

Research Article

Spiritual Holy Water Sites in Ethiopia: Unrecognized High-Risk Settings for Transmission of Pulmonary Tuberculosis

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Ethiopia is a high-tuberculosis (TB) burden country with 157 new cases per 100,000 people, with 23,800 TB-related deaths in 2020. In Ethiopia, TB patients have different healthcare-seeking behaviors. They frequently visit spiritual places, such as holy water sites (HWSs), to seek treatment for their illness spiritually. This study examined the prevalence of pulmonary TB (PTB) and drug susceptibility profiles of Mycobacterium tuberculosis (MTB) isolates among spiritual HWS attendees in Northwest Ethiopia. A cross-sectional study was conducted from June 2019 to March 2020. Sputum samples were collected, processed, and cultured using Löwenstein-Jensen (LJ) culture medium. Second-generation line probe assays (LPAs), GenoType®MTBDRplus VER2.0 and GenoType®MTBDRsl VER2.0, were used to detect anti-TB drug-resistant isolates. STATA 17 was utilized to perform descriptive statistics, bivariate, and multivariate regression analyses. Of 560 PTB-symptomatic participants, 21.8% ((95% confidence interval (95 CI): 18.4–25.2%)) were culture-positive, resulting in a point prevalence of 1,183/100,000 attendees. Amongst HWS attendees, culture-positive TB occurred most commonly in persons 18-33 years of age (28.5% (95 CI 23.4-34.3%)). Other participant characteristics significantly associated with culture-positive PTB were as follows: rural residents (adjusted odds ratio (aOR) 2.65; 95 CI 1.38-5.10), married participants (aOR 2.43; 95 CI 1.28-4.63), family members >5 per household (aOR 1.84; 95 CI 1.04-3.24), and sharing living space (aOR 10.57; 95 CI 3.60-31.13). Also, among 438 participants followed for 12 months after showing negative TB culture results while at the HWS, 6.8% (95 CI 4.4-9.4%) developed or contracted culture-positive TB postresidency at the HWSs. Of the 122 tested isolates, 20 (16.4%) were isoniazid (INH) and/or rifampicin (RIF) resistant. Multidrugresistant (MDR) TB was detected in 15 cases (12.3%), five of which were fluoroquinolones (FLQs) resistant. The findings from this study should raise a concern about HWSs as potential high-risk settings for TB transmission. It is recommended that appropriate control measures be instituted that include compulsory TB testing and tightened infection control at HWSs, where an increased risk exists for transmission of TB.

1. Introduction

Tuberculosis (TB) remains to be a major global health issue [1, 2]. Worldwide, "10.0 million people were infected with TB and over 1.5 million died" from it in 2020 [2]. Until the 2020 coronavirus disease (COVID-19) pandemic, TB was the major infectious agent-related cause of mortality globally [2]. Although TB is a global issue, its prevalence, public health, and economic impact vary greatly between nations [1, 2]. TB is the leading cause of mortality among infectious diseases in low- and middle-income countries (LMICs) due to several factors. Poor healthcare access, overcrowded living conditions, poor nutrition, and the HIV/AIDS pandemic all contribute to the high burden of TB in LMICs [2-4]. Ethiopia, like other LMICs, has a high burden of TB, which poses challenges to its public healthcare system [2, 5, 6]. Ethiopia's TB incidence was 157 per 100,000 persons, with 23,800 people dying from the disease in 2020 [1], suggesting that TB continues a significant cause of death in the country.

The emergence of drug-resistant TB (DR-TB) has posed a significant threat to global and national TB control efforts, as DR-TB has the potential to spread globally, emphasizing the need for additional prevention and care measures [1, 2]. A recent national report revealed that the prevalence of MDR or RIF-resistant TB (MDR/RR-TB) in Ethiopia was 0.71% among newly diagnosed TB cases and 12.0% in retreated TB patients [1]. Besides, "pre-extensively drugresistant (pre-XDR)" TB and "extensively drug-resistant (XDR)" TB were 5.7% and 0.6%, respectively [7]. Since the country lacks the facility to undertake universal drug susceptibility testing (DST) on all incident TB cases, early TB case detection and initiation of proper treatment are problematic in many public health facilities in Ethiopia [8].

Early TB case detection, successful TB patient treatment according to international standards, and TB prevention are global plans to halt TB and reduce deaths and transmission [2, 9]. However, "a key obstacle to achieving this goal has been that many people with TB are currently being" missed by healthcare systems [1]. Globally, over 2.9 million TB cases were anticipated to be underdiagnosed or diagnosed but unreported, most of which occurred in LMICs with weak healthcare systems [1]. Finding these cases, effectively diagnosing them, and commencing appropriate treatment are critical to containing the disease [1, 10]. In Ethiopia, about two-thirds of individuals with active TB in the community remain undiagnosed and thus untreated [11, 12]. This could be because the Ethiopian TB prevention and control program relies mostly on passive case finding, which requires TB suspects to self-present and visit public healthcare facilities [12]. However, studies showed that individuals with TB symptoms in LMICs, such as Ethiopia, exhibit different healthcare-seeking behaviors and use other methods before seeking treatment at public healthcare facilities [13-15]. Besides, people from poor communities have insufficient knowledge about TB disease, and they often confuse the symptoms of TB as indicative of other infections [14]. Moreover, healthcare facilities are not nearby to give TB services, and people with TB symptoms often live in places where government services have a hard time reaching, due to travel costs and fear of stigma, TB patients may not visit public healthcare facilities for diagnosis and treatment [14, 15]. Thus, TB patients can delay the opportunity for early diagnosis and the initiation of appropriate treatments. This can intensify and threaten TB transmission in households and the community.

In Ethiopia, people use spiritual holy water as an alternative treatment for a variety of diseases, including respiratory ailments [13, 16-18]. Holy water sites (HWSs) are designated areas with springs that are believed to have the power to cure various types of illness [13]. Although the number of people who use holy water as a treatment option is not well documented in the country, it is known that attendees believe in its healing power and use it as an alternative treatment for various illnesses. Particularly, Ethiopian Orthodox Tewahedo Christians have unwavering faith in the curative power of spiritual holy water [13, 16]. Thus, individuals from different parts of the country travel to HWSs to seek the curative power of holy water blessed by Orthodox priests and they reside in shared living spaces (rooms) for a certain period. The rooms are built as temporary waiting spaces, and they are small, overcrowded, and not well-ventilated, which can increase the risk of TB and other respiratory disease transmissions [13, 19].

In the Amhara region, the focus of this study, over 82.5% of the population are Ethiopian Orthodox Tewahedo Christian followers [13, 20], and faith-based therapy with spiritual holy water for TB and other diseases is widely practiced [13]. Although studies show that TB patients seek care from traditional healers and spiritual HWSs, the burden of TB and the drug resistance pattern of MTB isolates among HWS attendees in Ethiopia have not been thoroughly investigated. To the best of our knowledge, one earlier study has shown that "the prevalence of PTB among HWS attendees was 7.4-fold higher than the prevalence in the general population in Ethiopia" [13]. However, this study covered a limited geographical area and used smear microscopy to diagnose TB in symptomatic persons; given the low sensitivity of this technique, its findings might not reflect the true TB burden among this cohort of populations in the region. Hence, identifying such special congregate settings in the indigenous communities, considering these settings as hotspot sites for TB transmission, and conducting a systematic TB screening would aid in reaching those unreachable, missed, or undiagnosed TB cases for timely diagnosis and care. Therefore, this study aimed to assess the prevalence of TB and drug susceptibility profiles of MTB isolates among individuals with symptoms of PTB attending spiritual HWSs in the Amhara region, Ethiopia.

2. Methods

2.1. Study Setting. The Amhara region is located in the northwestern parts of Ethiopia and comprises eleven zones and three administrative towns (Figure 1). A cross-sectional study was done between June 2019 and March 2020 in nine purposefully selected HWSs found across nine administrative zones in the region. One HWS was chosen from each of the study zones. The HWS in each zone was chosen based



FIGURE 1: The study area map.

on its consistent popularity for holy water treatment, its ability to accommodate many attendees, and where many people visit it throughout the year and stay for an extended time [13, 18] (Figure 1 and Table S1).

2.2. Population, Study Participants, and Recruitment. The study population comprised all attendees at the HWSs during the data collection period [18]. Following TB screening criteria, individuals with a persistent cough lasting two weeks or longer and other PTB-suggestive symptoms, such as productive cough or expectorating blood-containing sputum, fever, chest pain, shortness of breath, fatigue, night sweating, loss of appetite, unexplained weight loss, and contact history with active TB patients and history of TB disease, were screened [5]. A total of 10,313 attendees (≥18 years of age) were screened at nine selected HWSs during the study period; 560 of these individuals exhibited symptoms of PTB and participated. The study settings, total attendees screened for PTB-suggestive symptoms, anticipated PTB-symptomatic attendees from each study site, total participants, and laboratory test results are depicted in the additional files (Table S1).

