

Research Article

An Eco-Friendly, Interference, and Solvent Free Surfactant-Assisted Dual-Wavelength β -Correction Spectrometric Method for Total Determination and Speciation of Cu²⁺ Ions in Water

Hassan Alwael,¹ Doaa S. Al-Raimi,² Khairia M. Al-Ahmary,² Hany A. Nasef,³ Amal S. Alharthi,¹ Tharawat N. Abduljabbar,¹ Liyakat H. Mujawar,⁴ Ekram Y. Danish,¹ Mohammad T. Soomro,⁴ and Mohammad S. El-Shahawi ¹

¹Department of Chemistry, Faculty of Science, King Abdulaziz University, P.O. Box 80203, Jeddah 21589, Saudi Arabia ²Department of Chemistry, College of Science, University of Jeddah, Jeddah, Saudi Arabia ³Department of Depis Science, University for Engineering and Technology 25111, Empt

³Department of Basic Sciences, ta Higher Institute for Engineering and Technology, Mansoura 35111, Egypt ⁴Center of Fuerlance in Environmental Studies King Aladeriz University, D.O. Ben 80316, Indek 21580, Soudi A

⁴Center of Excellence in Environmental Studies, King Abdulaziz University, P.O. Box 80216, Jeddah 21589, Saudi Arabia

Correspondence should be addressed to Mohammad S. El-Shahawi; mohammad_el_shahawi@yahoo.co.uk

Received 8 May 2023; Revised 11 July 2023; Accepted 20 September 2023; Published 4 November 2023

Academic Editor: Daipayan Roy

Copyright © 2023 Hassan Alwael et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Spectral interference through the presence of uninformative variables, excess reagents, and complications in the refinement of the analyte signal is common in the quest to identify complex species in real samples. Therefore, an economical green, facile, and sensitive strategy has been developed for Cu²⁺ detection using the anionic surfactant sodium dodecylsulphate- (SDS-) assisted dual-wavelength β -correction spectrophotometric strategy combined with the chromogenic reagent zincon (ZI). The low limits of detection (LOD) and quantification (LOQ) of Cu²⁺ using ordinary (single wavelength) spectrophotometry were 0.19 (3.02) and 0.63 (10.0) μ gmL⁻¹, and these values were improved to 0.08 (1.27) and 0.26 μ gmL⁻¹ (4.12 μ M)) using β -correction (dual wavelength) spectrophotometry, respectively. The LOD and LOQ were improved from 0.08 (1.27) and 0.26 (4.12) μ gmL⁻¹ to 0.02 (0.32) and 0.08 μ gmL⁻¹ (1.27 μ M) using SDS-assisted dual- β -correction spectrometry, respectively. Ringbom, *s*, and the corrected absorbance (A_c) versus Cu²⁺ concentration plots were linear over the concentration range 1.10–2.4 (17.4–38.1) and 0.50–2.40 μ gmL⁻¹ (7.94–38.1 μ M), respectively. Sandell's sensitivity index of $3.0 \times 10^{-3} \mu$ g/cm² was achieved. The selectivity was further confirmed via monitoring the impact of common diverse ions and surfactants on the corrected absorbance. Total determination and Cu²⁺ speciation in water were favorably implemented and validated by ICP-OES at 95% (P = 0.05). Satisfactory Cu²⁺ recoveries in tap (92.2–98.0%) and mineral (105–111.0%) water samples were achieved. The sensing system is simple, reliable, sensitive, and selective for Cu²⁺ detection.

1. Introduction

Copper is a crucial micronutrient for phytoplankton and in the human body, and it is an important component of human proteins and enzymes, where the lack of Cu^{2+} will hinder the physiological activities of the human body and easily cause various diseases [1–3]. Chemical speciation (labile and chelated) of copper (II) also plays a strong role in defining the bioavailability and toxicity upon exposure of copper in marine environments. In addition, Cu is a heavy metal widely discovered in the environment and excessive Cu^{2+} ions in water causes severe environmental pollution and even risk of the health of organisms [4]. Furthermore, Cu^{2+} ions beyond the recommended levels of copper causes adverse health effects, e.g., Alzheimer's disease and numerous neurological sicknesses [5]. Industrial and agricultural anthropogenic activities were responsible for a dramatically impact on the environment and human health. Maximum allowable limit (MAL) of copper contamination has been set at $1.3 \text{ mg} \cdot \text{kg}^{-1}$ (~20 μ M) by the United States-Environmental Protection Agency (US-EPA) and the World Health Organization (WHO) in drinking water and food staffs' regulations [5, 6]. Thus, searching for low-cost, sensitive, and precise reagent for total determination and speciation of Cu²⁺ in water is of great concern [7, 8].

Nowadays, numerous methods based on zincon reagent, i.e., 2-carboxy-2'-hydroxy-5'sulfo-formazyl benzene monosodium salt (Electronic Supporting Information's, ESI. 1) [9, 10], solid phase [11-20], cloud point [21-23], ionic liquid-based aqueous two-phase system [24], and dualligand Eu-MOF for fluorescence sensing [25] have been reversed-phase reported. Ligandless switchablehydrophilicity solvent liquid-liquid microextractions combined with atomic absorption spectrometry [26] have also been reported for the detection of Cu^{2+} and other metal ions. Electrothermal atomic absorption spectrometry [27, 28], inductively coupled plasma optical emission spectrometry (ICP-OES) [29], colorimetric [30-33], atomic fluorescence spectrometry [34-37], lighting up of carbon dots [38], and stripping voltammetry [39, 40] have also been reported for Cu²⁺ detection. Most of these techniques have several limitations such as time-consuming, required special sample preparation, highly sophisticated instruments, and high-skill operators [23-27, 33-40].

Ordinary spectrophotometric methods for the detection of metal ions including Cu²⁺ have many advantages such as simplicity, e, applicability, availability, easy to use, and low cost [41-51]. However, the excess of the chromogenic reagent minimizes the sensitivity and precision and limits the linear range of concentration of these methods because of the substantial interfering of the extra concentration of the colored reagent with the analyte at λ_{max} [52, 53]. In contrast, the dual-wave β -correction spectrophotometric technique has gained great attention and has a promising impact as an alternative approach due to its simplicity, low cost, portability, and elimination of the interference of the excess colorant chromogenic reagent [53, 54]. In the dual-wave β -correction spectrophotometric technique, quantification of the correct absorbance equivalent to the fractions of the chromogenic reagent that reacted with the analyte in the presence of excess of the chromogenic reagent is very much possible for precise analysis of the analyte [52-54]. β -Correction approach offers simple, rapid, costeffectiveness, and selective over most of the accessible modern instrumentation and also improves the sensitivity, precision, and accuracy of the ordinary single-wavelength spectrophotometry by solving the problem arising from the interference due to the excess chromogenic reagent [54–58]. Thus, dual-wave β correction spectrophotometry is the most significant and well-defined aspect for measuring the correct (real) absorbance of the formed colored Cu²⁺-zincon complex.

Recently, speciation of copper species forms of copper is essential for analytical laboratories in the copper industry because of the technological importance of such analytical information [59, 60]. Kumar et al. [59] have reported that,

ordinary natural organic compounds such as humic acid (HA), fulvic acid (FA), phenols, and surfactants can complex with copper, influencing its speciation and decreasing its bioavailability. To the best of our knowledge, zincon reagent has been used for detection of Cu²⁺ and other metal ions in water samples using only ordinary single-wavelength spectrophotometric methods [6, 15, 40-43]. Therefore, the current study was aimed to: (i) developing a low cost, and selective surfactant assisted β -correction spectrophotometric assay for total determination and copper speciation in water using zincon (ESI. 1) and (ii) assigning the stoichiometry, stability, and thermodynamic behavior of Cu²⁺ -ZI chelate. A cohesive collaboration of industry and academic institutes will be desired to miniaturize and automate the developed assay, where it has the advantages of miniaturization, automation, simplicity, and sensitivity.

2. Experimental

2.1. Apparatus. The electronic spectra and absorbance of the reagent and its Cu²⁺-zincon complex were recorded using UV-Vis spectrophotometer (Shimadzu UV-Vis 1800, Japan) connected to a Shimadzu TCC-ZUOA temperature controller unit. A Perkin-Elmer ICP MS (Sciex model Elan DRC II, USA) was also employed as a standard technique for copper analysis at the optimized operational parameters summarized in ESI. 2. A Perkin Mattson 5000 FTIR spectrometer was used for recording the FTIR. A Volac digital micropipette (10–100 μ L) and a Jenway pH-meter (model 3510) were used for preparation of diluted solutions in deionized water and pH measurements, respectively.

