

Research Article

Evaluation of NH_4^+ Adsorption Capacity in Water of Coffee Husk-Derived Biochar at Different Pyrolysis Temperatures

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Ammonium (NH_4^+) is a pollutant that can be harmful to the water environment. The purpose of this study is to assess NH_4^+ removal capacity from water by coffee husk-derived biochar. The properties of biochar prepared at different temperatures (300, 450, and 600°C) were determined including TOC, and pH, pH_{pzc} , functional groups of H^+/OH^- , cation-exchange capacity (CEC), and the characteristics of groups of organic matter (FT-IR spectrum) were identified and evaluated. The trend of NH_4^+ adsorption equilibrium and kinetics of biochar have been studied. The experimental design of adsorption equilibrium was carried out by exposing biochar to a NH_4^+ solution at different concentrations, ranging from 0 to 50 mg NH_4^+/L for 12 hours. Kinetic surveys were carried out when biochar was exposed to a solution containing 8.3 mg NH_4^+/L for a varying length of time. The results showed that Langmuir and Freundlich models and the pseudo-second-order kinetic model are suitable to explain the NH_4^+ adsorption equilibrium and kinetics on the biochar forms derived from coffee husk. Biochar derived from coffee husk prepared at lower pyrolysis temperature has a higher adsorption capacity. The results suggest that the biochar could be used as an adsorbent ammonium from water.

1. Introduction

Nitrogen is present in natural water in the forms of organic nitrogen, ammonia, nitrate, and nitrite. In most cases of raw wastewater, nitrogen is normally present in the organic nitrogen and NH_4^+ . Ammonia also can be naturally generated in the environment [1]. The presence of NH_4^+ at high concentrations can contribute to eutrophication and the subsequent devastation of aquatic life. The residual amount of ammonium in water causes ecological issues in relation to eutrophication, acidification of freshwater ecosystems, and in anoxic conditions, poisoning benthic organisms and fish [2]. According to Eddy, the ammoniac tolerance in freshwater fishes ranges from 0.07 to 2.00 mg/L [3]. At a dose of more than 33.7 mg of ammonium ion per kg of body weight per day, it influences the metabolism by shifting the

acid-base equilibrium, disturbing the glucose tolerance, and reducing the tissue sensitivity to insulin [4].

Therefore, removal of nitrous ammonium in polluted water is necessary. Many methods of removing NH_4^+ from water have been used, including ion adsorption and exchange, which is more effective than the others, yet of high cost [5]. Looking for new renewable materials that can be used in ion adsorption and exchange is a crucial research topic. Biochar prepared from agricultural waste is a useful material due to its efficiency, low cost, environment friendliness, and availability in large quantity [6]. Adsorption of inorganic pollutants by biochar is the result of (i) ion exchange, (ii) electrostatic attraction, or (iii) surface precipitation [7]. In the case of ammonium, mostly electrostatic exchange and interaction is used; moreover, Khalil et al. stated that biochar's surface chemistry is more important

than the material's surface area [5, 8]. However, surface chemistry properties are controlled by the pyrolysis condition, especially temperatures, heating, and heat-keeping time [9]. Ammonium adsorption capability by biochar which was prepared with varying pyrolysis temperatures among research is not always alike [10]. There have been studies which use biochar derived from corn plants, red oak (*Quercus rubra*), maple trees, and wheat plants to adsorption of ammonium [10].

Vietnam is the No. 1 producer of robusta coffee in the world, accounting for more than 40% of the global output in the 2019-2020 marketing year [11]. Coffee husks are the major solid residues from the processing of coffee, for which there are no current profitable uses, and their adequate disposal constitutes a major environmental problem. Dak Lak is a leading coffee-producing province in Vietnam with over 200,000 ha of coffee fields, and its annual coffee production reaches 450,000 tons. Together with about 250,000 tons of coffee husk to be discharged, it is a valuable material, yet has not been used efficiently [12]. Thus, innovative techniques and products for the profitable and adequate use of this type of residue are being sought. We have been also interested in the adsorption of metal ion, textile dyes, etc. on different materials [13–15]. In this work, we are reporting the assessment of NH_4^+ in the water adsorption capability of biochar derived from coffee husk at different pyrolysis temperatures.

2. Materials and Methodology

2.1. Sampling Method. The coffee husk was collected in January 2019 from a household at Hamlet 8, 9A village, Pong Drang Commune, Krong Buk Town, Dak Lak Province. The location of the sampling site is $12^\circ 34' 43.8''\text{N}$ $108^\circ 01' 39.3''\text{E}$. The composite samples were obtained from the coffee peeling process. Coffee husk was dried at 60°C for 24 hours and stored in polyethylene bags [16].

2.2. Chemicals. All chemicals used in the study were of analytical grade. Storage solution concentration was NH_4^+ 1000 mg/L. Sample water was distilled water and was purified by the model EASYpure II RF from Thermo Scientific, USA. Instruments must be cleaned by being filled with nitric acid for 24 hours and then cleaned by demineralized water [17].

