

Research Article

Imidazole and Azo-Based Schiff Bases Ligands as Highly Active Antifungal and Antioxidant Components

Siham Slassi,^{1,2} Adeline Fix-Tailler,³ Gérald Larcher,³
Amina Amine ,² and Abdelkrim El-Ghayoury ¹

¹Université d'Angers, CNRS UMR 6200, Laboratoire MOLTECH-Anjou, 2 bd Lavoisier, 49045 Angers Cedex, France

²LCBAE, Equipe Chimie Moléculaire et Molécules Bioactives, Université Moulay Ismail, Faculté des Sciences Meknès, Morocco

³Université d'Angers, Groupe d'Etude des Interactions Hôte-Pathogène (GEIHP, EA 3142), Institut de Biologie en Santé-IRIS, CHU, 4 rue Larrey, 49933 Angers Cedex, France

Correspondence should be addressed to Amina Amine; amine_7a@yahoo.fr
and Abdelkrim El-Ghayoury; abdelkrim.elghayoury@univ-angers.fr

Received 12 October 2018; Accepted 4 November 2018; Published 3 January 2019

Academic Editor: Jacek Skarzewski

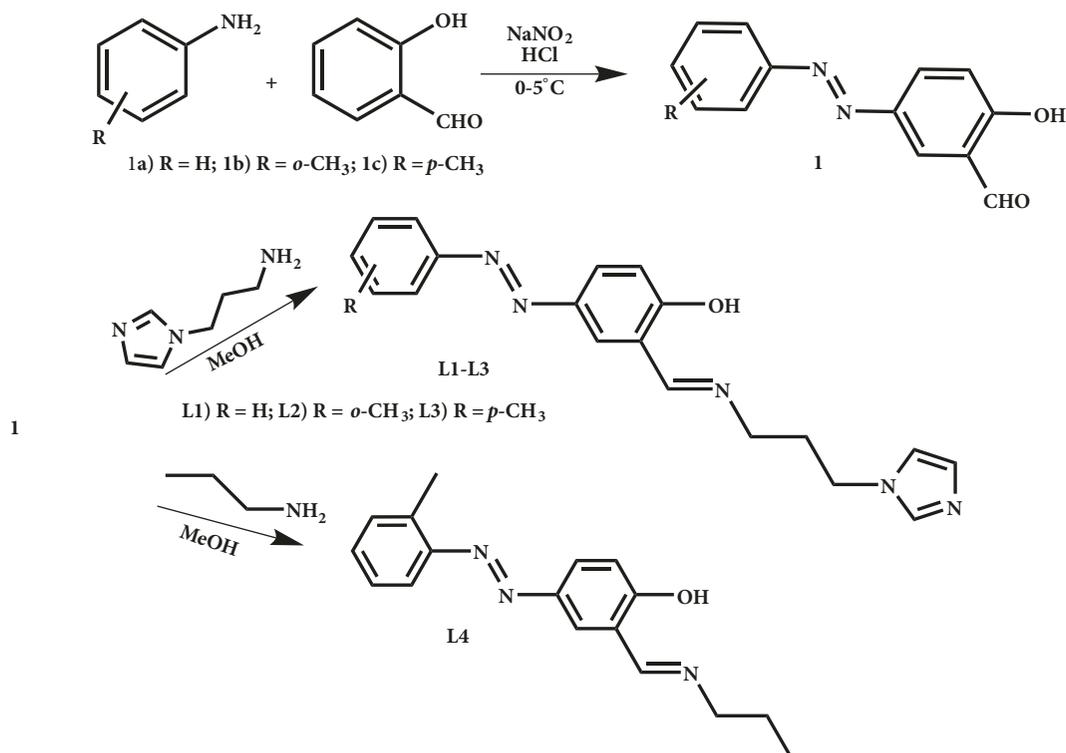
Copyright © 2019 Siham Slassi et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

We describe, herein, the synthesis, full characterization, and optical properties of four different ligands **L1-L4** which associate an azo group, an imidazole unit, and a Schiff base fragment. The UV-visible absorption bands are characteristic of $\pi \rightarrow \pi^*$ and $n \rightarrow \pi^*$ transitions with an additional charge transfer between the azobenzene moiety and the imino group. Finally the determination of MIC₈₀ values against pathogenic fungi such as *S. apiospermum*, *A. fumigatus*, and *C. albicans* revealed that these ligands have effective antifungal properties with highest activities (MIC₈₀) on *C. albicans* for the azole based ligands **L1-L3**. DPPH radical scavenging of the studied ligands was also tested.

1. Introduction

Imidazole derivatives constitute an important class of heterocycles being the core fragment of different natural products and biological systems. They occupy a unique place in the field of medicinal chemistry owing to their potent biological activity [1]. They are well known to possess many pharmacological properties and to play a very important role in diverse biochemical processes [2, 3]. Many substituted imidazoles display wide range of biological applications such as antiprotozoal, antifungal, and antihypertensive agents [4–6]. In addition, imidazoles constitute an entire or partial part of the binding sites of various transition metal ions such as Ni²⁺, Cu²⁺, or Zn²⁺ in a large number of metalloproteins and have thus been utilized as effective ligands to chelate these transition metals. For example, imidazole based ligands have been used to afford tetranuclear copper(II) and nickel(II) metal complexes, [7] copper(II) or zinc(II) coordination polymers, [8] 1D coordination chains, [9] tripodal metal complexes, [10] and many other complexes used for treatment of Wilson and Menkes diseases [11]. On the other hand,

azobenzene [12, 13] and its numerous derivatives are currently used as dyes and represent 60–70 % of the world production of “absorbing molecules” [14]. They are known to possess a unique photoisomerization process which leads to two stable *cis* and *trans* geometries. The *trans*-to-*cis* photoisomerization occurs by UV light irradiation while the reverse *cis*-to-*trans* takes place under blue light irradiation [15, 16]. This process is the key principle of various applications such as chemosensors, [17] optical storage media, [18] optical switches, [19, 20] nonlinear optics, [21] as well as trigger for protein folding [22]. Furthermore, azo compounds have also a variety of biological activities including antibacterial, [23] antifungal, [24] pesticidal, [25] antiviral, and anti-inflammatory activities [26]. Schiff bases ligands with chelating abilities have been recognized as privileged ligands to form stable complexes with a large variety of transition metals [27]. They have been used, for example, in metallo-ligand self-assembly to generate discrete supramolecular assemblies such as metallo-supramolecular helicates, [28–30] cages [31], and capsules, [32] and have also been used with a variety of transition metals in catalysis [33, 34]. In addition, Schiff base derivatives