2.2. Chemicals and Reagents. Analytical reagent (A.R.) grade chemicals and reagents were used as received. All laboratory glassware's including high-density polyethylene (HDPE) bottles were soaked in hot detergent, soaked in HCl solution (50% v/v)-conc. HNO₃ (2.0 M) at 1:1 v/v ratio rinsed with deionized water and finally dried at 80°C in an oven. Sodium salts of humic acid, fulvic acid, and phenol HA were purchased from Sigma-Aldrich. HDPE sample bottles were soaked overnight, washed with HNO₃ (10%, v/v) solution, and rinsed with deionized water prior to use, and finally placed in precleaned HDPE. A stock solution of Cu²⁺ (1 mg/ mL) was prepared by dissolving the appropriate mass of Cu (NO₃)₂. 3H₂O (BDH, Poole, England) chemicals) in Milli-Q water. Zincon reagent and other metal salts were purchased from Shanghai Macklin Biochemical Co., Ltd. (Shanghai, China). More diluted standard Cu²⁺solutions were prepared by dilution with deionized water. A stock solution of zincon $(1.0 \times 10^{-3} \text{M})$ was prepared in minimal amount of ethanol, and the solution was completed to the mark with Milli-Q water. Stock solutions (1000 μ g/mL) of the salts AgNO₃, MgCl₂, BaCl₂, CaCl₂, CdCl₂, Co (NO₃)₂, Fe₂ (SO₄)₃, HgCl₂, KCl, MnSO₄, KMnO₄, NiSO₄, Pb (NO₃)₂, ZnSO₄, NaCl⁻, and KIO₃ were prepared individually by dissolving the required weights of their salts in Milli-Q water. More diluted concentrations of these salts were also prepared by proper dilution with deionized water to give varying concentrations of diverse ion. A series of Britton–Robinson (B-R) buffer of pH 2–11) was prepared as reported [61]. The surfactants benzyldimethyltetradecylammonium chloride (BTAC), sodium dodecyl sulfate (SDS), and Triton X-114 (T-X 114) were purchased from Sigma (Oregon, MO, US). Stock solutions (100 μ g/mL) of the surfactants were prepared individually in water as model "surfactants." Millipore water (18.25 M Ω cm⁻¹) was provided from the Milli-Q Plus system (Millipore, Bedford, MA, USA) in all experiments. Stock and more diluted Cu²⁺, HCl (0.1 M), and NaOH (0.1 M) solutions were prepared from deionized water.

2.3. Recommended Procedures for Cu^{2+} Assay. A series of measuring flasks (10 mL) containing standard concentrations (0.05–2.4 µgmL⁻¹) of Cu²⁺ ions was mixed with an excess known concentration (5.0×10^{-5} M) of zincon in the presence of BR buffer of pH 3. The solution mixtures were completed to the mark with Milli-Q water in the presence and absence of SDS ($10 \mu g m L^{-1}$). The reaction mixtures were allowed to stand for 3–5 min at room temperature. The absorbance of the solutions was measured at 465 nm (λ_1) and 625 nm (λ_2) against reagent blank. The true absorbance (A_c) of the Cu²⁺-zincon in the test solutions was then computed successfully for the construction of the calibration plot.

2.4. Analytical Applications

2.4.1. Determination of Cu^{2+} in Water Samples. In dark precleaned glass HDPE bottles glass, tap water samples (200 mL) were collected as real samples of Cu²⁺ ions from the domestic tap water, which was left to run for 20–25 min prior to sampling at the Chemistry Laboratory of the Center of the Excellence in the Environmental Studies, King Abdulaziz University, Jeddah city, Saudi Arabia. The samples were spiked with known concentrations of Cu²⁺(0.4–2.4 µg mL⁻¹) and instantly filtered through a 0.45 µm pore-size cellulose membrane filter (Millipore Corporation) to remove all suspended particles and stored in HDPE sample bottles at 4°C prior to analysis. Cu²⁺ ions in the sample solutions were determined from the linear calibration plot of the developed dual-wave β -correction procedures using the following equation:

$$\left[\mathrm{Cu}^{2+}\right] = b \frac{C_{\mathrm{std}}}{m} V_x,\tag{1}$$

where *b* and *m* are the intercept and slope of the linear calibration plot, respectively, C_{std} is the known Cu²⁺ concentration, and V_x is the sample volume. Alternatively, the standard addition method was performed as follows: The corrected absorbance was computed in the absence and in the presence of known fractions (20.0–100 μ L) of the standard Cu²⁺ under the optimized parameters. The corrected absorbance (A_c) of each solution was subsequently evaluated, and the Cu²⁺ content was then calculated via the extrapolated abscissa of the linear plot of the standard addition, employing the following equation:

$$Cu^{2+}] = C_{std} \times \frac{Ac_{sam.}}{Ac_{std.}},$$
 (2)

where C_{std} is the known Cu²⁺ concentration, Ac_{sam} and Ac_{std} are the real absorbance exhibited by the unknown and after adding known Cu²⁺ concentrations, respectively.

2.4.2. Total Determination and Speciation of Cu^{2+} in Water. An approximate volume (0.3–0.5 L) of the tap water (TW) and mineral water (MW) samples were collected in HDPE bottles, filtered through a $0.45 \,\mu m$ pore size cellulose membrane filter, and stored in precleaned HDPE sample bottles (0.5 L) at 4°C. Known volumes (5.0 mL) of the water sample adapted to pH 3 were transferred to measuring flasks (25.0 mL) containing zincon and standard Cu²⁺ concentrations at the optimized conditions. The solutions mixtures were completed to the mark with Milli-Q water, and the absorbance at λ_1 and λ_2 was recorded. The corrected absorbance's were calculated before and after spiking of standard Cu²⁺ concentrations from the standard addition linear plot. Another water sample was exposed to UV radiation at 254 nm for 4 h in the HCl (10% v/v), stored in HDPE bottles, and subjected to Cu²⁺ analysis within one day of collection as follows: Transfer an accurate volume (5.0 mL) of prefiltered water sample onto a series of measuring flasks containing zincon at the optimal parameters. Based on these bases, the Ac of the first aliquot (Ac_1) will be a measure of labile Cu^{2+} ions in the mixture, while the Ac of the second aliquot (Ac₂) is a measure of the sum of labile and chelated Cu²⁺ with organic matter in the aliquot. The difference of corrected absorbance (Ac_2-Ac_1) is a measure of the complexed fractions of Cu^{2+} in water samples.

3. Results and Discussion

3.1. Electronic Spectra of Zincon and Its Cu^{2+} -Chelate Omitted. Zincon reagent (ESI. 1) contains four protonating groups: two acidic, sulfonic (pKa1) and carboxylic (pKa2); and two basic, a secondary amine (pKa3) and a phenolic one (pKa4). The most acidic group of zincon is the sulfonic group, which is usually omitted since zincon is commercially available as a monosodium salt and because of its rapid decomposition in acidic pH [16, 49]. Spectrophotometric measurements of zincon display significant change of the absorption at 565 nm around pH 4, which is characteristic for the carboxylic group rather than the sulfonic one. The UV-visible spectrum of zincon vs. water displayed distinct peak at 465 nm (λ_1) and was safely assigned to $n \longrightarrow \pi^*$ electronic transition (Figure 1, curve A) [62, 63]. The UV-visible spectrum of the reaction product of zincon with Cu²⁺ at pH = 3 (Figure 1 curve B) revealed broad and ill-defined peak like shoulder in the range 563-569 nm and a strong absorption peak at 600 nm. These peaks were safely assigned to charge transfer (L \longrightarrow MCT) and electronic $d \longrightarrow d$ transitions from shorter to longer wavelength in tetrahedral environment, respectively 16, [63]. The spectrum of Cu^{2+} -



FIGURE 1: Absorption UV-visible spectra of zincon $(1.0 \times 10^{-4} \text{ (M)})$ (reference water) (A); Cu²⁺-ZI complex (47.62 μ M) (reference water) (B) and Cu²⁺-ZI complex (reference zincon reagent) (C).

zincon chelate vs. zincon (Figure 1, curve C) displayed strong peak at 625 nm (λ_2). The experiential bathochromic (red) shift and the high value of the molar absorptivity ($\varepsilon = 2.5 \times 10^4$ L M⁻¹ cm⁻¹) suggested the suitability of the produced colored Cu²⁺-zincon chelate for establishing simple, cost-effectiveness, and reliable β -correction spectrophotometry approach for Cu²⁺ detection and speciation of Cu²⁺ species in water.

