

Review Article

Moringa oleifera: Recent Insights for Its Biochemical and Medicinal Applications

Aaser M. Abdelazim ¹, Mohamed Afifi ^{2,3}, Mohammed H. Abu-Alghayth ⁴,
and Dima H. Alkadri ⁵

¹Department of Basic Medical Sciences, College of Applied Medical Sciences, University of Bisha, Bisha 67714, P.O. Box 255, Saudi Arabia

²Department of Biochemistry, College of Science, University of Jeddah, Jeddah, Saudi Arabia

³Department of Biochemistry, Faculty of Veterinary Medicine, Zagazig University, Zagazig, Egypt

⁴Department of Medical Laboratory Sciences, College of Applied Medical Sciences, University of Bisha, Bisha 67714, P.O. Box 255, Saudi Arabia

⁵Department of Nutrition and Food Science, Faculty of Agriculture, Jerash University, Jerash, Jordan

Correspondence should be addressed to Mohamed Afifi; mafifi@uj.edu.sa

Received 19 October 2023; Revised 25 December 2023; Accepted 28 December 2023; Published 5 January 2024

Academic Editor: Sanju Bala Dhull

Copyright © 2024 Aaser M. Abdelazim et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Plants could be used for multiple medicinal purposes. *Moringa oleifera* (MO) is considered the most famous plant used for this purpose. The present review aimed to spot the light on the recent medicinal, biochemical, and nutritional applications of MO. The plant contains a huge number of nutrients such as fatty acids, amino acids, proteins, polysaccharides, minerals, and vitamins. It has been used to control glucose, lipids, proteins, minerals, vitamins, hormones, and antioxidants in many animals and human models. Its medicinal applications are also varied and wide; it could be used to control and manage lots of disorders. Extracts and isolated ingredients from the plant open the way for many researchers all over the world to study its biochemical and medicinal impact on many experimental and clinical models. The present review spots the light on the recent medicinal and biochemical significance of MO opening the discussion and demonstrating the strengths and weaknesses in the MO research area. Regardless of the contrary concept, we considered MO a promising plant that could be studied for its medicinal applications on both biochemical and molecular levels. We recommended further research on the molecular effects of MO in healthy and diseased models.

1. Introduction

1.1. History of *Moringa oleifera*. Medicinal plants and their extracts are widely used for multiple medicinal purposes all over the world [1–3]. Due to their huge extracts and bioactive ingredients, they could be utilized in the management, control, and treatment of many disorders [4–8]. A medicinal plant could be defined as any plant that contains bioactive ingredients used in therapy or contains a precursor for drug manufacture [9]. *Moringa oleifera* Lam. (MO) is a medicinal plant belonging to the *Moringaceae* family (Table 1). Its bioactive material is utilized in the treatment of a huge number

of disorders with magical and amazing results to the extent that it is called a miracle tree [10]. Its bioactive components have antipyretic, antioxidant, anti-inflammatory, antiaging, antidiabetic, antihypertensive, immunomodulatory, hepatoprotective, and diuretic as shown in Table 2. Furthermore, recently, the plant has been approved to have antiparasitic activity [67], neuroprotective and cerebroprotective [68, 69], and anti-proliferative activity against cancers [70]. On the other hand, the plant has a potential use in regenerative dentistry [71] and has the ability to control and attenuate osteoporosis [72]. Nowadays, its anticonstipation [73], anti-inflammatory [74], antioxidant [75], and neuroplasticity activities [76] have been

confirmed. The plant is natively discovered in Indian forests and was consumed as a food additive [77]. The medicinal applications of MO were recorded in ancient medical books in many countries. In China, the first record of *Moringa* back to the bower script (volume II), about the 4th–6th century A.D. Thousands of years old, the medicinal purposes of *Moringa* plant parts have been recorded in the Ayurvedic Pharmacopoeia of India (API) [78]. The ancient Egyptians used *Moringa* oil as a sunscreen. Ancient Greeks discovered many medicinal applications for *Moringa*, and they introduced it to Romans where it spread everywhere in Europe while, in modern history in 1817, the Jamaican introduced *Moringa* oil for many food purposes till it reached to British Empire from where *Moringa* has been expanded all over the world [79]. Although the plant has great medicinal purposes, there is a need for further studies to elaborate and explore more details about its isolated compounds of synergistic action [80]. We and authors recommended further studies and advised us to continue research in this field.

