

Research Article

Investigating the Fish Assemblages of the Neosho River System

Ethan J. Rasset ¹, Hae H. Kim,² Ben C. Neely,³ and Quinton E. Phelps²

¹United States Fish and Wildlife Service, Arizona Fish and Wildlife Conservation Office–Parker, 60911 Highway 95 Parker, AZ 85344, USA

²Applied Fisheries Management Lab, Missouri State University, 901 S National Ave, Springfield, MO 65897, USA

³Kansas Department of Wildlife and Parks, 1830 Merchant St., Emporia, KS 66801, USA

Correspondence should be addressed to Ethan J. Rasset; ejrasset@gmail.com

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Barrier presence in river systems has been demonstrated to impair fish assemblages. Low head dams specifically are frequently occurring barriers in riverine environments. Well-supported impacts of these structures on fishes include diminished movement, reproduction, and habitat availability. Longitudinal patterns in riverine fish assemblages have long been researched to ascertain dynamics and display interactions. The need for research becomes more critical when factoring in impacts of barriers and detrimental invasive species. Knowledge of fish assemblages can inform fisheries biologists and aid in improved management practices for recreational and ecologically important species, as well as invasive species. The Neosho River system in Kansas has 14 barriers present. Little fisheries sampling has been done in the Kansas portion of this river system from the John Redmond Dam to the Oklahoma border; therefore, sampling was conducted to inform questions posed about the fish assemblages. We sought to document the fish assemblages of the system in Kansas and examine for assemblage composition distinctions by geographic region along a longitudinal gradient. The fish assemblage dataset from this research generated a wealth of knowledge on sportfish infiltration from reservoirs, imperiled fishes, and apparent impacts from low-head dams. Information from this study will aid in future management and direct new research investigating imperiled fishes.

1. Introduction

Freshwater ecosystems in North America have been modified by human activities for perceived economic and recreational benefits for hundreds of years [1–3]. Human alteration of freshwater environments has generated impacts such as pollution, habitat alteration, channelization, and introduced invasive or non-native species [2, 4–9]. One of the most common forms of anthropogenic modifications has been damming of lotic waters [2, 4, 6, 7, 9]. Dams can impair natural processes in freshwater ecosystems by creating hydrologic alterations, reducing discharge, altering sediment transport, modifying depth profiles, and homogenizing aquatic habitats [2, 3, 10, 11]. Limnological and ecological characteristics of lotic systems are altered following dam placement; more dam placement leads to more environmental alterations and can result in population fragmentation [11]. This is potentially harmful for lotic specialist

fishes which have evolved best suited for unaltered systems [2, 12–16]. Proliferation of impounded rivers and streams in the past century has led to imperilment of many affected fish species [2]. One of the more deleterious impacts of dams on native lotic specialists has been impeded movement, including spawning migrations, and has led to population fragmentation [2, 3, 5, 8, 17–20].

Dams are often constructed to create reservoirs for perceived benefits such as flood control, hydroelectric power, mine wastes retention, and recreation [2, 3, 21]. These reservoirs alter habitat and hydrology, and in turn, the fish assemblages above, below, and within created reservoirs [15, 16, 20, 22, 23]. Fisheries biologists manage reservoirs for recreational angling and stock piscivorous fishes to provide opportunities to target “sportfish” at artificially high abundances, using barriers as population controls [20]. However, these recreationally important piscivorous sportfish can move (e.g., one-way and sometimes two-way)

from reservoirs into lotic environments, subsequently disrupting native aquatic communities [20]. Negative impacts of dams on native riverine fishes are multifaceted; barriers impede movement and subsequent completion of life cycles when recreationally important reservoir sportfish bypass barriers, enter river ecosystems, and then compete with or consume native riverine species [2, 8, 10, 16, 20, 24].

Impacts from barriers can be further investigated by documenting fish assemblages along the longitudinal gradient of impaired aquatic systems [25–28]. As such, fish assemblages are often studied with a multigear approach to reduce sampling biases and obtain a representative sample [29–33]. Specifically, assemblages are often documented above and below barriers on longitudinal gradients and can allow assessment of impacts [2, 5, 16, 20, 34].

One aquatic system impaired by anthropogenic modification is the Neosho River system in Kansas. In 2023, there were 14 barriers (e.g., low head dams and earth-fill embankment dams) present that were constructed from 1870 to 1964 for flood control, hydroelectric, and municipal water supply purposes. Habitat and anthropogenic impacts vary longitudinally on this system because of the present barriers [2, 3]. Research on fish assemblages has taken place upstream of John Redmond Dam in the past few decades, but formal fish assemblage surveys are historically sparse in the stretch of river downstream of John Redmond Dam [35–38]. However, informal surveys have identified a broad fish assemblage that includes imperiled fishes (i.e., species in need of conservation (SINC) and threatened and endangered species (T&E)) and non-native species including Bighead Carp *Hypophthalmichthys nobilis* and Grass Carp *Ctenopharyngodon idella* [39–42]. The Neosho River system is also connected to reservoirs managed for recreational angling, including John Redmond Reservoir in Kansas and Grand Lake O’ the Cherokees in Oklahoma. These reservoirs contain recreationally important piscivorous fishes defined as “sportfish” in Kansas (Table 1; [43–47]). Meanwhile, the invasive carp present in this system have perceived low population densities with unknown impacts [48, 49]. As such, establishing a baseline fish assemblage prior to potential invasion would allow biologists to measure effects in the future if invasive carp populations increase [50–55]. Influences listed above create a need to systematically describe fish assemblages across the gradient of the Neosho River.

We employed a suite of gears and conducted fisheries sampling above or below dams (where feasible) on the Neosho River system to document the fish assemblage at each location [33, 56]. Sites were grouped geographically to assess potential assemblage distinctions along a longitudinal gradient [25–28, 57, 58]. Our objectives for this project included establishing a protocol for long-term monitoring of the fish assemblages in the Neosho River basin, quantifying Neosho River system fish assemblages (i.e., establish baseline demographics), documenting and assessing imperiled fishes, investigating potential reservoir sportfish escapement, and examining for longitudinal distinctions in fish assemblages by the geographic region. We anticipated fish assemblage to change with river distance and expected sportfish and

imperiled fishes abundance to relate to barriers and reservoirs. Results from this study (i.e., [59]) provide a previously undocumented assemblage dataset to fisheries biologists and insight on barrier impacts.

2. Methodology

2.1. Study Area. The Neosho River system is over 725 km long. We conducted fish assemblage sampling on approximately 375 km of the rivers from John Redmond Dam in Kansas to the Oklahoma border at 12 sites on the Neosho River system corresponding to access feasibility, landowner permissions, and barriers (Figure 1). We geographically divided the Neosho River into three regions (i.e., upper, middle, and lower) based on barrier concentration or isolation to assess possible fish assemblage distinction across a longitudinal gradient. Grand Lake O’ the Cherokees and John Redmond Reservoir are reservoirs in closest proximity to lower and upper sites, respectively. In addition, upper, middle, and lower regions were differentiated a priori due to suppositions about each region. We perceived upper sites as tailrace influenced, middle sites as relatively uninfluenced, and lower sites as being influenced by Grand Lake O’ the Cherokees.

2.2. Sampling. We sampled similarly to the long-term resource monitoring (LTRM) element used on the Upper Mississippi River system [33]. The number and placement of sets or runs varied at each site (Table 2). Set and run location remained fixed across seasons [33]. Location of set or run was modified corresponding to river condition with alternative placement locations established for adverse conditions (e.g., net moved to opposite riverbank to avoid deposited woody debris and net moved downstream 100 to 1,000 m to avoid dam turbulence) [33]. We also recorded coordinates at each gear and noted substrate type, macrohabitat type, the presence or absence of wing dams, large woody debris (i.e., snag), tributaries, and rip rap (i.e., boulders) [33]. Mini gill net and gill net sets were short-term (i.e., four hrs) when water temperature was above 16°C and overnight when water temperature was below 16°C. We differentiated sampling into seasons from June 2021 through November 2022 to assess possible seasonal variations. All observed fishes were netted regardless of species. Effort varied at some sites because of seasonal variation in river condition and expanse of the sampling area [33].

Effort of all passive gears ranged from one to four sets per site. We deployed AFS experimental gill nets either entirely parallel to shore with both ends of the net offshore, or with one end (small mesh portion) staked to shore, with the net stretched downstream. River conditions resulted in the determination of set type (e.g., net staked to shore in high current velocity) [33]. Similarly, we staked mini gill nets to shore and stretched either perpendicularly or downstream. These nets were 4.6 m in length and had one panel of 3.8 cm mesh. Hoop nets were set with the mouth (open end) facing downstream, were 1.1 m wide, were three m in length, and

TABLE 1: Continued.

| Common name | John Redmond Dam | Burlington | Neosho Falls | Iola | Humboldt | Chanute 2 | Chanute 3 | Parsons | Oswego | Chetopa | Baxter Springs | Riverton-Empire Dam |
|-----------------------|------------------|------------|--------------|------|----------|-----------|-----------|---------|--------|---------|----------------|---------------------|
| *Redear Sunfish | | | | | | X | | | | | X | X |
| *Rock Bass | | | | | | | | | | | X | X |
| Redspot Chub | | | | | | | | | | | X | |
| River Carpsucker | X | X | X | X | X | X | X | X | X | X | X | X |
| River Redhorse | | | | | | | | | X | X | X | X |
| *Striped x White Bass | | | | | | | | | X | X | X | X |
| Spotfin Shiner | | | | | | | | | X | X | | |
| *Sauger x Walleye | | | | | | | | | X | X | | |
| Slenderhead Darter | X | | | | | | | | X | X | | |
| Slim Minnow | | X | X | X | | X | X | X | X | X | X | X |
| Smallmouth Buffalo | X | X | X | X | X | X | X | X | X | X | X | X |
| *Smallmouth Bass | | | | | | | | | X | X | | |
| Suckermouth Minnow | | X | X | X | X | X | X | X | X | X | X | |
| Shortnose Gar | X | X | X | X | | X | X | X | X | X | X | X |
| Sand Shiner | | | | | | | | | | | X | X |
| Spotted Sucker | | | | | | | | | | X | X | X |
| *Spotted Bass | X | X | | X | | X | X | X | | X | X | X |
| Stonecat | | | X | | | | | | | | | |
| Spotted Gar | | | | | | X | | | X | X | X | X |
| Threadfin Shad | | | | | | | X | | | X | X | |
| Tadpole Madtom | | X | | | X | | | | X | X | | |
| *Warmouth | | | | | | | | | X | X | X | X |
| *White Bass | X | X | X | X | | X | X | X | X | X | X | X |
| *White Crappie | X | X | X | X | | X | X | X | X | X | X | X |

Kansas sportfish are denoted with an asterisk.