In the aqueous reaction media, the interference popping up from extra zincon reagent can be eliminated and the true (corrected) absorbance (Ac) of the formed Cu^{2+} -zincon chelate can be computed employing the equation [53–59]:

$$Ac = \frac{\Delta A - \Delta A'}{1 - \alpha \beta},$$
(3)

where ΔA is the absorbance of the Cu²⁺-zincon complex at λ_2 , whereas $\Delta A'$ is its absorbance at λ_1 versus zincon (reference blank). The spectral parameters α and β were computed using the following equations [56, 57]:

$$\beta = \frac{A_0}{A'_0} = \frac{\varepsilon_L^{\lambda 2}}{\varepsilon_L^{\lambda 1}},\tag{4}$$

$$\alpha = \frac{A'_{\alpha}}{A_{\alpha}} = \frac{\varepsilon_{\rm ML}^{\lambda 1}}{\varepsilon_{\rm ML}^{\lambda 2}},\tag{5}$$

where A_{α} is the absorbance of the produced Cu²⁺-zincon chelate in solution at λ_2 while A'_{α} is it's absorbance at λ_1 versus water (blank). A'_0 and A_0 are the absorbance of the blank solution at λ_1 and λ_2 , against water, respectively. Thus, the planned β -correction spectrophotometry assay offered good sensitivity over ordinary spectrophotometry by suitable choice of λ_1 and λ_2 at the sink and the peak of the visible spectrum of Cu²⁺-zincon chelate versus reagent blank [53–59], respectively. The lowest and the highest absorbance of Cu²⁺-zincon chelate at λ_1 (465 nm) and λ_2 (625 nm) versus zincon (Figure 1, curve C) was used for calculating the corrected absorbance. Employing single wavelength spectrophotometry, the absorbance of the developed Cu²⁺zincon chelate at λ_2 versus zincon was lower than the corrected absorbance (Ac) computed via dual-wave β -correction spectrophotometry [53, 54] in consistence with the data reported [58, 59]. The value of β computed from Figure 1 (curve A) and equation (4) was 0.18, whereas the value of α calculated from Figure 1 (curve B) and equation (5) was 2.18 in close consistence with the data published [53–59].

3.2. Programing of the Analytical Parameters. To explore the impact of pH (pH 1.0–10.0) of the aqueous solution on the developed Cu²⁺-zincon colored chelate, the electronic spectra and the actual absorbance of Cu²⁺ ($3.0 \mu \text{gmL}^{-1}$) solution containing zincon (1.0×10^{-4} M) were recorded at various pH. The absorbance of the produced colored Cu²⁺-zincon complex reached its maximum value at pH 3.0 (Figure 2(a), dotted line). The binding sites of phenolic OH and azo (-N=N-) groups of zincon reagent at pH 3 are capable to coordinate with Cu²⁺ [54] in consistence with the data published for the Cu²⁺-zincon complex [54, 55]. Therefore, the solution pH was adopted at pH 3 in the following study.

An aqueous solution of Cu^{2+} (3.0 μ g mL⁻¹) at pH 3 was allowed to react with various known concentrations of zincon (2.5×10⁻⁵-2.5×10⁻⁴ M). The results are demonstrated in Figure 2(a) (column), where zincon concentration of 1.0×10⁻⁴ M was found enough for detection of Cu²⁺ up to 5.0 μ g mL⁻¹ in the solution. The calculated molar ratio of ligand to Cu²⁺ was found 2:1 suggesting formation of Cu²⁺-ZI complex of the formula Cu (L)₂, where L = ZI (ESI. 1).

The impact of the solution temperature $(10-50^{\circ}\text{C})$ on the absorbance of the Cu²⁺-zincon $\{3.0 \ \mu\text{g mL}^{-1} \text{ Cu}^{2+}\}$ at pH 3 was examined. The data are displayed in Figure 3. The absorbance of the formed Cu²⁺-zincon chelate increased on growing temperature up to 25°C, followed by a gradual decrease in the absorbance. The degradation of the formed Cu²⁺-zincon chelate and/or the decrease in the interaction between Cu²⁺ ions and ZI at a temperature higher than 25°C are most likely accounted for the observed trend. Hence, a room temperature (25 ± 1°C) was selected as a suitable condition for the formation of the Cu²⁺-ZI complex.

The absorbance of Cu^{2+} -ZI at known Cu^{2+} (2.0 µg mL⁻¹) and zincon (1.0×10⁻⁴ M) concentrations after mixing was recorded immediately over a wide range of time (0.0–80 min at pH 3. The absorbance and the corrected absorbance (A_c) of the produced Cu^{2+} -ZI complex were measured at several shaking time intervals (0.0–85 min) employing single wavelength and β -correction spectrophotometry, respectively. The colored Cu^{2+} -ZI complex was established within 1-2 min of shaking, and the absorbance remained constant up to a standing time of 85 min (ESI. 3). These data added further provision to the analytical utility of the established Cu^{2+} -ZI complex for developing a solvent-free β -correction spectrophotometry assay for Cu^{2+} detection in water. Thus, in the next study, the absorbance of the Cu^{2+} -Zi complex was measured within 80 min of mixing at pH 3.

The influence of various proportions $(0.0-1000 \,\mu\text{L})$ of NaCl $(1.0 \times 10^3 \,\mu\text{gmL}^{-1})$ and standing time $(0.0-70 \,\text{min})$ on the absorbance of the tested Cu²⁺-Zincon complex at pH 3 under the optimal parameters was also studied. The plots of



FIGURE 2: Plots of the corrected absorbance of Cu^{2+} (47.62 μ M) versus solution pH and zincon concentrations (2.5×10^{-5} - 2.5×10^{-4} (M) at 625 nm at room temperature.



FIGURE 3: Plots of the corrected absorbance of Cu²⁺ (47.62 μ M)zincon complex versus solution temperature (10–50°C) at 625 nm at pH 3. Inset plot of ln K_c vs. 1000/T (K⁻¹) of the Cu²⁺-ZI complex. Conditions: [Cu²⁺] = 47.62 μ M and [ZI] = 1.0 × 10⁻⁴ M.

the absorbance of the formed Cu^{2+} -ZI complex in the presence of various volumes of the salt added and time are shown in Figure 4. In the absence of the salt added, the corrected absorbance of the Cu-ZI complex computed for the added salt concentration was about 4.2% (Figure 4). These results, clearly simplifies that Cu^{2+} complexation with ZI was less influenced by the salt added to the medium. Thus, in the next study, no salt was added to the reaction medium.

3.3. Thermodynamic Parameters of Cu^{2+} -ZI Complex. The thermodynamic features of the developed Cu^{2+} -ZI complex in the temperature range of 293–323 K were determined. The Cu^{2+} species present as neutral species at pH 3 only one species of Cu^{2+} -ZI chelate existed, and no precipitation obtained. On rising the solution temperature from 293 to 323 K, the equilibrium constant (K_c) decreased signifying that, the complex formation is exothermic process [64, 65]. The slope and intercept of the linear plot of ln K_C versus 1000/T (Figure 3, inset) were used for calculating the ΔH , ΔS , and ΔG of the formed Cu^{2+} -ZI complex. The ΔH , ΔS ,

and ΔG were found -5.3 kJmol^{-1} , 72.75 J mol $^{-1}$ K $^{-1}$, and $-26.97 \text{ kJ mol}^{-1}$ (at 298 K), respectively. The value of the ΔH (-5.3 kJ mol^{-1}) reveals the exothermic nature and the bond

energy difference between zincon and its Cu²⁺ complex. Growing temperature minimizes Cu²⁺ interaction with ZI, resulting in decrease in the percent yield of the complex. The ΔG value (-26.98 kJ mol⁻¹) at 298 K decreased on increasing temperature, supporting the spontaneous nature of the complex.