2.3. Experimental Design. The biochar modulation refers to the research of Yoo et al., where processed coffee husk was furnace in a Nabertherm P330 furnace at 300, 450, and 600°C [18]. The heating rate was set to $10^\circ\text{C}\cdot\text{min}^{-1}$. Once the desired temperature was reached, the temperature was kept constant for 2 hours and the samples were let to cool in the oven overnight. The biochar was then pressed through a plastic sieve (hole diameter of 1 mm) to make it homogeneous and was stored separately in polyethylene (PE) containers in dark at 4°C [18]. Biochar samples were analyzed and used to conduct equilibrium and kinetics experiments. Parameters including recovery efficiency and

surface functional groups were determined. Analyses were conducted on these biochar samples to determine characteristics of surface functional groups such as pH and pH_{pzc} [19], total organic carbon (TOC) [20], functional group H^+/OH^- [21], and cation-exchange capacity (CEC) based on the Walkley Black method. Changes in biochar's functional groups were analyzed by reflectance spectroscopy FT-IR-4700 type A with $350\text{--}4000\text{ cm}^{-1}$ resolution.

Equilibrium experiment of NH_4^+ ion adsorption on biochar refers to the work of Khalil et al. and Xue et al. [5, 22]. The experiment was performed in 50 mL polypropylene tubes, and 0.3 g biochar was mixed with 30 mL of NH_4^+ solution. The concentration of the diluted NH_4^+ solution varied between 0 and $50\text{ mg}\cdot\text{L}^{-1}$. Two drops of chloroform were added to prevent microbial activities. The initial pH of the solutions was adjusted to 5.0–5.5 by adding either dilute HCl or NaOH solutions (pH value is close to that of natural surface water sources). The mixtures were then shaken by using a GFL3015 orbital shaker with frequency 150 round-per-minute (rpm) in 12 hours (which is the time for the NH_4^+ adsorption to reach the equilibrium, determined by preliminary experiments.). After that, the solutions were not adjusted during this experiment. The solid settlement was separated from the mixture by using a DLAB DM0636 centrifuge at 4000 rpm for 15 min. Then, the remaining solution was filtered through a $0.22\text{ }\mu\text{m}$ filter. NH_4^+ was determined according to ISO 7150-1: 1984 (E). Langmuir and Freundlich adsorption isotherms models were used in evaluating the suitability of experimental data.

Ion adsorption kinetics survey was conducted by mixing 0.3 g biochar and 30 mL of $8.3\text{ mg}\text{NH}_4^+/\text{L}$ solution. The mixture was then shaken at 150 rpm. All the samples went through shaking intervals 5; 10; 15; 20; 30; 40; 60; 90; and 120 minutes and then were filtered to conduct NH_4^+ analysis. Pseudo-first- and second-order kinetic models were used to consider experiment data and to estimate kinetic parameters.

2.4. Data Processing

2.4.1. Calculation Methods

(i) Productivity efficiency of biochar is

$$\% \text{ production efficiency} = \frac{m_b}{m_0} * 100, \quad (1)$$

where m_0 (g) is the initial coffee mass before being furnace; m_b (g) is the biochar mass after being furnace.

(ii) pH_{pzc} of biochar is

$$\Delta\text{pH} = (\text{pH}_f - \text{pH}_i), \quad (2)$$

where pH_i : initial pH value; pH_f : pH value after biochar is added to the 0.01 M KCl solution (they were shaken for 2 h and allowed to settle for 48 h). Plotting ΔpH according to the initial pH, pH_{pzc} is where the pH curve overcomes $\Delta\text{pH} = 0$ [19].

(iii) Adsorption equilibrium:

Adsorption capacity, mg/g:

$$q_i = \frac{(C_0 - C_i) \times V}{m}, \quad (3)$$

where C_0 (mg/L) is the initial NH_4^+ ion concentration, C_i (mg/L) is the adsorbed NH_4^+ concentration at equilibrium, V (L) is the NH_4^+ solution volume, m (g) is the adsorbent mass (biochar), And q_i (mg/g) is the NH_4^+ adsorption capacity at equilibrium.

Langmuir isothermal equation:

$$\frac{1}{q_i} = \frac{1}{K_L q_0} \frac{1}{C_i} + \frac{1}{q_0}, \quad (4)$$

where q_0 (mg.g^{-1}): NH_4^+ maximum adsorption capacity at equilibrium; K_L (L.mg^{-1}): Langmuir adsorption constant.

The abovementioned equation has the form of $y = ax + b$, and therefore, it could be solved by curve fitting $y = 1/q_i$ and $x = 1/C_i$ to yield R^2 , q_0 . Freundlich isothermal equation:

$$q = \frac{y}{m} = K_F C^{(1/n_F)} \quad (5)$$

or

$$\log q_i = \frac{1}{n_F} \log C_i + \log K_F, \quad (6)$$

where n_F is the Freundlich isothermal constant for NH_4^+ adsorption intensity; K_F is the Freundlich isothermal adsorption constant for adsorption capacity.