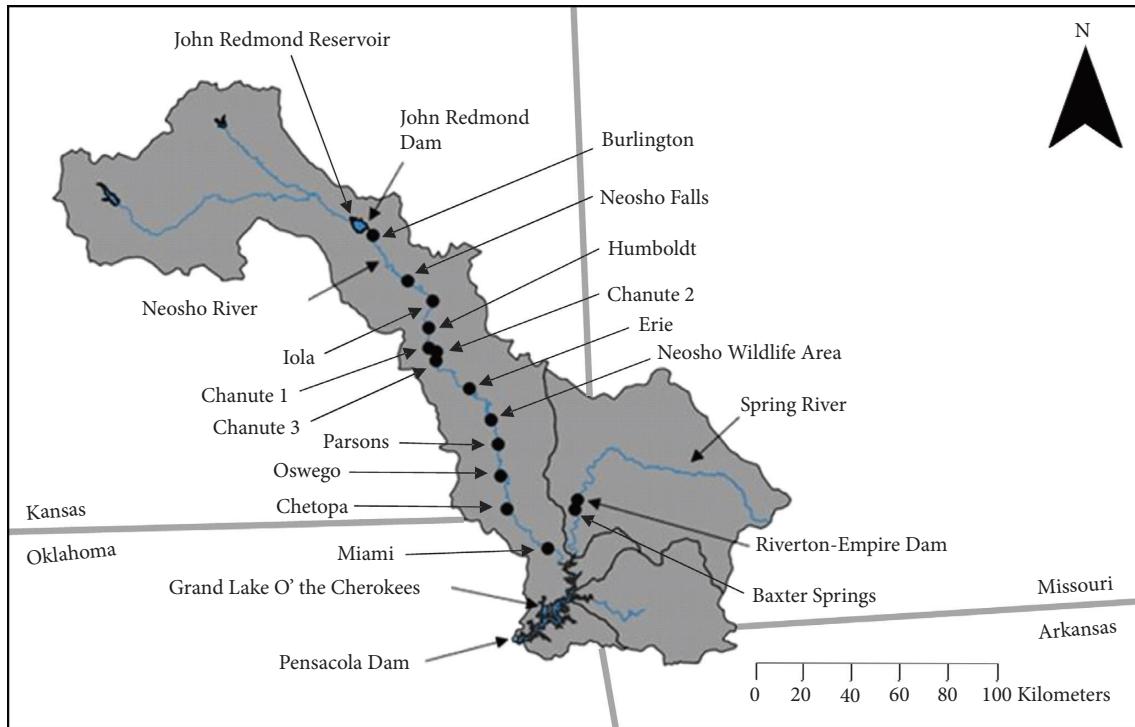


FIGURE 1: Map of the Neosho River system study area (adapted from [59]). Upper Neosho sites consist of John Redmond Dam, Burlington, Neosho Falls, and Iola. Middle Neosho sites include Humboldt, Chanute 2, and Chanute 3. Lower Neosho sites are Parsons, Oswego, and Chetopa.

had mesh that was 5.1 cm in width. We staked modified fyke nets to shore and extended nets perpendicularly to shore [33]. These nets had a lead that was 12.2 m in length and a frame that was 0.9 m by 1.5 m. Like that of modified fyke nets, we set mini fyke nets via stacking and subsequent stretching perpendicular to shore [33]. Mini fyke nets had mesh that was 0.3 cm in width, frames which were 0.6 m by 1.2 m, and a lead that was 6.1 m in length.

We used daytime pulsed-DC boat electrofishing via an ETS electrofishing control box (ETS Electrofishing Systems, LLC; Madison, WI); sampling effort consisted of 900 sec (15 min) “runs” [33]. All observed fishes were sought to be netted regardless of species. We set a goal of four or eight runs per site for standardization and achieved a power goal based on water temperature and conductivity [33]. Subsequently, sampling area availability and conditions contributed to electrofishing run variability [33]. While electrofishing, the boat was maneuvered according to LTRM, working downstream and into shorelines perpendicularly, and fishing low head dams as feasible [33]. Throughout sampling we sought to begin at a low head dam and work downstream or begin upstream and work down to a low head dam [33]. Runs of electrofishing either occurred by alternating bank position and maneuvering downstream (remaining on either river right or left for the entirety of one run before switching position) or were consecutively carried out while working downstream (sampling entirely on river right or left for half of the sampling effort before returning to the opposite bank of the initial starting point to additional shocking) [33].

2.3. Data Analyses. Catch per unit of effort (CPUE; the number of fish captured per unit of effort) was standardized using multigear mean standardization (MGMS) to assess fish assemblage structure and composition. We used MGMS to incorporate CPUE from all gears into one metric because of complexities associated with using CPUE from multiple gears for assemblage analyses [61, 62]. Multigear mean standardization was calculated by determining a total CPUE of all species at each site for a given gear [61, 62]. Subsequently, the mean of each total CPUE was calculated and used for determining MGMS [61, 62]. The MGMS value for a species was obtained by dividing the CPUE for that species at a given site by the mean total CPUE [61, 62]. Values of MGMS were averaged across gears to account for potential variation and are a unitless value that serves as a proportion [61, 62].

Nonmetric multidimensional scaling (nMDS) (via program PAST version 4.03) was used to quantify variation in fish assemblages among sites or regions [26–28, 57, 61, 63, 64]. Dimensions (i.e., species) and samples (i.e., sites and regions) were accounted for nMDS on a two-dimensional plane with distance-related indices. Distance between sites or regions and species was ranked and preserved within the scope of nMDS. While nMDS was used to visually display potential similarities or distinctions by site or region via corresponding drivers (i.e., species), classical clustering provided another means for visual delineation. As such, classical clustering expanded on nMDS by defining which sites were most similar outside of regional associations not necessarily addressed in nMDS by

TABLE 2: Sampling effort by gear type (i.e., total sets for net gears, hrs for electrofishing) from all sites on the Neosho River system from 2021 to 2022.

| Gear | John Redmond Dam | Burlington | Neosho Falls | Iola | Humboldt | Chanute 2 | Chanute 3 | Parsons | Oswego | Chetopa | Baxter Springs | Riverton-Empire Dam |
|---------------------------|------------------|------------|--------------|------|----------|-----------|-----------|---------|--------|---------|----------------|---------------------|
| Overnight gill sets | 5 | 3 | | | | 6 | 6 | 6 | 9 | 12 | 3 | 3 |
| Short-term gill sets | 9 | 5 | 5 | 7 | | 6 | | | | | 13 | 5 |
| Overnight mini gill sets | 2 | 4 | | | | 4 | 4 | 6 | 4 | 8 | 2 | 2 |
| Short-term mini gill sets | 6 | 2 | 2 | 6 | | 4 | 4 | 4 | 6 | 8 | 10 | 2 |
| Hoop sets | 11 | 6 | 2 | 5 | | 4 | 4 | 4 | 6 | 8 | 9 | 4 |
| Modified fyke sets | 9 | 5 | 2 | 5 | | 4 | 4 | 4 | 6 | 8 | 9 | 5 |
| Mini fyke sets | 18 | 14 | 8 | 11 | | 8 | 8 | 8 | 12 | 16 | 18 | 10 |
| Electrofishing hrs | 7.3 | 3.3 | 2.0 | 6.5 | 1.0 | 4.0 | 2.3 | 6.0 | 4.0 | 6.8 | 7.3 | 3.8 |

Short-term sets were four hrs; all other sets were overnight.