3.4. Selectivity Study

3.4.1. Impact of Diverse Ions. The analytical utility of the established β -correction spectrophotometry for Cu²⁺ (3.0 μ g mL^{-1} (47.62 μ M) in the presence of excess of various potentially interfering major ions with Cu²⁺ at concentrations representative of many fresh waters, tap, and mineral water at the optimal pH 3 and ZI $(1.0 \times 10^{-4} \text{M})$ is critical. Thus, the interference of of ions Cl⁻, Br⁻, OH⁻, NO₃⁻, SO₄²⁻, and metal ions, e.g., Na⁺, K⁺, Cd²⁺, Ni²⁺, Sr²⁺, Mn²⁺, Fe³⁺, Co²⁺, Pb^{2+} , Hg^{2+} , and Ag^+ , and the oxy ions (MnO₄⁻, IO₃⁻, and WO_4^{2-}) was studied individually in the presence of Cu^{2+} -ZI complex at the optimized parameters of pH and zincon concentrations. The absorbance of Cu²⁺-ZI complex was compared with that in the presence of the interfering species. The tolerance limit (acceptance edge) (m/m) was distinct as the added concentration of the interfering species producing a relative standard deviation (RSD) of ±5% of the true absorbance of Cu²⁺-ZI chelate. The results obtained are summarized in Table 1. The ions Cl⁻, Br⁻, NO₃⁻, SO₄²⁻, IO_3^{-} , Na⁺, K, and Ca²⁺ revealed negligible change in the corrected absorbance of Cu²⁺-ZI complex at 1:1000 molar excess of Cu^{2+} to the diverse species. The ions Cd^{2+} , Ni^{2+} , Sr^{2+} , Mn^{2+} , Cr^{3+} , Hg^{2+} , and Pb^{2+} were tolerable up to 50-fold excess to Cu^{2+} . The ions Cr^{3+} , Hg^{2+} , and Pb^{2+} at level up to 20 fold greater than Cu²⁺ ions did not interfered on the absorbance of Cu²⁺-zincon chelate, whereas Fe³⁺, Co²⁺, and Ag⁺ ions interfered extremely with the complex. The interference of Fe³⁺ and Co²⁺ was masked by adding a few drops of NaF (0.1% m/v) via the formation of colorless $[FeF_6]^{3-}$ complex, whereas Co²⁺ interference was minimized by adding ethanolamine (0.01%) to form a colorless complex in the test aqueous solution. The oxyanions MnO_4^- and WO_4^{2-} were tolerable by adding a few drops of sodium azide (NaN_3) .

3.4.2. Effect of Surfactants. The impact of surface-active agents (0.1–10 ppm) on the stability and the corrected absorbance of Cu^{2+} -zincon chelate in the developed procedure are critical. Thus, the impact of all kinds of surfactant such as cationic (BTAC), anionic (SDS) and nonionic (Triton X-100) on the selectivity of the established assay for Cu^{2+} was studied at the optimized condition. Cationic and nonionic surfactants have nonsignificant changes on the absorbance of the developed colored Cu^{2+} -ZI chelate, whereas in the presence of SDS, a synergistic increase in the value of the absorbance was only seen (Figure 5). The impact of SDS may be due to its ability to form versatile interactions including



FIGURE 4: Influence of standing time (0.0-70 min) and various volumes $(0.0-1000 \,\mu\text{L})$ of NaCl concentration $(1.0 \times 10^3 \,\mu\text{g mL}^{-1})$ on the absorbance of Cu-ZI complex. Conditions: $[\text{Cu}^{2+}] = 47.62 \,\mu\text{M}$, $[\text{ZI}] = 1.0 \times 10^{-4} \,\text{M}$ and solution pH 3 at 625 nm.

TABLE 1: Selectivity data and tolerance limit of diverse ions in Cu^{2+} determination by the developed dual-wave β -correction spectrophotometry.

Diverse ions	Tolerance limit $\mu g mL^{-1}$ (μM)
Cl ⁻ , Br ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ , IO ₃ ⁻ , Ca ²⁺ , Na ⁺ , K ⁺	1000 (15873.02)
Cd ²⁺ , Ni ²⁺ , Sr ²⁺ , Mn ²⁺ , Cr ³⁺ , Hg ²⁺ , Pb ²	50 (793.65)
Fe^{3+} , Co^{2+} , Ag^+	20 (317.46)
MnO_{4}^{-}, WO_{4}^{2-}	100 (1587.3)

electrostatic, hydrophobic, bi-bi interaction, complex ion association, and/or H bonding with the Cu²⁺ complex. The change in the effective microenvironment by SDS micellar solution around Cu²⁺ in the aqueous solution and their contribution to the physicochemical features such as rate constant and spectral characteristics may also participated in the trend observed. The possible association between the cationic Cu²⁺-zincon complex and SDS as a bulky anion by forming ternary complex ion associate between SDS may also enhanced the molar absorptivity of the produced ternary complex ion associate. The available hydroxy groups and water molecules may also screened by SDS at the boundary and subsequently resulting in a worthy association between ZI and Cu^{2+} ions. Thus, it is worthy to note that, the use of SDS is attractive in developing surfactant assisted β -correction spectrophotometry for Cu²⁺ detection owing to its low cost, toxicity, and reduced environmental impact. Thus, the effect of SDS in the absorbance was continual to improve the detection of Cu^{2+} .

3.5. Stoichiometry and Stability of Cu^{2+} -ZI Complex. The stoichiometry and stability of the formed Cu²⁺-ZI chelate are critical in testing the analytical utility of the colored Cu²⁺-zincon chelate for Cu²⁺ detection [59–62]. The impact of standing time (0.0–100 min) on absorbance of the Cu²⁺-ZI



FIGURE 5: Absorption UV-visible spectra of Cu²⁺-ZI chelate at zero and after addition of known concentrations ($10.0 \,\mu \text{gmL}^{-1}$) of SDS, BTAC, and Triton114 individually. Condition: [Cu²⁺] = 4.7 μ M, [ZI] = 1.0×10^{-4} M and solution pH 3 at 625 nm.

chelate at 625 nm using Cu^{2+} (3.0 μ gmL⁻¹) and 1.0 × 10⁻⁴ M ZI at pH 3 was recorded (ESI. 3). The data revealed good stability over a period up to 100 min, revealing the good stability of the produced copper (II) chelate. Assuming the existence of one complex species of Cu²⁺-ZI, Job's and mole ratio methods at pH 3 [62, 63] were used to determine the stoichiometry and stability of the formed Cu²⁺-ZI chelate. The results of Job's (ESI. 4) suggested formation of 1:2 stoichiometry of Cu²⁺-zincon complex. The stoichiometry of Cu²⁺: zincon was also supported from the mole ratio plot (ESI. 5) [62]. Reflectance electronic spectrum of Cu²⁺-zincon complex also displayed broad peak cantered at 17241cm⁻¹ interpretable in terms of square planar stereochemistry [16, 66, 67]. The FTIR spectra of the free zincon reagent and its copper (II) chelate are displayed in ESI. 6. The v(C=N)band in the complex was found in the same position of the FTIR of free zincon indicating no participation of the azomethine in the complex formation with Cu²⁺ ions [16, 66]. The spectra added further support to the participation of the azo (-N=N-) group N and the involvement of the phenolic OH in the complex formation with Cu²⁺ ions (see ESI. 6) [66]. Thus, it can be concluded that, the zincon coordinated to Cu²⁺ via two N and two O of the azo and phenolic OH groups, and the structure of the produced copper-zincon complex can be postulated as [CuL₂], where L = zincon and the most probable structure of Cu^{2+} -zincon chelate, can be proposed as shown in ESI. 1.

The stability constant (K) of the Cu^{2+} -ZI complex was further successfully computed from Job's plot (ESI. 4), where the extrapolated absorbance ($A_{extp.}$) near to the equivalence point corresponds to the absorbance of the Cu^{2+} -ZI complex. Based on the Job's plot (ESI. 4), the Cu^{2+} -ZI complex is dissociated in the area of extrapolation, and the true absorbance of the produced Cu^{2+} complex is to some extent lower than the hypothetical value. Thus, the produced Cu^{2+} -ZI complex can be stated by the following chemical equilibrium [62]: International Journal of Analytical Chemistry

$$Cu^{2+} + 2L^{-} \rightleftharpoons [CuL_2], \qquad (6)$$

where L = zincon ligand (L) in the complex formation, and [CuL₂] refer to the formed Cu²⁺ chelate, respectively. The fraction of the true corrected absorbance (Ac) at a given value on the X-axis of Job's plot (ESI. 4) to the extrapolated absorbance (A_{exp}) determined from the extrapolated lines corresponding to the same point in the X-axis is equivalent to the mole fraction of the formed chelate [CuL₂]. The K_s value of the formed [CuL₂] complex was also computed from Job's plot (ESI 4) using the following equation (7) [62]:

$$K = \frac{\left(A/A_{\text{expt}}\right).C}{\left(C_m - \left(A/A_{\text{expt}}\right).C\right)\left[C_x - \left(A/A_{\text{expt}}\right).C\right)},\tag{7}$$

where C_m is the molar analytical concentration of the metal, C_x is the total molar analytical concentration of the ligand depending on the controlling reactant at the end point and *n* is ligand to metal ratio in the Cu²⁺-zincon chelate, respectively. The average computed K_s value (2.0×10^6) displayed acceptable stability and the suitability of the Cu²⁺-ZI complex for developing spectrophotometric method for Cu²⁺ determination.