2.3. Eligibility Criteria. Inclusion criteria: Individuals who were \geq 18 years of age and fulfilled the screening criteria were included in the study [5, 21]. Exclusion criteria: Individuals who were seriously ill and unable to provide sputum samples and other relevant clinical or demographic information were excluded. Moreover, individuals who were receiving anti-TB treatment during data collection, and those whose permanent address was outside of the study region were excluded.

2.4. Screening and Socio-Demographic Data Collection. All attendees were screened for symptoms suggestive of PTB, following the guidelines [5, 21]. Trained nurses and medical laboratory technologists with experience in TB screening and similar field data collection perform the data collection. Descriptive demographic data were collected from eligible HWS attendees by interviewer-administered questionnaire. Participants' socio-demographic data included their sex, age, marital status, educational status, household size, and place of residence, as well as risk factors for PTB infection such as a history of TB disease, contact with active TB patients or person with a chronic cough, family history of TB disease, previous HWS visits (the last one year), the number of days 2.5. Culture and Identification Test. Sputum specimens from attendees with PTB symptoms were obtained using sterile and leak-proof collection containers [18]. The sputum specimen was placed in an icebox and delivered to the Amhara Public Health Institute, a regional public health referral laboratory center, Bahir Dar, Ethiopia [18]. Sputum prepared and inoculated specimens were into a Löwenstein-Jensen (LJ) culture medium following standard laboratory procedures. Ziehl-Neelsen (ZN) staining was done to confirm all LJ culture-positive isolates [22]. The MPT64 antigen test (Capilia TB-Neo, TAUNS Laboratories, Inc., Japan) was used to differentiate MTB complex species from non-TB mycobacteria (NTM) [23].

2.6. Specimen Preparation. After subculturing, suspensions were prepared from LJ-positive specimens and transferred into 1.5 ml of PrimeStore Molecular Transport Medium (PS-MTM; Longhorn Vaccine and Diagnostics, San Antonio, Texas, USA). Preparing suspensions of MTB colonies from LJ culture media depended on the culture state [24]. In brief, for intact slopes, colonies were gently scraped off using an inoculation loop and suspended (washed down) in 1 ml of sterile water in the original culture bottle. After pipetting off the suspension, it was transferred into a 1.5-ml Eppendorf tube and then transferred to the 1.5 ml PS-MTM [24]. Mycobacterial suspensions prepared in PS-MTMP tubes were transported to South Africa by air at ambient temperature for other genotyping procedures.

2.7. DNA Extraction. The MTB DNA was extracted from all 122 LJ-positive isolates using the PrimeXtract[™] kit (Longhorn Vaccines and Diagnostics, San Antonio, TX, USA) following the manufacturer's instructions [25]. Briefly, $200 \,\mu\text{L}$ of 100% ethanol, $200 \,\mu\text{L}$ of lysis buffer, and $200 \,\mu\text{L}$ of MTB inoculum (preserved in PS-MTM) were transferred into a 1.5-mL microcentrifuge tube, then vortexed and centrifuged. The entire supernatant was transferred to a microextraction column and centrifuged at 13,000 rpm for 1 minute, and the flow-through material was discarded. Wash buffer 1 (500 μ L) was applied to the extraction column and centrifuged at 13,000 rpm for 1 minute, followed by further addition of wash buffer 2 (500 μ L) to the extraction column and subsequent centrifuging as described above, discarding the flow-through material. Then, DNA was eluted by 1 minute of centrifugation at 13,000 rpm using $50 \,\mu\text{L}$ of preheated (60-70°C) elution solution [25]. For future use, the extracted MTB DNA was preserved at -20°C. The concentration and quality of extracted genomic DNA were assessed using a spectrophotometer at the optimal densities of 280 nm and 260 nm [18].

2.8. Drug Susceptibility Testing. Following MTB genomic DNA extraction using the PrimeXtractTM kit instructions [25], the second-generation line probe assays (LPAs),

MTBDR*plus* VER2.0 (to detect RIF and INH resistance), and MTBDR*sl* VER2.0 kit strips (to detect FLQs and aminoglycosides/peptide resistance) were performed following the manufacturer's protocol (Hain Lifescience GmBH, Nehren, Germany) [26, 27]. All INH and RIF-resistant TB isolates were tested using MTBDR*sl* VER 2.0 for detecting FLQs and second-line injectable drugs (SLIDs)-resistant isolates. For each run of LPAs, MTB strains H37Rv susceptible to all anti-TB drugs tested and molecular-grade water were used as a positive and negative control, respectively.

2.9. Prospective Follow-Up Study. A prospective follow-up study on 438 PTB-symptomatic individuals with culturenegative test results while at HWSs was done to determine the prevalence of developing active TB disease subsequent to residing at HWS. The duration of follow-up was 12 months, starting on the date that a negative culture result was obtained. TB status was confirmed via telephone or in-person contact, and any reports of active TB were confirmed by reviewing the patient's medical records at the diagnosing public healthcare facility. The date of diagnosis, the diagnostic method (acid-fast bacilli (AFB) smear microscopy, GeneXpert®MTB/RIF assay, LPAs, or other diagnoses, like chest X-rays, and clinical diagnoses), the diagnostic public healthcare facility, and the location (zone) were recorded from participants who reported developing active TB disease during the follow-up period.

2.10. Data Analysis. Laboratory test results and sociodemographic data were recorded in a Microsoft Excel spreadsheet, coded, and entered into STATA 17 (STATA; StataCorp) for analysis. The data were checked for completeness and consistency by running the frequencies of each variable. The results were summarized using descriptive statistics. Bivariate and multivariate logistic regression analysis models were employed to identify associated risk factors for culture-positive TB and developing active TB disease post-residency at the HWSs. Predictors having a p < 0.2 in the bivariable analysis were included in the multivariate analysis. In multivariate logistic regression analysis, the odds ratio (OR) and 95% confidence intervals were retrieved and variables with a p value of less than 0.05 were considered statistically significant.

2.11. Research Ethics. The Human Research Ethics Committee of the Faculty of Health Sciences, University of Pretoria, South Africa (Ethics Ref. No: 600/2018), and the Ethiopian National Research Ethics Review Committee (Ethics Ref. No: SHE/SM/14.4/708/2019) approved the study. Moreover, a signed official permission letter was sought from the Orthodox Tewahedo Church, Patriarchate Head Office, Addis Ababa, Ethiopia (Ref. No. 2478/6275/ 2011). A verbal or written consent declaration with details about the study was given or explained to participants and signed, and the study was conducted following the Helsinki Declaration. Participants who tested positive for PTB were transferred/linked to a nearby public healthcare facility for treatment and further patient management.

3. Results

3.1. Socio-Demographic Profiles of Participants. A total of 10,313 HWS attendees were screened for PTB-suggestive symptoms, and 560 (5.4%) were found to have PTB symptoms and participated. Of 560 participants, 308 (55.0%) were males and 263 (47.0%) were between the ages of 18 and 33 with a median age of 35 years. Most of the study participants (356; 63.6%) were married, whereas 302 (53.9%) were rural residents (Table 1).

3.2. Prevalence of Culture-Positive Pulmonary Tuberculosis. Of 560 PTB-symptomatic participants, 122 (21.8%) (95 CI: 18.4–25.2%) were culture-positive, resulting in a point prevalence of 1,183 per 100,000 attendees. The proportion of males and females with culture-positive PTB was nearly the same (21.8% each). The majority of bacteriologically confirmed cases (76/302; 25.2%) were rural residents. Moreover, participants aged 18–33 years (75/263; 28.5%) had a higher rate of culture-positive PTB (Table 2 and Table S2).

3.3. Risk Factors for Culture-Positive Pulmonary Tuberculosis. The bivariate logistic regression analysis revealed that participants aged 34–49 years ((crude odds ratio (cOR) 0.44; 95CI: 0.28–0.70)), and rural residents (cOR 1.55; 95CI: 1.03–2.34) were statistically associated with culture-positive PTB. Additionally, the analysis revealed that few independent variables were statistically associated with culture-positive PTB (p < 0.05), including a history of TB disease, contact with chronic coughers or active TB patients, having had close contact with a family member who has TB, the number of days (>21) spent at HWS, and sharing living spaces (rooms) at HWS (Table S3).

In the final multivariate logistic regression model, place of residence, marital status, family size per household, and sharing a living space (rooms) at HWSs were positively associated with culture-positive PTB. Rural residents were two times more likely to develop culture-positive PTB compared to urban residents (aOR 2.65; 95CI: 1.38–5.10). Married participants were more likely to have culturepositive PTB than single participants (aOR 2.43; 95CI: 1.28–4.63). On the other hand, participants with more than five family members per household were 1.84 times more likely to have culture-positive PTB than those with less than five (aOR 1.84; 95CI:1.04–3.24). Besides, sharing a living space at HWS increased the risk of developing culturepositive PTB by tenfold (aOR 10.57; 95CI: 3.60–31.13) (Table 3).