TABLE 3: Relative abundance (%) and frequency of occurrence (%) of 67 species observed at 12 sampling sites on the Neosho River system from 2021 to 2022.

| Common name | N | Sites present | Relative abundance (%) | Frequency of occurrence (%) |
|--------------------------|-------|---------------|------------------------|-----------------------------|
| Gizzard Shad | 7,183 | 12 | 25.9 | 100.0 |
| Red Shiner | 4,750 | 12 | 17.2 | 100.0 |
| Bullhead Minnow | 1,573 | 12 | 5.7 | 100.0 |
| Freshwater Drum | 1,499 | 12 | 5.4 | 100.0 |
| Mimic Shiner | 1,420 | 11 | 5.1 | 91.7 |
| Smallmouth Buffalo | 1,279 | 12 | 4.6 | 100.0 |
| Carmine Shiner | 1,121 | 6 | 4.0 | 50.0 |
| White Bass | 1,097 | 10 | 4.0 | 83.3 |
| Bluegill | 916 | 11 | 3.3 | 91.7 |
| White Crappie | 700 | 10 | 2.5 | 83.3 |
| Longnose Gar | 599 | 12 | 2.2 | 100.0 |
| Orangespotted Sunfish | 565 | 11 | 2.0 | 91.7 |
| Channel Catfish | 553 | 11 | 2.0 | 91.7 |
| River Carpsucker | 481 | 12 | 1.7 | 100.0 |
| Bluntnose Minnow | 456 | 11 | 1.6 | 91.7 |
| Common Carp | 429 | 12 | 1.5 | 100.0 |
| Black Buffalo | 418 | 12 | 1.5 | 100.0 |
| *Flathead Catfish | 415 | 12 | 1.5 | 100.0 |
| Shortnose Gar | 351 | 10 | 1.3 | 83.3 |
| Bigmouth Buffalo | 194 | 12 | 0.7 | 100.0 |
| Slim Minnow | 178 | 10 | 0.6 | 83.3 |
| Longear Sunfish | 150 | 10 | 0.5 | 83.3 |
| Pealip Redhorse | 144 | 6 | 0.5 | 50.0 |
| Brook Silverside | 113 | 8 | 0.4 | 66.7 |
| Spotted Bass | 108 | 9 | 0.4 | 75.0 |
| Emerald Shiner | 105 | 5 | 0.4 | 41.7 |
| Spotted Gar | 103 | 6 | 0.4 | 50.0 |
| *Blue Catfish | 91 | 8 | 0.3 | 66.7 |
| Fathead Minnow | 85 | 6 | 0.3 | 50.0 |
| Logperch | 75 | 8 | 0.3 | 66.7 |
| Spotfin Shiner | 68 | 2 | 0.2 | 16.7 |
| Largemouth Bass | 65 | 10 | 0.2 | 83.3 |
| Green Sunfish | 59 | 11 | 0.2 | 91.7 |
| Highfin Carpsucker | 51 | 3 | 0.2 | 25.0 |
| Western Mosquitofish | 45 | 6 | 0.2 | 50.0 |
| Spotted Sucker | 23 | 3 | 0.1 | 25.0 |
| Threadfin Shad | 23 | 3 | 0.1 | 25.0 |
| Blue Sucker | 18 | 5 | 0.1 | 41.7 |
| Black Redhorse | 18 | 1 | 0.1 | 8.3 |
| Redear Sunfish | 16 | 3 | 0.1 | 25.0 |
| Warmouth | 15 | 4 | 0.1 | 33.3 |
| Grass Carp | 14 | 5 | 0.1 | 41.7 |
| *Black Crappie | 12 | 3 | <0.1 | 25.0 |
| Suckermouth Minnow | 10 | 5 | <0.1 | 41.7 |
| Bigeye Shiner | 9 | 2 | <0.1 | 16.7 |
| Paddlefish | 8 | 3 | <0.1 | 25.0 |
| Banded Darter | 8 | 2 | <0.1 | 16.7 |
| Golden Redhorse | 8 | 2 | <0.1 | 16.7 |
| River Redhorse | 7 | 2 | <0.1 | 16.7 |
| Orangethroat Darter | 6 | 3 | <0.1 | 25.0 |
| Striped x White Bass | 6 | 2 | <0.1 | 16.7 |
| Quillback | 6 | 1 | <0.1 | 8.3 |
| Bluegill x Green Sunfish | 5 | 4 | <0.1 | 33.3 |
| *Smallmouth bass | 5 | 2 | <0.1 | 16.7 |
| Blackstripe Topminnow | 4 | 3 | <0.1 | 25.0 |
| Tadpole Madtom | 3 | 3 | <0.1 | 25.0 |
| *Rock Bass | 3 | 2 | <0.1 | 16.7 |
| Sauger x Walleye | 3 | 2 | <0.1 | 16.7 |

TABLE 3: Continued.

| Common name | N | Sites present | Relative abundance (%) | Frequency of occurrence (%) |
|---------------------------|--------|---------------|------------------------|-----------------------------|
| Sand Shiner | 3 | 2 | <0.1 | 16.7 |
| Redear Sunfish x Bluegill | 3 | 1 | <0.1 | 8.3 |
| Central Stoneroller | 2 | 2 | <0.1 | 16.7 |
| Black Bullhead | 1 | 1 | <0.1 | 8.3 |
| Golden Shiner | 1 | 1 | <0.1 | 8.3 |
| Northern Hog Sucker | 1 | 1 | <0.1 | 8.3 |
| Redspot Chub | 1 | 1 | <0.1 | 8.3 |
| Slenderhead Darter | 1 | 1 | <0.1 | 8.3 |
| Stonecat | 1 | 1 | <0.1 | 8.3 |
| Total | 27,683 | | | |

Kansas sportfish not historically native to the Neosho River system are denoted with an asterisk.

TABLE 4: Netting effort, catch, and catch per unit of effort (CPUE) from the Neosho River from 2021 to 2022.

| | Spring 2022 | Summer 2021-2022 | Fall 2021-2022 | Total |
|--------------------------------|-------------|------------------|----------------|--------------|
| <i>Netting effort</i> | | | | |
| Overnight gill sets | 6 | 2 | — | 8 |
| Short-term gill sets | — | 38 | 27 | 65 |
| Overnight mini gill sets | 4 | 2 | — | 6 |
| Short-term mini gill sets | — | 24 | 18 | 42 |
| Hoop sets | 4 | 28 | 18 | 50 |
| Modified fyke sets | 4 | 26 | 17 | 47 |
| Mini fyke sets | 8 | 53 | 42 | 103 |
| Total samples (sets) | 26 | 173 | 122 | 321 |
| <i>Netting catch</i> | | | | |
| Overnight gill | 148 | 83 | — | 231 |
| Short-term gill | — | 120 | 113 | 233 |
| Overnight mini gill | 10 | 18 | — | 28 |
| Short-term mini gill | — | 14 | 13 | 27 |
| Hoop | 8 | 44 | 22 | 74 |
| Modified fyke | 6 | 79 | 100 | 185 |
| Mini fyke | 228 | 4,122 | 6,334 | 10,684 |
| Total (N) | 400 | 4,480 | 6,582 | 11,462 |
| Total species | 21 | 39 | 38 | 44 |
| <i>Netting CPUE (fish/net)</i> | | | | |
| Overnight gill | 24.7 (5.2) | 41.5 (6.5) | — | 28.9 (4.8) |
| Short-term gill | — | 3.2 (0.6) | 4.2 (0.7) | 3.6 (0.4) |
| Overnight mini gill | 2.5 (1.0) | 9.0 (7.0) | — | 4.7 (2.4) |
| Short-term mini gill | — | 0.6 (0.2) | 0.7 (0.3) | 0.6 (0.2) |
| Hoop | 2.0 (2.0) | 1.6 (0.3) | 1.2 (0.6) | 1.5 (0.3) |
| Modified fyke | 1.5 (0.6) | 3.0 (1.0) | 5.9 (1.4) | 3.9 (0.8) |
| Mini fyke | 28.5 (9.4) | 77.8 (34.5) | 150.8 (74.9) | 103.7 (35.3) |
| Total | 15.4 (3.9) | 25.7 (10.8) | 54.0 (26.4) | 35.7 (11.6) |

Standard error is in parentheses.

organizing sites in a dendrogram. We performed classical clustering via a paired group (UPGMA) algorithm in program PAST (version 4.03) with MGMS values for this purpose [63]. We used the Bray-Curtis similarity index to compare sites and regions to species within both analyses and removed the Humboldt site because of a small sample size.

3. Results

We observed a total of 67 fish species from 13 families constituting 27,683 individual captures during all sampling on the Neosho River system (Table 1). Subsequently, species

found at all 12 sampling sites included Bullhead Minnow, *Pimephales vigilax*; Black Buffalo, *Ictiobus niger*; Bigmouth Buffalo, *Ictiobus cyprinellus*; Common Carp, *Cyprinus carpio*; Flathead Catfish, *Pylodictis olivaris*; Freshwater Drum, *Aplodinotus grunniens*; Gizzard Shad, *Dorosoma cepedianum*; Longnose Gar, *Lepisosteus osseus*; Red Shiner, *Cyprinella lutrensis*; River Carpsucker, *Carpiodes carpio*; and Smallmouth Buffalo, *Ictiobus bubalus*. The two most prevalent species we observed in terms of relative abundance (%) were Gizzard Shad (25.9) and Red Shiner (17.2) (Table 3).

Effort and catch were combined for both years of summer and fall sampling via netting and electrofishing. We

TABLE 5: Electrofishing effort, catch, and catch per unit of effort (CPUE) from the Neosho River from 2021 to 2022.

| | Spring 2022 | Summer 2021-2022 | Fall 2021-2022 | Total |
|------------------------------|---------------|------------------|----------------|--------------|
| <i>Electrofishing effort</i> | | | | |
| Samples (runs) | 30 | 85 | 57 | 172 |
| Electrofishing hrs | 7.5 | 21.3 | 14.3 | 43.0 |
| <i>Electrofishing catch</i> | | | | |
| Total (N) | 4,316 | 2,496 | 3,578 | 10,390 |
| Total species | 36 | 33 | 43 | 47 |
| CPUE (fish/hr) | 575.5 (387.4) | 117.5 (9.2) | 251.1 (68.4) | 241.6 (71.6) |

Standard error is in parentheses.

TABLE 6: Electrofishing effort, catch, and catch per unit of effort (CPUE) from upper, middle, and lower portions of the Neosho River from 2021 to 2022.

| | Upper | Middle | Lower |
|------------------------------|---------------|--------------|--------------|
| <i>Electrofishing effort</i> | | | |
| Samples (runs) | 76 | 29 | 67 |
| Electrofishing hrs | 19.0 | 7.3 | 16.8 |
| <i>Electrofishing catch</i> | | | |
| Total (N) | 6,572 | 1,024 | 2,794 |
| Total species | 36 | 34 | 41 |
| CPUE (fish/hr) | 345.9 (158.9) | 141.2 (30.3) | 166.8 (32.2) |

Standard error is in parentheses.

