cardiovascular Pharmacology*, vol. 75, no. 5, pp. 460–474, 2020.
- [27] W. Qin, N. Du, L. Zhang et al., "Genistein alleviates pressure overload-induced cardiac dysfunction and interstitial fibrosis in mice," *British Journal of Pharmacology*, vol. 172, no. 23, pp. 5559–5572, 2015.
- [28] J. Thorburn and A. Thorburn, "The tyrosine kinase inhibitor, genistein, prevents α-adrenergic-induced cardiac muscle cell hypertrophy by inhibiting activation of the ras-MAP kinase signaling pathway," *Biochemical and Biophysical Research Communications*, vol. 202, no. 3, pp. 1586–1591, 1994.
- [29] Y. Meng, Y. Zhang, Z. Ma et al., "Genistein attenuates pathological cardiac hypertrophy in vivo and in vitro," *Herz*, vol. 44, no. 3, pp. 247–256, 2019.
- [30] H. Matori, S. Umar, R. D. Nadadur et al., "Genistein, a soy phytoestrogen, reverses severe pulmonary hypertension and prevents right heart failure in rats," *Hypertension*, vol. 60, no. 2, pp. 425–430, 2012.

- [31] E. Grossini, C. Molinari, D. A. Mary et al., "Intracoronary genistein acutely increases coronary blood flow in anesthetized pigs through β -adrenergic mediated nitric oxide release and estrogenic receptors," *Endocrinology*, vol. 149, no. 5, pp. 2678–2687, 2008.
- [32] B. Deodato, D. Altavilla, G. Squadrito et al., "Cardioprotection by the phytoestrogen genistein in experimental myocardial ischaemia-reperfusion injury," *British Journal of Pharmacology*, vol. 128, no. 8, pp. 1683–1690, 1999.
- [33] C. De Gregorio, H. Marini, A. Alibrandi et al., "Genistein supplementation and cardiac function in postmenopausal women with metabolic syndrome: results from a pilot strainecho study," *Nutrients*, vol. 9, no. 6, p. 584, 2017.
- [34] S. Nabavi, M. Daglia, R. Tundis et al., "Genistein: a boon for mitigating ischemic stroke," *Current Topics in Medicinal Chemistry*, vol. 15, no. 17, pp. 1714–1721, 2015.
- [35] E. Sbarouni, E. K. Iliodromitis, A. Zoga, G. Vlachou, I. Andreadou, and D. T. Kremastinos, "The effect of the phytoestrogen genistein on myocardial protection, preconditioning and oxidative stress," *Cardiovascular Drugs and Therapy*, vol. 20, no. 4, pp. 253–258, 2006.
- [36] L. Leung, J. B. Martin, T. Lawmaster, K. Arthur, T. L. Broderick, and L. Al-Nakkash, "Sex-dependent effects of dietary genistein on echocardiographic profile and cardiac GLUT4 signaling in mice," *Evidence-based Complementary* and Alternative Medicine, vol. 2016, Article ID 1796357, 10 pages, 2016.
- [37] Y. Meng, Y. Zhang, Z. Ma et al., "Genistein attenuates pathological cardiac hypertrophy in vivo and in vitro," *Herz*, vol. 44, 2019.
- [38] K. K. Hintz and J. Ren, "Phytoestrogenic isoflavones daidzein and genistein reduce glucose-toxicity-induced cardiac contractile dysfunction in ventricular myocytes," *Endocrine Re*search, vol. 30, no. 2, pp. 215–223, 2004.
- [39] C.-E. Chiang, H.-N. Luk, T.-M. Wang, J. R. Sheu, and P. Y. A. Ding, "Genistein and tyrphostin AG 556 block the action potential shortening in septic shock," *Zhonghua yi xue za zhi= Chinese Medical Journal; Free China ed*, vol. 65, no. 12, pp. 570–579, 2002.
- [40] Z. Bai and Z. Wang, "Genistein protects against doxorubicininduced cardiotoxicity through Nrf-2/HO-1 signaling in mice model," *Environmental Toxicology*, vol. 34, no. 5, pp. 645–651, 2019
- [41] S. K. Gupta, S. Dongare, R. Mathur et al., "Genistein ameliorates cardiac inflammation and oxidative stress in streptozotocin-induced diabetic cardiomyopathy in rats," *Molecular and Cellular Biochemistry*, vol. 408, no. 1-2, pp. 63–72, 2015.