3.6. Analytical Performance and Figures of Merits. At pH = 3 and zincon concentration $(1.0 \times 10^{-4} \text{ M})$, Beer's-Lambert plots of the absorbance versus Cu²⁺ concentration were found linear over the range $0.50-2.4 \,\mu\text{g/mL}$ (7.9–38.1 μM) and $1.6-3.0 \,\mu\text{g/mL}$ (25.4–47.62 μM) employing the developed β -correction and ordinary spectrophotometry, respectively. The obtainable results are illustrated in Figure 6(a). The calibration plots using single wavelength and the established β -correction spectrophotometry were conveyed by the following regression equations (8) and (9), respectively:

$$A = 1.6 \times 10^{-1} \,\mathrm{Cx} - 7.5 \times 10^{-3} \big(R^2 = 0.9907 \big), \tag{8}$$

Ac =
$$3.9 \times 10^{-1}$$
Cx - 1.7×10^{-1} ($R^2 = 0.9567$). (9)

The molar absorptivity (ε) and Sandal's sensitivity index [67, 68] computed from Beer's–Lambert plots of Cu²⁺-ZI complex were 2.5×10^4 L mol⁻¹ cm⁻¹ and $0.003 \,\mu g$ cm⁻² using the β -correction assay and 1.0×10^4 L mol⁻¹ cm⁻¹ and $0.006 \,\mu g$ cm⁻² utilizing single wavelength spectrophotometry. Thus, the established β -correction spectrophotometric assay presented a worthwhile approach for improving the sensitivity and selectivity of Cu²⁺ in sensing over the ordinary one.

Ringbom's plot [67] of log Cu²⁺ concentration versus percent transmittance (T%) was assumed to quantify the optimal concentration range of Cu²⁺ for the system that follows Beer's–Lambert equation (Figure 6(b)). The effective linear concentration range for Cu²⁺ by the developed β -correction was in the range 1.1–2.4 µg mL⁻¹ (17.5–38.1 µM). The LOD and LOQ values of Cu²⁺ [68] using ordinary spectrophotometry were 0.19 µg mL⁻¹ (3.0 µm) and 0.63 µg mL⁻¹ (10.0 µm) and significantly improved to 0.08 µg mL^{-1} (1.27 µm) and 0.26 µg mL^{-1} (4.13 µM) Cu^{2+} ions employing the established dual-wavelength β -correction spectrophotometry, respectively. These results reflected the significant improvement of the sensitivity by about 50%. The linear dynamic range (LDR) for the β -correction spectrophotometry embedded in Figure 6(a) (right inset) is higher than that corresponding to the uncorrected absorbance. The LOD of the established assay based on corrected absorbance (Ac) was significantly improved. The β -correction assay exhibited better recoveries (91–108%) with a relative standard deviation (RSD) in the range 3-5% lower than the ordinary single wavelength method. The β -correction assay has also displayed great advantages in improving the peak shapes and reducing matrix effects compared to ordinary spectrophotometry (Figure 6(a)). The established assay took the advantage of the reduced matrix effects without complex preprocessing of the samples, greatly simplifying the experimental process on comparing with the existing methods for Cu^{2+} (Table 1) [41–51].

Further, the LOD and LOQ of the surfactant- (SDS-) assisted β -correction spectrophotometry were enhanced from 0.08 μ g mL⁻¹ (1.27 μ M) and 0.26 μ g mL⁻¹ (4.13 μ M) to 0.02 μ g mL⁻¹ (0.32 μ M) and 0.078 μ g/mL (1.24 μ M), respectively. The change in the effective microenvironment by micelles solution around Cu²⁺ ions may also improve formation of ternary ion associate of Cu²⁺-zincon complex and SDS. The hydroxy groups and the available free water molecules may also screened by SDS at the boundary and successively resulting in better organization between ZI and Cu²⁺ions.

The precision and accuracy of the proposed β -correction spectrophotometry assay were further computed from the recovery of three replicate measurements of known Cu²⁺ concentrations in water. The results demonstrated in Figures 6(a) (Table inset) revealed good performance of the planned β -correction spectrophotometric assay and support the current protocol for Cu²⁺ detection in water. Further, a judgment between the efficacies of the established β -correction spectrophotometry with several reported spectrometric methods is summarized in Table 2. The LOD, LOQ, and LDR of developed surfactant assisted-correction spectrophotometry assay are favorably associated with most of the established spectrophotometric protocols (Table 2). The LOD of the planned β -correction assay was higher than the LOD $(0.018 \,\mu g \ L^{-1})$ using paper-based chip for fluorescence Cu²⁺ detection (Table 2). However, the measured value by the established assay below the acceptable limit of Cu²⁺ fixed by WHO and US-EPA in water. The proposed assay frees from the interfering of various anions and cations present in water samples. Thus, the proposed assay could be suitable for Cu²⁺ detection in water and it has the benefits of low cost, simple, practical, and eco-friendly.

3.7. Analytical Applications and Validation of the Established Methodology. Due to the unavailability of certified reference materials (CRM) for Cu^{2+} to check the reliability and validity of the established assay for Cu^{2+} detection, known concentrations $(0.4-2.4\,\mu\text{gmL}^{-1})$ of Cu^{2+} were spiked into



FIGURE 6: Calibration plot for Cu²⁺ detection using ordinary single wavelength and β -correction spectrophotometry (a). Inset calibration curve using the corrected (A) and Ringbom's plot using β -correction spectrophotometry of Cu²⁺-ZI complex (b). Conditions: [ZI] = 1.0×10^{-4} M and solution pH 3 at λ_{max} 625 nm.

TABLE 2: A comparison between the figures of merits (μ M) of the developed β -correction and some of the reported spectrophotometric methods for Cu²⁺ detection^{*}.

Matrix	LDR (µM)	LOD (µM)	Remarks	Ref.
Vegetables and tea	0.0-16.25	0.111	Accurate, reliable, and time consuming	[41]
Soil and vegetables	2.54-20.63	0.841	pH range 8–10, interference from	[44]
	6.98-16.67	2.333	polyhydroxy species	[1 1]
Synthetic mixture and water	16.19-128.89	0.508	Required heating and time consuming	[45]
Natural water and pharmaceutical samples	1.59–158.73	0.159	High molar absorptivity and tolerant to many foreign species	[46]
Alloy, pharmaceutical, fertilizer, and environmental samples	Up to 277.78	6.587	Use hazard organic solvents	[47]
Food, leafy vegetables, and fertilizers environmental samples	Up to 277.78	9.524	Efficient and selective	[48]
Pharmaceutical and alloy samples	75.40-256.03	_	Long time of equilibration	[50]
Binary, synthetic mixtures, and various real samples	31.75-222.22	—	Less sensitive	[51]
Water	22.22-38.10	19.048		Present study without
	6.35-38.10	3.016		β -correction
Water	9.52-38.10	9.524	Precise, sensitive, and short analytical	Present work with
	6.35-38.10	1.270	time	β -correction

*Spect = spectrophotometry; LDR = linear dynamic range, LOD = limit of detection.

mineral water (MW) and tap water (TW) samples as mentioned before and analyzed using the established β -correction spectrophotometric assay. The results of Cu²⁺ determined in tap (TW) and mineral (MW) water samples are summarized in Table 3. Representative plots for measuring Cu²⁺ in tap water and mineral (MW) water samples are also shown in Figure 7. The results were further validated by determination of Cu²⁺ by the official ICP-OES at the optimal operation parameters. Acceptable percentage recoveries of Cu²⁺ in tap (90.4–113.9%) and mineral (97.5–110.7%) water samples were attained. The "added," "found" and recovery percentage (91–102%) of Cu²⁺ concentrations were found comparable and acceptable. At 95% confidence (n = 5, P < 0.05) [66], the tabulated Student t_{tab} and *F* values were greater than the experimental Student t_{exp} (1.96–2.1) and F_{exp} (2.3–2.8), respectively, revealing acceptable consequence between the tabulated and experimental values for the detection of Cu²⁺ in water.