TABLE 7: Netting effort, catch, and catch per unit of effort (CPUE) from upper, middle, and lower portions of the Neosho River from 2021 to 2022.

| | Upper | Middle | Lower |
|--------------------------------|-------------|---------------|-------------|
| <i>Netting effort</i> | | | |
| Overnight gill sets | 8 | — | — |
| Short-term gill sets | 26 | 12 | 27 |
| Overnight mini gill sets | 6 | — | — |
| Short-term mini gill sets | 16 | 8 | 18 |
| Hoop sets | 24 | 8 | 18 |
| Modified fyke sets | 21 | 8 | 18 |
| Mini fyke sets | 51 | 16 | 36 |
| Total samples (sets) | 152 | 52 | 117 |
| <i>Netting catch</i> | | | |
| Overnight gill | 231 | — | — |
| Short-term gill | 122 | 56 | 55 |
| Overnight mini gill | 28 | — | — |
| Short-term mini gill | 15 | 8 | 4 |
| Hoop | 41 | 7 | 26 |
| Modified fyke | 82 | 58 | 45 |
| Mini fyke | 3,041 | 5,246 | 2,397 |
| Total (N) | 3,560 | 5,375 | 2,527 |
| Total species | 39 | 30 | 31 |
| <i>Netting CPUE (fish/net)</i> | | | |
| Overnight gill | 28.9 (4.8) | — | — |
| Short-term gill | 4.7 (0.9) | 4.7 (0.8) | 2.0 (0.3) |
| Overnight mini gill | 4.7 (2.4) | — | — |
| Short-term mini gill | 0.9 (0.4) | 1.0 (0.5) | 0.2 (0.2) |
| Hoop | 1.7 (0.5) | 0.9 (0.4) | 1.4 (0.5) |
| Modified fyke | 3.9 (1.3) | 7.3 (2.7) | 2.5 (0.6) |
| Mini fyke | 59.6 (23.3) | 327.9 (191.3) | 66.6 (39.8) |
| Total | 23.4 (8.1) | 103.4 (61.3) | 21.6 (12.4) |

Standard error is in parentheses.

TABLE 8: Netting effort, catch, and catch per unit of effort (CPUE) from the Spring River from 2021 to 2022.

| | Spring 2022 | Summer 2021-2022 | Fall 2021-2022 | Total |
|--------------------------------|-------------|------------------|----------------|-------------|
| <i>Netting effort</i> | | | | |
| Overnight gill sets | 6 | — | — | 6 |
| Short-term gill sets | — | 12 | 6 | 18 |
| Overnight mini gill sets | 4 | — | — | 4 |
| Short-term mini gill sets | — | 8 | 4 | 12 |
| Hoop sets | 4 | 5 | 4 | 13 |
| Modified fyke sets | 4 | 6 | 4 | 14 |
| Mini fyke sets | 8 | 12 | 8 | 28 |
| Total samples (sets) | 26 | 43 | 26 | 95 |
| <i>Netting catch</i> | | | | |
| Overnight gill | 73 | — | — | 73 |
| Short-term gill | — | 33 | 25 | 58 |
| Overnight mini gill | 3 | — | — | 3 |
| Short-term mini gill | — | 3 | 1 | 4 |
| Hoop | 9 | 5 | 6 | 20 |
| Modified fyke | 14 | 61 | 38 | 113 |
| Mini fyke | 154 | 818 | 71 | 1,043 |
| Total (N) | 253 | 920 | 141 | 1,314 |
| Total species | 29 | 34 | 25 | 42 |
| <i>Netting CPUE (fish/net)</i> | | | | |
| Overnight gill | 12.2 (4.6) | — | — | 12.2 (4.6) |
| Short-term gill | — | 2.8 (1.7) | 4.2 (1.6) | 3.2 (1.2) |
| Overnight mini gill | 0.8 (0.5) | — | — | 0.8 (0.5) |
| Short-term mini gill | — | 0.4 (0.2) | 0.3 (0.3) | 0.3 (0.1) |
| Hoop | 2.3 (1.1) | 1.0 (0.3) | 1.5 (0.7) | 1.5 (0.4) |
| Modified fyke | 3.5 (2.4) | 10.2 (3.8) | 9.5 (4.1) | 8.1 (2.1) |
| Mini fyke | 19.3 (13.2) | 68.2 (31.2) | 8.9 (2.6) | 37.3 (14.5) |
| Total | 9.7 (4.3) | 21.4 (9.6) | 5.4 (1.2) | 13.8 (4.5) |

Standard error is in parentheses.

TABLE 9: Electrofishing effort, catch, and catch per unit of effort (CPUE) from the Spring River from 2021 to 2022.

| | Spring 2022 | Summer 2021-2022 | Fall 2021-2022 | Total |
|------------------------------|--------------|------------------|----------------|--------------|
| <i>Electrofishing effort</i> | | | | |
| Samples (runs) | 8 | 23 | 13 | 44 |
| Electrofishing hrs | 2.0 | 5.8 | 3.3 | 11.0 |
| <i>Electrofishing catch</i> | | | | |
| Total (N) | 450 | 1,955 | 2,112 | 4,517 |
| Total species | 35 | 44 | 48 | 52 |
| CPUE (fish/hr) | 225.0 (21.8) | 340.0 (58.4) | 649.9 (99.1) | 410.6 (48.3) |

Standard error is in parentheses.

TABLE 10: Species in need of conservation (SINC) and threatened species (i.e., Redspot Chub) according to the Kansas Department of Wildlife and Parks in terms of number encountered during boat electrofishing and netting and their respective relative abundance (%).

| | Neosho River | | Spring River | | Total | |
|---------------------|--------------|------------------------|--------------|------------------------|-------|------------------------|
| | N | Relative abundance (%) | N | Relative abundance (%) | N | Relative abundance (%) |
| Banded Darter | 2 | <0.1 | 6 | 0.1 | 8 | <0.1 |
| Bigeye Shiner | 0 | — | 9 | 0.2 | 9 | <0.1 |
| Black Redhorse | 0 | — | 18 | 0.3 | 18 | 0.1 |
| Blue Sucker | 18 | 0.1 | 0 | — | 18 | 0.1 |
| Highfin Carpsucker | 7 | <0.1 | 44 | 0.8 | 51 | 0.2 |
| Northern Hog Sucker | 0 | — | 1 | <0.1 | 1 | <0.1 |
| Redspot Chub | 0 | — | 1 | <0.1 | 1 | <0.1 |
| River Redhorse | 0 | — | 7 | 0.1 | 7 | <0.1 |
| Spotfin Shiner | 0 | — | 68 | 1.2 | 68 | 0.2 |
| Spotted Sucker | 6 | <0.1 | 17 | 0.3 | 23 | 0.1 |
| Tadpole Madtom | 3 | <0.1 | 0 | — | 3 | <0.1 |

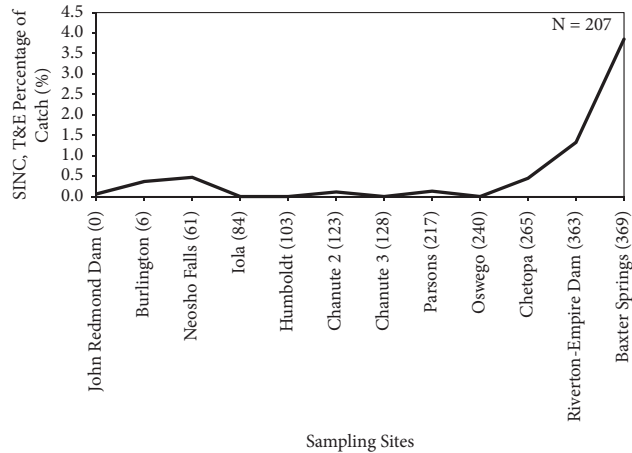


FIGURE 2: Species in need of conservation (SINC) and threatened and endangered (T&E) species percentage of catch from sampling sites on the Neosho River system. River distance in kilometers from John Redmond Dam is displayed in parentheses at each sampling location (i.e., barrier).

