

Research Article

Exploring the Functional Disorder and Corresponding Key Transcription Factors in Intraductal Papillary Mucinous Neoplasms Progression

Guiying Bai,^{1,2} Chenxuan Wu,¹ Yingtang Gao,² and Guiming Shu³

¹Department of Oncology, Tianjin Third Central Hospital, Tianjin 300179, China

²Key Laboratory of Artificial Cell Institute of Hepatobiliary Disease, Tianjin Third Central Hospital, Tianjin 300179, China

³Department of Hepatobiliary Surgery, Tianjin Third Central Hospital, Tianjin 300179, China

Correspondence should be addressed to Guiying Bai; guiyingbai@126.com

Received 18 June 2015; Accepted 11 August 2015

Academic Editor: Giulia Piaggio

Copyright © 2015 Guiying Bai et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

This study has analyzed the gene expression patterns of an IPMN microarray dataset including normal pancreatic ductal tissue (NT), intraductal papillary mucinous adenoma (IPMA), intraductal papillary mucinous carcinoma (IPMC), and invasive ductal carcinoma (IDC) samples. And eight clusters of differentially expressed genes (DEGs) with similar expression pattern were detected by *k*-means clustering. Then a survey map of functional disorder in IPMN progression was established by functional enrichment analysis of these clusters. In addition, transcription factors (TFs) enrichment analysis was used to detect the key TFs in each cluster of DEGs, and three TFs (FLI1, ERG, and ESRI) were found to significantly regulate DEGs in cluster 1, and expression of these three TFs was validated by qRT-PCR. All these results indicated that these three TFs might play key roles in the early stages of IPMN progression.

1. Introduction

Intraductal papillary mucinous neoplasms (IPMN) are pancreatic precancerous lesions composed of dilated main and branch ducts and mixed duct type (main and branch ducts are both involved) lined by mucin producing atypical epithelium, which usually proliferates in a papillary fashion [1]. Based on their increasing architectural and nuclear atypia, IPMN are divided into four stages: IPMN low-grade dysplasia (alternatively intraductal papillary mucinous adenoma, IPMA), IPMN intermediate grade dysplasia (alternatively borderline IPMN, IPMB), and IPMN high-grade dysplasia (alternatively intraductal papillary mucinous carcinoma, IPMC) and about 20% to 40% IPMC were invasive ductal carcinoma (IDC) [1, 2]. Moreover, on the basis of the histological and immunohistochemical characteristics, IPMN could be divided into intestinal, pancreatobiliary, oncocytic, and gastric subtypes [3, 4].

The genetic mechanism of IPMN progression has not been completely defined. IPMN are known to harbor mutations of KRAS [5, 6], p16 [6], p53 [5], GNAS [7], RNF43

[8], and DPC4/SMAD4/MADH4 [9], but the prevalence of the mutations is lower than in pancreatic ductal adenocarcinoma (PDAC). With development of high-throughput molecular techniques such as gene microarrays, lots of differentially expressed genes (DEGs) have been found with IPMN progression, such as BRCA1 [10], SOX9 [11], and MUC6 [12]. Meanwhile, some cellular signaling pathways were found to be involved in the IPMN carcinogenesis, such as phosphoinositide 3-kinases-protein kinase B/Akt pathway [13], the DNA damage checkpoint pathway [14], inactivation of tumor suppressor pathways, p53 pathway [15], activation of oncogenic pathways, and K-ras pathway [16]. However, the comprehensive integrated analysis of genes and biological processes and pathways leading to development and progression to malignancy of IPMN were still not clearly demonstrated.

In this study, for comprehensive understanding, the mechanism of IPMN, a microarray dataset including normal main pancreatic ductal tissue (NT), IPMA, IPMC, and IDC originating in IPMN were collected from National Center

for Biotechnology Information (NCBI) Gene Expression Omnibus (GEO). Time-series expression analysis was used to screen the multiclass differentially expressed genes (DEGs) during four stages of carcinogenesis. And the DEGs were clustered into different expression pattern clusters using k -means clustering. Functional enrichment analysis was used to explore the biological processes and pathways related to each cluster. Then, transcription factors (TFs) enrichment analysis was used to detect the key TFs in each cluster of DEGs. At last, qRT-PCR assay was used to validate the expression of TFs. Finally, an overall survey map of functional disorder and corresponding TFs for the progress of the IPMN were established.

2. Materials and Methods

2.1. Datasets Used. We selected the gene expression profile data GSE19650 [17] from NCBI GEO, executed on Affymetrix Human Genome U133 Plus 2.0 Array (platform number GPL570); the dataset contains 22 samples including 7 normal main pancreatic ducts (NT), 6 IPMA, 6 IPMC, and 3 IDC originating in IPMN.

