

Research Article

Limited Applicability of GW9662 to Elucidate PPAR γ -Mediated Fatty Acid Effects in Primary Human T-Helper Cells

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Synthetic antagonists of the nuclear receptor PPAR γ such as GW9662 are widely used to elucidate receptor-mediated ligand effects. In addition and complementary to recent work, we examined whether GW9662 is suitable to serve for mechanistic investigation in T-helper cells. Human peripheral blood mononuclear cells (PBMC) were preincubated with increasing concentrations of GW9662 (0, 0.4, 2, and 10 $\mu\text{mol/L}$) 30 min before adding the *c9,t11*-isomer of conjugated linoleic acid (*c9,t11*-CLA) as representative of PPAR γ -activating fatty acids with immunomodulatory properties. Corresponding cultures were incubated with GW9662 in the absence of the fatty acid. After 19 h, cells were mitogen stimulated for further 5 h. Subsequently, intracellular IL-2 was measured in CD3⁺CD4⁺ lymphocytes by means of flow cytometry. 100 $\mu\text{mol/L}$ *c9,t11*-CLA reduced the number of T-helper cells expressing IL-2 by 68%. GW9662 failed to abrogate this fatty acid effect, likely due to the fact that the compound exerted an own inhibitory effect on IL-2 production. Moreover, GW9662 dose-dependently induced cell death in human leukocytes. These results suggest that application of GW9662 is not conducive in this experimental setting.

1. Introduction

During the last decades, the scientific knowledge about the role of peroxisome proliferator-activated receptors (PPARs) in controlling metabolic and inflammatory processes has increased steadily. Among the three isoforms of the PPAR family, designated PPAR α (NR1C1), PPAR β/δ (NR1C2; NUC1), and PPAR γ (NR1C3), the latter has been specifically implicated in the regulation of immune cell function, for example, in macrophages [1] and T-helper cells [2]. In T-helper cells, the predominately expressed splice variant $\gamma 1$ is inducible by agonist ligation [3]. Its activation by ligand binding antagonizes the proinflammatory capability of several transcription factors such as nuclear factor- κB (NF- κB), signal transducer and activator of transcription (STAT) [4, 5], and nuclear factor of activated T cells (NFAT) to control

the expression of immunostimulatory cytokines such as IL-2 and IL-4 [6, 7].

Due to their ability to activate PPAR γ with micromolar affinity [8], conjugated linoleic acids (CLA), naturally occurring fatty acids in ruminant fats, aroused scientific interest as potentially anti-inflammatory agents. For instance, we have previously shown that the predominant natural isomer *c9,t11*-CLA reduces expression of the chemokine IL-8 in airway epithelial cells [9], inhibits IL-2 and TNF- α in T-helper cells [10], and prevents experimentally induced airway inflammation in mice at least in part via a PPAR γ -dependent mechanism [11].

GW9662 is widely used to elucidate PPAR γ -dependent anti-inflammatory mechanisms *in vitro* [12, 13] and *in vivo* [14–16]. This molecule covalently modifies the ligand-binding domain by arylation on the cysteine residue Cys²⁸⁵ [17] and

thereby inhibits irreversibly ligand binding to and activation of PPAR γ .

In the present study, which is complementary to previously published work of our group [10], we examined whether GW9662 is suitable to explain PPAR γ -mediated effects of *c9,t11*-CLA in primary human T-helper cells.

2. Materials and Methods

2.1. Chemicals. Lyophilized 2-chloro-5-nitrobenzanilide (GW9662) was solubilized in sterile dimethylsulfoxide (DMSO) according to the manufacturer's instruction (Enzo, Lörrach, Germany, and Sigma, Taufkirchen, Germany) and stored in aliquots at -20°C . *c9,t11*-CLA (Matreya LLC, Pleasant Gap, USA) in free fatty acid form, phorbol 12-myristate 13-acetate (PMA), ionomycin, and brefeldin A (all Enzo) were likewise dissolved in DMSO, aliquoted, and stored at -20°C .

2.2. Purification of PBMC. Mononuclear cells were isolated from buffy coats obtained from peripheral blood of healthy volunteers who gave their written consent for blood donation. Buffy coat blood was 1:1 diluted with PBS (PAA, Cölbe, Germany), layered onto Lymphocyte Separation Medium (LSM) 1077 (1.077 g/mL; PAA; ratio 1:1), and centrifuged at $700 \times g$ for 20 min at 20°C . The PBMC interphase was collected, washed three times with PBS, and resuspended in RPMI 1640 medium supplemented with 10% FBS Gold (PAA).