4. Conclusion, Advantages, Limitations, and Outlooks

In summary, our research demonstrates the potential for the development of significant surfactant simple, effective, cost-effectiveness, interference, and solvent free β -correction spectrophotometric approach for total determination and speciation (labile and chelated) of Cu²⁺. The molar absorptivity of the assay reveals good sensitivity and linear

Samples	Added (µM)	Found (µM)	Recovery (%)	Error (%)
Tap water	0.00	3.71		_
	7.94	7.17	90.4	-9.6
	15.87	18.08	113.9	+13.9
	23.81	22.75	95.6	-4.4
	31.75	30.29	95.4	-4.6
	39.68	40.75	102.7	+2.7
Mineral water	0.00	-0.11		_
	7.94	7.81	98.4	-1.6
	15.87	14.29	90.0	-10.0
	23.81	26.35	110.7	+10.7
	31.75	31.92	100.6	+0.6
	39.68	38.67	97.5	-2.5

TABLE 3: Analytical results of Cu^{2+} determination in tap and mineral water samples by the developed dual-wave β -correction spectrophotometric method*.

*Average of three measurements (n = 3).



FIGURE 7: Plot of the standard addition for Cu^{2+} detection as Cu^{2+} -ZI complex in mineral (MW) and in tap (TW) water samples.

range, LOD, reliability, rapid analysis, no extra costly material necessary, and free from interference of common metal ions. The planned methodology may also substitute the common analytical methods (AAS and ICP-OES) that suffered from time consuming, complicated instrumentation and multiple preconcentration steps. Further, the developed assay can be drawn-out for detection and speciation of Cu²⁺ at ultra-trace low levels in water via online enrichment from water samples by dispersive liquidliquid microextraction [69] and/or on nano-sized sorbent packed column and succeeding elution prior analysis [70]. Thus, the assay can set the trend for coupling sorbent packed column that can assist as a new dimension in β -correction spectroscopy. The accuracy and applicability of the proposed assay were proved by recovery studies for water samples and the results were close to 100%. The absence of the interactive effects of the analytical parameters using one issue at a time represent the main drawback and might decrease the analytical utility of the current study. Accepting the positive impact of SDS in the absorbance will be studied properly in more detail to advance Cu²⁺ detection and to assign the most probable mechanism. The method could also be extended for detection of Cu^{2+} ions in natural waters with high complexing capacity of organic matter, e.g., humic, fulvic acids, phenols, surfactants, etc. Design experiment is also extremely suggested for advance the present approach for attaining effective and perfect Cu^{2+} detection.

Data Availability

Electronic Supplementary data are available upon request.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

Authors' Contributions

All authors made significant contributions for data acquisition, suggestions, revisions, manuscript editing during its preparation, and approved the final version. D. Al-Raimi performed all experiment, data acquisition, and wrote the original draft. M.S. El-Shahawi supervised the study, provided idea, contributed to data acquisition, performed data analysis, and reviewed and edited the article. K. M. Al-Ahmary performed cosupervision and reviewed and edited the article. Other authors provided resources and contributed to funding acquisition. Tharawat N. Abduljabbar and Mohammad S. El-Shahawi equally contributed to this study.

Acknowledgments

This project was funded by the Saudi Basic Industries Corporation (SABIC), Corporation (SABIC), and the Deanship of Scientific Research (DSR) at King Abdulaziz University, Jeddah, under grant no. S-31-247-1440. The authors therefore acknowledge with thanks to SABIC and DSR for technical and financial support.

Supplementary Materials

The following figures and tables will be displayed under the SI download link online as a guide for readers. ESI. 1. Chemical structures of zincon (I) and its copper (II) complex (II). ESI. 2. ICP-OES operational parameters for copper

determination. ESI. 3. Effect of standing time on absorbance of the Cu²⁺-ZI complex at 625 nm using Cu²⁺ ($3.0 \mu \text{gmL}^{-1}$) and 1.0×10^{-4} M ZI at pH 3. ESI.4. The Job method of Cu²⁺zincon chelate; Cu²⁺ and zincon 1×10^{-3} M. ESI.5. Moleratio method of Cu²⁺-zincon complex; [Cu²⁺] = 1×10^{-3} M and [zincon] = 1×10^{-3} M. ESI.6. FTIR spectra of zincon and its Cu²⁺ complex. (*Supplementary Materials*)

References

- J. Krause, M. J. Hopwood, J. Höfer et al., "Trace element (Fe, Co, Ni and Cu) dynamics across the salinity gradient in arctic and antarctic Glacier Fjords," *Frontiers in Earth Science*, vol. 9, Article ID 725279, 2021.
- [2] S. Tang, Q. Liu, J. Hu et al., "A simple colorimetric assay for sensitive Cu^{2+} detection based on the glutathione-mediated etching of MnO₂ nanosheets," *Frontiers in Chemistry*, vol. 9, Article ID 812503, 2021.
- [3] S. Chowdhury, B. Rooj, A. Dutta, and U. Mandal, "Review on recent advances in metal ions sensing using different fluorescent probes," *Journal of Fluorescence*, vol. 28, no. 4, pp. 999–1021, 2018.
- [4] X. Zhou, J. Nie, and B. Du, "Functionalized ionic microgel sensor array for colorimetric detection and discrimination of metal ions," ACS Applied Materials and Interfaces, vol. 9, no. 24, pp. 20913–20921, 2017.
- [5] A. Pal, "Copper toxicity induced hepatocerebral and neurodegenerative diseases: an urgent need for prognostic biomarkers," *Neurotoxicology*, vol. 40, pp. 97–101, 2014.
- [6] G. S. Tomasin, W. R. Silva, B. E. dos Santos Costa, and N. M. M. Coelho, "Highly sensitive determination of Cu(II) ions in hemodialysis water by F AAS after disposable pipette extraction (DPX) using *Moringa oleifera* as solid phase," *Microchemical Journal*, vol. 161, Article ID 105749, 2021.
- [7] N. Aksuner, E. Henden, I. Yilmaz, and A. Cukurovali, "A highly sensitive and selective fluorescent sensor for the determination of copper(II) based on a schiff base," *Dyes and Pigments*, vol. 83, no. 2, pp. 211–217, 2009.
- [8] R. Khani, F. Shemirani, and B. Majidi, "Combination of dispersive liquid-liquid microextraction and flame atomic absorption spectrometry for preconcentration and determination of copper in water samples," *Desalination*, vol. 266, no. 1-3, pp. 238–243, 2011.
- [9] R. M. Liu, D. J. Liu, and A. L. Sun, "Simultaneous determination of multicomponents by flow injection analysis: determination of copper and zinc in serum by using zincon as colouring reagent," *Talanta*, vol. 40, no. 4, pp. 511–514, 1993.
- [10] M. A. Taher, A. Mostafavi, and S. Z. M. Mobarakeh, "Flame atomic absorption spectrometric determination of cobalt (II) after extraction and preconcentration onto modified analcime loaded with zincon," *Asian Journal of Chemistry*, vol. 17, p. 1456, 2005.
- [11] R. M. Rush and J. H. Yoe, "Colorimetric determination of zinc and copper with 2-carboxy-2'-hydroxy-5'-sulfoformazylbenzene," *Analytical Chemistry*, vol. 26, no. 8, pp. 1345– 1347, 1954.
- [12] J. Ghasemi, S. Ahmadi, and K. Torkestani, "Simultaneous determination of copper,nickel, cobalt, and zinc using Zincon as a metallochromic indicator with partial least squares," *Analytica Chimica Acta*, vol. 487, no. 2, pp. 181–188, 2003.
- [13] Z. Li, X. Chang, Z. Hu et al., "Zincon-modified activated carbon for solid-phase extraction and preconcentration of trace lead and chromium from environmental samples,"

Journal of Hazardous Materials, vol. 166, no. 1, pp. 133–137, 2009.