3.4. Drug Resistance Profiles of Mycobacterium tuberculosis Isolates. Of 122 isolates tested, 83.6% (102) were susceptible to both RIF and INH, while 16.4% (20) were resistant to RIF and/or INH. Any INH and RIF-resistant TB was detected in 16.4% (20) and 12.3% (15) isolates, respectively. Multidrugresistant TB (MDR-TB) (resistant to both RIF and INH) was found in 12.3% (15) of culture-positive cases, five (4.1%) of which were fluoroquinolones- (FLQs-) resistant TB isolates.

TABLE 1: Socio-demographic characteristics of PTB-symptomatic HWS attendees (n = 560).

Socio-demographic characteristics Frequence			
Sov	Male	308 (55.0)	
Sex	raphic characteristics Frequency, n (% Male 308 (55.0) Female 252 (45.0) 18–33 263 (47.0) 34–49 208 (37.1) ≥50 89 (15.9) Urban 258 (46.1) Rural 302 (53.9) Married 356 (63.6) Single* 204 (36.4) Can't read and write 256 (45.7) Primary school 165 (29.5) Secondary school and above 139 (24.8) 1-5 294 (52.5) >5 266 (47.5) Farmer 235 (42.0) Employed ^a 24 (4.3) Unemployed ^b 117 (20.9) Housewife 94 (16.8) Students and others** 90 (16.1)		
	18-33	263 (47.0)	
Age (year)	34-49	208 (37.1)	
0 1	≥50	89 (15.9)	
Desidence	Urban	258 (46.1)	
Residence	Rural	302 (53.9)	
Marital status	Married	356 (63.6)	
Warnar status	Single*	204 (36.4)	
	Can't read and write	256 (45.7)	
Educational	Primary school	165 (29.5)	
status	Secondary school and above	139 (24.8)	
Hawaahald size	1-5	294 (52.5)	
Household size	>5	266 (47.5)	
	Farmer	235 (42.0)	
	Employed ^a	24 (4.3)	
Occupation	Unemployed ^b	117 (20.9)	
	Housewife	94 (16.8)	
	Students and others**	90 (16.1)	

Notes. *Single, divorced, and widowed; **religious leaders and deacons; HWS: holy water site; PTB: pulmonary tuberculosis; ^aconstruction worker, administrative worker, healthcare worker, public transport worker; ^bbusinessman, trader, daily laborer.

These five FLQs-resistant strains were identified at the South Wello zone study site. Furthermore, 7.1% (3/42 retreated cases) of RIF-resistant/MDR (RR/MDR) TB isolates were detected in previously treated TB cases (Table 4).

3.5. Risk Factors Associated with Drug Resistance. The multivariate logistic regression analysis revealed that a history of TB disease and study area has a significant association with the occurrence of some form of drug-resistant TB. Attendees who had a history of TB disease were nine times more likely to suffer from drug-resistant TB as compared with other HWS attendees who had no history of TB (aOR 9.22; 95 CI: 1.55-54.82). On the other hand, participants who attended HWS in the South Wello zone were three times more likely to develop any drug-resistant TB as compared with those who attended HWS in Central Gondar (aOR 3.06; 95 CI: 0.21-6.72). Similarly, the analysis revealed that attendees' occupations and study areas have a significant association with the occurrence of MDR-TB. Thus, farmers and housewives were nine (aOR 9.78; 95 CI: 1.55-61.59) and fifteen (aOR 15.68; 95 CI: 1.46-168.10) times more likely to develop MDR-TB as compared with students, respectively. Moreover, participants who attended HWS in the South Wello zone were twice as likely to develop MDR-TB as compared to those who attended HWS in Central Gondar (aOR 2.08; 95 CI: 0.31-3.92) (Table 5).

In addition, the proportion of any drug-resistant and MDR-TB in each study zone was estimated based on the participant's age category and found that participants aged 18–33 years appeared to be the most affected. Thus,

Socia demographic characteristics		Prevalence of cu	6 l		
Socio-dellic	graphic characteristics	Positive, n (%)	Negative, n (%)	<i>p</i> value	
Sex	Male Female	67 (21.8) 55 (21.8)	241 (78.2) 197 (78.2)	1.00	
Age group (year)	18-33 34-49 ≥50	75 (28.5) 31 (14.9) 16 (18.0)	188 (71.5) 177 (85.1) 73 (82.0)	0.001	
Residence	Urban Rural	46 (17.8) 76 (25.2)	212 (82.2) 226 (74.8)	0.040	
Marital status	Married Single*	85 (23.9) 37 (18.1)	271 (76.1) 167 (81.9)	0.136	
Educational status	Can't read and write Primary school Secondary school and above	59 (23.0) 30 (18.2) 33 (23.7)	197 (77.0) 135 (81.8) 106 (76.3)	0.396	
Household size	1–5 >5	57 (19.4) 65 (24.4)	237 (80.6) 201 (75.6)	0.153	
Occupation	Farmer Employed ^a Unemployed ^b Housewife Students and others**	45 (19.1) 6 (25.0) 23 (19.7) 24 (25.5) 24 (26.7)	190 (80.9) 18 (75.0) 94 (80.3) 70 (74.5) 66 (73.3)	0.495	

TABLE 2: Proportion of bacteriologically confirmed PTB with socio-demographic characteristics of participants (n = 560).

Notes. *Single, divorced, and widowed; **religious leaders and deacons; PTB: pulmonary tuberculosis; ^aconstruction worker, administrative worker, healthcare worker, public transport worker; ^bbusinessman, trader, daily laborer.

TABLE 3: Multivariate logistic regression analysis of potential associated risk factors for culture-positive TB among PTB-symptomatic HV	WS
attendees $(n = 560)$.	

Variables		Culture-p	ositive PTB		. 1	
v allables		Positive (n)	Negative (n)	aOR (95% CI)	<i>p</i> value	
Residence	Urban Rural	46 76	212 226	Ref 2.65 (1.38–5.10)	0.003	
Marital status	Married Single*	85 37	271 167	2.43 (1.28–4.63) Ref	0.007	
Household size	1-5 >5	57 65	237 201	Ref 1.84 (1.04–3.24)	0.036	
Sharing drinking cups at the HWS	Yes No	91 31	268 170	10.59 (3.60–31.13) Ref	<0.001	
Sharing a living space at the HWS	Yes No	118 4	320 118	10.57 (3.60–31.13) Ref	< 0.001	
Do you know your HIV status?	Yes No	34 88	179 259	Ref 2.69 (1.50–4.84)	0.001	

Notes. *Single, divorced, and widowed; aOR: adjusted odds ratio; CI: confidence interval; HIV: human immunodeficiency virus; HWS: holy water sites; PTB: pulmonary tuberculosis; ref: reference.

attendees aged 18–33 years who attended HWS in the South Wello zone had the highest rate of any drug-resistant TB, with a rate of $47.1\% \pm 12.1$ SE (95 CI: 24.3-71.1%), and MDR-TB with a rate of $58.3\% \pm 14.2$ SE (95 CI: 28.5-83.1%) (Table S4). On the other hand, the logistic regression analysis showed that the odds of developing any drug-resistant TB and MDR-TB were 14.54 (95 CI: 1.65-128.44) and 12.00 (95% CI: 1.35-106.80) times higher in the South Wello zone compared to the North Wello zone (Table S5).

3.6. The Prospective Follow-Up Study Results. A prospective follow-up study on 438 PTB-symptomatic individuals with culture-negative tests while at HWS was done to determine the proportion of developing active TB disease subsequent to residing at HWS. The duration of follow-up was 12 months, starting on the date that a negative culture result was obtained. Among 438 participants who were on follow-up, 30 (6.8%) (95 CI 4.4–9.4%) developed active TB disease post-residency at the HWS. Male participants and those

TABLE 4: The proportio	n of drug-resistant	TB isolates and	characteristics o	f study	participants	(n = 122).
	<i>(</i>)					· · · · · ·