2.3. Cytokine Production. PBMC ($1 \times 10^6/\text{mL}$) were preincubated for 30 min without or with different concentrations of GW9662 (0.4, 2, and $10 \mu\text{mol/L}$) before $100 \mu\text{mol/L}$ *c9,t11*-CLA was added. After 19 h of incubation, cells were stimulated with PMA (2.5 ng/mL) and ionomycin (0.5 $\mu\text{g/mL}$) in the presence of brefeldin A (5 $\mu\text{g/mL}$) for another 5 h. Control cultures contained maximum 0.2% DMSO. Afterwards, aliquots were stained with anti-human CD3 mAb (PE-Dy647, clone MEM-57, Immunotools, Friesoythe, Germany) and anti-human CD4 mAb (FITC, clone MEM-241, Immunotools) before cells were fixed with 2% formaldehyde (Histofix, Roth, Karlsruhe, Germany). For intracellular cytokine quantification, cells were permeabilized by washing with PBS/0.1% BSA/0.1% saponin, stained with anti-human IL-2 mAb (PE, clone MQ1-17H12, eBioscience), and analyzed in reference to FMO-controls by means of flow cytometry. Nonspecific fluorescence was controlled by incubation with isotype-matched antibodies. Data were assessed and illustrated by WinMDI v.2.8 software (J. Trotter, Scripps Research Institute).

2.4. Cell Viability. To assess the impact of GW9662 on cell viability, PBMC ($1 \times 10^6/\text{mL}$) were incubated without or with 0.4, 2, and $10 \mu\text{mol/L}$ of this compound for 19 h, followed by 5 h stimulation with PMA (2.5 ng/mL) and ionomycin (0.5 $\mu\text{g/mL}$) in the presence of brefeldin A (5 $\mu\text{g/mL}$). Control cultures contained the according volume of DMSO. Cell viability was analyzed by annexin-V (Immunotech, Marseille, France) and propidium iodide (PI; Sigma-Aldrich, Munich,

Germany) exclusion double staining as previously described [10].

2.5. Statistics. Differences in the percentages of IL-2 positive cells were evaluated using a linear mixed model with the fixed factors "fatty acid treatment" (*c9,t11*-CLA and DMSO) and "PPAR γ antagonist treatment" (GW9662 and control) and the interaction of these two factors. The assumption of normality and homoscedasticity was justified by visual inspection of QQ-plots and predicted versus residual plots. A random intercept specific for each subject was included to control for interindividual differences. Tukey-Kramer was conducted as posthoc test and *P* values were adjusted for multiple comparisons. For evaluation of data obtained in the absence of *c9,t11*-CLA, the concentration of GW9662 was entered into the model as fixed factor while IL-2 positive cells, MFI, and viability were defined as dependent variables, respectively. Because the distribution of viability was skewed, a log-transform was applied. For the latter outcome, differences between concentrations $0 \mu\text{mol/L}$ and $0.4 \mu\text{mol/L}$ were additionally evaluated by defining posthoc contrasts between these two concentration levels. Significance of difference was set at $P < 0.05$. All calculations were carried out using SAS 9.3 (PROC MIXED).

3. Results

3.1. GW9662 Fails to Abrogate the Inhibitory Effect of *c9,t11*-CLA on IL-2 Expression in T-Helper Cells. In stimulated control cultures, $15 \pm 2\%$ of the T cells (CD3 $^{+}$) were identified as IL-2 positive T-helper cells (CD3 $^{+}$ CD4 $^{+}$; Figure 1(a)). Incubation with $100 \mu\text{mol/L}$ *c9,t11*-CLA for 24 h significantly reduced the intracellular content of IL-2 in stimulated T-helper cells by 68% to $5 \pm 1\%$. Preincubation with $0.4 \mu\text{mol/L}$ GW9662 did not result in reexpansion of the IL-2 positive T-helper cell population. This was unexpected as preincubation with $0.4 \mu\text{mol/L}$ of the PPAR γ antagonist T0070907, a compound with similar molecular structure to GW9662 except for one single N atom, did so in the aforementioned similar approach [10].

We further tested in a range of fivefold increases of the concentration of GW9662 whether a reversal of the fatty acid effect, in terms of blocked PPAR γ , was achieved. Interestingly, pretreatment with increasing concentrations of GW9662 did not lead to increased IL-2 production but even to a reduction. At $10 \mu\text{mol/L}$ and in the presence of *c9,t11*-CLA, GW9662 caused a drop in the percentage of IL-2 positive T-helper cells even stronger than did the *c9,t11*-CLA treatment alone (Figure 1(b)).