Firstly, the raw data were normalized by RMA (robust multiarray average) background subtraction and quantile was normalized using Affymetrix Expression Console Software, and noise derived from absent genes, background, and nonspecific hybridizations was removed. A well-known cancer gene dataset including 547 genes was collected from Cancer Gene Census database [18]. And the significance of overlap between known cancer genes and DEGs was calculated using the cumulative hypergeometric function ($P < 0.05$) (see details in Section 2.4), and calculations were implemented in R statistical package using function `phyper` (<http://www.r-project.org/>).

2.2. DEGs Screening and Clustering. DEGs among the four stages of IPMN progression were identified by significance analysis of microarrays (SAM) algorithm (multiclass SAM, at least one [fold change] ≥ 2 , Q values < 0.05). Then these DEGs were clustered by k -means clustering. Cluster 3.0 software was used to cluster the DEGs clusters and draw cluster picture.

2.3. Gene Ontology and Pathway Enrichment Analysis. The functional significance of the DEGs clusters was investigated by performing functional enrichment and clustering analysis using The Database for Annotation, Visualization and Integrated Discovery (DAVID) [19]. One of the DAVID's features is functional annotation cluster in those places similar to GO categories, based on the parent/child GO term associations and the number of the shared genes, into a functional cluster. The GO cluster enrichment score is based on geometric mean of member's P values and used to rank their biological significance. So DAVID would report significance of enriched clusters. We used this feature to estimate the relationship between the GO terms (biological process of the GO database) and the KEGG pathways.

TABLE 1: The PCR primers of gene.

Gene	Sequence (5' → 3')	Length
β -actin	Forward primer	CATGTACGTTGCTATCCAGGC
	Reverse primer	CTCCTTAATGTACACGCACGAT
ESR1	Forward primer	GGGAAGTATGGCTATGGAATCTG
	Reverse primer	TGGCTGGACACATATAGTCGTT
ERG	Forward primer	CGTGCCAGCAGATCCTACG
	Reverse primer	GGTGAGCCTCTGGAAGTCG
FLI1	Forward primer	CCAACGAGAGGAGAGTCATCG
	Reverse primer	TTCCGTGTTGTAGAGGGTGGT

The threshold of enrichment score for enriched clusters was set as 1 artificially.

2.4. TFs Enrichment Analysis. In this study, we downloaded TFs and their regulation genes from TRANSFAC database (Release 3.4) [20]. All experimentally verified TFs and their target gene sets were used in this study, containing 106 TFs, 2921 target genes, and 6238 TF-target gene interactions. Among the 106 TFs, 22 TFs can be mapped to DEGs of IPMN progression. TFs enrichment analysis is a common statistical technique to reveal the significantly changed modules of the target gene sets against the background so as to elucidate the underlying TF regulation mechanisms. Then we enriched DEGs in each cluster with target genes of TFs. The score of each TF and target gene set was calculated using the cumulative hypergeometric function as follows:

$$P = 1 - \sum_{i=0}^{x-1} \frac{\binom{K}{i} \binom{M-K}{N-i}}{\binom{M}{N}}, \quad (1)$$

where M represents the total number of genes tested. N represents the number of genes regulated by a TF. K is the number of DEGs in certain cluster. x is the number of the DEGs in certain cluster that also appeared in certain TF target genes. The resulting P value ($P < 0.05$) reflects the probability of extracting up to x of possible K genes in N drawings. All of the above calculations were implemented in R statistical package using function `phyper` (<http://www.r-project.org/>).

2.5. qRT-PCR. The tissue samples of four stages of IPMN were collected from Tianjin Third Central Hospital, including five NT, one IPMA, five IPMC, and five IDC originating in IPMN. All IPMA and IPMC samples were classified into gastric type. This study was approved by the Experimental and Ethics Committees of Tianjin Third Central Hospital, and written informed consent from each participating subject had been obtained.

Three TFs (ESR1, ERG, and FLI1) enriched in cluster 1 were validated by quantitative (q) real-time (RT) polymerase chain reaction (PCR) assays using these samples. Primers for the qRT-PCR and annealing temperatures are shown in Table 1. The qRT-PCR reactions were performed on a Roche LightCycler Instrument 1.5, using a LightCycler Fast

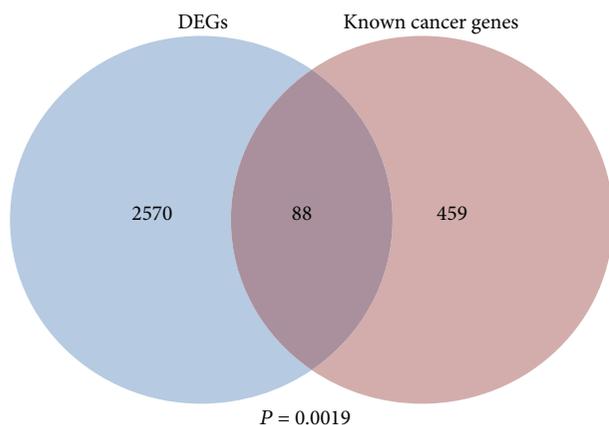


FIGURE 1: Overlap of DEGs and known cancer genes. The *P* value was calculated by hypergeometric test.