SCHEME 1: Synthesis of the azo-based salicylaldehydes (**1a-c**) and the ligands (**L1-L4**).

showed a variety of biological and pharmacological activities as antimicrobial, [35] antidepressant, [36] cytotoxic, [37] analgesic, [38] anti-HIV, [39] antileishmanial, [40] anticonvulsant, [41] fungicides, [42] anti-inflammatory [43], and anticancer [44]. In this present investigation work, we are interested by the combination of the azo group, the imidazole unit and the Schiff base fragment. Thus, we report herein on the synthesis, characterization, and optical properties of four different Schiff bases ligands **L1-L4**. We also report the possible use of such systems in biological applications, in particular due to their antifungal properties and antioxidant activities. Note that the X-ray crystal structure of ligand **L2** has been recently described by us [45].

2. Results and Discussions

2.1. Synthesis. The synthesis of the Schiff bases was started by the preparation of the azo-based salicylaldehyde (**1a-c**) [46] by a coupling reaction between 2-hydroxybenzaldehyde and the substituted anilines as it was previously reported in the literature (Scheme 1). The resulting salicylaldehydes (**1a-c**) were subjected to condensation reaction with one equivalent of N-(3-Aminopropyl)imidazole or one equivalent of propylamine to afford in good yields the Schiff bases **L1-L4**.

2.2. UV-Visible Absorption Spectroscopy. Figure 1 shows the normalized UV-visible absorption spectra of the Schiff bases ligands **L1-L4** that were recorded in dichloromethane solution ($\sim 2 \cdot 10^{-5}$ M) at room temperature. Ligands **L1-L3** exhibit two strong electronic absorption bands at around $\lambda = 275$ nm and 350 nm while the two absorption bands of **L4** are

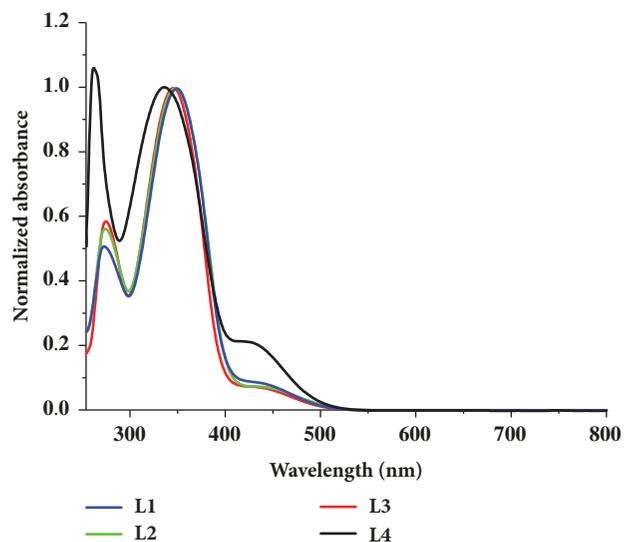


FIGURE 1: UV-visible absorption spectra of ligands **L1-L4** ($2 \cdot 10^{-5}$ M) in dichloromethane at room temperature.

blue shifted to $\lambda = 262$ nm and 335 nm. These absorption bands are assigned to $\pi \rightarrow \pi^*$ and $n \rightarrow \pi^*$ transitions in the azobenzene moiety and the imidazole rings for **L1-L3** and in the azobenzene fragment for **L4**. In the visible region the four ligands show an additional broad absorption band which might be assigned to a charge transfer band between the azobenzene moiety and the imino group.

TABLE 1: Antifungal activities determined on solid medium and expressed by the diameter of the growth inhibition zone (mm).

Compounds tested	Fungi		
	<i>S. apiospermum</i>	<i>A. fumigatus</i>	<i>C. albicans</i>
L1	32	28	17
L2	19	13	14.5
L3	21	33	14.5
L4	16.5	12	12.5
Miconazole	28.5	18.5	18.5

TABLE 2: Antifungal activities determined in liquid medium and expressed by ^aMIC₈₀ in $\mu\text{g}\cdot\text{mL}^{-1}$.

Compounds tested	Fungi		
	<i>S. apiospermum</i>	<i>A. fumigatus</i>	<i>C. albicans</i>
L1	4	1.5	0.8
L2	4	12	0.1
L3	4	3	0.2
L4	4	1.5	3
Amphotericin B	4	8	1

a: MIC₈₀ corresponds to the minimum inhibitory concentration of the compound that inhibits 80% of the fungal growth.

2.3. Biological Activities

2.3.1. Antifungal Activity. Antifungal activities of the ligands **L1** to **L4** were evaluated on three important fungal species pathogenic for humans: *Candida albicans*, *Aspergillus fumigatus*, and *Scedosporium apiospermum*, especially implicated in patients with cystic fibrosis [47]. First, for a rapid antifungal screening, we have used a disc diffusion method on solid medium. The results reported in Table 1 indicate that, concerning *C. albicans*, we notice that the ligands **L2**, **L3**, and **L4** exhibit an antifungal activity which is lower than the one of the reference antifungal miconazole, while ligand **L1** shows an antifungal activity with the same order of magnitude to the one of the reference miconazole. Against *S. apiospermum* all the ligands were active; three of them showed activities according to this ascending order **L4** < **L2** < **L3** but lower than the one of miconazole. Once again, **L1** had a growth inhibition zone diameter (32 mm) superior to the one of miconazole (28.5 mm). The best antifungal activities against *A. fumigatus* were found for two ligands **L1** and **L3** whose inhibition zone diameters of 28 and 33 mm, respectively, were much superior to the one of miconazole (18.5 mm). A weak activity on the growth of *A. fumigatus* was observed with **L2** and **L4**. In all the tested fungi, it appears clearly that ligand **L1** presents the best antifungal results, which clearly indicate that any additional substituent on the phenyl ring is not beneficial for the antifungal activity.