- [14] M. G. De Martino, G. T. Macarovscha, and S. Cadore, "The use of Zincon for preconcentration and determination of Zinc by flame atomic absorption spectrometry," *Analytical Methods*, vol. 2, no. 9, pp. 1258–1262, 2010.
- [15] A. M. El-Menshawy and A. A. El-Asmy, "Zincon polymer as a new modifier for selective separation and determination of copper and zinc from synthetic, water and drug samples," *Indian Journal of Science and Technology*, vol. 2, p. 78, 2009.
- [16] A. Kocyła, A. Pomorski, and A. Krężel, "Molar absorption coefficients and stability constants of Zincon metal complexes for determination of metal ions and bioinorganic applications," *Journal of Inorganic Biochemistry*, vol. 176, pp. 53–65, 2017.
- [17] B. Anilan, T. Gedikbey, and S. Tunali Akar, "Determination of copper in water samples after solid-phase extraction using dimethylglyoxime-modified silica," *Clean: Soil, Air, Water*, vol. 38, no. 4, pp. 344–352, 2010.
- [18] S. Şahan and U. Şahin, "Determination of copper(II) using atomic absorption spectrometry and eriochrome blue black R loaded amberlite XAD-1180 resin," *Clean: Soil, Air, Water*, vol. 38, no. 5-6, pp. 485–491, 2010.
- [19] M. Ghaedi, R. Shabani, M. Montazerozohori et al., "SDScoated Sepabeads SP70-modified by 4-[(E)-3-phenylallylidene) amino] benzenethiol as new efficient solid phase for enrichment and determination of copper, nickel, chromium, and zinc ions in soil, plants, and mint water samples," *Environmental Monitoring and Assessment*, vol. 174, no. 1-4, pp. 171–186, 2011.
- [20] Z. Fang, J. Růžička, and E. H. Hansen, "An efficient flowinjection system with on-line ion-exchange preconcentration for the determination of trace amounts of heavy metals by atomic absorption spectrometry," *Analytica Chimica Acta*, vol. 164, pp. 23–39, 1984.
- [21] V. A. Lemos, M. S. Santos, M. J. S. dos Santos, D. R. Vieira, and C. G. Novaes, "Determination of copper in water samples by atomic absorption spectrometry after cloud point extraction," *Microchimica Acta*, vol. 157, no. 3-4, pp. 215–222, 2007.
- [22] H. Shoaee, M. Roshdi, N. Khanlarzadeh, and A. Beiraghi, "Simultaneous preconcentration of copper and mercury in water samples by cloud point extraction and their determination by inductively coupled plasma atomic emission spectrometry," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 98, pp. 70–75, 2012.
- [23] L. Zhao, S. Zhong, K. Fang, Z. Qian, and J. Chen, "Determination of cadmium(II), cobalt(II), nickel(II), lead(II), zinc(II), and copper(II) in water samples using dual-cloud point extraction and inductively coupled plasma emission spectrometry," *Journal of Hazardous Materials*, vol. 239-240, pp. 206–212, 2012.
- [24] K. Bostanci and M. Merdivan, "Ionic liquid-based aqueous two-phase system coupled with flame atomic absorption spectrometry for the preconcentration and determination of copper in water samples," *Atomic Spectroscopy*, vol. 40, no. 5, pp. 193–198, 2019.
- [25] Y.-D. Xia, Y.-Q. Sun, Y. Cheng, Y. Xia, and X.-B. Yin, "Rational design of dual-ligand Eu-MOF for ratiometric fluorescence sensing Cu²⁺ ions in human serum to diagnose Wilson's disease," *Analytica Chimica Acta*, vol. 1204, Article ID 339731, 2022.
- [26] M. Hassan, Z. Erbas, U. Alshana, and M. Soylak, "Ligandless reversed-phase switchable-hydrophilicity solvent

liquid–liquid microextraction combined with flame-atomic absorption spectrometry for the determination of copper in oil samples," *Microchemical Journal*, vol. 156, Article ID 104868, 2020.

- [27] M. Soylak, Y. Akkaya, and L. Elci, "Flame Atomic Absorption spectrometric determination of Cu(II), Co(II), Cd(II), Fe(III) and Mn(II) in ammonium salts and industrial fertilizers after preconcentration/separation on Diaion HP-20," *International Journal of Environmental Analytical Chemistry*, vol. 82, no. 4, pp. 197–206, 2002.
- [28] J. Mierzwa, Y. C. Sun, Y. T. Chung, and M. H. Yang, "Comparative determination of Ba, Cu, Fe, Pb and Zn in tea leaves by slurry sampling electrothermal atomic absorption and liquid sampling inductively coupled plasma atomic emission spectrometry," *Talanta*, vol. 47, no. 5, pp. 1263–1270, 1998.
- [29] M. Faraji, Y. Yamini, A. Saleh, M. Rezaee, M. Ghambarian, and R. Hassani, "A nanoparticle-based solid-phase extraction procedure followed by flow injection inductively coupled plasma-optical emission spectrometry to determine some heavy metal ions in water samples," *Analytica Chimica Acta*, vol. 659, no. 1-2, pp. 172–177, 2010.
- [30] S. M. Z. Hossain and J. D. Brennan, "β-Galactosidase-based colorimetric paper sensor for determination of heavy metals," *Analytical Chemistry*, vol. 83, no. 22, pp. 8772–8778, 2011.
- [31] H.-Y. Zhou, T.-Q. Chai, L.-J. Peng et al., "Bisubstrate multicolorimetric assay based on the peroxidase-like activity of Cu²⁺-triethylamine complex for copper ion detection," *Dyes and Pigments*, vol. 210, no. 6, Article ID 111028, 2023.
- [32] L. Chen, Y. Li, P. Sun et al., "A facile colorimetric method for ultra-rapid and sensitive detection of copper ions in water," *Journal of Inorganic and Organometallic Polymers and Materials*, vol. 32, no. 7, pp. 2473–2481, 2022.
- [33] S. Tang, Q. Liu, J. Hu et al., "A Simple colorimetric assay for sensitive Cu²⁺ detection based on the glutathione-mediated etching of MnO₂ Nanosheets," *Frontiers in Chemistry*, vol. 9, Article ID 812503, 2021.
- [34] L. Chen, X. Tian, C. Yang et al., "Highly selective and sensitive determination of copper ion based on a visual fluorescence method," *Sensors and Actuators B: Chemical*, vol. 240, pp. 66–75, 2017.
- [35] Q. Miao, J. Qi, Y. Li et al., "Anchoring zinc-doped carbon dots on a paper-based chip for highly sensitive fluorescence detection of copper ions," *Analyst*, vol. 146, no. 20, pp. 6297– 6305, 2021.
- [36] C. Sun, S. Du, T. Zhang, and J. Han, "A novel Calix [4] crownbased 1, 3, 4-oxadiazole as a fluorescent chemosensor for copper (II) ion detection," *Frontiers in Chemistry*, vol. 9, Article ID 766442, 2021.
- [37] M. Mohebbi, R. Heydari, and M. Ramezani, "Determination of Cu, Cd, Ni, Pb and Zn in edible oils using reversed-phase ultrasonic assisted liquid-liquid microextraction and flame atomic absorption spectrometry," *Journal of Analytical Chemistry*, vol. 73, no. 1, pp. 30–35, 2018.
- [38] L. C. Zhang, Y. M. Yang, L. Liang et al., "Lighting up of carbon dots for copper (ii) detection using an aggregation-induced enhanced strategy," *Analyst*, vol. 147, no. 3, pp. 417–422, 2022.
- [39] E. Tesarova, L. Baldrianova, S. B. Hocevar, I. Svancara, K. Vytras, and B. Ogorevc, "Anodic stripping voltammetric measurement of trace heavy metals at antimony film carbon paste electrode," *Electrochimica Acta*, vol. 54, no. 5, pp. 1506–1510, 2009.
- [40] A. L. Squissato, E. M. Richter, and R. A. A. Munoz, "Voltammetric determination of copper and tert-

butylhydroquinone in biodiesel: a rapid quality control protocol," *Talanta*, vol. 201, pp. 433–440, 2019.