Start DNA Master PLUS SYBR Green I kit (Roche Cat. 03515885001, Castle Hill, Australia). Briefly, 15 μ L reactions: 7.5 μ L Master Mix, 0.1 μ L forward primer and reverse primer, 1 μ L cDNA sample, and 6.3 μ L ddH₂O were prepared. Each sample was run in triplicate. The RT-PCR program was set to 95°C for 5 minutes and then 45 cycles of 95°C for 10 seconds, 55–60°C for 35 seconds, and 72°C for 40 seconds. At the end of each program, a melting curve analysis was performed. Also, the data were automatically analyzed by the system, and an amplification plot was generated for each cDNA sample at the end of each RT-PCR run. The internal control gene was β -actin.

3. Results

3.1. Identification of DEGs among Four Stages of IPMN Progression. To investigate the gene expression patterns in IPMN progression, multiclass SAM algorithm was used to screen the DEGs among four stages of carcinogenesis, and 2658 DEGs were identified. Then a well-known cancer gene dataset called Cancer Gene Census was used to evaluate the relationship between DEGs and tumorigenesis. Cancer Gene Census is an ongoing effort to catalogue those genes for which mutations have been causally implicated in cancer [18], and many studies used it as a well-known cancer gene dataset [21, 22]. Particularly, there were 88 DEGs (hypergeometric $P = 0.0019$) that overlap with the known cancer genes (see Figure 1), including PDAC related genes such as MYC, BRCA1, and KRAS [23–25], which indicates that the DEGs of this study have close bond with the cancer progression indeed.

3.2. Clustering DEGs by Expression Pattern Using *k*-Means Clustering. To thoroughly investigate the genomic expression pattern among four stages of IPMN progression, *k*-means clustering was used to cluster the DEGs by expression pattern similarity. As in the results shown in Figure 2, all DEGs were clustering into 8 clusters (named as C1–C8). C1 contained 992 DEGs, which was downregulated in IPMA stage and then kept low expression level in IPMC and IDC

stages; C2 contained 397 DEGs, which was upregulated in IPMA stage and then slightly downregulated during IPMC and IDC stages; C3 contained 476 DEGs, which was upregulated in IPMA stage, slightly downregulated in IPMC stage, and then slightly upregulated in IDC stage; C4 contained 244 DEGs, which kept low expression level in IPMA stage and was upregulated in IPMC stage and then slightly upregulated in IDC stage; C5 contained 97 DEGs, which was downregulated in IPMA stage, slightly upregulated in IPMC stage, and then upregulated in IDC stage; C6 contained 83 DEGs, which was downregulated in IPMA and IPMC stages and then kept low expression level in IDC stage; C7 contained 205 DEGs, which was continuously upregulated during the three stages of IPMN; C8 contained 164 DEGs, which was upregulated in IPMA stage and kept high expression level in IPMC stage and was then downregulated in IDC stage. For all these clusters of DEGs, different clusters were regulated in different cancer stages, which indicates that each cluster may play roles in one or several specific cancer stages.

3.3. Functional Enrichment Analysis of 8 DEGs Clusters.

Analysis of DEGs clusters could help pinpoint potential functional mechanism important for gene expression during IPMN progression [26]. Therefore, DAVID was used to evaluate the biological processes and pathways related to these 8 clusters of DEGs (enrichment score > 1). As shown in Figure 3 and Supplementary Tables in Supplementary Material available online at <http://dx.doi.org/10.1155/2015/197603>, DEGs in C1 that contains almost half of all DEGs were enriched in many biological processes, mainly involved in 5 aspects: “vasculature development,” “cell migration and locomotion,” “neuron development differentiation,” “phosphorylation,” and “endocytosis”; DEGs in C2 were enriched in “phosphorylation,” “pigmentation during development,” “cation homeostasis,” “protein localization and transport,” and “glycosylation”; DEGs in C3 were enriched in “acid biosynthetic process,” “immune cell differentiation,” “cytoskeleton organization,” “glycosylation,” “small molecule catabolic process,” and “nuclear transport”; DEGs in C4 were enriched in “cell cycle,” “mitosis,” “DNA and chromosome organization,” “protein ubiquitination,” and “phosphorylation”; DEGs in C5 were enriched in “cell growth, development, and differentiation,” “skeletal system development,” “cell communication,” “blood circulation,” and “response to nutrient, metal ion”; DEGs in C6 were enriched in “regulation of transcription”; DEGs in C7 were enriched in “phosphorylation,” “proteolysis,” “cell proliferation,” and “regulation of binding”; DEGs in C8 were enriched in “RNA splicing,” “protein localization and transport,” and “proteolysis.” Among these various biological processes, most of them were involved in multiple cancer stages such as “phosphorylation,” “glycosylation,” and “proteolysis”; for comparison, some biological processes played a role in specific cancer stage; for example, “cell cycle” and related functions play a main role in C4, in other words, between IPMA and IPMC stage.