Characteristics		N	Anti-TB drug resistance patterns			
C		IN	INH^{r} , n (%)	RIF ^r , <i>n</i> (%)	RIF ^r /MDR-TB, n (%)	FLQs-resistant, n (%)
Corr	Male	67	14 (20.9)	9 (13.4)	9 (13.4)	2 (3.0)
Sex	Female	55	6 (10.9)	6 (10.9)	6 (10.9)	3 (5.5)
	18-33	75	17 (22.7)	12 (16.0)	12 (16.0)	5 (6.7)
Age (year)	34-49	N Anti-TB drug resistance patterns INH ^r , n (%) RIF ^r , n (%) RIF ^r /MDR-TB, n (%) FLC le 67 14 (20.9) 9 (13.4) 9 (13.4) ale 55 6 (10.9) 6 (10.9) 6 (10.9) 33 75 17 (22.7) 12 (16.0) 12 (16.0) 49 31 0 (0.0) 0 (0.0) 0 (0.0) 0 16 3 (18.8) 3 (18.8) 3 (18.8) an 46 10 (21.7) 7 (15.2) 7 (15.2) al 76 10 (13.2) 8 (10.5) 8 (10.5) ied 85 11 (12.9) 7 (8.2) 7 (8.2) e* 37 9 (24.3) 8 (21.6) 8 (21.6) and write 59 6 (10.2) 4 (6.8) 4 (6.8) school 30 8 (26.7) 7 (23.3) 7 (23.3) ol and above 33 6 (18.2) 4 (12.1) 4 (12.1) 5 57 13 (22.8) 9 (15.8) 9 (15.8)	0 (0.0)			
	≥50	16	3 (18.8)	3 (18.8)	3 (18.8)	0 (0.0)
D: J	Urban	46	10 (21.7)	7 (15.2)	7 (15.2)	2 (4.3)
Residence	Rural	76	10 (13.2)	8 (10.5)	8 (10.5)	3 (3.9)
	Married	85	11 (12.9)	7 (8.2)	7 (8.2)	2 (2.4)
Marital status	Single*	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	3 (8.1)			
	Can't read and write	59	6 (10.2)	4 (6.8)	4 (6.8)	0 (0.0)
Educational status	Primary school	30	8 (26.7)	7 (23.3)	7 (23.3)	4 (13.3)
Educational status Secor	Secondary school and above	33	6 (18.2)	4 (12.1)	4 (12.1)	1 (3.0)
TT 1 11 :	1-5	57	13 (22.8)	9 (15.8)	9 (15.8)	3 (5.3)
Household size	>5	65	7 (10.8)	6 (9.2)		2 (3.1)
	Farmer	45	4 (8.9)	2 (4.4)	2 (4.4)	1 (2.2)
	Employed ^a	6	3 (50.0)	2 (33.3)	2 (33.3)	1 (16.7)
Age (year) Residence Marital status Educational status Household size Occupation Study area (zone) C Types of PTB cases	Unemployed ^b	23	5 (21.7)	3 (13.0)	3 (13.0)	1 (4.3)
	Housewife	24	1 (4.2)	1 (4.2)	1 (4.2)	0 (0.0)
	Students and others**	24	7 (29.2)	7 (29.2)	7 (29.2)	2 (8.3)
	North Wello	22	1 (4.5)	1 (4.5)	1 (4.5)	0 (0.0)
	South Wello	22	9 (40.9)	8 (36.4)	8 (36.4)	5 (22.7)
Study area (zone)	North Shewa	33	3 (9.1)	3 (9.1)	3 (9.1)	0 (0.0)
	South Gondar	28	6 (21.4)	2 (7.1)	2 (7.1)	0 (0.0)
	Central Gondar and others***	17	1 (5.9)	1 (5.9)	1 (5.9)	0 (0.0)
Turnes of DTD areas	Previously treated	42	3 (7.1)	3 (7.1)	3 (7.1)	1 (2.4)
Types of PID cases	Newly diagnosed	80	17 (21.3)	12 (15.0)	12 (15.0)	4 (5.0)

Notes. *Single, divorced, and widowed; **others: religious leaders and deacons; ***others: Awi zone, West Gojjam, East Gojjam, and Wag-Hamra FLQs: fluoroquinolones; HWS: holy water site; INH^r: isoniazid resistance; MDR: multidrug-resistant; *N*: total number of culture-positive cases; PTB: pulmonary tuberculosis; RIF^r: rifampicin resistance; TB: tuberculosis; ^aconstruction worker, administrative worker, healthcare worker, public transport worker; ^bbusinessman, trader, daily laborer.

who were over 50 years of age had a great share of developing active TB disease post-residency at the HWSs, with a proportion of 5.3% (n = 23) and 3.3% (n = 14), respectively (Table S6).

3.7. Risk Factors for Developing Active TB Disease Post-Exposure to HWS. The multivariate logistic regression analysis revealed that gender and educational status of participants were associated risk factors for developing active TB disease post-residency at the HWSs. Thus, the odds of developing active TB disease were eight times higher among females (aOR 8.43; 95 CI: 1.90–36.70), whereas it was sixfold higher among those who cannot read and write (aOR 6.09; 95 CI: 1.25–29.55). Furthermore, the odds of developing active TB disease were higher among participants who had a history of contact with TB patients in their family members (aOR 25.20; 95 CI: 3.01–206.50), shared living space at HWS (aOR 37.19; 95 CI: 2.46–561.20), and those who had recently been hospitalized (aOR 14.50; 95 CI: 1.16–180.90) compared to their counterparts (Table S7).

4. Discussion

In Ethiopia, HWSs are traditional places of healing where people travel all over to seek the restorative benefits of holy water blessed by Orthodox priests [13, 16, 19]. In the study region, people use faith-based therapy with spiritual holy water for different diseases [13, 16, 17]. Although studies have shown that TB patients seek treatment from traditional healers and HWSs, the burden of TB among HWS attendees in Ethiopia has not been thoroughly studied.

In this study, the prevalence of culture-positive PTB was 21.8%. Thus, the point prevalence was 1,183 per 100,000 attendees, which was 4.3 times higher than a national TB prevalence study in Ethiopia, which reported 277 per 100,000 bacteriologically confirmed TB cases and 108 per 100,000 smear-positive TB cases [28]. Similarly, our finding was 1.5-fold higher than a previous study that reported 795 per 100,000 HWS attendees had TB in the study region [13]. The same study reported that the prevalence of smear-positive PTB among adult HWS attendees was 7.4-fold higher than Ethiopia's national TB prevalence [13]. This

Variables		Ν	Any drug resistance (RIF ^r and/or INH ^r)		aOR (95% CI)	p value
			Yes, <i>n</i> (%)	No, n (%)		-
Histomy of TP diagona	Yes	41	2 (4.9)	39 (95.1)	9.22 (1.55-54.82)	0.015
mistory of 1 b disease	No	81	18 (22.2)	63 (77.8)	Ref	
	Employed	6	3 (50.0)	3 (50.0)	0.74 (0.06-9.27)	0.817
	Unemployed	23	5 (1.7)	18 (78.3)	1.19 (0.26-5.36)	0.823
Occupation	Farmer	45	4 (8.9)	41 (91.1)	4.57 (0.99-21.19)	0.052
	Housewife	24	1 (4.2)	23 (95.8)	10.18 (0.95-109.55)	0.056
	Student and others*	24	7 (29.2)	17 (70.8)	Ref	
	North Wello	22	1 (4.5)	21 (95.5)	0.96 (0.05-20.16)	0.980
	South Wello	22	9 (40.9)	13 (59.1)	3.06 (0.21-6.72)	0.026
The study area (zone)	North Shewa	33	3 (9.1)	30 (90.9)	0.82 (0.07-9.36)	0.872
· · · ·	South Gondar	28	6 (21.4)	22 (78.6)	0.23 (0.02-2.34)	0.212
	Central Gondar and others**	17	1 (5.9)	16 (94.1)	Ref	
		RIF ^r /MDR-TB				
			Yes, <i>n</i> (%)	No, n (%)	aOR (95% CI)	<i>p</i> value
Manifal status	Married	85	7 (8.2)	78 (91.8)	_	_
Marital status	Single***	37	8 (21.6)	29 (78.4)	aOR (95% CI) 9.22 (1.55–54.82) Ref 0.74 (0.06–9.27) 1.19 (0.26–5.36) 4.57 (0.99–21.19) 10.18 (0.95–109.55) Ref 0.96 (0.05–20.16) 3.06 (0.21–6.72) 0.82 (0.07–9.36) 0.23 (0.02–2.34) Ref aOR (95% CI) 	_
	Employed ^a	6	2 (33.3)	4 (66.6)	2.84 (0.27-29.59)	0.383
	Unemployed ^b	23	3 (13.0)	20 (87.0)	3.42 (0.62-18.84)	0.157
Occupation	Farmer	45	2 (4.4)	43 (95.6)	9.78 (1.55-61.59)	0.015
-	Housewife	24	1 (4.2)	23 (95.8)	15.68 (1.46-168.10)	0.023
	Student and others*	24	7 (29.2)	17 (70.8)	Ref	
	North Wello	22	1 (4.5)	21 (95.5)	0.83 (0.04-16.71)	0.901
Study area (zone)	South Wello	22	8 (36.4)	14 (63.6)	2.08 (0.31-3.92)	0.043
	North Shewa	33	3 (9.1)	30 (90.9)	0.61 (0.05-6.96)	0.688
	South Gondar	28	2 (7.1)	26 (92.9)	0.85 (0.07-11.13)	0.901
	Central Gondar and others**	17	1 (5.9)	16 (94.1)	Ref	

TABLE 5: Factors associated with any drug-resistant TB (RIF^r and/or INH^r) and RIF^r/MDR-TB.