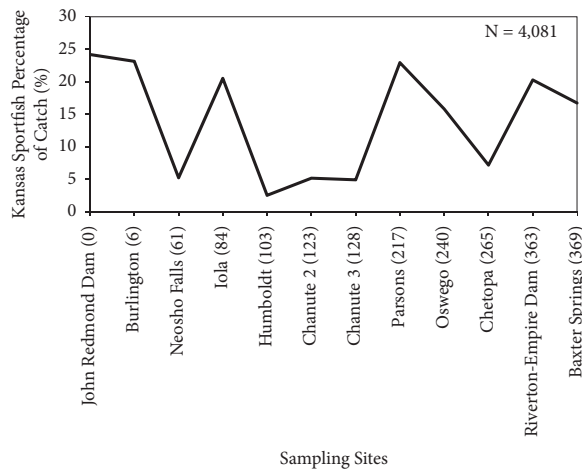


FIGURE 3: Kansas sportfish percentage of catch from sampling sites (i.e., dams) on the Neosho River system. River distance in kilometers from John Redmond Dam is displayed in parentheses at each sampling location. John Redmond Reservoir is directly upstream of John Redmond Dam. Grand Lake O' the Cherokees is 36 km downstream of Baxter Springs Dam and 61 km downstream of Chetopa Dam.

set 321 nets on the Neosho River and captured 11,462 fishes and 44 species (Table 4). Our overall effort and CPUE in terms of total fish/net (standard error (SE) in parentheses) included eight overnight gill nets (28.9 (4.8)), 65 short-term gill nets (3.6 (0.4)), six overnight mini gill nets (4.7 (2.4)), 42 short-term mini gill nets (0.6 (0.2)), 50 hoop nets (1.5 (0.3)), 47 modified fyke nets (3.9 (0.8)), and 103 mini fyke nets (103.7 (35.3)). Notably, mini fyke nets accounted for 93.2% of the total netting catch. Our most encountered fish species and the number of their captures by gear type included 121 Shortnose Gar *Lepisosteus platostomus* via gill netting, 26 Longnose Gar via mini gill netting, 21 Flathead Catfish from hoop nets, 43 White Crappie *Pomoxis annularis* from modified fyke nets, and 3,810 Red Shiners from mini fyke nets. We conducted 43.0 hrs (172 runs) of pulsed-DC boat electrofishing on the Neosho River and observed 10,390 fishes and 47 species (241.6 (71.6 SE) fish/hr) (Table 5). Electrofishing generated the greatest fish diversity

among gears used on the Neosho River; Gizzard Shad, Freshwater Drum, and Smallmouth Buffalo were the three most encountered species. Spring electrofishing resulted in the greatest electrofishing CPUE (575.5 (384.4 SE) fish/hr), while fall produced the greatest number of captures via netting (i.e., 6,582 fishes). In addition, Upper Neosho had the greatest CPUE from electrofishing (345.9 (158.9 SE) fish/hr), while Middle Neosho generated the most observations via netting (i.e., 5,375 fishes; Tables 6 and 7).

We deployed 95 nets on the Spring River and observed 1,314 fishes (42 species) (Table 8). Gears used across all seasons and the corresponding effort and CPUE (total fish/net; SE in parentheses) included six overnight gill nets (12.2 (4.6)), 18 short-term gill nets (3.2 (1.2)), four overnight mini gill nets (0.8 (0.5)), 12 short-term mini gill nets (0.3 (0.1)), 13 hoop nets (1.5 (0.4)), 14 modified fyke nets (8.1 (2.1)), and 28 mini fyke nets (37.3 (14.5)). Mini fyke netting comprised a large portion of the total catch (i.e., 79.4%) similar to

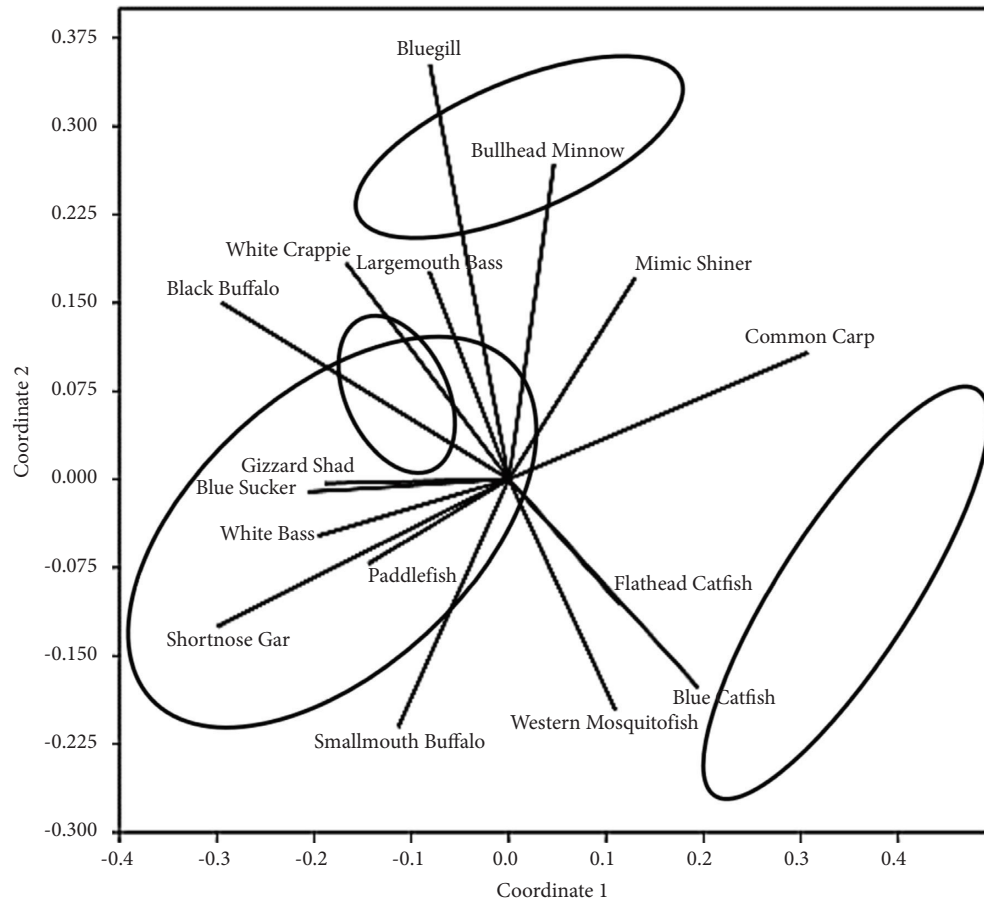


FIGURE 4: Nonmetric multidimensional scaling (nMDS) analysis with the Bray-Curtis similarity index with calculated multigear mean standardization (MGMS) values for 67 species of fishes from all gears at sampling sites and regions of the Neosho River system (stress = 0.239). Regions are represented by ellipses (i.e., upper Neosho = lower left, middle Neosho = upper middle, lower Neosho = lower right, and Spring river = middle left). Species with higher abundances in specific regions thus contributed to distinct spatial patterns.

results from the Neosho River. Longnose Gar (gill net, mini gill net, hoop net, and modified fyke net) and Bluegill *Lepomis macrochirus* (mini fyke net) were the most encountered species within select respective gears. We also conducted 11.0 hrs (44 runs) of pulsed-DC boat electrofishing on the Spring River and observed 4,517 fishes and 52 species (410.6 (48.3 SE) fish/hr; Table 9). Of these fishes, Gizzard Shad, Smallmouth Buffalo, and Channel Catfish *Ictalurus punctatus* were the most observed. Subsequently, summer sampling resulted in the highest netting catch (i.e., 920 fishes), whereas fall electrofishing produced the highest CPUE (649.9 (99.1 SE) fish/hr).

We encountered 207 fishes (11 species) listed as species in need of conservation (SINC) or threatened and endangered species (T&E) by the state of Kansas [65, 66]; Table 10. Of these, Spottfin Shiner *Cyprinella spiloptera* (0.2), Highfin Carpsucker *Carpododes velifer* (0.2), and Spotted Sucker *Minytrema melanops* (0.1) were the three most prevalent SINC or T&E fishes in terms of relative abundance (%). Species in need of conservation and T&E fishes constituted 0.7% of the total catch, ranging by sampling location from 0.0% (at both Iola and Humboldt) to 2.2% (at Chetopa and Baxter Springs, with SINC and T&E

catch combined and total catch combined for both sites) (Figure 2). We examined SINC and T&E catch at the lowermost dams on each river in Kansas; 159 of 207 SINC and T&E fishes (i.e., 76.8%) were observed in this area. Total observations rose to 187 (i.e., 90.3%) when including the two most downstream dams on each river.

We observed 20 species (including hybrids) and 4,081 total fishes categorized as Kansas sportfish in the Neosho River system. These recreationally important species constituted 14.7% of the total catch (Figure 3). In addition, sportfish percentage of catch at upper and lower sites was 24.2% at John Redmond Dam, 23.1% at Burlington, 5.2% at Neosho Falls, 20.5% at Iola, 18.9% between Oswego and Riverton-Empire Dam (sportfish catch combined and total catch combined for both sites), and 12.1% between Chetopa and Baxter Springs (sportfish catch combined and total catch combined for both sites). Overall, White Bass *Morone chrysops* were the most encountered sportfish (4.0% relative abundance).

We obtained patterns in assemblage of similarity and distinction via nMDS for sites and regions of the Neosho River system based on species' MGMS values (stress = 0.239). The stress value we obtained from nMDS modeling indicates that

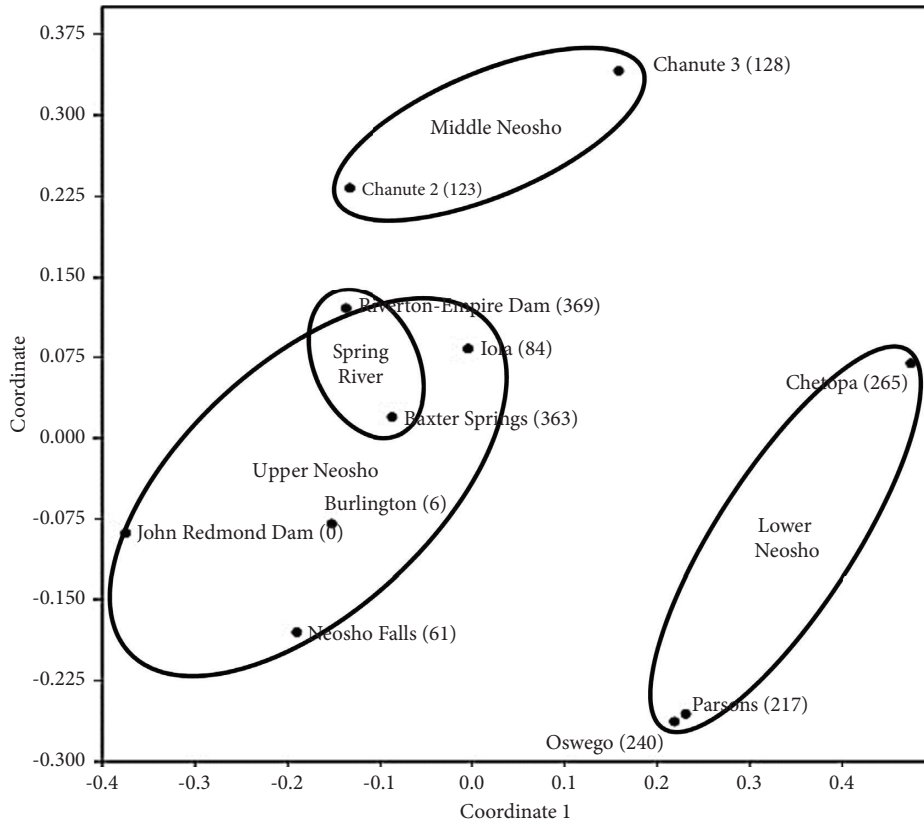


FIGURE 5: Nonmetric multidimensional scaling (nMDS) analysis with the Bray-Curtis similarity index with calculated multigear mean standardization (MGMS) values for 67 species of fishes from all gears at sampling sites and regions of the Neosho River system (stress = 0.239). River distance in kilometers from John Redmond Dam is displayed in parentheses at each sampling location (i.e., barrier).