3.4. TFs Enrichment Analysis of 8 Clusters of DEGs. The cluster analysis of DEGs identified groups of genes that

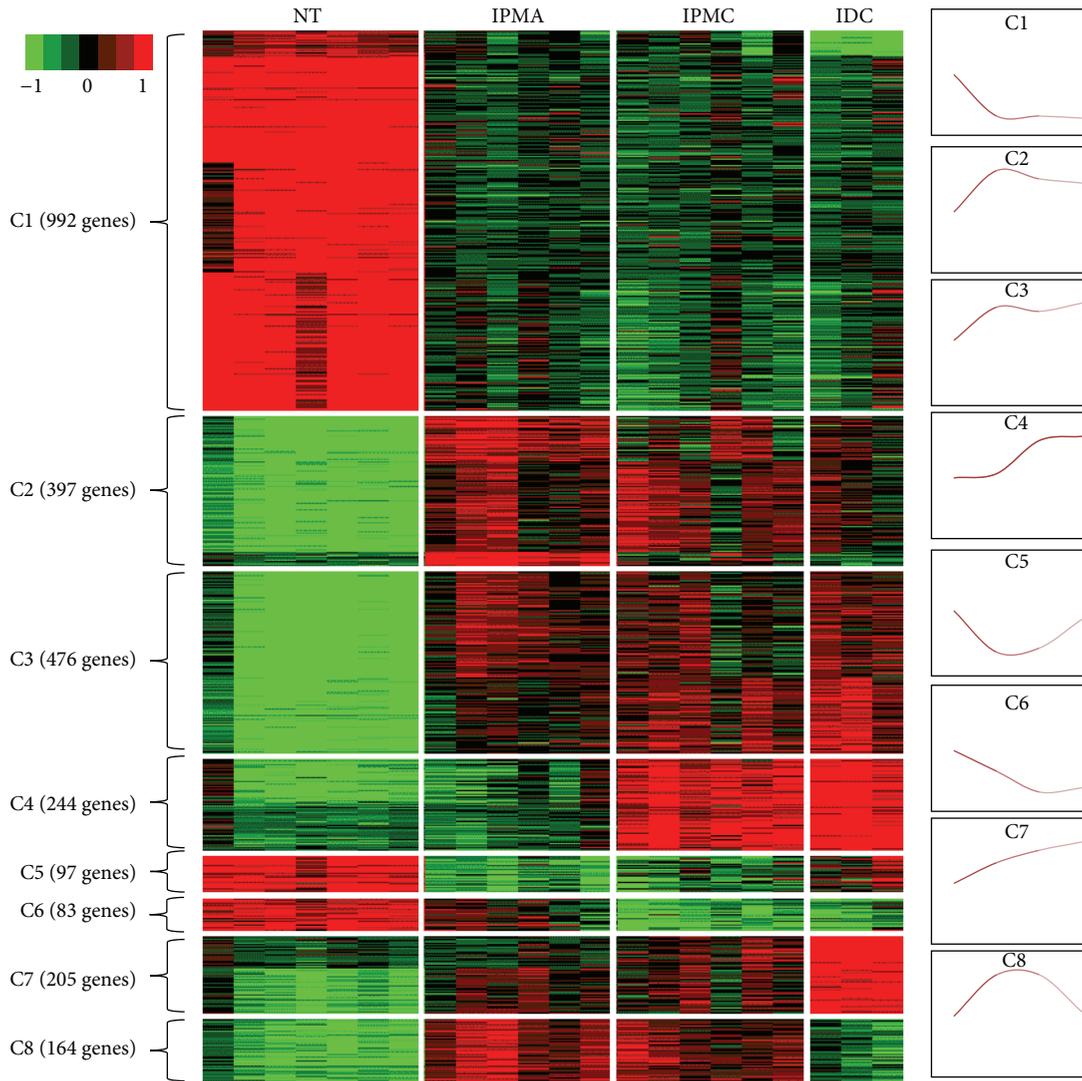


FIGURE 2: The clustering map of DEGs clusters and expression patterns.

exhibit similar expression profiles and thus might be coregulated. Analysis of such coregulated gene sets could help pinpoint potential TFs important for gene expression during cancer progression [27]. Therefore, to gain an initial understanding of the regulatory mechanisms of genes differentially expressed during cancer progression, a total of experimentally validated TFs network data containing 106 TFs, 2921 target genes, and 6238 TF-target gene interactions were gathered from TRANSFAC database [20]. Genes in each cluster were screened for overrepresentation of known TFs (cumulative hypergeometric; see Section 2 for details). 14 TFs which have target genes in these clusters were detected, and TFs enrichment analysis showed that three TFs (ESR1, ERG, and FLI1) were selectively enriched in cluster 1 (as shown in Table 2 and Figure 4), suggesting that these three TFs and their corresponding target genes may play important roles in early stages of IPMN progression, but these three TFs in

IPMN progression have not yet received much attention and research. In addition, the rest of 11 TFs such as SAMD4, MYC, PPARG, and PPARGA have also been proved to play key roles in various cancer progressions [28–30].