Notes. *Others: religious leaders and deacons; **others: Awi, West Gojjam, Wag-Hamra, East Gojjam; ***single, widowed, and divorced; aOR: adjusted odds ratio; CI: confidence interval; INH^r: isoniazid resistance; ref: reference; RIF^r/MDR-TB: rifampicin-resistant/multidrug-resistant tuberculosis; RIF^r: rifampicin resistance; TB: tuberculosis; ^aconstruction worker, administrative worker, healthcare worker, public transport worker; ^bbusinessman, trader, daily laborer.

discrepancy may be due to the study population and laboratory diagnosis methods. The national TB prevalence survey report in Ethiopia excluded congregate settings and high-risk groups found in the HWSs [28]. On the other hand, Derseh and his colleagues used sputum smear microscopy, which has low sensitivity for TB detection, resulting in a low-prevalence finding report [13]. The high prevalence of culture-positive PTB at HWSs may be due to overcrowding, close contact, inadequate ventilation of shared living spaces, and long stays, which increase exposure and TB transmission [13]. People who visit HWS choose to treat their diseases religiously, including TB and HIV/AIDS, and perceive spiritual HWS as their best treatment choice [13, 16, 18]. In addition, people in rural areas and poor communities lack knowledge of TB and often misinterpret TB symptoms as signs of other diseases [14]. Another possible explanation for the high prevalence of PTB in our study and the report of a previous study of a similar nature [13] could be due to study setting selection biases since persons with TB symptoms are more likely to visit HWS to treat the disease spiritually [18, 29, 30]. The prevalence of culture-positive PTB (21.8%) in the current study was also higher than that found in other high-risk settings in Ethiopia, including prisons, homeless shelters, and university students [31-38]. The difference may be due to the fact that these studies used sputum smear microscopy,

whereas we used conventional culture methods, which are more sensitive and specific for TB diagnosis. Another probable explanation is that everyone who attends HWSs is prone to TB and other infectious diseases due to their various health conditions. In addition, the availability of TB diagnosis and treatment services in prisons and universities may enable the early diagnosis and treatment of TB cases. Our study result was also higher than community-based studies in Southern Ethiopia's rural districts, which reported 3.0 to 6.3% [39-41]; Amhara region (3.8 to 4.9%) [11, 12]; Oromia region (7.6%) [42]; Tigray, Northern Ethiopia (8.6%) [43]; and central Ethiopia, Addis Ababa (13.3%) [44]. The difference may be due to differences in the study populations (subnational versus HWS attendees), TB diagnostic methods (sputum smear microscopy versus culture technique), the study period, and study setting selection biases.

The current study found that 18–33 years of age participants had the highest rate of culture-positive PTB. This may be because 47.0% of study participants were 18–33 years of age. Another possible explanation is that young people are more likely to regularly attend HWS and are hence likely to be exposed to TB infection. Most HWSs are in remote places, making it difficult for older adults to attend regularly. A similar study found that 68.6% of spiritual HWS attendees were 15–45 years of age [13]. The burden of TB among young individuals has major health consequences, as these individuals are economically active, and their travel for employment and high social interactions within the community exacerbate TB transmission in the general population [45].

This study found that rural residents are more likely to have culture-positive TB than urban residents. This was consistent with other studies' reports [46, 47]. According to global statistics, urban areas have a higher TB burden; however, TB is also prevalent among rural inhabitants in countries where a large portion of the population resides in rural areas and has a low income [48]. Due to inadequate public healthcare facilities, poor TB services, individuals' poor healthcare-seeking behavior, and a lack of knowledge and information about TB, early diagnosis and treatment are especially difficult in rural areas [40]. On the other hand, due to travel costs, fear of stigma, and other sociocultural and socioeconomic factors, people with TB in rural areas are unable to access healthcare facilities for early diagnosis and treatment [14, 15]. However, further research is required to investigate this interaction.

The majority of participants with culture-positive TB in the current study were married. This was consistent with a previous study [48]. In contrast to our findings, a few studies have revealed that unmarried people have a higher risk of TB infection than married people [46, 49, 50]. This might be because single people have a different lifestyle than married individuals. However, further study is necessary to better understand the factors that influence marital status as a predictor of active TB.

Household size (>5 family members) was statistically associated with culture-positive TB in the current study. This was consistence with a previous similar study [13] and institution-based studies in southeast Ethiopia [49, 51]. This may be because poverty, malnutrition, and overcrowded living conditions all increase the risk of TB transmission [5]. Since TB is mainly transmitted indoors, having a large family size per household results in overcrowding and increases the risk of TB transmissions. Another possible explanation is that people who live in rural areas and have many family members are more likely to be of lower socioeconomic status.

In our study, participants who shared living space at HWSs were tenfold more likely to have culture-positive PTB. This is because rooms at the HWSs are built as temporary waiting spaces, and they are very small, overcrowded, and poorly ventilated, which can intensify the risk of TB transmission [13, 18, 52]. The overcrowded and poorly ventilated waiting rooms at the HWS, prolonged stays at the HWS, and poor healthcare-seeking behaviors of individuals worsen the active transmission of TB at the HWS. Our findings were comparable to those of prior studies [32, 34, 35]. Congregate settings, such as HWSs, are places where people live together, and it is common for people to share living space [13, 18, 52]. Hence, poor living conditions and overcrowding in the shared room can increase TB transmission.

Drug-resistant TB (DR-TB) threatens national TB prevention and care efforts. In the current study, the prevalence of any DR-TB (resistance to RIF and/or INH) was 16.4%, indicating that it is a significant concern among HWS attendees. This is likely because many people with TB (infected with DR-TB strains) attended HWSs, resulting in DR-TB transmission [13, 18, 52]. This is comparable with previous reports from the study region (10.3 to 20.2%) [53–59], and other parts of Ethiopia (11.1 to 18.4%) [60–65]. However, our finding was lower than the finding reported in Ethiopia [66–69]. The discrepancy may be due to the study population (PTB-symptomatic HWS attendees versus TB patients who visited public healthcare facilities), and different drug susceptibility testing (DST) methods (phenotypic and/or PCRbased), which have varied sensitivity for diagnosing DR-TB strains.

Moreover, our study found 12.3% MDR-TB (resistance to both RIF and INH), suggesting that MDR-TB is also a major concern in these high-risk groups. Furthermore, 7.1% (3/42 retreated cases) of RR/MDR-TB isolates were identified in previously treated, while 15% (12/80) were isolated from newly diagnosed TB cases. Our result was higher than earlier studies in the study region (1.0 to 8.4%) [53, 54, 58, 59, 70, 71] and elsewhere in Ethiopia (1.2% to 8.3%) [64, 67-69, 72, 73]. It was also higher than 4.4% in the Ethiopian national survey [66], 0.71% in a recent national report on MDR-TB among new cases [1], and prison settings in Ethiopia (9.5%) [74]. However, it was comparable to previous studies from central Ethiopia, Addis Ababa [61, 75], Eastern Ethiopia [63], and a recent Ethiopian national report on MDR-TB among retreated TB patients (12.0%) [1]. The discrepancy may be due to differences in study populations, study settings, and DST methods used to diagnose MDR-TB. The high prevalence of DR-TB, particularly MDR-TB, among HWS attendees in the study region indicated that many TB patients, including those infected with DR-TB strains, attended the sites and that DR-TB strain transmission is common. However, molecular epidemiology studies are necessary to understand resistant TB strain transmission in HWS attendees and the community.

In this study, 4.1% of isolates were FLQs-resistant. Interestingly, all five FLQs-resistant and/or pre-XDR-TB isolates were identified at the South Wello zone. This suggests that these pre-XDR-TB strains may be circulating in this study site and have recently disseminated, although a molecular epidemiology analysis with strong discriminatory power would be required to confirm genotypic similarities, and recent transmission [18, 52]. Consistent with our result, an earlier study conducted in the same study region found that 5.7% of TB patients had pre-XDR-TB strains [7]. Similarly, a multicenter study revealed that the prevalence of pre-XDR-TB in Ethiopia was 5.0% [76]. However, both studies and ours had quite different study populations. Shibabaw and his colleagues included all RIF-resistant or MDR-TB patients who attended MDR-TB treatment centers before starting anti-TB therapy [7], while Dagne and his colleagues included new and retreated TB cases who attended TB treatment centers in different Ethiopian settings [76]. Thus, our study demonstrated that DR-TB, particularly MDR-TB, and pre-XDR-TB, among PTB-symptomatic HWS attendees in Ethiopia is a major issue that necessitates urgent prevention intervention measures and more studies on the same high-risk groups.

In the present study, participants between the ages of 18 and 33 years appeared to be at the highest risk of any DR-TB and MDR-TB infection, with rates of 47.1% and 58.3%, respectively. Although the comparison is difficult since different studies use different age cutoff points, an earlier study in the same study region found a strong link between anti-TB drug resistance and the age range of 25 to 34 years [58]. Similarly, a study conducted in South Africa revealed a strong link between DR-TB and the age groups 35 to 54 years and over 55 years [77]. Young age groups' increased exposure to the external environment, high-risk behavior, high workload, and broad range of mobility might contribute to TB and DR-TB infection. The high rate of DR-TB among younger folks has major health repercussions, as these individuals are economically active, and their travel for employment and high social contacts within the community exacerbates TB transmission in the general population [45].

In addition, in the current study, the South Wello zone study area appears to have the highest rate of any DR-TB and MDR-TB, with the odds of 14.5- and 12.0-fold higher than other study sites, respectively. Interestingly, five FLQsresistant and/or pre-XDR-TB strains were identified at the South Wello zone study site. This could be because DR-TB is prominent in this study area and there is recent TB transmission, although further molecular epidemiology analysis with strong discriminatory power would be required to confirm genotypic similarities and transmission patterns of the resistant strains [18, 52].