This equation also has the form of $y = ax + b$, and as all the parameters are known, the plot of $\log q_i$ against $\log C_i$ can be drawn.

(iv) Adsorption kinetics:

Pseudo-first- and second-order (PFO and PSO) reaction models were usually used to study the adsorption mechanism, in order to assess adsorption kinetics parameters.

Pseudo-first-order kinetics equation:

$$\ln(q_e - q_t) = -k_1 t + \ln q_e. \quad (7)$$

$\ln(q_e - q_t)$ is plotted according to t .

Pseudo-second-order kinetics equation:

$$\frac{1}{q_t} = \frac{1}{t} \frac{1}{k_2 q_e^2} + \frac{1}{q_e}. \quad (8)$$

The abovementioned equation has the form of $y = ax + b$, and therefore, it could be solved by curve fitting $y = 1/q_t$ and $x = 1/t$ to yield R^2 , q_e is the NH_4^+ adsorption capacity at equilibrium (mg/g), q_t is the NH_4^+ adsorption capacity at time t , k_1 (1/minute) and k_2 ($\text{g/mg} \cdot \text{minute}$) are the pseudo-first- and second-order kinetics constants, and t (minute) is the adsorption time.

2.4.2. Data Processing. Control and replicate samples ($n = 3$) were employed in order to minimize error sources. The analysis evaluated the accuracy and precision of the parameters calculated using the methods described in the previous section. SPSS 20.0 was used to determine variance's homogeneity, and root mean square of the results was then calculated with p value <0.05 using Tukey's test post hoc if $\text{Sig} >0.05$ or Tamhane if $\text{Sig} <0.05$.

3. Results and Discussion

3.1. Influences of Chemical Synthesis Temperatures on Productivity Efficiency and Surface Chemistry of Biochar. The study showed that chemical synthesis temperatures affect productivity efficiency and surface chemistry of coffee husk biochar; Table 1 illustrates that, with increasing temperatures of 300, 450, and 600°C, recovery efficiency (%H) decreased at 51.4, 34.9, and 30.7%, respectively. This was due to the reduction of volatile substances in biochar [8]. The results were consistent with those from the work of Yavari et al. which reported a decrease in recovery efficiency when biochar pyrolysis temperature rises from 300 to 700°C [23, 24]. Biochar acquired at 300°C was statistically significantly different to the ones at 450 and 600°C, while biochar produced at 450 and 600°C was not statistically significantly different.

Total Organic Carbon (%TOC) in Table 1 shows that, with increasing temperatures of 300, 450, and 600°C, %TOC decreased at 25.5, 10.4, and 1.5%, respectively, which was consistent with the findings by the previous studies [8, 23]. This was related to the production of organic acids and phenolic compounds due to thermal decomposition of cellulose and hemicellulose compounds in the pyrolysis temperature range of the study [23] and the removal of water, hydrocarbons, H_2 , CO, and CO_2 during the carbonization process [8]. %TOC of biochar samples had statistically significant differences. pH values at 7.59, 9.16, and 9.69, respectively, to the biochar pyrolysis temperatures 300, 450, and 600°C, the pH values were also statistically significantly different (Table 1). This is due to the partial transformation of organic matters into ashes, which releases alkali salts and increases the pH level of biochar [25].

Similar to pH, pH_{pzc} values at 7.2, 8.2, and 9.5, respectively, were also statistically significantly different. With the increase in biochar pyrolysis temperatures, the number of acid functional groups (H^+) decreased and the values of H^+ functional groups were statistically significantly different. The number of base functional groups (OH^-) increased at 11.17, 11.24, and 12.50 $\text{mmolOH}^- \cdot \text{g}^{-1}$, respectively, to the increasing biochar pyrolysis temperatures, and the value at 600°C was statistically significantly different from the ones at 300 and 450°C, whereas the values at 300 and 450°C were not. This observation matched the conclusion reached by Mukherjee et al. and Yang et al. in regard to biochar originating from plant [26, 27]. CEC values were 309, 290, and 266 mmol.kg^{-1} , respectively. Decreasing CEC values were statistically significantly different with increasing biochar pyrolysis temperatures, and a similar conclusion was stated by Fidel et al. [10].

TABLE 1: Productivity efficiency and surface physical chemistry properties of biochar.

$t^{\circ}\text{C}$	%H	pH	pH_{pzc}	$\text{mmolH}^{+}\cdot\text{g}^{-1}$	$\text{mmolOH}^{-}\cdot\text{g}^{-1}$	%TOC	CEC, $\text{mmol}\cdot\text{kg}^{-1}$
300	51, 4 ^b	7, 59 ^a	7, 2 ^a	1, 73 ^c	11, 17 ^a	25, 5 ^c	309 ^c
SD	0, 8	0, 16	0, 1	0, 11	0, 15	1, 0	11
450	34, 9 ^a	9, 16 ^b	8, 2 ^b	0, 50 ^b	11, 24 ^a	10, 4 ^b	290 ^b
SD	0, 6	0, 02	0, 0	0, 10	0, 04	0, 3	3
600	30, 7 ^a	9, 69 ^c	9, 5 ^c	0, 17 ^a	12, 50 ^b	1, 5 ^a	266 ^a
SD	4, 0	0, 02	0, 3	0, 06	0, 04	0, 3	1

^{a,b,c} in a column illustrate statistically significant differences ($p < 0.05$). SD: standard deviation.