3.5. Expression of Three TFs. The qRT-PCR experiments were used to verify the expression of these three TFs in different stages of IPMN. As in the results showed in Figure 5, the mRNA levels of ESR1, ERG, and FLI1 were remarkably decreased between NT and IPMA (t -test, $P < 0.05$) and then continuously and slightly decreased during IPMN progression, which confirmed the results of the microarray analysis and TFs enrichment analysis. These results suggested that these three TFs may play important roles at the beginning of IPMN and still have some effect in the process of IPMN progress to IDC.

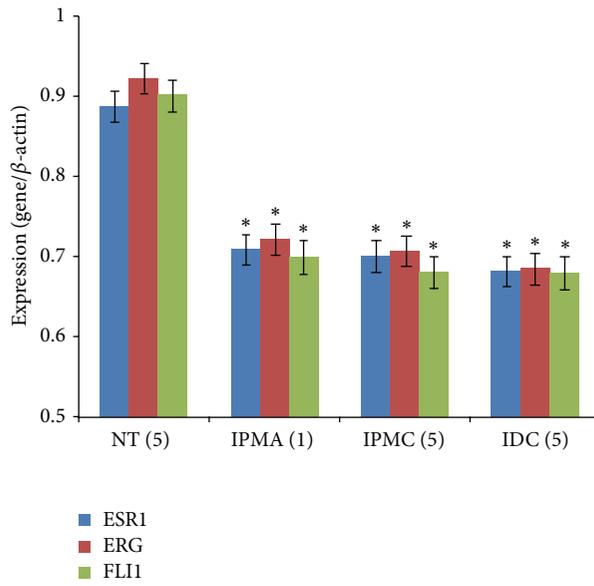


FIGURE 5: Expression of three TFs in different stages of IPMN samples. * $P < 0.05$ as determined by a Student's t -test compared with the NT samples.