In the prospective follow-up study, a significant proportion of participants reported the development of active TB disease following exposure to HWS. Although our prospective follow-up method has limitations (unable to collect specimens for further laboratory confirmation), it suggests that HWS is a high-risk setting for TB transmission. Thus, these individuals can spread the disease to the community and their household members. We found that the attendee's educational status and sex, having had close contact with active TB patients in their family members, sharing living space at HWS, and recent hospitalization were potential risk factors for developing active TB disease postresidency at the HWS. TB transmission will be enhanced in environments where social mixing is more likely (along with overcrowding). Similarly, factors that prolong an infectious patient's exposure time will enhance TB transmission to other individuals [78].

4.1. Limitations. There were a few limitations to our study. First, there is a population selection bias and likely information bias when using self-reported data on risk factors. Second, the confirmatory tests were not done on those who reported active TB progression post-residency at the HWSs. Besides, in the follow-up study cohort, a comparison group of people who never attended or had no HWS exposure is crucial to determine the level of risk of HWS exposure to acquiring active TB post-residency at the HWS. Third, due to financial constraints, we were unable to perform BD BACTEC MGIT 960 for the initial MTB isolation process, and no phenotypic DST (pDST) was done on these isolates because liquid culture techniques, especially MGIT 960, have a significant advantage over solid culture techniques in terms of turnaround time and better recovery of MTB isolates. Last, further molecular epidemiology analysis would be warranted to confirm the transmission patterns of TB strains among HWS attendees.

5. Conclusions

Given that the prevalence of PTB among HWS attendees in this study population was seen to be higher than in the general population, hence, proactive preventive measures are recommended. The current study revealed that rural residents, being married, having >5 family members, and sharing a living space at HWSs were predictors of culturepositive PTB. The follow-up study also revealed that a higher proportion of attendees developed active TB disease postresidency at the HWS. Furthermore, the study showed a higher rate of DR-TB, especially MDR-TB, and pre-XDR-TB, among HWS attendees. Participants aged 18-33 years and the South Wello zone study site appeared to be more affected by DR-TB strains. Thus, regional and national TB prevention and control programs should recognize HWSs as high-risk settings for TB transmission and implement regular systematic TB screening, detection of DR-TB strains, and routine drug resistance surveillance among HWS attendees. Robust collaboration between the Ethiopian Orthodox Tewahedo Church and the regional and national TB control program is essential to develop locally appropriate, culturally accepted, and effective policy interventions to halt TB transmission among HWS attendees and the community.

Data Availability

The data sets analyzed during this study are available from the corresponding author upon reasonable request.

Disclosure

A poster presentation was made at the Union World Conference on Lung Health 2022, and the corresponding abstract has been published and is available at https:// theunion.org/sites/default/files/2022-11/Abstract_Book_ 2022-compressed.pdf [79], University of Pretoria, and Woldia University (only for scholarship and laboratory fees). The funders had no role in study design, data collection, and interpretation, or the decision to submit the work for publication.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

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Supplementary Materials

Table S1 illustrates the administrative zones that were included during the study, the spiritual holy water sites that were selected from each administrative zone, the total number of attendees who were screened for pulmonary TB (PTB) symptoms, the number of attendees who had PTBsuggestive symptoms, the number of bacteriologically confirmed cases, and the number of individuals who were Löwenstein-Jensen culture-negative test result. Table S2 shows the proportion of PTB positivity by gender and age group. The proportion estimation analysis revealed that the prevalence of culture-positive PTB was nearly equal for males and females (21.8% ± 2.35SE Vs 21.8% ± 2.6SE). Also, participants aged 18 to 33 years had a higher proportion of culture-positive PTB (28.5% ± 2.8SE). Table S3 summarizes the bivariate logistic regression analysis of sociodemographic characteristics of participants and associated risk factors for culture-positive PTB. The analysis revealed that participants aged 34-49 years and rural residents were statistically associated with culture-positive PTB (p < 0.05). Furthermore, the analysis revealed that few independent variables were statistically associated with culture-positive PTB ($p \le 0.01$), including a history of TB disease, contact with chronic coughers or active TB patients, having had close contact with a family member who had TB, the number of days spent (>21) at HWS, and sharing living spaces at HWS. In Table S4, we compute the proportion of any drugresistant TB and MDR-TB among each age group of participants in each study area. This allows us to assess the extent to which each age group appears to be affected by any drug-resistant TB and MDR-TB strains. Similarly, as illustrated in Table S5, we used a logistic regression analysis model to determine the odds of developing any drugresistant TB and MDR-TB among participants in each study site. Tables S6 and S7 provide detailed data about participants who had culture-negative test results while at HWS. Table S6 illustrates the profiles of participants and the proportion of developing active TB disease post-residency at the HWS among those who had culture-negative results

while at HWS. Besides, Table S7 shows the bivariate and multivariate logistic regression analysis of the sociodemographic characteristics of participants and associated factors among those who reported contracting active TB disease post-exposure to HWS. (*Supplementary Materials*)

References

- [1] Who, *Global Tuberculosis Report 2020*, World Health Organization, Geneva, Switzerland, 2020.
- [2] Who, *Global Tuberculosis Report 2021*, World Health Organization, Geneva, Switzerland, 2021.
- [3] G. J. Churchyard, L. D. Mametja, L. Mvusi et al., "Tuberculosis control in South Africa: successes, challenges and recommendations," *South African Medical Journal*, vol. 104, no. 3, pp. 244–248, 2014.
- [4] A. A. Mitku, T. Zewotir, D. North, P. Jeena, and R. N. Naidoo, "The differential effect of maternal dietary patterns on quantiles of Birthweight," *BMC Public Health*, vol. 20, no. 1, p. 976, 2020.
- [5] Ethiopia-Ministry of Health, Guidelines for Clinical and Programmatic Management of TB, Leprosy and TB/HIV in Ethiopia, Federal Ministry of Health, Addis Ababa, Ethiopia, 2012.
- [6] Who, *Global Tuberculosis Report 2019*, World Health Organization, Geneva, Switzerland, 2019.
- [7] A. Shibabaw, B. Gelaw, W. Gebreyes, R. Robinson, S. H. Wang, and B. Tessema, "The burden of pre-extensively and extensively drug-resistant tuberculosis among MDR-TB patients in the Amhara region, Ethiopia," *PLoS One*, vol. 15, no. 2, Article ID e0229040, 2020.
- [8] Ethiopia Ministry of Health, National Guidelines for Management of Tuberculosis, Drug Resistant-Tuberculosis and Leprosy in Ethiopia, Ethiopia Ministry of Health, Addis Ababa, Ethiopia, 6th edition, 2017.
- [9] M. Raviglione, Global Strategy and Targets for Tuberculosis Prevention, Care, and Control after 2015, World Health Organization, Geneva, Switzerland, 2013.
- [10] M. J. A. Reid, N. Arinaminpathy, A. Bloom et al., "Building a tuberculosis-free world: the Lancet Commission on tuberculosis," *The Lancet*, vol. 393, no. 10178, pp. 1331–1384, 2019.
- [11] T. Tadesse, M. Demissie, Y. Berhane, Y. Kebede, and M. Abebe, "Two-thirds of smear-positive tuberculosis cases in the community were undiagnosed in Northwest Ethiopia: population based cross-sectional study," *PLoS One*, vol. 6, no. 12, Article ID e28258, 2011.
- [12] S. Yimer, C. Holm-Hansen, T. Yimaldu, and G. Bjune, "Evaluating an active case-finding strategy to identify smearpositive tuberculosis in rural Ethiopia," *International Journal* of *Tuberculosis & Lung Disease*, vol. 13, no. 11, pp. 1399–1404, 2009.
- [13] D. Derseh, F. Moges, and B. Tessema, "Smear positive pulmonary tuberculosis and associated risk factors among tuberculosis suspects attending spiritual holy water sites in Northwest Ethiopia," *BMC Infectious Diseases*, vol. 17, no. 1, p. 100, 2017.
- [14] E. A. Dodor, "The feelings and experiences of patients with tuberculosis in the Sekondi-Takoradi Metropolitan district: implications for TB control efforts," *Ghana Medical Journal*, vol. 46, no. 4, pp. 211–218, 2012.
- [15] Core Group Tb Working Group, "Community-based tuberculosis prevention and care: why and how to get involved," An International Handbook for Nongovernmental

Organizations and Civil Society Organizations, CORE Group, Washington, DC, USA, 2013.