Analyzing the relationship between the parameters, Table 2 showed that the biochar pyrolysis temperatures were correlating and proportional to pH, pH_{pzc} , and mmolOH^{-} and reversely proportional to %H, mmolH^{+} , %TOC, and CEC.

Analysis results by FT-IR reflectance spectroscopy, Figure 1, showed that the featured through series at 3251 cm^{-1} and 1575 cm^{-1} of biochar 300°C clearly indicated the presence of OH^{-} and COO^{-} groups, the results showed that the carboxylase group decreased with increasing biochar pyrolysis temperatures, and similar conclusions were stated by Fidel et al. [10] and Thuy and Do [28]. This suggested a decrease in the polar organic functional groups with an increase in pyrolysis temperature [8]. Series at 1076 cm^{-1} were assumed to be due to the decrease of C-O in connection with the increasing pyrolysis temperatures [29]. An aromatic hydrocarbon peak (789 cm^{-1}) was observed at 450 and 600°C , which was in accordance with the study by Lugovoy et al. showing that lignin decomposition occurs at $280\text{--}500^{\circ}\text{C}$ [30]. The interchangeable results were concluded by Park et al. who also studied biochar derived from coffee husk produced at 300, 400, 500, and 600°C [31].

3.2. Adsorption Equilibrium NH_4^{+} in Water of Biochar.

Results of the NH_4^{+} adsorption equilibrium experiments of biochar derived from coffee husk produced at varying temperatures showed that adsorption capacity increased with increasing the initial NH_4^{+} concentration in all 3 biochar forms and they all reached saturation in the experimental condition (Figure 2). Increasing the initial NH_4^{+} concentration resulted in competition among cation in the solution, which promoted adsorption chance and capacity [5], specifically when raising C_0 from 0 to $50\text{ mgNH}_4^{+}\cdot\text{L}^{-1}$, the maximum capacity at equilibrium (saturated) was 1.8, 1.4, and $1.1\text{ mg}\cdot\text{g}^{-1}$, respectively, to biochar produced at 300, 450, and 600°C (Figure 2). One-way ANOVA on data of the adsorption showed that ammonium adsorption values varied and there were statistically significant differences in initial ammonium concentration ranging from 0 to $25\text{ mg}\cdot\text{L}^{-1}$. Ammonium concentrations from 25 to $50\text{ mg}\cdot\text{L}^{-1}$ were statistically insignificantly different. This indicated the adsorption process which reached saturation. The same explanation is also used for the case of biochar at 450 and 600°C (Figure 2).

The data (Figure 2, Table 3) showed that Langmuir and Freundlich models were suitable to describe the NH_4^{+} adsorption, with R^2 greater than 0.95 in all the 3 biochars,

and maximum adsorption values are 1.5, 1.2 and $0.9\text{ mg}\cdot\text{g}^{-1}$, respectively, to 300, 450, and 600°C biochar, which fitted the experiment values. Other studies also reported that the ammonium adsorption onto some biochars followed both the Langmuir and Freundlich models [32, 33]. R^2 values of the Langmuir model were 0.97, 0.95, and 0.96, respectively, which were slightly less than those of the Freundlich model, 0.97, 0.99, and 0.97, respectively. Therefore, the Freundlich adsorption isotherm appeared to be more favorable in describing the adsorption of ammonium onto the coffee husk-derived biochar surface than the Langmuir model. The Freundlich model indicated the heterogenous nature of the biochar surfaces. In light of the heterogeneity nature of the biochar surface, ammonium adsorption would occur at multiple layers onto sites where nonuniform energy distribution could occur. Similar observations were also found in the study of Fan et al. [8, 34]. The adsorption capacity decreased with increasing biochar pyrolysis temperatures [8, 28]. The decrease in adsorption capacity may be due to reduction of organic groups such as COOH and OH on the surface of biochar samples (Figure 1). The study used biochar produced from maple trees at 500°C and generated a Langmuir maximum adsorption capacity of $0.99\text{ mg NH}_4^{+}\cdot\text{g}^{-1}$ at pH 5.9 [35]. In the study of Thuy and Do, ammonium adsorption capacity on biochar derived from coffee husk was relatively high after 6 hours with maximum $2.8\text{ mg}\cdot\text{N}\cdot\text{g}^{-1}$ biochar [28]. The higher adsorption capacity can be attributed to the biochar being prepared at 350°C , but the heat retention time is much shorter, only 1 hour.

Langmuir adsorption constantly follows a downward trend with increasing temperatures, and the same conclusion is found in the work of Fidel et al., on certain biochar [10]. In terms of the Freundlich model, n_F greater than 1 indicates exposure to the adsorption (for n_F represents the surface exchange intensity or surface heterogeneity) [5].