1.2. Taxonomy, Synonyms, and Distribution. *Moringaceae* family is globally distributed especially in tropical areas. The plant is distributed in subtropical and tropical areas and includes about 13 species [81]. Globally, MO is widely distributed over Africa, Asia, Central America, and the Caribbean islands [82, 83]. MO in English is called the horseradish tree and drumstick tree; in Hindi, it is called Saijan; in Sanskrit, it is called Shigru [84]. The taxonomical classification and plant tree synonyms are illustrated in Tables 1 and 3 [85].

2. Bioactive Components and Their Medicinal Importance of *Moringa oleifera*

The tree of MO has many medicinal importance. It contains a huge number of bioactive compounds that could be used as it is or included in the formulation of drugs [86]. Organic agents such as protein, lipids, minerals, vitamins, tannin, flavonoids, saponins, phenolic acids, isothiocyanate, and others are active components isolated from MO illustrated in Figure 1. The pharmacological and medicinal aspects of these compounds are listed in Table 2. The cultivation processes and storage of plant parts have a great influence on the contents of these bioactive compounds which may affect its application and use [87].

3. Nutritional and Biochemical Significance of *Moringa oleifera*

3.1. Nutritional Significance of Moringa oleifera. A wide range of nutritinal components have been islated from MO seeds, roots, stems, leaves, flowers and pods as; proteins, carbohydrates, fats, glycosids and phenolic compounds [88]. These components are illustrated in Table 4. The plant parts contain a high amount of sulphur-containing amino acids, and they contain higher amounts of β -carotenes than carrots, higher amounts of L-ascorbate than oranges, higher levels of calcium than milk, higher potassium levels than that

TABLE 1: Taxonomy of *Moringa oleifera*.

Plant kingdom	Plantae
Plant subkingdom	Tracheobionta
Plant super division	Spermatophyta
Plant division	Magnoliophyta
Plant class	Magnoliopsida
Plant subclass	Dilleniidae
Plant order	Capparales
Plant family	Moringaceae
Plant genus	<i>Moringa</i>
Plant species	<i>Oleifera</i>

in bananas, nine times more iron than that in spinach, and four times more fiber than that in oats [110]. Other functional chemical groups have been extracted from leaves such as aldehydes, acids, amides, alcohols, phenols, vitamins, and phytosterols [111]. Due to their great nutritional properties, plant parts have been included in the formulation of both animal and human diets. Laboratory studies on the blood of models fed on MO investigated an increase in serum levels of calcium, proteins, phosphorus, and antioxidants but a decrease in glucose, triglycerides, and cholesterol levels [112].

3.1.1. Moringa oleifera as a Source for Animal Diets. Dietary preparations from MO were included in many animal diets such as chicken [113], fishes [114, 115], sheep [116], cows [117], and rabbits [118]. For chicken, *Moringa* diets formulated from leaves and stems improved the growth performance and carcass trait [119], and productivity and the quality of egg production [120]. MO extracts added to chicken diets lead to support and potentiate the immune response and introduce protection against infectious diseases [121]. MO diets are introduced to cows to improve colostrum quality, immunity, and milk production [117], protect the mammary epithelium from oxidants [122], and increase milk yield, rumen fermentation, and digestibility [123]. Diets supplemented with MO for pigs improved the reproduction performances, increased the protein contents of colostrum, and improved the antioxidant activities [124].

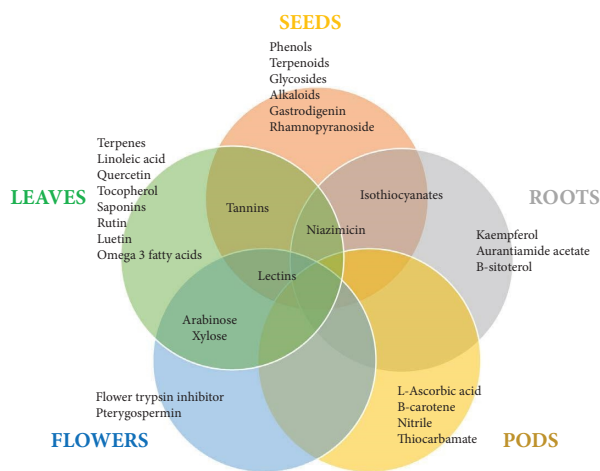
3.1.2. Moringa oleifera as a Source for Human Diets. MO has been also included in the formulation of human diets for many nutritional and medicinal purposes [125]. *Moringa* plant parts have been used as a natural agent for food fortification which is used to improve various aspects of food deficiencies, especially in children with micronutrient malnutrition [126]. It has been used to supplement humans with proteins and lipids of high biological values as well as a potent source for supplementation of iron, zinc, copper, and calcium [127]. It has been included in the diets of young women (teenagers) to improve their nutritional status and educational performance [128]. The addition of MO leaves powder to cookies or as an herbal drink improved the glycemic index, lowered blood glucose, improved appetite, and gastrointestinal health, and lowered both diastolic and systolic high pressure in highly consumed salt models [49, 129, 130]. MO dried leaves have been added to the

TABLE 2: Medicinal applications of *Moringa oleifera*.