- [16] Z. Berhanu, "Holy water as an intervention for HIV/AIDS in Ethiopia," *Journal of HIV*, vol. 9, no. 3, pp. 240–260, 2010.
- [17] A. Kebede and Z. Shewangizaw, "Prevalence of use of holy water as a complementary treatment among PLWHA at debrebrihan referral hospital and health Centre, North East, Ethiopia: cross-sectional study," *International Journal of Medical Science Research and Practice*, vol. 2, no. 2, pp. 85–89, 2015.
- [18] M. A. Reta, H. M. Said, N. E. Maningi, G. Y. Wubetu, M. Agonafir, and P. B. Fourie, "Genetic diversity of *Myco-bacterium tuberculosis* strains isolated from spiritual holy water site attendees in Northwest Ethiopia. A cross-sectional study," *New Microbes and New Infections*, Article ID 101235, 2024.
- [19] L. Fekadu, C. Hanson, M. Osberg, J. Makayova, P. Mingkwan, and D. Chin, "Increasing access to tuberculosis services in Ethiopia: findings from a patient-pathway analysis," *The Journal of Infectious Diseases*, vol. 216, no. suppl_7, pp. S696–s701, 2017.
- [20] Ethiopian Population and Census, Summary and statistical report of the 2007 population and housing census, Federal Democratic Republic of Ethiopia Population Census Commission, Addis Ababa, Ethiopia, 2008.
- [21] Who consolidated guidelines on tuberculosis, Module 2: Screening – Systematic Screening for Tuberculosis Disease, World Health Organization, Geneva, Switzerland, 2021.
- [22] K. Weyer, I. Kantor, S. Kim et al., Laboratory Services in Tuberculosis Control. Part II: Microscopy, World Health Organization, Geneva, Switzerland, 1998.
- [23] G. Shen, C. Chen, C. Hung et al., "Combining the Capilia TB assay with smear morphology for the identification of *Mycobacterium tuberculosis* complex," *International Journal of Tuberculosis & Lung Disease*, vol. 13, no. 3, pp. 371–376, 2009.
- [24] N. E. Maningi, L. T. Daum, J. D. Rodriguez et al., "Multi-and extensively drug-resistant *Mycobacterium tuberculosis* in South Africa: a molecular analysis of historical isolates," *Journal of Clinical Microbiology*, vol. 56, no. 5, Article ID e01214, 2018.
- [25] Longhorn Vaccines & Diagnostics, "PrimeXtractTM, total nucleic acid isolation for molecular applications and NEXT-GENERATION sequencing," *Vertex*, vol. 5, 2017.
- [26] Hain-Lifescience, GenoType MTBDRplus VER 2.0, Molecular Genetic Assay for Identification of the M. tuberculosis Complex and its Resistance to Rifampicin and Isoniazid from Clinical Specimens and Cultivated Samples, Hain-Lifescience, Tübingen, Germany, 2015.
- [27] Hain Lifescience, GenoType MTBDRsl VER 2.0; Molecular Genetic Assay for Identification of the M. tuberculosis Complex and its Resistance to Fluoroquinolones and Aminoglycosides/ cyclic Peptides from Sputum Specimens or Cultivated Sample, Hain-Lifescience, Tübingen, Germany, 2017.
- [28] A. H. Kebede, Z. Alebachew, F. Tsegaye et al., "The first population-based national tuberculosis prevalence survey in Ethiopia, 2010-2011," *International Journal of Tuberculosis* and Lung Disease, vol. 18, no. 6, pp. 635–639, 2014.
- [29] M. M. Mesfin, J. N. Newell, J. D. Walley, A. Gessessew, and R. J. Madeley, "Delayed consultation among pulmonary tuberculosis patients: a cross-sectional study of 10 DOTS districts of Ethiopia," *BMC Public Health*, vol. 9, no. 1, p. 53, 2009.
- [30] M. Senbeto, S. Tadesse, T. Tadesse, and T. Melesse, "Appropriate health-seeking behavior and associated factors"

among people who had cough for at least two weeks in northwest Ethiopia: a population-based cross-sectional study," *BMC Public Health*, vol. 13, no. 1, p. 1222, 2013.

- [31] K. Adane, M. Spigt, S. Ferede, T. Asmelash, M. Abebe, and G. J. Dinant, "Half of pulmonary tuberculosis cases were left undiagnosed in prisons of the Tigray region of Ethiopia: implications for tuberculosis control," *PLoS One*, vol. 11, no. 2, Article ID e0149453, 2016.
- [32] D. S. Abebe, G. Bjune, G. Ameni, D. Biffa, and F. Abebe, "Prevalence of pulmonary tuberculosis and associated risk factors in Eastern Ethiopian prisons," *International Journal of Tuberculosis and Lung Disease*, vol. 15, no. 5, pp. 668–673, 2011.
- [33] S. Ali, A. Haileamlak, A. Wieser et al., "Prevalence of pulmonary tuberculosis among prison inmates in Ethiopia, a cross-sectional study," *PLoS One*, vol. 10, no. 12, Article ID e0144040, 2015.
- [34] T. G. Fuge and S. Y. Ayanto, "Prevalence of smear-positive pulmonary tuberculosis and associated risk factors among prisoners in Hadiya Zone prison, Southern Ethiopia," *BMC Research Notes*, vol. 9, no. 1, p. 201, 2016.
- [35] T. Gebrecherkos, B. Gelaw, and B. Tessema, "Smear-positive pulmonary tuberculosis and HIV co-infection in prison settings of North Gondar Zone, Northwest Ethiopia," *BMC Public Health*, vol. 16, no. 1, p. 1091, 2016.
- [36] B. Moges, B. Amare, F. Asfaw et al., "Prevalence of smearpositive pulmonary tuberculosis among prisoners in North Gondar Zone Prison, northwest Ethiopia," *BMC Infectious Diseases*, vol. 12, no. 1, p. 352, 2012.
- [37] T. Semunigus, B. Tessema, S. Eshetie, and F. Moges, "Smear positive pulmonary tuberculosis and associated factors among homeless individuals in Dessie and Debre Birhan towns, Northeast Ethiopia," *Annals of Clinical Microbiology and Antimicrobials*, vol. 15, no. 1, p. 50, 2016.
- [38] A. Mekonnen, J. M. Collins, A. Aseffa, G. Ameni, and B. Petros, "Prevalence of pulmonary tuberculosis among students in three eastern Ethiopian universities," *International Journal of Tuberculosis and Lung Disease*, vol. 22, no. 10, pp. 1210–1215, 2018.
- [39] D. G. Datiko, E. A. Guracha, E. Michael et al., "Sub-national prevalence survey of tuberculosis in rural communities of Ethiopia," *BMC Public Health*, vol. 19, no. 1, p. 295, 2019.
- [40] Y. Merid, Y. W. Mulate, M. Hailu et al., "Population-based screening for pulmonary tuberculosis utilizing community health workers in Ethiopia," *International Journal of Infectious Diseases*, vol. 89, pp. 122–127, 2019.
- [41] E. B. Shargie, M. A. Yassin, and B. Lindtjørn, "Prevalence of smear-positive pulmonary tuberculosis in a rural district of Ethiopia," *International Journal of Tuberculosis and Lung Disease*, vol. 10, no. 1, pp. 87–92, 2006.
- [42] S. Hamusse, M. Demissie, D. Teshome, M. S. Hassen, and B. Lindtjørn, "Prevalence and incidence of smear-positive pulmonary tuberculosis in the hetosa district of arsi zone, Oromia regional state of Central Ethiopia," *BMC Infectious Diseases*, vol. 17, no. 1, p. 214, 2017.
- [43] G. Berhe, F. Enqueselassie, E. Hailu et al., "Population-based prevalence survey of tuberculosis in the Tigray region of Ethiopia," *BMC Infectious Diseases*, vol. 13, no. 1, p. 448, 2013.
- [44] M. Demissie, B. Zenebere, Y. Berhane, and B. Lindtjorn, "A rapid survey to determine the prevalence of smear-positive tuberculosis in Addis Ababa," *International Journal of Tuberculosis and Lung Disease*, vol. 6, no. 7, pp. 580–584, 2002.
- [45] K. Middelkoop, L. G. Bekker, H. Liang et al., "Force of tuberculosis infection among adolescents in a high HIV and TB

prevalence community: a cross-sectional observation study," *BMC Infectious Diseases*, vol. 11, no. 1, p. 156, 2011.