According to the work in [6], better adsorption of the NH_4^{+} of biochar with lower pyrolysis temperatures was assumed to be related to cation-exchange capacity (CEC) with greater quantity carboxylase groups on the biochar's surface, where adsorbing positions locate, and this finding fits the results of this study. The results showed that NH_4^{+} adsorption on coffee husk biochar occurred mainly by cation H^{+} on functional groups containing surface oxygen; therefore, high adsorption capacity means low pyrolysis temperature. Additionally, the adsorption capacity shrinks with increasing temperatures because of the increasing competition among ions in organic minerals (K, Fe, Zn, Ca, and Mg) in biochar

TABLE 2: Correlations between physical chemistry properties of biochar.

	$t^{\circ}\text{C}$	%H	pH	pH_{pzc}	$\text{mmolH}^{+}\cdot\text{g}^{-1}$	$\text{mmolOH}^{-}\cdot\text{g}^{-1}$	%TOC	CEC, $\text{mmol}\cdot\text{kg}^{-1}$
$t^{\circ}\text{C}$	1	-0.924**	0.958**	0.984**	-0.943**	0.883**	-0.988**	-0.954**
% H	-0.924**	1	-0.970**	-0.854**	0.962**	-0.673*	0.958**	0.875**
pH	0.958**	-0.970**	1	0.919**	-0.990**	0.730*	-0.985**	-0.904**
pH_{pzc}	0.984**	-0.854**	0.919**	1	-0.905**	0.907**	-0.960**	-0.922**
$\text{mmolH}^{+}\cdot\text{g}^{-1}$	-0.943**	0.962**	-0.990**	-0.905**	1	-0.692*	0.976**	0.863**
$\text{mmolOH}^{-}\cdot\text{g}^{-1}$	0.883**	-0.673*	0.730*	0.907**	-0.692*	1	-0.800**	-0.876**
%TOC	-0.988**	0.958**	-0.985**	-0.960**	0.976**	-0.800**	1	0.940**
CEC, $\text{mmol}\cdot\text{kg}^{-1}$	-0.954**	0.875**	-0.904**	-0.922**	0.863**	-0.876**	0.940**	1

** The correlation is significant at 0.01; * the correlation is significant at 0.05.

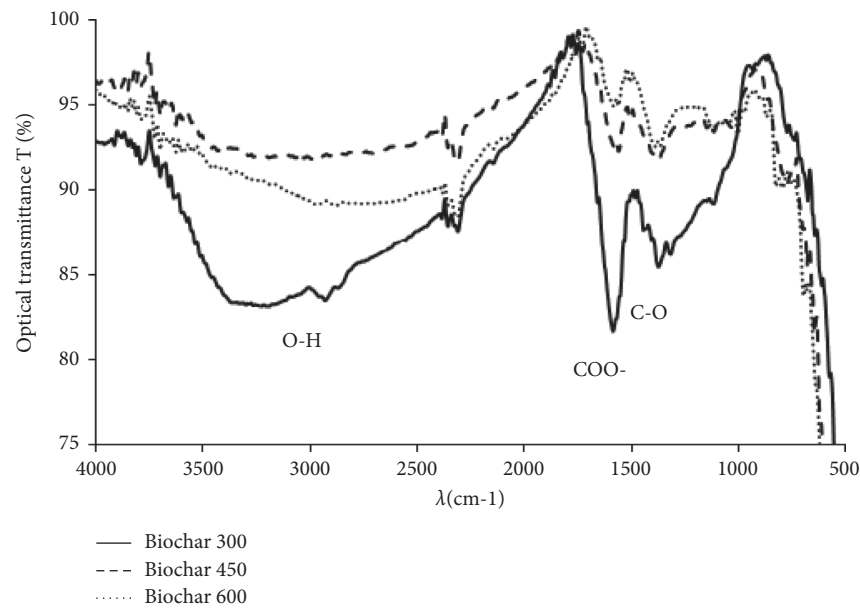


FIGURE 1: Biochar's FT-IR spectra at 300, 450, and 600°C.

with increasing temperatures [36]. Similar results were also found in the study of Fidel et al., which suggested that, with increasing pyrolysis temperature, the NH_4^+ adsorption capacity of biochar decreased [10]. This finding was also mentioned by Gao et al. who studied the ammonium adsorption capability of biochar from peanut shells, corn cobs, and cotton tree trunks at 300, 450, and 600°C and kept in 2 hours [33]. The study's result matched with the work of Begum et al. of pyrolyzed woodchips at 700°C whose result was $0.96 \text{ mg}\cdot\text{g}^{-1}$ [32].