In liquid medium, the antifungal activities measured in triplicate were confirmed (Table 2). Against *S. apiospermum*, the four ligands had the same MIC₈₀ (4 $\mu\text{g}/\text{mL}$) which was equal to the one of the reference antifungal amphotericin B. For *A. fumigatus*, as observed on solid medium, **L1** and **L3** exerted significant antifungal activities (1.5 and 3 $\mu\text{g}/\text{mL}$, respectively) which were superior to the one of amphotericin B (8 $\mu\text{g}/\text{mL}$). It was also the case for **L4** (1.5 $\mu\text{g}/\text{mL}$) in contrary to the result obtained on solid medium. However, in

correlation with the solid medium, **L2** had a weaker effect on the growth of *A. fumigatus* in liquid medium. High antifungal activities were detected against *C. albicans* for three ligands in opposite ascending order of the MIC₈₀ values: **L1** (0.8 $\mu\text{g}/\text{mL}$) < **L3** (0.2 $\mu\text{g}/\text{mL}$) < **L2** (0.1 $\mu\text{g}/\text{mL}$). All of these values were lower than the MIC₈₀ value (1 $\mu\text{g}/\text{mL}$) of amphotericin B. **L4** had a weaker antifungal effect with a MIC₈₀ value three times more the value of amphotericin B. As for solid media **L1** had an excellent antifungal activity, although it is less than **L2** and **L3**. The fact that **L4** shows a weak effect clearly indicates that the presence of the imidazole unit is of great importance for such biological activities.

As previously described for thiosemicarbazone ligands, [48] we noticed certain discordance for few results between the solid and liquid media. For example, it was the case for **L4** against *A. fumigatus* and *S. apiospermum*. But we can notice that **L4** has a structural formula very different from the other three ligands **L1-L3**. In fact, the first three ligands contain an imidazole ring which is absent in ligand **L4**. As the disk diffusion method consists of test compounds in solid agar medium, we may hypothesize that steric hindrance and/or lipophilic character of the molecules alter their diffusion in such medium. All these effects favor the fungal growth. These results strengthen the interest of the broth microdilution method for a large scale evaluation of compounds, disk diffusion method being suitable for a rapid antifungal screening.

The weaker antifungal activities of **L4** observed against *C. albicans* may be explained by the lack of azole group in its formula. This group found in antifungals used in therapy is known to interact with the lanosterol 14 α -demethylase (or CYP51A1) which is implicated in the biosynthesis of the ergosterol, important component of the fungal plasmic membrane. Curiously in liquid medium, **L4** exerted a good activity against the other fungi *S. apiospermum* and *A. fumigatus* suggesting antifungal targets different from the ergosterol synthesis. As related in previous paper concerning

TABLE 3: The antioxidant activity of L1-L4 in DPPH.

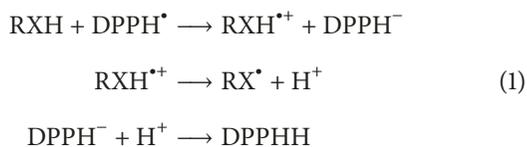
Compound	Concentration ($\mu\text{g/ml}$)						
	10	50	100	200	400	600	IC50
L1	39,12	44,48	51,62	64,28	76,29	88,63	95,98
L2	41,07	43,83	51,13	63,14	77,75	91,07	94,48
L3	35,87	46,1	51,13	64,61	83,44	91,55	96,01
L4	33,60	37,01	49,83	71,42	83,27	88,31	116,39
Ascorbic Acid	49,18	52,07	55,31	65,04	82,34	98,73	25,88
Blank	-	-	-	-	-	-	-

thiosemicarbazones, [48] there is the possibility that the activity may be linked to the relative lipophilicity of the molecule that makes the penetration in the fungal cell through the cell membrane and finally its destabilization easier. More generally for all the molecules tested, we may hypothesize that the capacity of these heterocyclic ligands to chelate metal ions which are essentials in numerous biological processes may impact the physiology of the fungi [49, 50].

Considering the need for newazole antifungals able to overcome the increase of life-threatening fungal infections and the emergence of resistant fungal isolates, our newazole ligands may be worthy of interest to develop new therapeutic agents.

2.3.2. Antioxidant Activity. Antioxidants are important bioactive species since they are capable of capturing free radicals responsible for many diseases such as cancer [51]. Due to the mobility of a proton (OH, NH, ...) in their structure, they act by direct scavenging of reactive oxygen species (ROS).

DPPH (2,2-Diphenyl-1-picrylhydrazyl) radical scavenging capacities of the Schiff bases L1-L4 were evaluated using UV-visible spectroscopy at different concentrations and compared to that of the well-known antioxidant ascorbic acid. Data summarized in Table 3 show that all the tested compounds exhibit an antioxidant activity although it is less than the ascorbic acid. This promising activity is probably due to the hydroxyl group that is commonly known to be the active group for DPPH scavenging [52]. Indeed, several studies [53] have established the relation between antiradical properties and the ability to transfer the H-atom of the OH group from antioxidants to free radicals according to the proposed three steps mechanism drawn below: [54]



We observed also that the tested ligands L1-L3, which structures differ only by the presence of a methyl group, gave similar inhibitory concentrations and that they are more active than the ligand L4 whose formula lacks the imidazole ring. This result led us to conclude that the presence of the imidazole ring significantly contributes to the bioactivity of the studied compounds.

3. Experimental

3.1. General Methods and Materials. All organic solvents were commercially available, distilled, and dried by appropriate methods.

^1H and ^{13}C NMR spectra were obtained from a Bruker Avance DRX 300 spectrometer. Chemical shifts are expressed in parts per million (ppm) downfield from external TMS. Perkin Elmer spectrophotometer was used to record the UV-visible absorption spectra. Mass spectra were measured on Bruker Biflex-III TM. IR spectra were measured on a Bruker vertex 70. A Thermo-Scientific Flash 2000 Analyzer was used to obtain (C, H and N) elemental analyses.

3.2. Synthesis of the Precursors and the Ligands

3.2.1. General Procedure for the Synthesis of 2-Hydroxy-5-((aryl) diazenyl)benzaldehyde. A diazonium solution was prepared by dissolving 0.01 mol of amine in 8 mL of water and 5 mL of concentrated hydrochloric acid was cooled to 0°C , treated with 15 mL of aqueous 1.0 M sodium nitrate dropwise, and stirred for 15 min. The resulting solution was added dropwise to a solution of salicylaldehyde (0.01 mol) dissolved in 50 mL of 10 % aqueous sodium hydroxide. After the resulting mixture had been stirred for an hour at $0-5^{\circ}\text{C}$, the precipitate was filtered and the products were obtained by recrystallized from ethanol.