- [42] Z. Fang, G. Wu, D. Zhang et al., "Genistein protects against burn-induced myocardial injury via Notch1-mediated suppression of oxidative/nitrative stress," *Shock*, vol. 54, no. 3, pp. 337–346, 2020.
- [43] R. Lu, Z. Zheng, Y. Yin, and Z. Jiang, "Effect of genistein on cholesterol metabolism-related genes in HepG2 cell," *Journal of Food Science*, vol. 84, no. 8, pp. 2330–2336, 2019.
- [44] H. Zhang, X. Yang, X. Pang, Z. Zhao, H. Yu, and H. Zhou, "Genistein protects against ox-LDL-induced senescence through enhancing SIRT1/LKB1/AMPK-mediated autophagy flux in HUVECs," *Molecular and Cellular Biochemistry*, vol. 455, no. 1-2, pp. 127–134, 2019.
- [45] A. Yousefinejad, F. Siassi, M. H. Javanbakht et al., "Effect of genistein and L-carnitine and their combination on lipid profile and inflammatory cytokines in experimental nephrotic

- syndrome," Reports of biochemistry and molecular biology, vol. 7, no. 1, pp. 1-8, 2018.
- [46] S. Amanat, F. Ashkar, M. H. Eftekhari et al., "The effect of genistein on insulin resistance, inflammatory factors, lipid profile, and histopathologic indices in rats with polycystic ovary syndrome," *Clinical and experimental reproductive* medicine, vol. 48, no. 3, pp. 236–244, 2021.
- [47] E. Agacayak, S. Basaranoglu, S. Y. Tunc et al., "Oxidant/antioxidant status, paraoxonase activity, and lipid profile in plasma of ovariectomized rats under the influence of estrogen, estrogen combined with progesterone, and genistein," *Drug Design, Development and Therapy*, vol. 9, pp. 2975–2982, 2015.
- [48] J.-S. Lee, "Effects of soy protein and genistein on blood glucose, antioxidant enzyme activities, and lipid profile in streptozotocin-induced diabetic rats," *Life Sciences*, vol. 79, no. 16, pp. 1578–1584, 2006.
- [49] L. L. Legette, W.-H. Lee, B. R. Martin et al., "Genistein, a phytoestrogen, improves total cholesterol, and Synergy, a prebiotic, improves calcium utilization, but there were no synergistic effects," *Menopause (New York, NY)*, vol. 18, no. 8, pp. 923–931, 2011.
- [50] B. Sosić-Jurjević, B. Filipović, V. Ajdzanović et al., "A brief communication: subcutaneously administrated genistein and daidzein decrease serum cholesterol and increase triglyceride levels in male middle-aged rats," *Experimental Biology and Medicine*, vol. 232, no. 9, pp. 1222–1227, 2007.
- [51] C. Tang, K. Zhang, Q. Zhao, and J. Zhang, "Effects of dietary genistein on plasma and liver lipids, hepatic gene expression, and plasma metabolic profiles of hamsters with diet-induced hyperlipidemia," *Journal of Agricultural and Food Chemistry*, vol. 63, no. 36, pp. 7929–7936, 2015.
- [52] K. Borgwardt, S. Bonifatius, and A. Gardemann, "Acidic peptides enhanced genistein-dependent inhibition of human platelet aggregation: potential protective effect of digestible peptides plus genistein against atherosclerosis," *Nutrition Research*, vol. 28, no. 8, pp. 523–531, 2008.
- [53] G. Ferretti, T. Bacchetti, F. Menanno, and G. Curatola, "Effect of genistein against copper-induced lipid peroxidation of human high density lipoproteins (HDL)," *Atherosclerosis*, vol. 172, no. 1, pp. 55–61, 2004.
- [54] R. Usategui-Martín, M. Pérez-Alonso, L. Socorro-Briongos et al., "Estrogen receptor genes polymorphisms determine serum lipid profile in healthy postmenopausal women treated with calcium, vitamin D, and genistein," *Journal of Cellular Biochemistry*, vol. 120, no. 8, pp. 13115–13120, 2019.
- [55] H. Marini, A. Bitto, D. Altavilla et al., "Efficacy of genistein aglycone on some cardiovascular risk factors and homocysteine levels: a follow-up study," *Nutrition, Metabolism, and Cardiovascular Diseases*, vol. 20, no. 5, pp. 332–340, 2010.