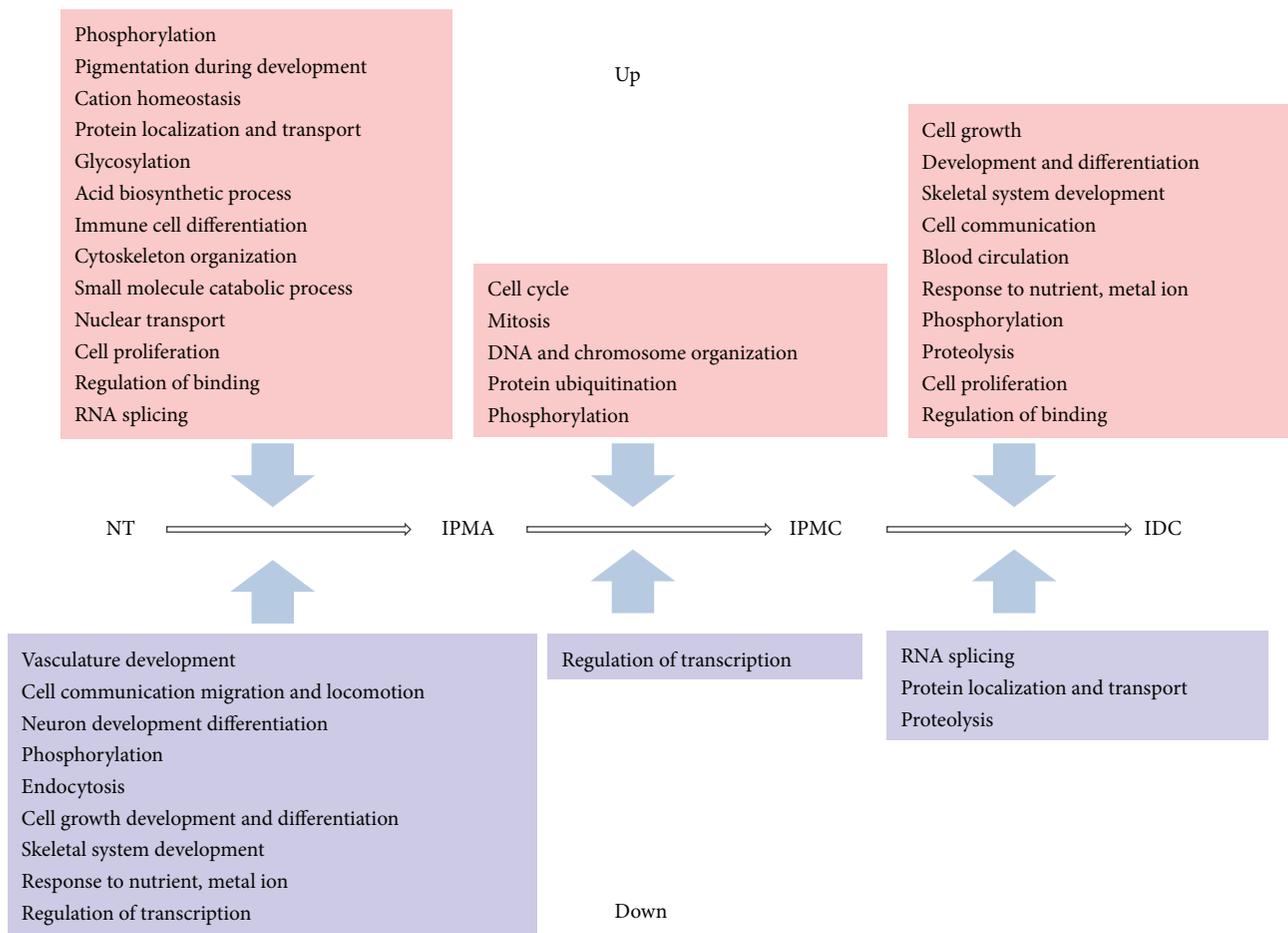


FIGURE 6: The overall survey map of functional disorder for IPMN progression.

target genes dataset was gathered from TRANSFAC database for enrichment analysis. Three TFs (ESR1, ERG, and FLI1) were detected that not only significantly regulate the DEGs in cluster 1, but also have regulatory relationships between each other. What is more, the results of qRT-PCR experiments validated that these three TFs were remarkably decreased between NT and IPMA and then continuously and slightly decreased during IPMN progression. ERG and FLI1 genes are closely related members of the erythroblast transformation-specific (ETS) family of transcription factors, and studies have proved that these two TFs were usually involved in various types of cancer jointly [38, 39]. Meanwhile, the ESR1, estrogen receptor encoding gene, has been proved to be involved in pathological processes including breast cancer and endometrial cancer [40]. But until now no studies have focused on these 3 TFs in IPMN. Moreover, most of the regulated genes (18/25) of these three TFs could be found in Pancreatic Cancer Database [41]. Nevertheless, our study indicates that these three TFs may be the key regulators in early stage of IPMN and are definitely worth further exploration. In addition, the rest of TFs such as PPAR γ , PPAR α , SMAD3, MYC, and STAT1 have been proved to be related to cancer progression [28–30, 42].

5. Conclusion

Our study provides a comprehensive perspective for the progression of pancreatic carcinogenesis by distinct bioinformatics analysis. A number of DEGs clusters and specific biological processes were detected in progression, and then an overall survey map of functional disturbance for the progression has been established. Then, the TFs enrichment analysis and qRT-PCR experiments found that three TFs may be playing key roles in early stages of IPMN progression. We believe that these results have a certain value in development of IPMN mechanism study and based medicine research.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgment

This research was supported by National Natural Sciences Foundation of China (no. 81273420).