Medicinal applications	Plant parts	Major findings	References
Analgesic	Roots	Inhibited the production of TNF-alpha and IL-2	[11]
Antiallergic	Pods and seeds	Inhibited beta-hexosaminidase, histamine, IL-4, and TNF- α release	[12]
Antiatherosclerotic	Leaves	Reduced cholesterol levels and reduced atherosclerotic plaque to 50% and 86%, respectively	[13]
Anticonstipation	Flowers and leaves	Adjusted stool number, weight and water contents of feces, and recovered the thickness of colon muscles and mucus	[14]
Anthelmintic	Seeds	Delayed the development of <i>Aedes aegypti</i> larvae, <i>Anopheles stephensi</i> Liston, and helminth eggs in irrigation water	[15–18]
Anti-inflammatory	Seeds, leaves, and roots	Downregulated TNF- α and interleukin-1 β , and improved IL-6	[19–23]
Anticlastogenic	Leaves and pods	Decreased number of micronucleated peripheral reticulocytes	[24, 25]
Anticonvulsant	Leaves and roots	Enhanced the inhibitory mechanism through the release γ -amino butyric acid (GABA)	[26]
Antinociceptive	Leaves	Reduced the protein levels of ICAM-1 (intercellular adhesion molecule 1) and CD55	[27]
Antioxidant	Leaves and seeds	Included antioxidant agents such as vitamins, minerals, and phenols	[28]
Antipyretic	Leaves	Decreased body temperature	[29]
Antispasmodic	Leaves and seeds	Inhibited the release of acetylcholine	[30]
Antitumor	Seeds, leaves, and roots	Reduced the tumor weight and progression	[31]
Antilicerogenic	Leaves and seeds	Increased the volume of gastric juice, PGE ₂ , IL-10, and GSH	[32–34]
Bactericidal	Leaves, stems, pods, and seeds	Showed antibacterial effect against <i>Staphylococcus aureus</i> , <i>Vibrio cholerae</i> , and <i>Escherichia coli</i>	[35–37]
Diuretic	Seeds	Increased urine output and increased urine volume and concentration	[30, 38]
Fungicide	Leaves, seeds, and roots	Showed antifungal activities against <i>Trichophyton rubrum</i> , <i>Trichophyton mentagrophytes</i> , <i>Epidermophyton floccosum</i> , and <i>Microsporum canis</i>	[39, 40]
Hepatoprotective	Seeds and leaves	Enhanced plasma protein levels, reduced hepatic dysfunction markers, and regenerated hepatic tissue	[41–45]
Hypocholesterolemic	Leaves, seeds, and stem	Lowered plasma LDL cholesterol, VLDL cholesterol, and total cholesterol	[46–48]
Hypotensive	Seeds and pods	Decreased both systolic blood pressure (SBP) and diastolic blood pressure (DBP) and modulated angiotensin-1 converting enzyme (ACE) activity and expression	[49–54]
Immunomodulatory	Seeds, leaves, and flowers	Increased the proliferation of splenocytes, activated macrophages, increased NO production, and increased WBC counts and thymus weight	[55–61]
Potentiate memory	Leaves	Prevented memory impairment and errors	[62]
Radioprotective	Leaves	Prevented the electromagnetic and gamma radiation deleterious consequences	[63–66]

TABLE 3: Synonyms of *Moringa oleifera*.

Region	Name
Arabian	Rawag
Ayurvedic	Haritashaaka, Raktaka, and Akshiva
Chinese	La ken
English	Drumstick tree and Horseradish tree
French	Morungue
Gujarati	Suragavo
Hindi	Saguna and Sainjna
Latin	Moringa oleifera
Malayalam	Murinna and Sigrum
Punjabi	Sainjna and Soanjna
Sanskrit	Subhanjana
Spanish	Angela, Ben, and Moringa
Tamil	Mulaga and Munaga
Unani	Sahajan

FIGURE 1: Bioactive compounds isolated from different plant parts of *Moringa oleifera*.

dishes and snacks of children to compete their malnutrition and deficiencies of β -carotenes or/and minerals and improve energy, iron, and zinc requirements [131–133]. In the same line, MO was effective to compete malnutrition in a clinical study performed on malnourished girls [134]. MO leaf powder could be added to cereals meals or sprinkled on infants' foods for improving micronutrient contents and competing vitamin A deficiency [135, 136]. Interestingly, Moringa was added to the diets of healthy overweight human models lead to improve the overweight and their lipid profiles [137].