To investigate the main factors influencing biochar's ability to adsorb $\text{NH}_4^+ - \text{N}$, correlations between q_0 , some properties of biochar, biochar pyrolysis temperature, and CEC of biochars were analyzed, Table 4. Maximum NH_4^+ adsorption capacity q_0 was positively correlated with the total organic content in the biochar (r 0.986) (Table 4). This indicated that the removal of the organic functional groups with increasing pyrolysis temperature induced the decreasing NH_4^+ adsorption capacity of biochar. CEC seemed to be the dominating factor influencing the NH_4^+ adsorption capacity of biochar. The q_0 values were positively correlated with CEC (r 0.954). That is to say, the biochar with higher CEC values had larger NH_4^+ adsorption capacity. The q_0

values were negatively correlated with biochar pyrolysis temperature, $t^{\circ}\text{C}$ (r -1, 000) so that NH_4^+ adsorption capacity of the biochar also decreased with increasing pyrolysis temperatures.

3.3. The NH_4^+ Adsorption Kinetics of Biochar. The NH_4^+ adsorption in the water process of biochar with varying pyrolysis temperatures and exposure time periods, Figure 3, showed that adsorption kinetics occurs in 3 kinetic phases: the fast phase, slow phase, and ultraslow phase. The adsorption was dramatic within the first 15 minutes in 300, 450, and 600°C biochars. The adsorption capacity reached 82.0, 83.8, and 84.2%, respectively, to the 3 biochars and then slowed down and reached the equilibrium after 45 minutes. The speed of the reaction can be explained in the fast phase, where some anionic groups with negative charges were located on the surface of the adsorbent such as $-\text{COO}^-$. These groups rapidly interacted with ammonium ions. Then, the NH_4^+ went into a slow process, which represented the adsorption in the internal part of the biochar [5]. So, the NH_4^+ adsorption on biochar goes through (hoặc chia thì quá khứ) two reaction processes.