References

- [1] M. Distler, D. Aust, J. Weitz, C. Pilarsky, and R. Grützmann, "Precursor lesions for sporadic pancreatic cancer: PanIN, IPMN, and MCN," *BioMed Research International*, vol. 2014, Article ID 474905, 11 pages, 2014.
- [2] F. T. Bosman, F. Carneiro, R. H. Hruban, and N. Thise, *Classification of Tumours of the Digestive System*, WHO, Lyon, France, 2010.
- [3] M. Distler, S. Kersting, M. Niedergethmann et al., "Pathohistological subtype predicts survival in patients with intraductal papillary mucinous neoplasm (IPMN) of the pancreas," *Annals of Surgery*, vol. 258, no. 2, pp. 324–330, 2013.
- [4] T. Furukawa, G. Klöppel, N. V. Adsay et al., "Classification of types of intraductal papillary-mucinous neoplasm of the pancreas: a consensus study," *Virchows Archiv*, vol. 447, no. 5, pp. 794–799, 2005.
- [5] K. Zgraggen, J. A. Rivera, C. C. Compton et al., "Prevalence of activating K-ras mutations in the evolutionary stages of neoplasia in intraductal papillary mucinous tumors of the pancreas," *Annals of Surgery*, vol. 226, no. 4, pp. 491–500, 1997.
- [6] F. Sessa, E. Solcia, C. Capella et al., "Intraductal papillary-mucinous tumours represent a distinct group of pancreatic neoplasms: an investigation of tumour cell differentiation and K-ras, p53 and c-erbB-2 abnormalities in 26 patients," *Virchows Archiv*, vol. 425, no. 4, pp. 357–367, 1994.
- [7] J. Wu, H. Matthaei, A. Maitra et al., "Recurrent GNAS mutations define an unexpected pathway for pancreatic cyst development," *Science Translational Medicine*, vol. 3, no. 92, Article ID 92ra66, 2001.
- [8] J. Wu, Y. Jiao, M. Dal Molin et al., "Whole-exome sequencing of neoplastic cysts of the pancreas reveals recurrent mutations in components of ubiquitin-dependent pathways," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 52, pp. 21188–21193, 2011.
- [9] P. S. Moore, S. Orlandini, G. Zamboni et al., "Pancreatic tumours: molecular pathways implicated in ductal cancer are involved in ampullary but not in exocrine nonductal or endocrine tumorigenesis," *British Journal of Cancer*, vol. 84, no. 2, pp. 253–262, 2001.
- [10] P. Jiang, W. Zang, L. Wang, Y. Xu, Y. Liu, and S.-X. Deng, "Protein-protein interaction and SNP analysis in intraductal papillary mucinous neoplasm," *Gene*, vol. 513, no. 1, pp. 219–224, 2013.
- [11] T. Tanaka, T. Kuroki, T. Adachi et al., "Evaluation of SOX9 expression in pancreatic ductal adenocarcinoma and intraductal papillary mucinous neoplasm," *Pancreas*, vol. 42, no. 3, pp. 488–493, 2013.
- [12] O. Basturk, S. Khayyata, D. S. Klimstra et al., "Preferential expression of MUC6 in oncocytic and pancreatobiliary types of intraductal papillary neoplasms highlights a pyloropancreatic pathway, distinct from the intestinal pathway, in pancreatic carcinogenesis," *American Journal of Surgical Pathology*, vol. 34, no. 3, pp. 364–370, 2010.
- [13] S. Semba, T. Moriya, W. Kimura, and M. Yamakawa, "Phosphorylated Akt/PKB controls cell growth and apoptosis in intraductal papillary-mucinous tumor and invasive ductal adenocarcinoma of the pancreas," *Pancreas*, vol. 26, no. 3, pp. 250–257, 2003.
- [14] Y. Miyasaka, E. Nagai, H. Yamaguchi et al., "The role of the DNA damage checkpoint pathway in intraductal papillary mucinous neoplasms of the pancreas," *Clinical Cancer Research*, vol. 13, no. 15, pp. 4371–4377, 2007.
- [15] T. Furukawa, R. Fujisaki, Y. Yoshida et al., "Distinct progression pathways involving the dysfunction of DUSP6/MKP-3 in pancreatic intraepithelial neoplasia and intraductal papillary-mucinous neoplasms of the pancreas," *Modern Pathology*, vol. 18, no. 8, pp. 1034–1042, 2005.
- [16] M. Kitago, M. Ueda, K. Aiura et al., "Comparison of K-ras point mutation distributions in intraductal papillary-mucinous tumors and ductal adenocarcinoma of the pancreas," *International Journal of Cancer*, vol. 110, no. 2, pp. 177–182, 2004.