- [46] T. Ephrem, B. Mengiste, F. Mesfin, and W. Godana, "Determinants of active pulmonary tuberculosis in ambo hospital, west Ethiopia," *African journal of primary health care and family medicine*, vol. 7, no. 1, pp. e1–e8, 2015.
- [47] Who, Stop TB Partnership: The Paradigm Shift 2016-2020. Genebra, World Health Organization, Geneva, Switzerland, 2015.
- [48] M. Beyanga, B. R. Kidenya, L. Gerwing-Adima, E. Ochodo, S. E. Mshana, and C. Kasang, "Investigation of household contacts of pulmonary tuberculosis patients increases case detection in Mwanza City, Tanzania," *BMC Infectious Diseases*, vol. 18, no. 1, p. 110, 2018.
- [49] B. Tulu, N. Dida, Y. Kassa, and B. Taye, "Smear positive pulmonary tuberculosis and its risk factors among tuberculosis suspect in South East Ethiopia; a hospital-based crosssectional study," *BMC Research Notes*, vol. 7, no. 1, p. 285, 2014.
- [50] F. Rodriguez and S. Agbo, "An assessment of the risk factors for pulmonary tuberculosis among adult patients suffering from human immunodeficiency virus attending the Wellness Clinic at Themba Hospital," *South African Family Practice*, vol. 57, no. 2, pp. 1–6, 2015.
- [51] A. Yohanes, S. Abera, and S. Ali, "Smear positive pulmonary tuberculosis among suspected patients attending metehara sugar factory hospital; eastern Ethiopia," *African Health Sciences*, vol. 12, no. 3, pp. 325–330, 2012.
- [52] M. A. Reta, N. E. Maningi, and P. B. Fourie, "Patterns and profiles of drug resistance-conferring mutations in *Mycobacterium tuberculosis* genotypes isolated from tuberculosissuspected attendees of spiritual holy water sites in Northwest Ethiopia," *Frontiers in Public Health*, vol. 12, Article ID 1356826, 2024.
- [53] F. Gashaw, B. Erko, Y. Mekonnen et al., "Phenotypic and genotypic drug sensitivity profiles of *Mycobacterium tuberculosis* infection and associated factors in northeastern Ethiopia," *BMC Infectious Diseases*, vol. 21, no. 1, p. 261, 2021.
- [54] A. Alelign, A. Zewude, T. Mohammed, S. Tolosa, G. Ameni, and B. Petros, "Molecular detection of *Mycobacterium tuberculosis* sensitivity to rifampicin and isoniazid in South Gondar Zone, northwest Ethiopia," *BMC Infectious Diseases*, vol. 19, no. 1, p. 343, 2019.
- [55] W. Mulu, B. Abera, M. Yimer, T. Hailu, H. Ayele, and D. Abate, "Rifampicin-resistance pattern of *Mycobacterium tuberculosis* and associated factors among presumptive tuberculosis patients referred to Debre Markos Referral Hospital, Ethiopia: a cross-sectional study," *BMC Research Notes*, vol. 10, no. 1, p. 8, 2017.
- [56] K. N. Jaleta, M. Gizachew, B. Gelaw, H. Tesfa, A. Getaneh, and B. Biadgo, "Rifampicin-resistant *Mycobacterium tuberculosis* among tuberculosis-presumptive cases at University of Gondar Hospital, northwest Ethiopia," *Infection and Drug Resistance*, vol. 10, pp. 185–192, 2017.
- [57] M. Maru, S. Haile Mariam, T. Airgecho, E. Gadissa, and A. Aseffa, "Prevalence of tuberculosis, drug susceptibility testing, and genotyping of mycobacterial isolates from pulmonary tuberculosis patients in dessie, Ethiopia," *Tuberc Res Treat*, vol. 10, 2015.
- [58] K. Adane, G. Ameni, S. Bekele, M. Abebe, and A. Aseffa, "Prevalence and drug resistance profile of *Mycobacterium tuberculosis* isolated from pulmonary tuberculosis patients attending two public hospitals in East Gojjam zone, northwest Ethiopia," *BMC Public Health*, vol. 15, no. 1, p. 572, 2015.

- [59] B. Tessema, J. Beer, F. Emmrich, U. Sack, and A. Rodloff, "First-and second-line anti-tuberculosis drug resistance in Northwest Ethiopia," *International Journal of Tuberculosis & Lung Disease*, vol. 16, no. 6, pp. 805–811, 2012.
- [60] B. Haile, K. Tafess, A. Zewude, B. Yenew, G. Siu, and G. Ameni, "Spoligotyping and drug sensitivity of *Mycobacterium tuberculosis* isolated from pulmonary tuberculosis patients in the Arsi Zone of southeastern Ethiopia," *New Microbes and New Infections*, vol. 33, Article ID 100620, 2020.
- [61] D. Damena, S. Tolosa, M. Hailemariam et al., "Genetic diversity and drug susceptibility profiles of *Mycobacterium tuberculosis* obtained from Saint Peter's TB Specialized Hospital, Ethiopia," *PLoS One*, vol. 14, no. 6, Article ID e0218545, 2019.
- [62] B. Wondale, G. Medhin, G. Abebe et al., "Phenotypic and genotypic drug sensitivity of *Mycobacterium tuberculosis* complex isolated from South Omo Zone, Southern Ethiopia," *Infection and Drug Resistance*, vol. 11, pp. 1581–1589, 2018.
- [63] M. Brhane, A. Kebede, and Y. Petros, "Molecular detection of multidrug-resistant tuberculosis among smear-positive pulmonary tuberculosis patients in Jigjiga town, Ethiopia," *Infection and Drug Resistance*, vol. 10, pp. 75–83, 2017.
- [64] T. A. Lobie, Y. Woldeamanuel, D. Asrat, D. Beyene, M. Bjørås, and A. Aseffa, "Genetic diversity and drug resistance pattern of *Mycobacterium tuberculosis* strains isolated from pulmonary tuberculosis patients in the Benishangul Gumuz region and its surroundings, Northwest Ethiopia," *PLoS One*, vol. 15, no. 4, Article ID e0231320, 2020.
- [65] G. Worku, B. Gumi, M. Girma et al., "Drug sensitivity of clinical isolates of *Mycobacterium tuberculosis* and its association with bacterial genotype in the Somali region, eastern Ethiopia," *Frontiers in Public Health*, vol. 10, p. 2705, 2022.
- [66] M. Getahun, G. Ameni, A. Kebede et al., "Molecular typing and drug sensitivity testing of *Mycobacterium tuberculosis* isolated by a community-based survey in Ethiopia," *BMC Public Health*, vol. 15, no. 1, p. 751, 2015.
- [67] M. Tilahun, G. Ameni, K. Desta et al., "Molecular epidemiology and drug sensitivity pattern of *Mycobacterium tuberculosis* strains isolated from pulmonary tuberculosis patients in and around Ambo Town, Central Ethiopia," *PLoS One*, vol. 13, no. 2, Article ID e0193083, 2018.
- [68] Z. Bedewi, Y. Mekonnen, A. Worku et al., "Mycobacterium tuberculosis in central Ethiopia: drug sensitivity patterns and association with genotype," New Microbes and New Infections, vol. 17, pp. 69–74, 2017.
- [69] S. D. Hamusse, D. Teshome, M. S. Hussen, M. Demissie, and B. Lindtjorn, "Primary and secondary anti-tuberculosis drug resistance in hitossa district of arsi zone, Oromia regional state, Central Ethiopia," *BMC Public Health*, vol. 16, no. 1, p. 593, 2016.
- [70] S. A. Yimer, M. Agonafir, Y. Derese, Y. Sani, G. A. Bjune, and C. Holm-Hansen, "Primary drug resistance to antituberculosis drugs in major towns of Amhara region, Ethiopia," *Apmis*, vol. 120, no. 6, pp. 503–509, 2012.
- [71] F. Mekonnen, B. Tessema, F. Moges, A. Gelaw, S. Eshetie, and G. Kumera, "Multidrug resistant tuberculosis: prevalence and risk factors in districts of metema and west armachiho, Northwest Ethiopia," *BMC Infectious Diseases*, vol. 15, no. 1, p. 461, 2015.
- [72] O. Zewdie, A. Mihret, T. Abebe et al., "Genotyping and molecular detection of multidrug-resistant *Mycobacterium tuberculosis* among tuberculosis lymphadenitis cases in Addis Ababa, Ethiopia," *New Microbes and New Infections*, vol. 21, pp. 36–41, 2018.

- [73] H. Bedru, M. Fikru, W. Niguse et al., "Drug resistance pattern of *M. tuberculosis* complex in Oromia region of Ethiopia," *Infection and Drug Resistance*, vol. 14, pp. 1679–1689, 2021.
- [74] S. Ali, P. Beckert, A. Haileamlak et al., "Drug resistance and population structure of *M.tuberculosis* isolates from prisons and communities in Ethiopia," *BMC Infectious Diseases*, vol. 16, no. 1, p. 687, 2016.
- [75] W. Sinshaw, A. Kebede, A. Bitew et al., "Prevalence of tuberculosis, multidrug-resistant tuberculosis and associated risk factors among smear-negative presumptive pulmonary tuberculosis patients in Addis Ababa, Ethiopia," *BMC Infectious Diseases*, vol. 19, no. 1, p. 641, 2019.
- [76] B. Dagne, K. Desta, R. Fekade et al., "The Epidemiology of first and second-line drug-resistance *Mycobacterium tuberculosis* complex common species: evidence from selected TB treatment initiating centers in Ethiopia," *PLoS One*, vol. 16, no. 1, Article ID e0245687, 2021.
- [77] E. Green, C. L. Obi, M. Nchabeleng et al., "Drug-susceptibility patterns of *Mycobacterium tuberculosis* in Mpumalanga province, South Africa: possible guiding design of retreatment regimen," *Journal of Health, Population and Nutrition*, vol. 28, no. 1, pp. 7–13, 2010.
- [78] P. Narasimhan, J. Wood, C. R. Macintyre, and D. Mathai, "Risk factors for tuberculosis," *Pulmonary Medicine*, vol. 2013, Article ID 828939, 11 pages, 2013.
- [79] The Union World Conference On Lung Health and Abstract Book, "The international union against tuberculosis and lung disease (the union)," 2022, https://theunion.org/sites/default/ files/2022-11/Abstract_Book_2022-compressed.pdf.