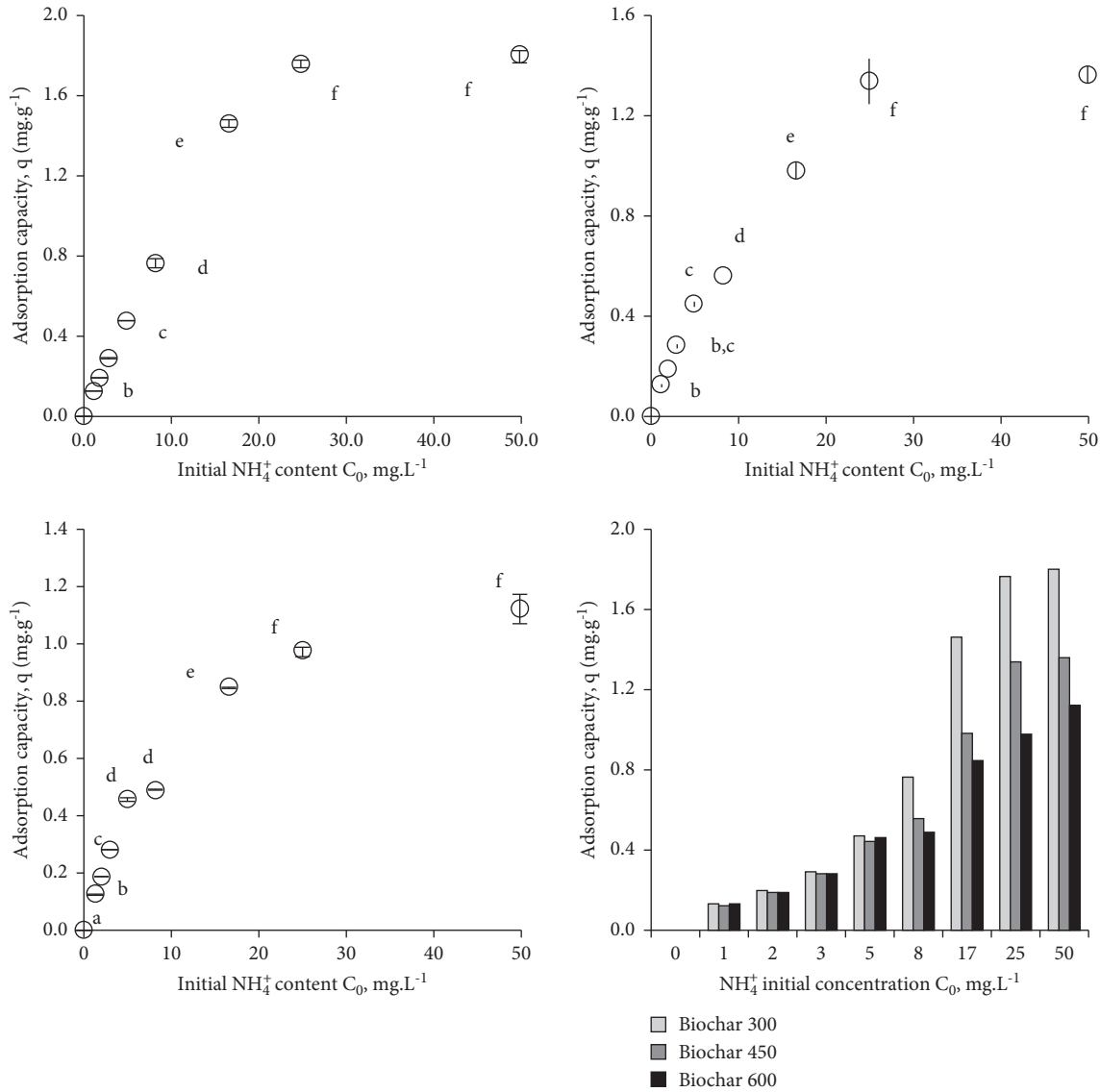


FIGURE 2: Adsorption capacity NH_4^+ ($\text{mg}\cdot\text{g}^{-1}$) vs. C_0 ($\text{mg}\cdot\text{L}^{-1}$) at different pyrolytic temperatures. Different letters represent statistically significant differences. (a) Biochar at 300°C , (b) biochar at 450°C , (c) biochar at 600°C , and (d) adsorption efficiency of 3 forms of biochar.

TABLE 3: Parameters of NH_4^+ adsorption isothermal models.

Form		Parameters of the Langmuir model				Parameters of the Freundlich model		
		K_L	R^2	q_0	q_{Exp}	K_F	n_F	R^2
Biochar 300°C	Value	2.5	0.97	1.5	1.80	0.9	1.5	0.97
	SD	0.1		0.1	0.03	0.2	0.9	
Biochar 450°C	Value	1.1	0.95	1.23	1.36	1.0	1.0	0.99
	SD	0.1		0.04	0.03	0.3	0.9	
Biochar 600°C	Value	2.3	0.96	0.88	1.12	0.8	1.7	0.97
	SD	0.5		0.06	0.05	0.4	1.3	

According in ANOVA analysis, differences in average experiment results showed, that after 30 minutes, the increasing NH_4^+ adsorption capacity was not statistically significant ($p < 0.05$) in all the 3 biochars (Figure 3). This showed that the process reached a saturated adsorption state.

Simulations from the first-order and second-order models described the experimental data fairly well with R^2 values larger than 0.89. However, only the pseudo-second-order kinetic model with q_e derived from the model and the experiment was satisfactory; specifically, the modelled

TABLE 4: Correlations between some properties of biochar and maximum NH_4^+ adsorption capacity.

	$t^\circ\text{C}$	%TOC	CEC, $\text{mmol}\cdot\text{kg}^{-1}$	$q_0 \text{NH}_4^+$, $\text{mg}\cdot\text{g}^{-1}$
$t^\circ\text{C}$	1	-0.988 **	-0.954 **	-1.000 **
%TOC	-0.988 **	1	0.940 **	0.986 **
CEC, $\text{mmol}\cdot\text{kg}^{-1}$	-0.954 **	0.940 **	1	0.954 **
$q_0 \text{NH}_4^+$, $\text{mg}\cdot\text{g}^{-1}$	-1.000 **	0.986 **	0.954 **	1