3.2.2. 2-Hydroxy-5-(phenyldiazenyl)benzaldehyde (1a). Yield 84%, mp: 128°C . ^1H NMR (300MHz, DMSO) δ /ppm: 11.6 (s,1H), 10.4 (s,1H), 8.22 (d, 1H, $J = 3$), 8.12 (q, 1H, $J = 3$), 7.85(dd,2H, $J = 6, J = 3$), 7.70 (m, 2H), 7.62 (m, 1H), 7.22 (d, 1H, $J = 9$). ^{13}C NMR (75 MHz, DMSO) δ /ppm: 190.6, 163.3, 151.8, 144.8, 131.1, 129.6, 129.4, 123.8, 122.58, 122.3, 118.4. MALDI TOF MS calcd: $m/z = 226.07$ Da. Found $m/z = 227.10$ $[\text{M}+1]^+$. Selected IR bands (cm^{-1}) 3185.84, 1960, 1620, 1570, 1446, 1301, 1281, 1154, 1019, 952, 905, 843, 809, 759, 735, 708, 682, 641.

3.2.3. 2-Hydroxy-5-(o-tolyldiazenyl)benzaldehyde (1b). Yield 88%, mp: 130°C . ^1H NMR (300 MHz, DMSO) δ /ppm: 11.55 (s, 1H), 10.40 (s,1H), 8.21 (d, 1H, $J = 3$), 8.12 (dd,1H, $J = 6, J = 3$), 7.59-7.45 (m, 3H), 7.34 (m, 1H), 7.24 (d, 1H, $J = 9$), 2.68 (s, 3H). ^{13}C NMR (75 MHz, DMSO) δ /ppm: 190.6, 163.1, 149.8, 145.3, 137.2, 131.4, 132.0, 129.4, 126.6, 124.1, 122.6, 118.4, 115.1, 17.1. MALDI TOF MS calcd: $m/z = 240.09$ Da. Found $m/z = 241.10$ $[\text{M}+1]^+$. Selected IR bands (cm^{-1}) 3100, 2928, 1650,

1621, 1581, 1486, 1373, 1277, 1140, 1094, 1038, 951, 896, 867, 848, 834, 766, 735, 697, 643.

3.2.4. 2-Hydroxy-5-(*p*-tolylidiazanyl)benzaldehyde (1c). Yield 89%, mp: 154°C. ¹H NMR (300MHz, DMSO) δ/ppm: 11.53 (s, 1H), 10.39 (s, 1H), 8.19 (d, 1H, *J* = 3), 8.10 (dd, 1H, *J* = 6, *J* = 3), 7.80 (d, 2H, *J* = 6), 7.42 (dd, 2H, *J* = 6, *J* = 3), 7.22 (d, 1H, *J* = 9), 2.42 (s, 3H). ¹³C NMR (75 MHz, DMSO) δ/ppm: 190.6, 163.1, 149.9, 144.8, 141.3, 129.9, 129.6, 123.5, 122.6, 122.4, 118.3, 21.0. MALDI TOF MS calcd: *m/z* = 240.09 Da. Found *m/z* = 241.10 [M+1]⁺. Selected IR bands (cm⁻¹) 3170, 2866, 1650, 1602, 1572, 1477, 1375, 1275, 1166, 1105, 1013, 951, 908, 850, 825, 777, 763, 739, 725, 688, 614.

3.2.5. General Procedure for the Synthesis of the Schiff Bases Based on Imidazole: LI-L3. N-(3-Aminopropyl)imidazole (4mmol) was added to a methanol solution (30 mL) of 2-Hydroxy-5-(*o*-tolylidiazanyl)benzaldehyde (4 mmol). The mixture was refluxed for 2 h and cooled to room temperature. The solvent was removed on a rotatory evaporator and the orange products were rinsed and recrystallized with mixture of methanol and ether. The products were obtained as orange crystals.

3.2.6. Schiff Base Ligand LI. Yield 84 %, m.p 110°C. ¹H NMR (300MHz, DMSO) δ/ppm: 8.65 (s, 1H), 8.03 (d, 1H, *J* = 2.4), 7.91(dd, 1H, *J* = 6.6, *J* = 2.4), 7.78 (dd, 2H, *J* = 7.2, *J* = 1.5), 7.65 (s, 1H), 7.55-7.5 (m, 2H), 7.46-7.48 (m, 1H), 7.2 (s, 1H), 6.92-6.89 (m, 2H), 4.04 (t, 2H, *J* = 6.9), 3.56 (t, 2H, *J* = 6.9), 2.12 (qd, 2H, *J* = 6.9). ¹³C NMR (75 MHz, DMSO) δ/ppm: 31.8, 44.3, 55.7, 118.0, 118.2, 118.7, 122.6, 127.1, 127.2, 131.0, 130.0, 137.1, 145.4, 152.6, 164.1, 165.91. MALDI TOF MS calcd: *m/z* = 333.16 Da. Found *m/z* = 334.4 [M+1]⁺. HR-MS(M): for C₁₉H₁₉N₅O: 333.1589 found: 347.1589. Selected IR bands (cm⁻¹) 3104, 2946, 1630, 1584, 1497, 1480, 1437, 1383, 1350, 1275, 1250, 1195, 977, 926, 906, 852, 837, 769, 745, 665, 641. Anal. Calc. for C₁₉H₁₉N₅O: C, 68.45%; H, 5.74%; N, 21.01%. Found C, 68.25%; H, 5.79%; N, 20.82 %.

3.2.7. Schiff Base Ligand L2. Yield 79 %, m.p. 82°C. ¹H MR (300MHz, DMSO) δ/ppm: 8.65 (s, 1H), 8.01 (d, 1H, *J* = 2.4), 7.91 (dd, 1H, *J* = 6.6, *J* = 2.4), 7.65 (s, 1H), 7.52 (dd, 1H, *J* = 7.8, *J* = 1.5), 7.35-7.34 (m, 1H), 7.33 (dd, 1H, *J* = 7.6, *J* = 1.5), 7.28-7.25 (m, 1H), 7.20 (s, 1H), 6.93-6.90 (m, 2H), 4.01 (t, 2H, *J* = 7.2), 3.55 (t, 2H, *J* = 6.9), 2.61 (s, 3H), 2.10 (qd, 2H, *J* = 6.9). ¹³C NMR (75 MHz, DMSO) δ/ppm: 17.6, 31.7, 44.2, 53.7, 115.4, 117.6, 119.8, 120.1, 126.5, 127.0, 130.4, 130.8, 131.7, 137.2, 137.8, 144.1, 150.4, 166.8, 168.6. MALDI TOF MS calcd: *m/z* = 347.17 Da. Found *m/z* = 348.4 [M+1]⁺. HR-MS(M): for C₂₀H₂₁N₅O: 347.1746 found: 347.1745. Selected IR bands (cm⁻¹): 3136, 2941, 1621, 1567, 1486, 1381, 1308, 1274, 1146, 1095, 1031, 961, 906, 830, 748, 703, 670. Anal. Calc. for C₂₀H₂₁N₅O: C, 69.14%; H, 6.09%; N, 20.16%. Found C, 68.84%; H, 6.13%; N, 19.95 %.