- [56] S. Zhang and Y. Wang, "Network pharmacology approach reveals the potential immune function activation and tumor cell apoptosis promotion of xia qi decoction in lung cancer," *Medical sciences (Basel, Switzerland)*, vol. 8, no. 1, pp. 1–8, 2019.
- [57] S. B. Cepeda, M. J. Sandoval, M. B. Rauschemberger, and V. L. Massheimer, "Beneficial role of the phytoestrogen genistein on vascular calcification," *The Journal of Nutritional Biochemistry*, vol. 50, pp. 26–37, 2017.
- [58] A. H. Lin, G. P. Leung, S. W. Leung, P. M. Vanhoutte, and R. Y. Man, "Genistein enhances relaxation of the spontaneously hypertensive rat aorta by transactivation of epidermal growth factor receptor following binding to membrane estrogen receptors- α and activation of a G protein-coupled,

- endothelial nitric oxide synthase-dependent pathway," *Pharmacological Research*, vol. 63, no. 3, pp. 181–189, 2011.
- [59] J.-W. Xu, K. Ikeda, and Y. Yamori, "Genistein inhibits expressions of NADPH oxidase p22phox and angiotensin II type 1 receptor in aortic endothelial cells from stroke-prone spontaneously hypertensive rats," *Hypertension Research*, vol. 27, no. 9, pp. 675–683, 2004.
- [60] W. Pan, K. Ikeda, M. Takebe, and Y. Yamori, "Genistein, daidzein and glycitein inhibit growth and DNA synthesis of aortic smooth muscle cells from stroke-prone spontaneously hypertensive rats," *The Journal of Nutrition*, vol. 131, no. 4, pp. 1154–1158, 2001.
- [61] N. Palanisamy and A. C. Venkataraman, "Beneficial effect of genistein on lowering blood pressure and kidney toxicity in fructose-fed hypertensive rats," *The British Journal of Nutrition*, vol. 109, no. 10, pp. 1806–1812, 2013.
- [62] Z. Zheng, S. Yu, W. Zhang et al., "Genistein attenuates monocrotaline-induced pulmonary arterial hypertension in rats by activating PI3K/Akt/eNOS signaling," *Histology and Histopathology*, vol. 32, no. 1, pp. 35–41, 2017.
- [63] L. Al-Nakkash, J. B. Martin, D. Petty et al., "Dietary genistein induces sex-dependent effects on murine body weight, serum profiles, and vascular function of thoracic aortae," *Gender Medicine*, vol. 9, no. 5, pp. 295–308, 2012.
- [64] A. Bermejo, A. Zarzuelo, and J. Duarte, "In vivo vascular effects of genistein on a rat model of septic shock induced by lipopolysaccharide," *Journal of Cardiovascular Pharmacology*, vol. 42, no. 3, pp. 329–338, 2003.
- [65] N. Polini, M. B. Rauschemberger, J. Mendiberri, J. Selles, and V. Massheimer, "Effect of genistein and raloxifene on vascular dependent platelet aggregation," *Molecular and Cellular En*docrinology, vol. 267, no. 1-2, pp. 55–62, 2007.
- [66] R. Nevala, M. Lassila, P. Finckenberg, K. Paukku, R. Korpela, and H. Vapaatalo, "Genistein treatment reduces arterial contractions by inhibiting tyrosine kinases in ovariectomized hypertensive rats," *European Journal of Pharmacology*, vol. 452, no. 1, pp. 87–96, 2002.
- [67] Y. Zhang, D. Wang, T. Zhu, and C. Li, "Genistein attenuates monocrotaline-induced pulmonary arterial hypertension in rats by up-regulating heme oxygenase-1 expression," Nan Fang yi ke da xue xue bao= Journal of Southern Medical University, vol. 32, no. 2, pp. 151–154, 2012.
- [68] T. M. Cho, N. Peng, J. T. Clark et al., "Genistein attenuates the hypertensive effects of dietary NaCl in hypertensive male rats," *Endocrinology*, vol. 148, no. 11, pp. 5396–5402, 2007.
- [69] R. Vera, M. Sánchez, M. Galisteo et al., "Chronic administration of genistein improves endothelial dysfunction in spontaneously hypertensive rats: involvement of eNOS, caveolin and calmodulin expression and NADPH oxidase activity," Clinical Science, vol. 112, no. 3, pp. 183–191, 2007.