3.2. GW9662 Dose-Dependently Downregulates IL-2 Expression in T-Helper Cells. We next examined whether the PPAR γ antagonist exerted a fatty acid independent effect itself. Indeed, with increasing concentrations of GW9662 we found a continuous reduction in the IL-2 expressing T-helper cell population. Simultaneously, mean fluorescence intensity (MFI) reflecting the cytokine levels on a per-cell basis dose-dependently decreased (Figure 2).

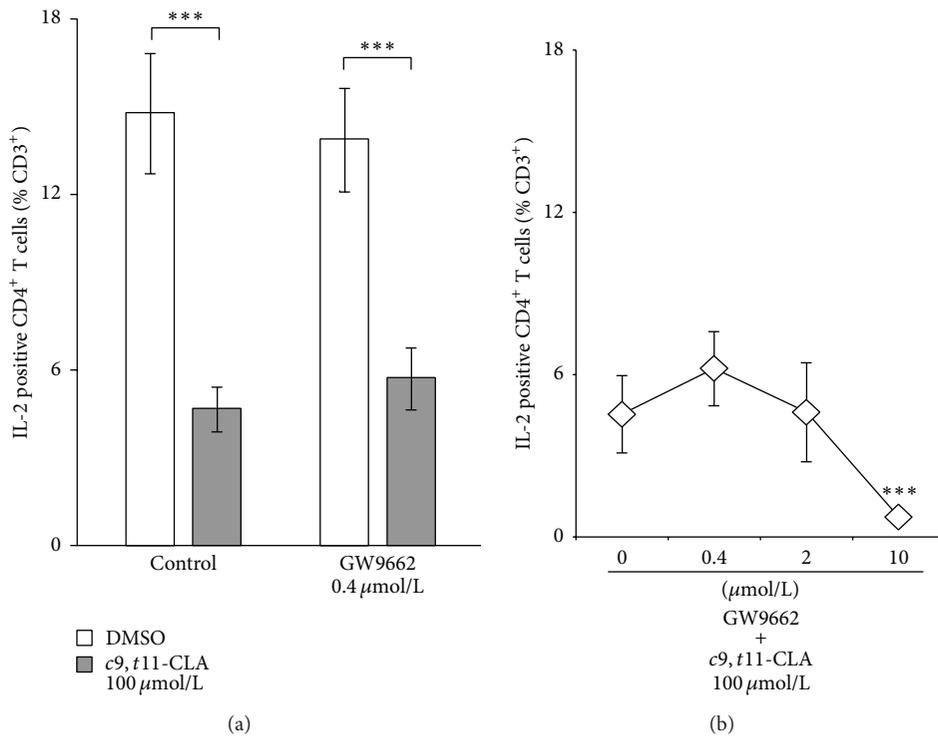


FIGURE 1: GW9662 exerts no effect up to 2 μmol/L and an additive effect on IL-2 inhibition in T-helper cells at 10 μmol/L. PBMC were pretreated for 30 min with GW9662 before 100 μmol/L c9,t11-CLA was added. After 19 h, cells were activated for subsequent 5 h. Intracellular IL-2 was flow cytometrically analyzed in lymphocytes gated for CD3 and CD4. ***P ≤ 0.001. Data are expressed as means ± SEM of n = 6 (a) and n = 5 (b).

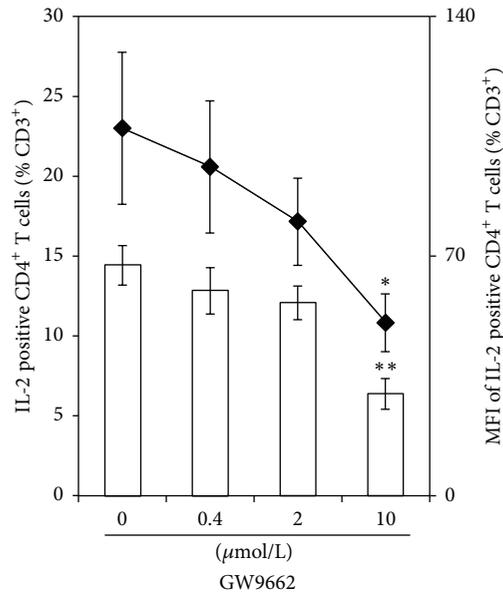


FIGURE 2: GW9662 dose-dependently downregulates IL-2 expression in T-helper cells. PBMC were incubated for a total of 24 h with increasing concentrations of GW9662. After 19 h, cells were activated for further 5 h. IL-2 expression of T-helper cells was flow cytometrically analyzed. Data are expressed as means ± SEM of n = 6. Right scales denote mean fluorescence intensity (MFI) depicted as aligned dots. The dose-dependent effect is statistically significant with **P < 0.01 and *P < 0.05.