3.2.8. Schiff Base Ligand L3. Yield 82%, m.p 95°C. ¹H MR (300MHz, DMSO) δ/ppm: 8.63 (s, 1H), 7.99 (d, 1H, *J* = 2.7), 7.88 (dd, 1H, *J* = 6.6, *J* = 2.4), 7.71 (d, 2H, *J* = 8.1), 7.65 (s, 1H), 7.32 (d, 2H, *J* = 8.1), 7.19 (s, 1H), 6.90-7.00 (m, 2H), 4.04 (t, 2H, *J* = 7.2), 3.55 (t, 2H, *J* = 6.6), 2.34 (s, 3H), 2.13 (qd, 2H,

J = 6.9). ¹³C NMR (75 MHz, DMSO) δ/ppm: 21.5, 31.8, 44.3, 55.7, 117.9, 118.2, 118.7, 122.6, 126.9, 127.1, 129.8, 129.9, 137.1, 141.1, 145.5, 150.7, 163.8, 166.0. MALDI TOF MS calcd: *m/z* = 347.17 Da. Found *m/z* = 348.4 [M+1]⁺. HR-MS(M): for C₂₀H₂₁N₅O: 347.1746 found: 347.1745. Selected IR bands (cm⁻¹) 3105, 2943, 1634, 1580, 1485, 1445, 1377, 1283, 1229, 1107, 999, 959, 905, 851, 756, 689, 662. Anal. Calc. for C₂₀H₂₁N₅O: C, 69.14%; H, 6.09%; N, 20.16%. Found: C, 69.23%; H, 6.01%; N, 20.07 %.

3.2.9. Schiff Base Ligand L4. Propylamine (4 mmol) was added to a methanol solution (30 mL) of 2-Hydroxy-5-(*o*-tolylidiazanyl)benzaldehyde (4 mmol). The mixture was refluxed for 2h and cooled to room temperature. The solvent was removed on a rotatory evaporator and the orange product solid was recrystallized with mixture of methanol and ether. Product was obtained as orange crystals. Yield 90 %. ¹H NMR(300MHz, DMSO) δ/ppm: ¹H NMR(300MHz, DMSO) δ/ppm: 14.42 (s, 1H), 8.45 (s, 1H), 8.02 (dd, 1H, *J* = 2.4, *J* = 9), 7.92 (d, 1H, *J* = 2.4), 7.65-7.63 (m, 1H), 7.37-7.33 (m, 2H), 7.32-7.27 (m, 1H), 7.09 (d, 1H, *J* = 9), 3.65 (t, 2H, *J* = 6), 2.74 (s, 3H), 1.8 (m, 2H), 1.06 (t, 3H, *J* = 9). ¹³CNMR (75 MHz, DMSO) δ/ppm: 11.2, 17.1, 23.2, 56.5, 115.04, 116.4, 120.6, 126.0, 126.5, 130.0, 131.1, 131.2, 136.6, 142.9, 150.0, 165.8, 170.7. MALDI TOF MS calcd: *m/z* = 281.15 Da. Found *m/z* = 282.4 [M+1]⁺. HR-MS(M): for C₁₇H₁₉N₃O: 281.1528 found: 281.1522. Selected IR bands (cm⁻¹): 3115, 2956, 1640, 1612, 1510, 1478, 1461, 1434, 1396.66, 1366, 1301, 1258, 1237, 1201, 1169, 1151, 1115, 1027, 944, 914, 887, 836, 761, 750, 715, 678, 628. Anal. Calc. for C₁₇H₁₉N₃O: C, 72.57%; H, 6.81%; N, 14.94%. Found C, 72.46%; H, 6.81%; N, 14.74 %.

3.2.10. Microorganisms. Antifungal activity was assayed on human pathogenic fungi, including *Candida albicans* (ATCC 1066), *Aspergillus fumigatus* (CBS 11326), and *Scedosporium apiospermum* (IHEM 15115). The yeast was obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA) and the opportunistic molds were furnished for *A. fumigatus* by the Centraalbureau voor Schimmelcultures (CBS, Delft, Netherlands) and for *S. apiospermum* by the Institute of Hygiene and Epidemiology-Mycolology section, Institute of Public Health (IHEM, Brussels, Belgium). All the strains were maintained on yeast extract-peptone-dextrose agar (YPDA) plates which contained 0.05 % chloramphenicol at 37°C during an incubation time of 48 h for yeast, 72 h for *A. fumigatus*, and 7 days for *S. apiospermum*.

3.2.11. Determination of Antifungal Activity. Disk Diffusion Method on Solid Medium. Antifungal activities of the different ligands were evaluated using a disk diffusion method adapted from routinely antifungal tests [55]. Casitone agar plates of 90 mm diameter were used for the experiences. Antifungal inocula were prepared for yeast by suspending one colony in 10 mL of sterile distilled water. For the filamentous fungi, the mycelium was recovered by scrapping the YPDA plates with 10 mL of sterile distilled water. Conidia were then harvested from the suspension after a centrifugation at 1500 g for 5 min. The supernatant was then adjusted by spectrophotometry at 630 nm to an absorbance of 0.1. Casitone Petri dishes were flooded with 10 mL of the spore suspensions. Excess of the

suspension was eliminated and the Petri dishes were dried 10 min at 37°C.

Compounds were dissolved in DMSO at a final concentration of 10 mg/mL and 25 μ L aliquots were applied on 12 mm diameter paper disks (rf 06234304, Prolabo 33173 Gradigan, France). Then, disks were deposited in the center of the casitone agar plates previously inoculated by fungal suspensions.