the data could be a better fit within the model. However, stress values alone should not determine decipherability of nMDS models [67]. The large number of observations encompassed within the model lends support that a low stress value was unlikely to occur [67]. In addition, each region of the Neosho River system (including the Spring River) separated into unique, mostly isolated clusters via their corresponding sites (Figures 4 and 5). Upper, Middle, and Lower Neosho all were visually identified by nMDS as unique regions from fish assemblage compositions. Species' MGMS values and subsequent distance rankings displayed distinctive upstream and downstream patterns. The Upper Neosho had comparatively higher abundances of Blue Sucker, *Cycleptus elongatus*; Shortnose Gar; and Paddlefish, *Polyodon spathula*; the Middle Neosho had higher abundances of Bluegill; Mimic Shiner, *Notropis volucellus*; and Bullhead Minnow; the Lower Neosho River had greater abundances of Flathead Catfish; Blue Catfish, *Ictalurus furcatus*; Common Carp; and Western Mosquitofish, *Gambusia affinis*; and the Spring River had higher abundances of White Crappie and Largemouth Bass, *Micropterus salmoides*. Black Redhorse, *Moxostoma duquesnei*; Golden Redhorse, *Moxostoma erythrurum*; Bigeye Shiner, *Notropis boops*; Spotfin Shiner; and Highfin Carpsucker also were concentrated in the Spring River. In addition, the Spring River, Upper Neosho, Middle Neosho, and Lower Neosho grouped distinctly within the classical clustering analyses (Figure 6). The cophenetic correlation coefficient associated with this analysis (i.e., 0.85) affirms that the

dendrogram validly displays association patterns between sites and regions, expanding on site-by-site similarity or distinction [68]. Classical clustering displayed that the Lower Neosho was the most distinct from Middle and Upper Neosho and the Spring River fell within the Upper Neosho. Chetopa was the most distinct from Parsons and Oswego within Lower Neosho. John Redmond Dam was the most distinct from Burlington within the Upper Neosho. Parsons and Oswego, Chanute 2 and Chanute 3, and Riverton-Empire Dam and Baxter Springs were the most similar sites, respectively. Classical clustering also expanded on nMDS results by defining which sites were most similar outside of regional associations. For example, the fish assemblage at John Redmond was more like Chanute 3 than Lower Neosho sites such as Oswego and Parsons, which was not evident via nMDS.

4. Discussion

Longitudinal differences in the Neosho River system fish assemblages were observed in this study and are likely the result of a combination of natural changes in assemblage with river distance, barrier placement, corresponding hydrologic modifications (e.g., altered flow), and subsequent fragmented interconnectivity [2, 3, 10, 11, 69]. Consequently, modifications may disproportionately influence rare and imperiled fishes that evolved in fluvial unaltered systems [2, 8, 12, 13, 15, 22]. Human activities

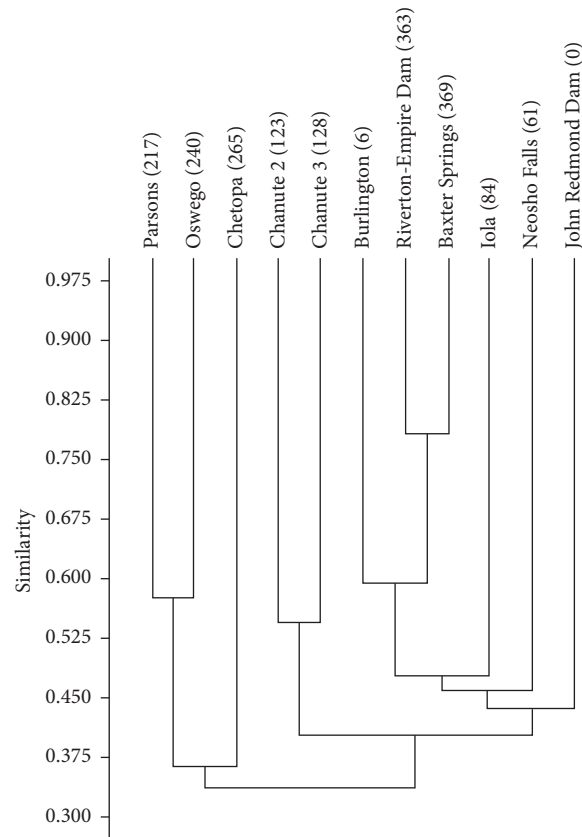


FIGURE 6: Classical clustering with a paired group (UPGMA) algorithm and the Bray-Curtis similarity index for sampling sites of the Neosho River system performed with calculated multigear mean standardization (MGMS) values for 67 species of fishes from all utilized gears (cophenetic correlation coefficient = 0.85). River distance in kilometers from John Redmond Dam is displayed in parentheses at each sampling location (i.e., barrier).

have homogenized river systems, altered hydrology, reduced habitat abundance, and heterogeneity and in general have caused serial discontinuity in lotic systems [2, 3, 10, 11]. Over 90% of SINC and T&E fishes found below the two most downstream dams on the Neosho River and Spring Rivers could be a product of dams causing serial discontinuity and impeding upstream movement.

Sportfish as defined by KDWP [70] were encountered more frequently at sites near large reservoirs (i.e., John Redmond Reservoir, Grand Lake O' the Cherokees). As such, these reservoirs are likely source populations for piscivorous sportfish populations within those associated river sites. Results from this study suggest that recreationally important piscivorous sportfish are entering the Neosho River system via upstream movement (i.e., Grand Lake O' the Cherokees) and downstream escapement (i.e., John Redmond Reservoir). Specifically, movement of fishes between Oklahoma into Kansas is likely occurring, suggesting interjurisdictional collaboration is necessary [71]. Management of recreationally important species with considerations for native riverine fishes should be prioritized, as sportfish angling in reservoirs and tailwaters provides recreational and economic benefits in North America [20, 72–74]. Previous studies have established that piscivorous sportfish can disrupt native

biological communities (e.g., competition and consumption) in lotic environments following reservoir escapement [20]. Our results suggest that sportfish are exhibiting passage from reservoirs into the Neosho River system and, as such, could be disrupting the native biota [20].

A comparatively greater number of SINC and T&E fishes observed at the Spring River suggest that the Spring River may be more suitable for these species than the Neosho River. We also observed greater species richness from the Spring River versus the Neosho River, perhaps suggesting the available habitat (e.g., presence of aquatic vegetation and low turbidity) may better support fish diversity. A higher number of species present at Upper and Lower Neosho versus Middle Neosho could be attributable in part to sportfish presence from reservoir escape as well as SINC and T&E fishes at the most downstream barriers.

Our results suggest that there are both similarities and distinctions in fish assemblages between sites and regions of the Neosho River system. The Spring River, Upper Neosho, Middle Neosho, and Lower Neosho supported unique fish assemblages, indicative of distinctions along a longitudinal gradient. The presence of recreationally important sportfish (e.g., Paddlefish, White Bass, White Crappie, and Ictalurids), SINC and T&E species, small-bodied minnow species, and species

found exclusive to a single site pattern dissimilarity between regions. Species responsible for assemblage differences are likely a product of reservoir escapement, subsequent integration, and barrier-induced passage inhibition. In addition, SINC and T&E species and recreationally important reservoir refugees (i.e., sportfish) occupy the same locations throughout the Neosho River system. As such, it is plausible that these reservoir refugees may not be a limiting factor for imperiled species ranges. Methods developed for fish assemblage sampling in this study can be applied to other rivers of interest or continued on this system to obtain a historical fish assemblage dataset. Future studies should investigate SINC and T&E fishes and their potential interactions with reservoir refugee piscivorous sportfish.

Data Availability

The data that support the findings of this study are available from the corresponding author, EJR, upon reasonable request.

Ethical Approval

Approval for research from the Missouri State University Institutional Animal Care and Use Committee was obtained on March 1st, 2021 (IACUC protocol 2020-14).

Disclosure

This manuscript was prepared from the master's thesis "Investigating the Fish Community of the Neosho River System."

Conflicts of Interest

Ben C. Neely is employed by the funding agency for the research associated with this manuscript and is the sole author with potential conflicts of interest.