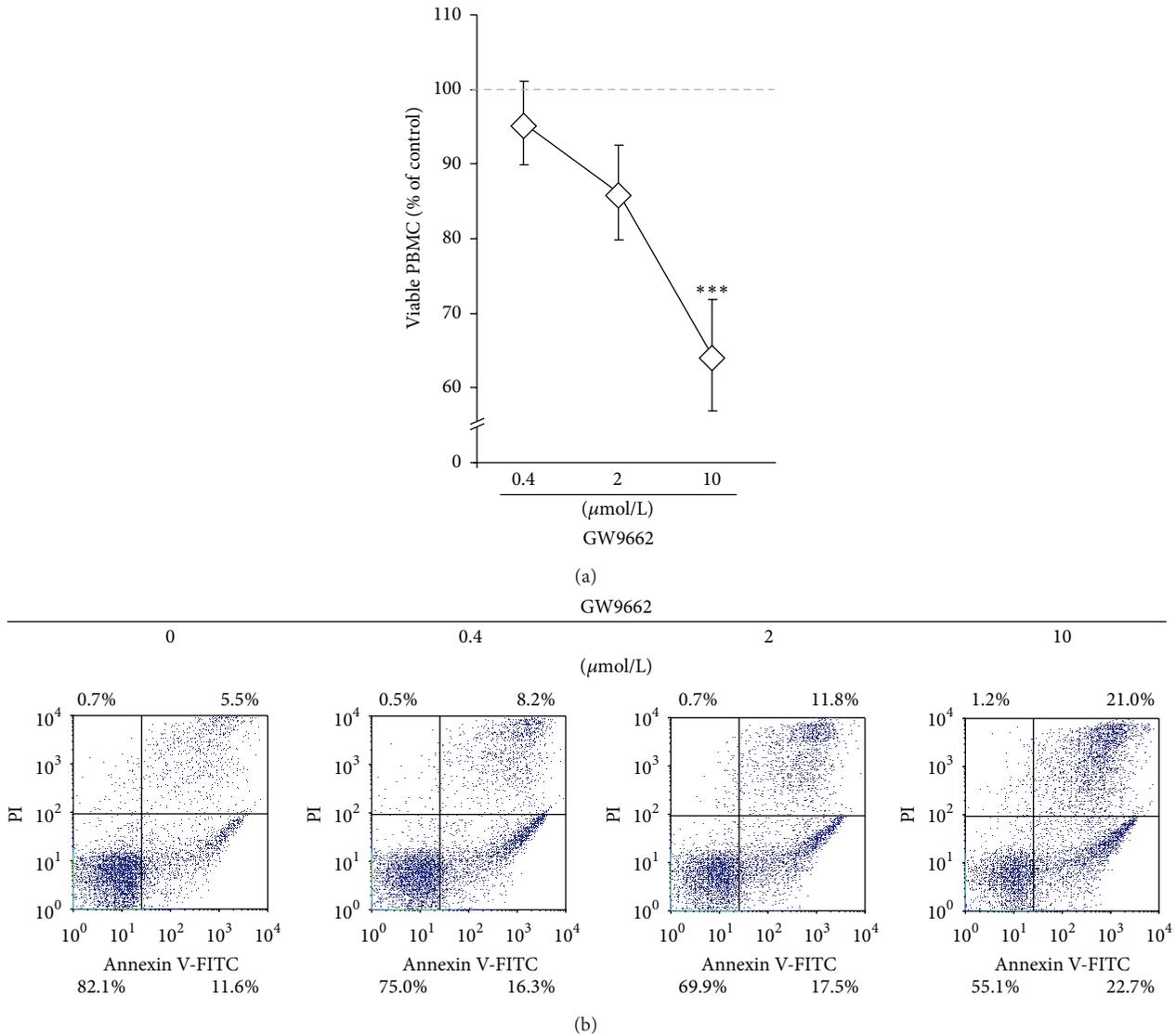


FIGURE 3: GW9662 dose-dependently causes cell death in leukocytes. PBMC were incubated for a total of 24 h with increasing concentrations of GW9662. After 19 h, stimulants were added for further 5 h. Cell viability was flow cytometrically assessed by annexin-V and propidium iodide exclusion double staining and is expressed as % of control without GW9662 (dotted line). Annexin-V positive and PI negative cells were defined as early apoptotic cells; annexin-V positive and PI positive cells were defined as late apoptotic and necrotic cells. (a) Data are expressed as means \pm SEM of $n = 4$. The dose-dependent effect is statistically significant with *** $P < 0.001$. (b) Representative dot plots of GW9662 treated PBMC, gated for lymphocytes.