After an incubation time of 48 h for *C. albicans*, 72 h for *A. fumigatus*, and 7 days for *S. apiospermum* at 37°C, diameters of the growth inhibition zones were measured. Growth control was performed using filter paper disks soaked with an equal volume of, respectively, drug-free solvent (DMSO), and positive control was made with miconazole (Neosentabs tablets, Rosco Diagnostic, Denmark).

Microdilutions Method in Liquid Medium. Antifungal tests were performed by following the guidelines of the Clinical Laboratory Standards Institute (CLSI) corresponding for yeasts to the reference method M27-A3 [56] and for filamentous fungi to the M38-A2 reference method [57]. Briefly, the fungal suspensions were prepared in RPMI-1640 culture medium (Sigma) added with 2 mM of L-glutamine, buffered with 0.165 M of morpholine propane-sulfonic acid (MOPS), and adjusted spectrophotometrically at 630 nm to final inocula concentrations of 0.5 to 2.5 $\times 10^3$ colony-forming units (CFU) per mL for *C. albicans* and 0.5 to 5.0 $\times 10^4$ CFU per mL for *A. fumigatus* and *S. apiospermum*. Broth microdilution tests were performed using sterile 96-wells flat-shaped microtitre plates. Each well was inoculated with 195 μ L of the corresponding fungal working suspension.

Serial twofold dilutions of ligand were made in DMSO, 40 times the strength of the final drug concentration (10-0.0025 mg.mL⁻¹), and were dispensed in triplicate at a volume of 5 μ L per well. Final concentrations for the drugs were from 250.00 to 0.061 μ g.mL⁻¹. According to this procedure, the final concentration of DMSO solvent was lower than 2.5 % which did not affect significantly the growth of any fungus. After a 48 h incubation time for the yeast, 72 h for *A. fumigatus*, and 7 days for *S. apiospermum* at 37°C, the spectrophotometric reading of each well was performed at 630 nm with a Dynatech Laboratories MRX_{TC} automatic plate spectrophotometric reader. The minimum inhibitory concentration of the compound that inhibits 80% (MIC₈₀) of the fungal growth was calculated from the turbidimetric data compared to that of the DMSO-free control as the lowest compound concentration giving rise to an inhibition of growth equal or greater than 80 % of the compound-free control.

3.2.12. Antioxidant Activity. DPPH free radical scavenging assays were performed according to the procedure described by Blois *et al.* [58]. A DPPH solution was prepared by dissolving 2 mg DPPH in 100 mL of methanol and then 250 μ L of this solution was mixed with 50 μ L of different concentrations of compounds **L1-L4** dissolved in methanol. After 30 min incubation in the dark and at room temperature, the absorbance was read against a blank at 517 nm. Inhibition

of free radical (DPPH) in percentage (I%) was calculated as follows:

$$I\% = \left(A_0 - \frac{A_i}{A_0} \right) \times 100 \quad (2)$$

where A₀ is the absorbance of the control (containing all reagents except the test compound), and A_i is the absorbance of the tested sample.

IC₅₀ values were also defined as the concentration of the sample that causes 50 % of DPPH radicals inhibition. These values were calculated from the plot of inhibition percentages against sample concentration. Ascorbic acid was used as a positive control. Each measurement was performed in triplicate.

4. Conclusion

In this paper, we describe the synthesis, full characterization, and optical properties of four different Schiff bases ligands **L1-L4** which associate an azo group, an imidazole unit, and a Schiff base fragment. The antifungal activities of the prepared ligands were evaluated and the results indicate two main tendency: (a) the presence of an azole group in this ligands is of great importance for the biological activities since **L1-L3** exhibit higher growth inhibition zone diameters against the studied *S. apiospermum*, *A. fumigatus*, and *C. albicans* fungi and lower MIC₈₀ values for *C. albicans* as compared with the azole free ligand **L4**; (b) the presence of an additional substituent on the phenyl ring of the azo group lowers the MIC₈₀ activity of the ligands. Note that **L1-L3** azole based ligands show excellent MIC₈₀ values against *C. albicans* with concentrations as low as 0.1 μ g/mL which is ten times lower than the reference concentration (1 μ g/mL). In addition the four ligands show a significant antioxidant activity. All these findings clearly indicate the great potential of these new azole ligands as valuable candidates for developing new therapeutic agents. The complexation ability of these novel Schiff bases ligands towards transition metal cations such as Cu(II), Fe(II), Co(II),... as well as their corresponding biological activities is in progress.

Data Availability

The experimental data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] A. Puratchikody and M. Doble, "Antinociceptive and anti-inflammatory activities and QSAR studies on 2-substituted-4,5-diphenyl-1H-imidazoles," *Bioorganic & Medicinal Chemistry*, vol. 15, p. 1083, 2007.
- [2] K. Shalini, P. K. Sharma, and N. Kumar, "Imidazole and its biological activities: a review," *Der Chemica Sinica*, vol. 1, p. 36, 2010.