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References

- [1] R. M. Hughes, J. N. Rinne, and B. Calamusso, "Historical changes in large river fish assemblages of the Americas," *American Fisheries Society Symposium*, vol. 45, pp. 603–612, 2005.
- [2] R. L. McLaughlin, E. R. B. Smyth, T. Castro-Santos et al., "Unintended consequences and trade-offs of fish passage," *Fish and Fisheries*, vol. 14, no. 4, pp. 580–604, 2012.
- [3] P. M. Vitousek, H. A. Mooney, J. Lubchenco, and J. M. Melillo, "Human domination of earth's ecosystems," *Science*, vol. 277, no. 5325, pp. 494–499, 1997.
- [4] D. J. Deegan, J. P. Foy, and L. A. Brabec, "The influence of dams on fish communities and associated habitat in the St. Joseph River watershed, Indiana," *Proceedings of the Indiana Academy of Science*, vol. 126, pp. 48–54, 2017.
- [5] H. L. Jelks, S. J. Walsh, N. M. Burkhead et al., "Conservation status of imperiled North American freshwater and diadromous fishes," *Fisheries*, vol. 33, no. 8, pp. 372–407, 2008.
- [6] O. Katano, T. Nakamura, S. Abe, S. Yamamoto, and Y. Baba, "Comparison of fish communities between above- and below-dam sections of small streams; barrier effect to diadromous fishes," *Journal of Fish Biology*, vol. 68, no. 3, pp. 767–782, 2006.
- [7] K. Kukula and A. Bylak, "Barrier removal and dynamics of intermittent stream habitat regulate persistence and structure of fish community," *Scientific Reports*, vol. 12, pp. 1512–1514, 2022.
- [8] A. Ricciardi and J. B. Rasmussen, "Extinction rates of North American freshwater fauna," *Conservation Biology*, vol. 13, no. 5, pp. 1220–1222, 1999.
- [9] K. Turgeon, C. Turpin, and I. Gregory-Eaves, "Dams have varying impacts on fish communities across latitudes: a quantitative synthesis," *Ecology Letters*, vol. 22, no. 9, pp. 1501–1516, 2019.
- [10] J. M. Holcomb, R. B. Nichols, and M. M. Gangloff, "Effects of small dam condition and drainage on stream fish community structure," *Ecology of Freshwater Fish*, vol. 25, no. 4, pp. 553–564, 2016.
- [11] J. V. Ward and J. A. Stanford, "The serial discontinuity concept of lotic ecosystems," *Dynamics of Lotic Ecosystems*, vol. 10, pp. 29–42, 1983.
- [12] J. D'Amours, S. Thibodeau, and R. Fortin, "Comparison of lake sturgeon (*Acipenser fulvescens*), Stizostedion spp., Catostomus spp., Moxostoma spp., quillback (*Carpiodes cyprinus*), and mooneye (*Hiodon tergisus*) larval drift in Des Prairies River, Quebec," *Canadian Journal of Zoology*, vol. 79, no. 8, pp. 1472–1489, 2001.
- [13] R. T. Muth and J. C. Schmulbach, "Downstream transport of fish larvae in a shallow prairie river," *Transactions of the American Fisheries Society*, vol. 113, no. 2, pp. 224–230, 1984.
- [14] A. T. Robinson, R. W. Clarkson, and R. E. Forrest, "Dispersal of larval fishes in a regulated river tributary," *Transactions of the American Fisheries Society*, vol. 127, no. 5, pp. 772–786, 1998.
- [15] V. H. Travnicek and M. J. Maceina, "Comparison of flow regulation effects on fish assemblages in shallow and deep water habitats in the Tallapoosa River, Alabama," *Journal of Freshwater Ecology*, vol. 9, no. 3, pp. 207–216, 1994.
- [16] J. M. Watson, S. M. Coghlan, J. Zydlewski, D. B. Hayes, and I. A. Kiraly, "Dam removal and fish passage improvement influence fish assemblages in the Penobscot River, Maine," *Transactions of the American Fisheries Society*, vol. 147, no. 3, pp. 525–540, 2018.

- [17] R. A. Abell, C. J. Loucks, P. Hedao et al., *Freshwater Ecoregions of North America: A Conservation Assessment*, Island Press, Washington, DC, USA, 1999.
- [18] Y. Abernethy and R. E. Turner, "US forested wetlands: 1940-1980," *BioScience*, vol. 37, no. 10, pp. 721-727, 1987.
- [19] G. S. Helfman, *Fish Conservation: A Guide to Understanding and Restoring Global Aquatic Biodiversity and Fishery Resources*, Island Press, Washington, DC, USA, 2007.
- [20] M. C. Lewis, W. R. Cope, T. P. Miles et al., "Reservoir fish escapement in North America: a historical review and future directions," *North American Journal of Fisheries Management*, vol. 43, no. 2, pp. 352-368, 2022.
- [21] R. M. Hughes, F. Amezcua, D. M. Chambers et al., "AFS position paper and policy on mining and fossil fuel extraction," *Fisheries*, vol. 41, no. 1, pp. 12-15, 2016.
- [22] C. R. Liermann, C. Nilsson, J. Robertson, and R. Y. Ng, "Implications of dam obstruction for global freshwater fish diversity," *BioScience*, vol. 62, no. 6, pp. 539-548, 2012.
- [23] G. N. Salvador, R. M. Hughes, F. Vieira, R. Ligeiro, and L. F. A. Montag, "Mine tailings storage dams modify upstream headwater fish assemblages," *Water in Biological Systems*, vol. 2, no. 2, Article ID 100136, 2023.
- [24] R. M. Hughes and A. T. Herlihy, "Patterns in catch per unit effort of native prey fish and alien piscivorous fish in 7 Pacific Northwest USA rivers," *Fisheries*, vol. 37, no. 5, pp. 201-211, 2012.
- [25] J. A. Garnett, D. P. Batzer, and C. Ramcharan, "Longitudinal variation in community structure of floodplain fishes along two rivers of the southeastern USA," *Canadian Journal of Fisheries and Aquatic Sciences*, vol. 71, no. 9, pp. 1291-1302, 2014.
- [26] M. A. McClelland, M. A. Pegg, and T. W. Spier, "Longitudinal patterns of the Illinois waterway fish community," *Journal of Freshwater Ecology*, vol. 21, no. 1, pp. 91-99, 2006.
- [27] C. P. Porter and T. M. Patton, "Patterns of fish diversity and community structure along the longitudinal gradient of the Kiamichi River in Southeastern Oklahoma," *Oklahoma Academy of Science*, vol. 95, pp. 104-118, 2015.
- [28] G. Seegert, J. Vondruska, E. Perry, and D. Dixon, "Longitudinal variation in the Ohio River fish community," *North American Journal of Fisheries Management*, vol. 33, no. 3, pp. 539-548, 2013.
- [29] M. H. Andersson, M. Gullström, M. E. Asplund, and M. C. Ohman, "Importance of using multiple sampling methodologies for estimating of fish community composition in offshore wind power construction areas of the Baltic Sea," *Ambio*, vol. 36, no. 8, pp. 634-636, 2007.
- [30] E. C. Boone, S. J. Meiners, L. D. Frankland, J. R. Laursen, and R. E. Colombo, "A comparison between fixed and random sampling of a low density spotted bass population in a large river," *Journal of Freshwater Ecology*, vol. 34, no. 1, pp. 533-540, 2019.
- [31] J. R. Fische, N. P. Johnson, R. D. Schultz, and M. C. Quist, "A comparison of modified fyke nets for evaluating fish assemblages and population structure," *Journal of Freshwater Ecology*, vol. 25, no. 4, pp. 555-563, 2010.
- [32] K. R. Holzgart, K. Deak, J. Miller et al., "Springs Coast long-term fish community assessment: the Rainbow River system," *Florida Scientist*, vol. 83, pp. 83-97, 2020.
- [33] E. N. Ratliff, E. J. Gittinger, T. M. O'Hara, and B. S. Ickes, "Long term resource monitoring program procedures: fish monitoring," *Program Report submitted to the US Army Corps of Engineers' Upper Mississippi River Restoration-Environmental Management Program*, 2014, <https://pubs.usgs.gov/mis/ltrmp2014-p001/>.
- [34] M. E. Herbert and F. P. Gelwick, "Spatial variation of headwater fish assemblages explained by hydrologic variability and upstream effects of impoundment," *Copeia*, vol. 2003, no. 2, pp. 273-284, 2003.
- [35] J. Falke and K. B. Gido, "Spatial effects of reservoirs on fish assemblages in Great Plains streams in Kansas; USA," *River Research and Applications*, vol. 22, no. 1, pp. 55-68, 2006.
- [36] D. P. Gillette, J. S. Tiemann, D. R. Edds, and M. L. Wildhaber, "Spatiotemporal patterns of fish assemblage structure in a river impounded by low-head dams," *Copeia*, vol. 2005, no. 3, pp. 539-549, 2005.
- [37] D. P. Gillette, J. S. Tiemann, D. R. Edds, and M. L. Wildhaber, "Habitat use by a midwestern U.S.A. riverine fish assemblage: effects of season, water temperature and river discharge," *Journal of Fish Biology*, vol. 68, no. 5, pp. 1494-1512, 2006.
- [38] J. S. Tiemann, D. P. Gillette, M. L. Wildhaber, and D. R. Edds, "Effects of lowhead dams on riffle-dwelling fishes and macroinvertebrates in a midwestern river," *Transactions of the American Fisheries Society*, vol. 133, no. 3, pp. 705-717, 2004.
- [39] J. Mounts and J. Tomelleri, *A Pocket Guide to Kansas Stream Fishes*, Great Plains Nature Center, Wichita, KS, USA, 2015, <https://gpnc.org/wp-content/uploads/sites/32/2017/12/SFPocketGuide.pdf>.
- [40] MSUBT (Michigan State University Board of Trustees), *Midwest Invasive Species Information Network (MISIN)*, Michigan State University, Department of Entomology-Applied Spatial Ecology and Technical Services Laboratory, East Lansing, MI, USA, 2010a, <https://nas.er.usgs.gov/queries/SpecimenViewer.aspx?SpecimenID=1553842>.
- [41] J. Pigg, J. Smith, and M. Ambler, "Additional records of bighead carp, *Hypophthalmichthys nobilis*, in Oklahoma waters," *Proceedings of the Oklahoma Academy of Science*, vol. 77, p. 123, 1997.
- [42] J. Pigg, J. Stahl, M. Ambler, and J. Smith, "Two potential sources of exotic fish in Oklahoma," *Proceedings of the Oklahoma Academy of Science*, vol. 73, p. 67, 1993.
- [43] P. Fuller and M. Neilson, *Ambloplites rupestris (Rafinesque, 1817)*, U.S. Geological Survey, Nonindigenous Aquatic Species Database, Gainesville, FL, USA, 2019.
- [44] P. Fuller, M. Cannister, and M. Neilson, *Micropterus dolomieu Lacepede, 1802*, U.S. Geological Survey, Nonindigenous Aquatic Species Database, Gainesville, FL, USA, 2019a.
- [45] P. Fuller, M. Cannister, and M. Neilson, *Pomoxis nigromaculatus (Lesueur in Cuvier and Valenciennes, 1829)*, U.S. Geological Survey, Nonindigenous Aquatic Species Database, Gainesville, FL, USA, 2019b.
- [46] P. Fuller and M. Neilson, *Ictalurus furcatus (Valenciennes in Cuvier and Valenciennes, 1840)*, U.S. Geological Survey, Nonindigenous Aquatic Species Database, Gainesville, FL, USA, 2021.
- [47] P. Fuller, M. Neilson, R. Sturtevant, and A. Bartos, *Pylodictis olivaris (Rafinesque, 1818)*, U.S. Geological Survey, Nonindigenous Aquatic Species Database, Gainesville, FL, USA, 2022.
- [48] ODWC (Oklahoma Department of Wildlife Conservation), *An Assessment of Impacts of Bighead Carp on Species of Greatest Conservation Need in the Neosho and Spring Rivers*, Oklahoma Department of Wildlife Conservation, State Wildlife Grant, Lincoln Blvd, OK, USA, 2018.
- [49] T. Patton and C. Tackett, "Status of silver carp (*Hypophthalmichthys molitrix*) and bighead carp (*Hypophthalmichthys nobilis*) in Southeastern Oklahoma," *Proceedings of the Oklahoma Academy of Science*, vol. 92, pp. 53-58, 2012.