3.3. GW9662 Dose-Dependently Induces Cell Death of Human Primary Leukocytes. We further assessed whether putative cytotoxic effects underlie the failure of GW9662 to restore the cytokine production inhibited by *c9,t11*-CLA. As revealed by annexin-V and PI exclusion double staining, GW9662 dose-dependently caused cell death in PBMC (Figures 3(a) and 3(b)). After 24 h in the presence of GW9662, viability decreased by up to $35 \pm 8\%$ at $10 \mu\text{mol/L}$. However, at $0.4 \mu\text{mol/L}$ GW9662 did not affect cell viability significantly ($>95\%$ of the control, $P = 0.531$).

4. Discussion

In line with previous work of our group [10], we demonstrated at first that *c9,t11*-CLA reduces the expression of the

immunostimulatory cytokine IL-2 in T-helper cells. We have previously shown that *c9,t11*-CLA acts at least in part via a PPAR γ -mediated pathway, since low-dose cotreatment with the PPAR γ inhibitor T0070907 largely reverted this fatty acid effect [10]. Though intended to be likewise applicable, GW9662 failed to abrogate the fatty acid effect at all tested concentrations in the present approach. This outcome was unexpected, as a large body of evidence exists that indicates suitability of GW9662 to elucidate PPAR γ -dependent mechanisms when used at concentrations within the single- to double-digit micromolar range, including own results from *in vitro* studies in human epithelial cells [9]. However, we have indications that GW9662 acts differently from T0070907 not only in primary lymphocytes but also in other cells

such as macrophages (unpublished findings). Nevertheless, in agreement with the literature, in a similar designed study like the one herein, GW9662 completely negated the modulating effects of *t*10,*c*12-CLA, a synthetic CLA isomer, on TNF- α expression in stimulated porcine PBMC [18]. However, corroborating our findings, Raman et al. recently reported in the Jurkat T-cell line that not only PPAR γ agonists but also its antagonists decreased the mitogen stimulated elevation in intracellular Ca²⁺, which could lead to IL-2 suppression via decreased transcriptional activity of NFAT [19].

In order to justify our data, we repeated the experiments with GW9662 purchased from different manufacturers (not shown). Since the results were comparable we can exclude that false-negative data have been produced. Besides PPAR γ , PPAR α , and PPAR β/δ are also expressed by PBMC [20, 21] and are bound and activated by CLA [22, 23]. However, it is not plausible that the fatty acid effects have been mediated through either of these isoforms, as Cys²⁸⁵, the modified residue in PPAR γ , is conserved among all three PPARs. Moreover, significantly higher concentrations of GW9662 are required for inhibition of ligand binding to PPAR α (factor ~10 over PPAR γ) and PPAR β/δ (factor ~600 over PPAR γ), respectively [17]. We clearly found that GW9662 dose-dependently exerts an own fatty acid independent diminishing effect on IL-2 production in primary T-helper cells. This finding is new and of significance since effects of the antagonist by its own might mask those which should be actually explained by its usage. Moreover, GW9662 is cell toxic in PBMC with increasing concentrations. GW9662 has previously been shown to cause apoptotic cell death in a concentration-dependent manner in oral squamous cells [24] and colon cells [25]. However, in these studies cancer cell lines were used and these cells underwent apoptosis also after treatment with T0070907 at concentrations higher than 10 μ mol/L. The cell death inducing effect of high doses of PPAR γ antagonists led to discuss them as potential therapeutic agents in the treatment of cancer [25, 26] but must be considered undesired in primary cells. However, as cell viability was not affected at 0.4 μ mol/L in our experiments, other effects than cytotoxic underlie the failure of GW9662 to serve for mechanistic exploration of the fatty acid effect that remains elusive.

In summary, and with the restriction that concentrations below 0.4 μ mol/L have not been tested, our data suggest that GW9662 is not valuable for determining the specific PPAR γ -mediated mode of fatty acid action in primary T-helper cells due to own regulatory and cytotoxic effects.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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