- [3] H. Bhawar, N. Dighe, P. Shinde, R. Lawre, and S. Bhawar, *Asian Journal of Pharmacy and Technology*, vol. 4, p. 189, 2014.
- [4] F. Bellina, S. Cauteruccio, and R. Rossi, "Synthesis and biological activity of vicinal diaryl-substituted 1H-imidazoles," *Tetrahedron*, vol. 63, no. 22, pp. 4571–4624, 2007.
- [5] R. Di Santo, A. Tafi, R. Costi et al., "Antifungal agents. II. N-substituted derivatives of 1-[(aryl)(4-aryl-1H-pyrrol-3-yl)methyl]-1H-imidazole: Synthesis, anti-Candida activity, and QSAR studies," *Journal of Medicinal Chemistry*, vol. 48, no. 16, pp. 5140–5153, 2005.
- [6] S. Dutta, "Synthesis and anthelmintic activity of some novel 2-substituted-4,5-diphenyl imidazoles," *Acta Pharmaceutica*, vol. 60, p. 229, 2010.
- [7] S. Mukherjee, T. Weyhermüller, E. Bill, and P. Chaudhuri, "Tetranuclear copper(II) and nickel(II) complexes incorporating a new imidazole-containing ligand," *European Journal of Inorganic Chemistry*, p. 4209, 2004.
- [8] R. Singh, M. Ahmad, and P. K. Bharadwaj, "Coordination polymers of copper and zinc ions with a linear linker having imidazole at each end and an azo moiety in the middle: pedal motion, gas adsorption, and emission studies," *Crystal Growth & Design*, vol. 12, p. 5025, 2012.
- [9] C. Y. Chen, P. Y. Cheng, H. H. Wu, and H. M. Lee, "Conformational effect of 2,6-bis(imidazol-1-yl)pyridine on the self-assembly of 1D coordination chains: spontaneous resolution, supramolecular isomerism, and structural transformation," *Inorganic Chemistry*, vol. 46, p. 5691, 2007.
- [10] G. Brewer, M. J. Olida, A. M. Schmiedekamp, C. Viragh, and P. Y. Zavalij, "A DFT computational study of spin crossover in iron(III) and iron(II) tripodal imidazole complexes. A comparison of experiment with calculations," *Journal of the Chemical Society, Dalton Transactions*, no. 47, pp. 5617–5629, 2006.
- [11] B. Sharkar, "Treatment of Wilson and Menkes diseases," *Chemical Reviews*, vol. 99, p. 2535, 1999.
- [12] G. S. Hartley, "The cis-form of azobenzene," *Nature*, vol. 140, p. 281, 1937.
- [13] G. S. Hartley, "The cis-form of azobenzene and the velocity of the thermal cis→trans-conversion of azobenzene and some derivatives," *Journal of the Chemical Society*, vol. 633, 1938.
- [14] H. Zollinger, *Color Chemistry, Syntheses, Properties and Applications of Organic Dyes and Pigments*, Wiley-VCH, Weinheim, Germany, 3rd edition, 2003.
- [15] N. Tamai and H. Miyasaka, "Ultrafast dynamics of photochromic systems," *Chemical Reviews*, vol. 100, p. 1875, 2000.
- [16] T. Schultz, J. Quenneville, B. Levine et al., "Mechanism and dynamics of azobenzene photoisomerization," *Journal of the American Chemical Society*, vol. 125, no. 27, pp. 8098–8099, 2003.
- [17] A. G. Cheelham, M. G. Hutchings, T. D. W. Claridge, and H. L. Anderson, "Enzymatic synthesis and photoswitchable enzymatic cleavage of a peptide-linked rotaxane," *Angewandte Chemie International Edition*, vol. 45, p. 1596, 2006.
- [18] R. Fernández, J. A. Ramos, L. Espósito, A. Tercjak, and I. Mondragon, "Reversible optical storage properties of nanostructured epoxy-based thermosets modified with azobenzene units," *Macromolecules*, vol. 44, no. 24, pp. 9738–9746, 2011.
- [19] H. Nishihara, "Combination of redox- and photochemistry of azo-conjugated metal complexes," *Coordination Chemistry Reviews*, vol. 249, p. 1468, 2005.
- [20] A. Bianchi, E. Delgado-Pinar, E. García-España, C. Giorgia, F. Pina, and E. García-España, "Highlights of metal ion-based photochemical switches," *Coord. Chem. Rev.*, vol. 260, p. 156, 2014.
- [21] A. Natansohn and P. Rochon, "Photoinduced motions in azo-containing polymers," *Chemical Reviews*, vol. 102, p. 4139, 2002.
- [22] S. Sporlein, H. Carstens, H. Satzger et al., "Ultrafast spectroscopy reveals subnanosecond peptide conformational dynamics and validates molecular dynamics simulation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 12, pp. 7998–8002, 2002.
- [23] S. Concilio, P. Iannelli, L. Sessa et al., "Biodegradable antimicrobial films based on poly(lactic acid) matrices and active azo compounds," *Journal of Applied Polymer Science*, vol. 132, no. 33, p. 42357, 2015.
- [24] S. Piotto, S. Concilio, L. Sessa et al., "Novel antimicrobial polymer films active against bacteria and fungi," *Polymer Composites*, vol. 34, no. 9, pp. 1489–1492, 2013.
- [25] S. Piotto, S. Concilio, L. Sessa et al., "Small azobenzene derivatives active against bacteria and fungi," *European Journal of Medicinal Chemistry*, vol. 68, p. 178, 2001.
- [26] M. Huang, S. Wu, J. Wang et al., "Biological study of naphthalene derivatives with antiinflammatory activities," *Drug Development Research*, vol. 60, p. 261, 2003.
- [27] A. Corma, H. García, F. X. Llabrés, and I. Xamena, "Engineering metal organic frameworks for heterogeneous catalysis," *Chemical Reviews*, vol. 110, p. 4606, 2010.
- [28] R. Ziesel, A. Harriman, A. El-Ghayoury et al., "First assembly of copper(I) naphthyridine-based helicates," *New Journal of Chemistry*, vol. 24, no. 10, pp. 729–732, 2000.
- [29] M. C. Young, A. M. Johnson, A. S. Gamboa, and R. J. Hooley, "Achiral endohedral functionality provides stereochemical control in Fe(II)-based self-assemblies," *Chemical Communications*, vol. 49, p. 1627, 2013.
- [30] S. E. Howson, A. Bolhuis, V. Brabec et al., "Optically pure, water-stable metallo-helical "flexicate" assemblies with antibiotic activity," *Nature Chemistry*, vol. 4, p. 31, 2012.
- [31] J. L. Bolliger, A. M. Belenguer, and J. R. Nitschke, "Enantiopure water-soluble [Fe₄L₆] cages: host-guest chemistry and catalytic activity," *Angewandte Chemie International Edition*, vol. 52, p. 7958, 2013.
- [32] R. A. Bilbeisi, T. K. Ronson, and J. R. Nitschke, "A self-assembled [Fe^{II}₁₂L₁₂] capsule with an icosahedral framework," *Angewandte Chemie International Edition*, vol. 52, p. 9027, 2013.
- [33] J. Cloete and S. F. Mapolie, "Functionalized pyridinyl-imine complexes of palladium as catalyst precursors for ethylene polymerization," *Journal of Molecular Catalysis A: Chemical*, vol. 243, p. 221, 2006.
- [34] P. Shejwalkar, N. P. Rath, and E. B. Bauer, "New iron(II) α -aminopyridine complexes and their catalytic activity in the oxidation of activated methylene groups and secondary alcohols to ketones," *Dalton Transactions*, vol. 40, p. 7617, 2011.
- [35] N. Mishra, K. Poonia, S. K. Soni, and D. Kumar, "Synthesis, characterization and antimicrobial activity of Schiff base Ce(III) complexes," *Polyhedron*, vol. 120, pp. 60–68, 2016.
- [36] A. B. Thomas, R. K. Nanda, L. P. Kothapalli, and S. C. Hamane, "Synthesis and biological evaluation of Schiff's bases and 2-azetidinones of isonicotinyl hydrazone as potential antidepressant and nootropic agents," *Arabian Journal of Chemistry*, vol. 9, p. S79, 2016.
- [37] E. S. Aazam and W. A. El-Said, "Synthesis of copper/nickel nanoparticles using newly synthesized Schiff-base metals complexes and their cytotoxicity/catalytic activities," *Bioorganic Chemistry*, vol. 57, pp. 5–12, 2014.