- [41] L. Yuan, S. H. Huo, X. N. Ren, and H. Chen, "A novel spectrophotometric determination of copper (II) with bromosulphonazo III," *Chinese Chemical Letters*, vol. 19, no. 1, pp. 92–94, 2008.
- [42] S. Suresha, M. F. Silwadi, and A. A. Syed, "Sensitive and selective spectrophotometric determination of Hg(II), Ni(II), Cu(II) and Co(II) using iminodibenzyl and 3 chloroiminodibenzyl as new reagents and their applications to industrial effluents and soil samples," *International Journal of Environmental Analytical Chemistry*, vol. 82, no. 5, pp. 275– 289, 2002.
- [43] D. Fu and D. Yuan, "Spectrophotometric determination of trace copper in water samples with thiomichlersketone," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 66, no. 2, pp. 434–437, 2007.
- [44] D. Admasu, D. N. Reddy, and K. N. Mekonnen, "Spectrophotometric determination of Cu(II) in soil and vegetable samples collected from Abraha Atsbeha, Tigray, Ethiopia using heterocyclic thiosemicarbazone," *Springer Plus*, vol. 5, no. 1, p. 1169, 2016.
- [45] Lutfullah, S. Sharma, N. Rahman et al., "UV Spectrophotometric determination of Cu(II) in synthetic mixture and water Samples," *Journal of the Chinese Chemical Society*, vol. 57, no. 4A, pp. 622–631, 2010.
- [46] O. Turkoglu and M. Soylak, "Spectrophotometric determination of copper in natural waters and pharmaceutical samples with Chloro(phenyl) glyoxime," *Journal of the Chinese Chemical Society*, vol. 52, no. 3, pp. 575–579, 2005.
- [47] A. V. Sadlapurkar, U. B. Barache, A. B. Shaikh, S. H. Gaikwad, and T. N. Lokhande, "2-chlorobenzaldehyde thiocarbohydrazone: a novel reagent for liquid-liquid extractive spectrophotometric determination of copper(II) from environmental and real samples," *International Journal of Environmental Analytical Chemistry*, vol. 103, no. 16, pp. 3683–3703, 2021.
- [48] U. B. Barache, A. B. Shaikh, T. N. Lokhande, G. S. Kamble, M. A. Anuse, and S. H. Gaikwad, "An efficient, cost effective, sensing behaviour liquid-liquid extraction and spectrophotometric determination of copper(II) incorporated with 4-(4'chlorobenzylideneimino)-3-methyl-5-mercapto-1, 2, 4-triazole: analysis of food samples, leafy vegetables, fertilizers and environmental samples," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 189, pp. 443–453, 2018.
- [49] C. E. Säbel, J. M. Neureuther, and S. Siemann, "A spectrophotometric method for the determination of zinc, copper, and cobalt ions in metalloproteins using Zincon," *Analytical Biochemistry*, vol. 397, no. 2, pp. 218–226, 2010.
- [50] D. S. Al-Raimi, K. M. Al-Ahmary, H. M. Harbi et al., "Varnish polish functionalized paper as a new solid platform for colorimetric determination of copper (II) in water," *International Journal of Environmental Analytical Chemistry*, pp. 1–20, 2021.
- [51] R. A. Nalawade, A. M. Nalawade, G. S. Kamble, and M. A. Anuse, "Rapid, synergistic extractive spectrophotometric determination of copper (II) by using sensitive chromogenic reagent N",N"'-bis [(E)-(4-fluorophenyl) methylidene]thiocarbonohydrazide]," Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, vol. 146, pp. 297–306, 2015.

- [52] A. Abbaspour and L. Baramakeh, "Dual-wavelength β -correction spectrophotometry for selective determination of Zr," *Talanta*, vol. 57, no. 4, pp. 807–812, 2002.
- [53] A. Abbaspour and L. Baramakeh, "Simultaneous determination of antimony and bismuth by beta-correction spectrophotometry and an artificial neural network algorithm," *Talanta*, vol. 65, no. 3, pp. 692–699, 2005.
- [54] A. Hamza, A. S. Bashammakh, A. A. Al-Sibaai, H. M. Al-Saidi, and M. S. El-Shahawi, "Part 1. Spectrophotometric determination of trace mercury (II) in dental-unit wastewater and fertilizer samples using the novel reagent 6-hydroxy-3-(2oxoindolin-3-ylidenea-mino)-2-thioxo-2H-1,3-thiazin-4(3H)-one and the dual-wavelength β-correction spectrophotometry," *Journal of Hazardous Materials*, vol. 178, no. 1-3, pp. 287–292, 2010.
- [55] A. Hamza, A. S. Bashammakh, A. A. Al-Sibaai, H. M. Al-Saidi, and M. S. El-Shahawi, "Dual-wavelength β-correction spectrophotometric determination of trace concentrations of cyanide ions based on the nucleophilic addition of cyanide to imine group of the new reagent 4-hydroxy-3-(2-oxoindolin-3-ylideneamino)-2-thioxo-2H-1,3-thiazin-6(3H)-one," *Analytica Chimica Acta*, vol. 657, no. 1, pp. 69–74, 2010.
- [56] A. Abbaspour and R. Mirzajani, "Application of spectral β-correction method and partial least squares for simultaneous determination of V(IV) and V(V) in surfactant media," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 64, no. 3, pp. 646–652, 2006.
- [57] G. I. Mohammed, W. Ahmad, H. Alwael, A. S. Bashammakh, Z. M. Saigl, and M. S. El-Shahawi, "Application of β-correction spectrophotometry for determination and speciation of bismuth (III) and (V) species in various water samples, soil, hair and drug formulations," *Journal of Molecular Liquids*, vol. 236, pp. 241–248, 2017.
- [58] A. H. Al-Bagawi, W. Ahmad, Z. M. Saigl, H. Alwael, E. A. Al-Harbi, and M. S. El-Shahawi, "A simple and low cost dualwavelength β-correction spectrophotometric determination and speciation of mercury(II) in water using chromogenic reagent 4-(2-thiazolylazo) resorcinol," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 187, pp. 174–180, 2017.
- [59] V. Kumar, S. Pandita, G. P. Singh Sidhu et al., "Copper bioavailability, uptake, toxicity and tolerance in plants: a comprehensive review," *Chemosphere*, vol. 262, Article ID 127810, 2021.
- [60] A. Bourgeault, P. Ciffroy, C. Garnier et al., "Speciation and bioavailability of dissolved copper in different freshwaters: comparison of modelling, biological and chemical responses in aquatic mosses and gammarids," *Science of the Total Environment*, vol. 452-453, pp. 68–77, 2013.
- [61] H. T. S. Britton, *Potentiometric Hydrogen Ion*", Chapman and Hall, London, UK, 4th edition, 1952.
- [62] D. T. Sawyer, W. R. Heineman, and J. M. Beebe, *Chemistry Experiments for Instrumental Methods*, John Wiley and Sons, New York, NY, USA, 1st edition, 1984.
- [63] A. B. P. Lever, Inorganic Electronic Spectroscopy, Theory and Applications, Elsevier Science, Amsterdam, Netherlands, 2nd edition, 1984.
- [64] J. M. Smith, H. C. Van Ness, M. M. Abbott, and M. T. Swihart, *Introduction to Chemical Engineering Thermodynamics*, McGraw-Hill Education, New York, NY, USA, 8th edition, 2018.
- [65] R. J. Silbey, R. A. Alberty, and M. G. Bawendi, *Physical Chemistry*, Wiley, Hoboken, NJ, USA, 4th edition, 2004.

- [66] K. Nakamoto, Infrared and Raman Spectra of Inorganic and Coordination Compounds, Part B: Applications in Coordination, Organometallic, and Bioinorganic Chemistry, Wiley, Hoboken, NJ, USA, 6th edition, 2009.
- [67] Z. Marczenko, M. Balcerzak, and E. Kloczko, Separation, Preconcentration and Spectrophotometry in Inorganic Analysis (Analytical Spectroscopy Library, Volume 10), Elsevier Science, Amsterdam, Netherlands, 1st edition, 2000.
- [68] J. C. Miller and J. N. Miller, *Statistics for Analytical Chemistry*", Ellis Horwood Ltd, New York, NY, USA, 6th edition, 2010.
- [69] W. Ahmad, A. A. Al-Sibaai, A. S. Bashammakh, H. Alwael, and M. S. El-Shahawi, "Recent advances in dispersive liquidliquid microextraction for pesticide analysis," *TrAC, Trends in Analytical Chemistry*, vol. 72, pp. 181–192, 2015.
- [70] O. Oribayo, X. Feng, G. L. Rempel, and Q. Pan, "Synthesis of lignin-based polyurethane/graphene oxide foam and its application as an absorbent for oil spill clean-ups and recovery," *Chemical Engineering Journal*, vol. 323, pp. 191–202, 2017.