** Correlation is significant at the 0.01 level (2 tailed). * Correlation is significant at the 0.05 level (2 tailed).

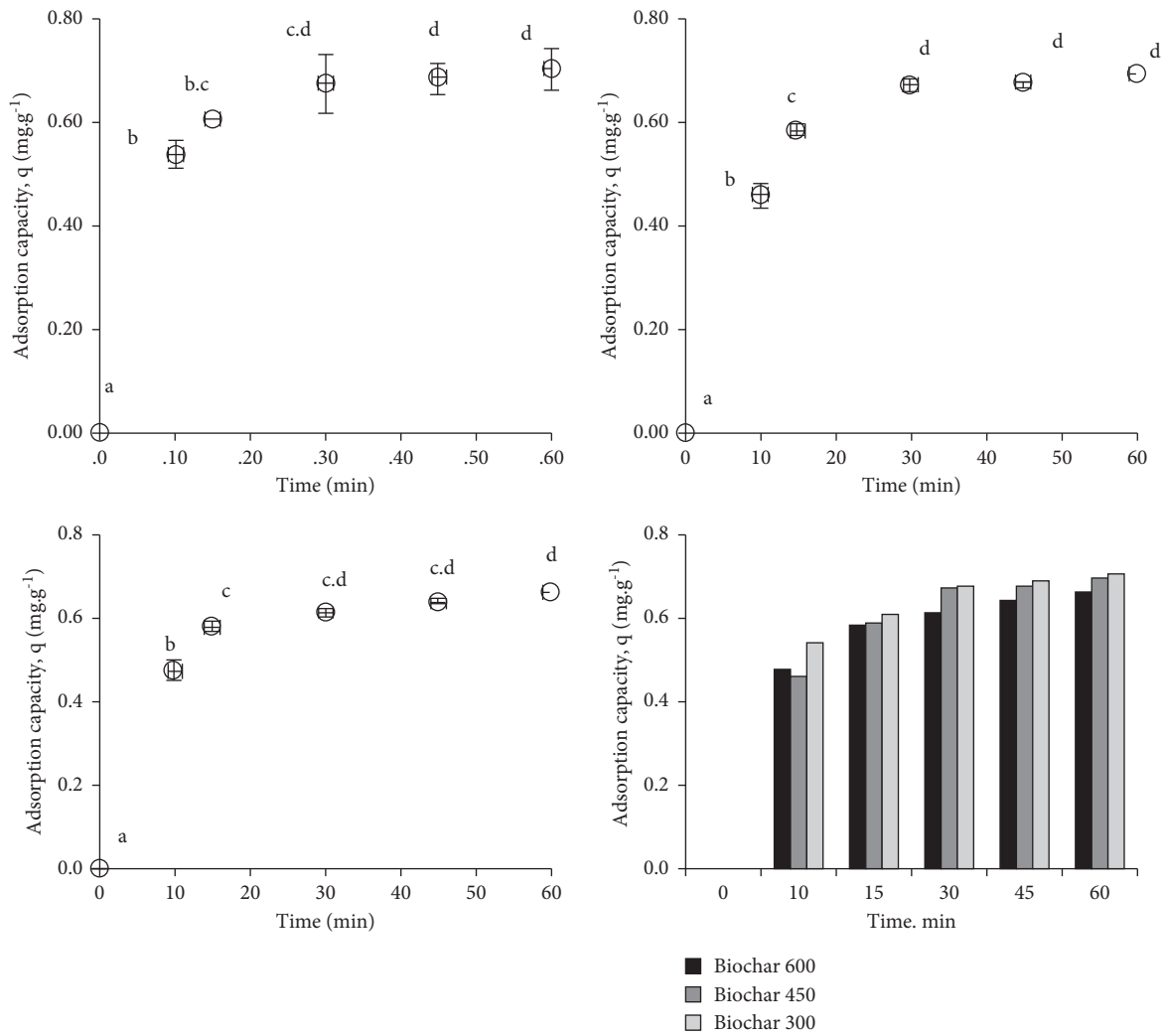


FIGURE 3: Illustration of NH_4^+ adsorption capacity (mg/g) by time t (min) of the biochar; a,b,c,d indicate statistically significant differences. (a) Biochar at 300°C, (b) biochar at 450°C, (c) biochar at 600°C, and (d) adsorption efficiency of 3 forms of biochar.

q_e of the 300, 450, and 600°C biochar was 0.77, 0.78, and 0.75 $\text{mg}\cdot\text{g}^{-1}$, respectively, while the experimented q_e was 0.74, 0.70, and 0.69 $\text{mg}\cdot\text{g}^{-1}$ (Table 5). Therefore, using the pseudo-second-order kinetic model to explain the NH_4^+ adsorption kinetic of biochar derived from coffee husks was suitable. The same was also confirmed from the

finding of Khalil et al. [5]. This also means that the kinetics was controlled by valency-related adsorptions by sharing or ion-exchanging between adsorbents and adsorbates [37]. The k_2 parameter did not behave in a particular trend when increasing the pyrolysis temperature from 300 to 600°C.

TABLE 5: NH_4^+ adsorption kinetics parameters of biochar.

Models	Pyrolytic temperature	q_e (mg·g ⁻¹)	Kinetic constant	q_e of experiment, mg·g ⁻¹	R^2
Pseudo-first-order	Biochar 300°C	-0.19	$k_1(1. \text{ min}^{-1}) = 0.025$	0.74	0.9
	Biochar 450°C	0.22	$k_1(1. \text{ min}^{-1}) = 0.051$	0.70	0.89
	Biochar 600°C	0.24	$k_1(1. \text{ min}^{-1}) = 0.065$	0.69	0.9
Pseudo-second-order	Biochar 300°C	0.77	$k_2 (\text{ g} \cdot \text{ mg}^{-1} \cdot \text{ min}^{-1}) = 0.314$	0.74	0.92
	Biochar 450°C	0.78	$k_2 (\text{ g} \cdot \text{ mg}^{-1} \cdot \text{ min}^{-1}) = 0.200$	0.70	0.93
	Biochar 600°C	0.75	$k_2 (\text{ g} \cdot \text{ mg}^{-1} \cdot \text{ min}^{-1}) = 0.247$	0.69	0.92

4. Conclusions

Coffee husks acquired from Krong Buk Town (Dak Lak Province, Vietnam) were used to transform into/produce biochar. Recovery efficiency and surface physical-chemical properties of the biochar (TOC, pH, pH_{pzc} , H^+ and OH^- groups, and CEC) at different temperatures were determined. The results showed that biochar pyrolysis temperatures were positively correlated with pH, pH_{pzc} , and mmolOH^- and negatively with %H, mmolH^+ , %TOC, and CEC. The study showed biochar derived from coffee husks with lower pyrolysis temperature had higher NH_4^+ adsorption capacity. Both Langmuir and Freundlich isotherm models fit the NH_4^+ adsorption process with R^2 in range 0.91–0.98. Time to reach adsorption equilibrium ranged from 15 to 30 minutes. The pseudo-second-order kinetic model was used to explain NH_4^+ adsorption kinetics of coffee husks to derive biochar. In conclusion, a new idea of researching on the application of biochar produced from coffee husks in water treatments by NH_4^+ adsorption is based.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflicts of interest.

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