- [38] S. Murtaza, M. S Akhtar, F. Kanwal, A. Abbas, S. Ashiq, and S. Shamim, "Synthesis and biological evaluation of schiff bases of 4-aminophenazone as an anti-inflammatory, analgesic and antipyretic agent," *Journal of Saudi Chemical Society*, vol. 21, p. S359, 2017.
- [39] P. Zhan, X. Liu, J. Zhu et al., "Synthesis and biological evaluation of imidazole thioacetanilides as novel non-nucleoside HIV-1 reverse transcriptase inhibitors," *Bioorganic & Medicinal Chemistry*, vol. 17, p. 5775, 2009.
- [40] N. Süleymanoğlu, R. Ustabaş, Ş. Direkel, Y. B. Alpaslan, and Y. Ünver, "1,2,4-triazole derivative with Schiff base; thiol-thione tautomerism, DFT study and antileishmanial activity," *Journal of Molecular Structure*, vol. 1150, pp. 82–87, 2017.
- [41] N. Karali and A. Gürsoy, "Synthesis and anticonvulsant activity of some new thiosemicarbazone and 4-thiazolidone derivatives bearing an isatin moiety," *Farmaco*, vol. 49, p. 819, 1994.
- [42] E. Gungor, S. Celen, D. Azaaz, and H. Kara, "Two tridentate Schiff base ligands and their mononuclear cobalt (III) complexes: Synthesis, characterization, antibacterial and antifungal activities," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 94, pp. 216–221, 2012.
- [43] A. M. Alafeefy, M. A. Bakht, M. A. Ganie, N. Ansari, N. N. El-Sayed, and A. S. Awaad, "Synthesis, analgesic, anti-inflammatory and anti-ulcerogenic activities of certain novel Schiff's bases as fenamate isosteres," *Bioorganic & Medicinal Chemistry Letters*, vol. 25, no. 2, pp. 179–183, 2015.
- [44] Z. D. Mou, N. Deng, F. Zhang, J. Zhang, J. Cen, and X. Zhang, "'Half-sandwich' Schiff-base Ir(III) complexes as anticancer agents," *European Journal of Medicinal Chemistry*, vol. 138, pp. 72–82, 2017.
- [45] S. Slassi, M. Aarjane, A. Amine, H. Zouihri, and K. Yamni, "2-((E)-{3-(1H-Imidazol-1-yl)propyl}imino)methyl)-4-[(E)-(2-methylphenyl)diazanyl]phenol," *IUCrData*, vol. 2, 2017, x171477.
- [46] R. Botros, US Patent 1977, 4051119.
- [47] M. Pihet, J. Carrere, B. Cimon et al., "Occurrence and relevance of filamentous fungi in respiratory secretions of patients with cystic fibrosis—a review," *Medical Mycology*, vol. 47, no. 4, pp. 387–397, 2009.
- [48] K. Alomar, A. Landreau, M. Allain, G. Bouet, and G. Larcher, "Synthesis, structure and antifungal activity of thiophene-2,3-dicarboxaldehyde bis(thiosemicarbazone) and nickel(II), copper(II) and cadmium(II) complexes: Unsymmetrical coordination mode of nickel complex," *Journal of Inorganic Biochemistry*, vol. 126, pp. 76–83, 2013.
- [49] P. I. S. Maia, F. R. Pavan, C. Q. F. Leite et al., *Metal Ions in Biology and Medicine and Aqueous Chemistry and Biochemistry of Silicon*, John Libbey Eurotext, Paris, France, 2011, 164.
- [50] K. Alomar, V. Gaumet, M. Allain, G. Bouet, and A. Landreau, "Synthesis, crystal structure, characterisation, and antifungal activity of 3-thiophene aldehyde semicarbazone (3STCH), 2,3-thiophene dicarboxaldehyde bis(semicarbazone) (2,3BSTCH₂) and their nickel (II) complexes," *Journal of Inorganic Biochemistry*, vol. 115, pp. 36–43, 2012.
- [51] P. L. de Sá, D. A. D. Junior, A. S. Porcacchia et al., "The Roles of ROS in Cancer Heterogeneity and Therapy," *Oxidative Medicine and Cellular Longevity*, vol. 2017, Article ID 2467940, 12 pages, 2017.
- [52] W. Brand-Williams, M. E. Cuperlier, C. Berset, and C. Food, "Use of a free radical method to evaluate antioxidant activity," *Science and technology*, vol. 28, p. 25, 1995.
- [53] D. Amic, V. Stepamic, B. Lucic et al., "PM6 study of free radical scavenging mechanisms of flavonoids: why does O–H bond dissociation enthalpy effectively represent free radical scavenging activity?" *Journal of Molecular Modeling*, vol. 9, p. 2593, 2013.
- [54] M. C. Foti, C. Daquino, and C. Geraci, "Electron-transfer reaction of cinnamic acids and their methyl esters with the DPPH(*) radical in alcoholic solutions," *The Journal of Organic Chemistry*, vol. 69, p. 2309, 2004.
- [55] A. L. Barry and D. Brown, "Fluconazole disk diffusion procedure for determining susceptibility of *Candida* species," *Journal of Clinical Microbiology*, vol. 34, pp. 2154–2157, 1996.
- [56] CLS. Institute, *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts*, Clinical Laboratory Standard Institute, M27-A3.Wayne, 2008.
- [57] Institute CLS, "Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi, M38-A2," Wayne, PA: Clinical Laboratory Standard Institute, 2008.
- [58] M. S. Blois, "Antioxidant determinations by the use of a stable free radical," *Nature*, vol. 181, p. 1119, 1958.



Hindawi

Submit your manuscripts at
www.hindawi.com