- [50] A. R. Cupp, M. K. Brey, R. D. Calfee et al., “Emerging control strategies for integrated pest management of invasive carps,” *Journal of Vertebrate Biology*, vol. 70, no. 4, Article ID 21057, 2021.
- [51] K. S. Irons, G. G. Sass, M. A. McClelland, and J. D. Stafford, “Reduced condition factor of two native fish species coincident with invasion of non-native Asian carps in the Illinois River, U.S.A. Is this evidence for competition and reduced fitness?” *Journal of Fish Biology*, vol. 71, pp. 258–273, 2007.
- [52] N. L. Kinlock, A. J. Laybourn, C. E. Murphy, J. J. Hoover, and N. A. Friedenber, “Modelling bioenergetic and population-level impacts of invasive bigheaded carps (*Hypophthalmichthys* spp.) on native paddlefish (*Polyodon spathula*) in backwaters of the lower Mississippi River,” *Freshwater Biology*, vol. 65, no. 6, pp. 1086–1100, 2020.
- [53] C. S. Kolar, D. C. Chapman, W. R. Courtenay, C. M. Housel, J. D. Williams, and D. P. Jennings, *Asian Carps of the Genus Hypophthalmichthys (Pisces, Cyprinidae): A Biological Synopsis and Environmental Risk Assessment*, U.S. Fish and Wildlife Service, Washington, DC, USA, 2005.
- [54] Q. E. Phelps, S. J. Tripp, K. R. Bales, D. James, R. A. Hrabik, and D. P. Herzog, “Incorporating basic and applied approaches to evaluate the effects of invasive Asian carp on native fishes: A necessary first step for integrated pest management,” *PLoS One*, vol. 12, no. 9, Article ID 184081, 2017.
- [55] L. E. Solomon, R. M. Pendleton, J. H. Chick, and A. F. Casper, “Long-term changes in fish community structure in relation to the establishment of Asian carps in a large floodplain river,” *Biological Invasions*, vol. 18, no. 10, pp. 2883–2895, 2016.
- [56] C. Dunn and C. P. Paukert, “A flexible survey design for monitoring spatiotemporal fish richness in nonwadeable rivers: optimizing efficiency by integrating gears,” *Canadian Journal of Fisheries and Aquatic Sciences*, vol. 77, no. 6, pp. 978–990, 2020.
- [57] A. Foubert, F. Lecomte, P. Legendre, and M. Cusson, “Spatial organisation of fish communities in the St. Lawrence River: a test for longitudinal gradients and spatial heterogeneities in a large river system,” *Hydrobiologia*, vol. 809, no. 1, pp. 155–173, 2018.
- [58] L. E. Miranda and K. J. Killgore, “Longitudinal distribution of uncommon fishes in a species-rich basin,” *Aquatic Conservation*, vol. 30, no. 3, pp. 577–585, 2020.
- [59] E. J. Rasset, “Investigating the fish community of the Neosho River system,” M.Sc. thesis, Missouri State University, Springfield, MI, USA, 2023.
- [60] G. W. Whitledge and J. D. Schooley, *Using Dentary Bone Microchemistry to Identify Natal River and Evaluate Natal Side Fidelity for Paddlefish Harvested from the Grand Lake Stock*, Oklahoma Department of Wildlife Conservation, Sport Fish Restoration Grant, Lincoln Blvd, OK, USA, 2020.
- [61] D. K. Gibson-Reinemer, B. S. Ickes, and J. H. Chick, “Development and assessment of a new method for combining catch per unit effort data from different fish sampling gears: multigear mean standardization (MGMS),” *Canadian Journal of Fisheries and Aquatic Sciences*, vol. 74, no. 1, pp. 8–14, 2017.
- [62] R. N. Hupfeld, G. Jones, K. Hansen, H. H. Kim, and Q. E. Phelps, “An assessment of gears for sampling shovelnose sturgeon in tributaries,” *Journal of Applied Ichthyology*, vol. 38, no. 2, pp. 165–169, 2022.
- [63] Ø. Hammer, D. A. T. Harper, and P. D. Ryan, “PAST: paleontological statistics software package for education and data analysis,” *Palaeontologia Electronica*, vol. 4, pp. 1–9, 2001.
- [64] A. L. Whitten and D. Gibson-Reinemer, “Tracking the trajectory of change in large river fish communities over 50 y,” *The American Midland Naturalist*, vol. 180, no. 1, pp. 98–107, 2018.
- [65] KDWP (Kansas Department of Wildlife and Parks), *Kansas Species in Need of Conservation (SINC)*, Kansas Department of Wildlife and Parks, Topeka, KS, USA, 2019a, <https://ksoutdoors.com/Services/Research-Publications/T-E-and-SINC-Species>.
- [66] KDWP (Kansas Department of Wildlife and Parks), *Kansas Threatened and Endangered Species (T&E)*, Kansas Department of Wildlife and Parks, Topeka, KS, USA, 2019b, <https://ksoutdoors.com/Services/Research-Publications/T-E-and-SINC-Species>.
- [67] E. Dexter, G. Rollwagen-Bollens, and S. M. Bollens, “The trouble with stress: a flexible method for the evaluation of nonmetric multidimensional scaling,” *Limnology and Oceanography: Methods*, vol. 16, no. 7, pp. 434–443, 2018.
- [68] D. D. Nanayakkara and D. T. Blumstein, “Defining yellow-bellied marmot social groups using association indices,” *Oecologia Montana*, vol. 12, pp. 7–11, 2003.
- [69] R. L. Vannote, G. W. Minshall, K. W. Cummins, J. R. Sedell, and C. E. Cushing, “The river continuum concept,” *Canadian Journal of Fisheries and Aquatic Sciences*, vol. 37, no. 1, pp. 130–137, 1980.
- [70] KDWP (Kansas Department of Wildlife and Parks), *2023 Kansas Fishing Regulations Summary*, Kansas Department of Wildlife and Parks, Topeka, KS, USA, 2023, <https://ksoutdoors.com/Services/Publications/Fishing/2023-Regulations-Summary-Fishing-ENG>.
- [71] S. J. Tripp, Q. E. Phelps, R. N. Hupfeld et al., “Sturgeon and paddlefish migration: evidence to support the need for interjurisdictional management,” *Fisheries*, vol. 44, no. 4, pp. 183–193, 2019.
- [72] K. R. Criddle, M. Herrmann, S. T. Lee, and C. Hamel, “Participation decisions, angler welfare, and the regional economic impact of sportfishing,” *Marine Resource Economics*, vol. 18, no. 4, pp. 291–312, 2003.
- [73] R. M. Hughes, “Recreational fisheries in the USA: economics, management strategies, and ecological threats,” *Fisheries Science*, vol. 81, pp. 1–9, 2015.
- [74] B. L. Tufts, J. Holden, and M. DeMille, “Benefits arising from sustainable use of North America’s fishery resources: economic and conservation impacts of recreational angling,” *International Journal of Environmental Studies*, vol. 72, no. 5, pp. 850–868, 2015.