- [17] N. Hiraoka, R. Yamazakiitoh, Y. Ino et al., "CXCL17 and ICAM2 are associated with a potential anti-tumor immune response in early intraepithelial stages of human pancreatic carcinogenesis," *Gastroenterology*, vol. 140, no. 1, pp. 310–321, 2011.
- [18] P. A. Futreal, L. Coin, M. Marshall et al., "A census of human cancer genes," *Nature Reviews Cancer*, vol. 4, no. 3, pp. 177–183, 2004.
- [19] D. W. Huang, B. T. Sherman, and R. A. Lempicki, "Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources," *Nature Protocols*, vol. 4, no. 1, pp. 44–57, 2009.
- [20] S. Gama-Castro, V. Jiménez-Jacinto, M. Peralta-Gil et al., "RegulonDB (version 6.0): gene regulation model of *Escherichia coli* K-12 beyond transcription, active (experimental) annotated promoters and Textpresso navigation," *Nucleic Acids Research*, vol. 36, no. 1, pp. D120–D124, 2008.
- [21] X. Gong, R. Wu, Y. Zhang et al., "Extracting consistent knowledge from highly inconsistent cancer gene data sources," *BMC Bioinformatics*, vol. 11, article 76, 2010.
- [22] W. Ma, D. Yang, Y. Gu, X. Guo, W. Zhao, and Z. Guo, "Finding disease-specific coordinated functions by multi-function genes: insight into the coordination mechanisms in diseases," *Genomics*, vol. 94, no. 2, pp. 94–100, 2009.
- [23] M. P. Di Magliano and C. D. Logsdon, "Roles for KRAS in pancreatic tumor development and progression," *Gastroenterology*, vol. 144, no. 6, pp. 1220–1229, 2013.
- [24] C. Huang, W.-M. Wang, J.-P. Gong, and K. Yang, "Oncogenesis and the clinical significance of K-ras in pancreatic adenocarcinoma," *Asian Pacific Journal of Cancer Prevention*, vol. 14, no. 5, pp. 2699–2701, 2013.
- [25] J. B. Greer and D. C. Whitcomb, "Role of BRCA1 and BRCA2 mutations in pancreatic cancer," *Gut*, vol. 56, no. 5, pp. 601–605, 2007.
- [26] J. K. Choi, U. Yu, O. J. Yoo, and S. Kim, "Differential coexpression analysis using microarray data and its application to human cancer," *Bioinformatics*, vol. 21, no. 24, pp. 4348–4355, 2005.
- [27] E. Breeze, E. Harrison, S. McHattie et al., "High-resolution temporal profiling of transcripts during Arabidopsis leaf senescence reveals a distinct chronology of processes and regulation," *Plant Cell*, vol. 23, no. 3, pp. 873–894, 2011.
- [28] Z. Chen, W. Holland, J. M. Shelton et al., "Mutation of mouse Samd4 causes leanness, myopathy, uncoupled mitochondrial respiration, and dysregulated mTORC1 signaling," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 111, no. 20, pp. 7367–7372, 2014.
- [29] C. V. Dang, "MYC, metabolism, cell growth, and tumorigenesis," *Cold Spring harbor perspectives in medicine*, vol. 3, no. 8, 2013.
- [30] K. Tachibana, D. Yamasaki, K. Ishimoto, and T. Doi, "The role of PPARs in cancer," *PPAR Research*, vol. 2008, Article ID 102737, 15 pages, 2008.
- [31] N. V. Adsay, K. Merati, A. Andea et al., "The dichotomy in the preinvasive neoplasia to invasive carcinoma sequence in the pancreas: differential expression of MUC1 and MUC2 supports the existence of two separate pathways of carcinogenesis," *Modern Pathology*, vol. 15, no. 10, pp. 1087–1095, 2002.
- [32] W.-Q. Chen, D. Liang, S.-W. Zhang, R.-S. Zheng, and Y.-T. He, "Pancreatic cancer incidence and mortality patterns in China, 2009," *Asian Pacific Journal of Cancer Prevention*, vol. 14, no. 12, pp. 7321–7324, 2013.
- [33] S. T. Chari, D. Yadav, T. C. Smyrk et al., "Study of recurrence after surgical resection of intraductal papillary mucinous neoplasm of the pancreas," *Gastroenterology*, vol. 123, no. 5, pp. 1500–1507, 2002.
- [34] T. Nakagohri, M. Konishi, K. Inoue, Y. Tanizawa, and T. Kinoshita, "Invasive carcinoma derived from intraductal papillary mucinous carcinoma of the pancreas," *Hepato-Gastroenterology*, vol. 51, no. 59, pp. 1480–1483, 2004.
- [35] G. Cheng, J. Zielonka, D. McAllister, S. Tsai, M. B. Dwinell, and B. Kalyanaraman, "Profiling and targeting of cellular bioenergetics: inhibition of pancreatic cancer cell proliferation," *British Journal of Cancer*, vol. 111, no. 1, pp. 85–93, 2014.
- [36] K. Ohmine, K. Kawaguchi, S. Ohtsuki et al., "Attenuation of phosphorylation by deoxycytidine kinase is key to acquired gemcitabine resistance in a pancreatic cancer cell line: targeted proteomic and metabolomic analyses in PK9 cells," *Pharmaceutical Research*, vol. 29, no. 7, pp. 2006–2016, 2012.
- [37] N. Bardeesy and R. A. DePinho, "Pancreatic cancer biology and genetics," *Nature Reviews Cancer*, vol. 2, no. 12, pp. 897–909, 2002.
- [38] E. A. Kruse, S. J. Loughran, T. M. Baldwin et al., "Dual requirement for the ETS transcription factors Fli-1 and Erg in hematopoietic stem cells and the megakaryocyte lineage," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 33, pp. 13814–13819, 2009.
- [39] D. L. Stockman, J. L. Hornick, M. T. Deavers, D. C. Lev, A. J. Lazar, and W.-L. Wang, "ERG and FLI1 protein expression in epithelioid sarcoma," *Modern Pathology*, vol. 27, no. 4, pp. 496–501, 2014.
- [40] D. R. Robinson, Y.-M. Wu, P. Vats et al., "Activating ESRI mutations in hormone-resistant metastatic breast cancer," *Nature Genetics*, vol. 45, no. 12, pp. 1446–1451, 2013.
- [41] J. K. Thomas, M.-S. Kim, L. Balakrishnan et al., "Pancreatic Cancer Database: an integrative resource for pancreatic cancer," *Cancer Biology and Therapy*, vol. 15, no. 8, pp. 963–967, 2014.
- [42] L. M. Hix, J. Karavitis, M. W. Khan, Y. H. Shi, K. Khazaie, and M. Zhang, "Tumor STAT1 transcription factor activity enhances breast tumor growth and immune suppression mediated by myeloid-derived suppressor cells," *Journal of Biological Chemistry*, vol. 288, no. 17, pp. 11676–11688, 2013.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