3.2. Biochemical Significance of *Moringa oleifera*

3.2.1. *Moringa oleifera* and Glucose Status

(1) *The Action of Moringa oleifera to Control Blood Glucose Level.* MO were widely used to control blood glucose and its metabolism through multiple mechanisms (Figure 2). First, the plant extracts have been approved to inhibit many enzymes that could control glucose absorption or advanced glycation as intestinal α -glucosidase and pancreatic

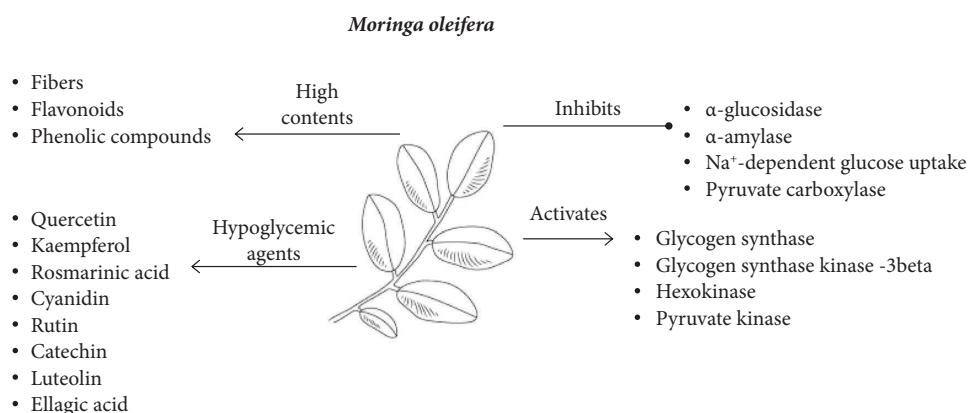
α -amylase [130, 138–140]. Another mechanism includes its ability to inhibit Na^+ -dependent glucose uptake [141, 142]. The plant's high fiber led to a delay in gastric emptying [142]. Also, its contents of flavonoids and many phenolic compounds induce hypoglycemic action [143, 144]. The plants contain a huge amount of hypoglycemic bioactive agents such as catechin, cyanidin, ellagic acid, luteolin, kaempferol, quercetin, rosmarinic acid, and rutin [145]. The potential hypoglycemic power of MO has been approved in many clinical and laboratory studies for both animals and human diabetic models [146–149]. A majority of these studies' results showed a significant improvement in fasting blood glucose levels or glucose tolerance. The plant action revealed the normalization of the gene expressions of enzymes of insulin signaling, glycolytic mechanisms, glycogen storage, and decline in hepatic gluconeogenesis [150–152]. Aqueous extract of MO has been approved to increase the expression levels of glycogen synthase, hexokinase, and pyruvate kinase and lower pyruvate carboxylase expression in the liver of diabetic mice model [153] while seed extract activated glycogen synthase kinase-3 β (GSK-3 β) [154]. The insulin-like proteins isolated from the MO plant contributed to the improvement of glucose uptake, and induced blood glucose reduction could be another way for its hypoglycemic action [155, 156]. MO stimulated insulin secretion through the activation of the insulin-dependent Akt pathway and increased GLUT-4 in skeletal muscle [157].

(2) *Moringa oleifera as a Diabetic Control Plant.* Some animal studies provided evidence for the potential use of MO as a hypoglycemic plant in diabetics and prediabetics [158]. While there is an existing viewpoint that these studies remain inconclusive and necessitate additional clarification and refinement [159]. Regarding human clinical studies, it is difficult to obtain scientific consensus about the antidiabetic action of MO as there are only few approvals with variable results [160]. The plant has been approved to improve blood glucose levels through its ability to increase insulin secretion sensitivity with a noticed inhibition of β -glucosidase and α -amylase activities and increases muscles and liver glucose uptake with noticed inhibition in intestinal uptake and decreases liver gluconeogenesis [161].

(3) *The Hypoglycemic Evidences for Moringa oleifera.* Pre-diabetics supplemented with MO showed an improvement in fasting blood glucose (FBG) and glycated hemoglobin (HbA1c) levels [162], while, postprandial blood glucose levels (PBG) declined in diabetics [130], the same results obtained from studies on diabetic rats [163]. Fermented MO improved the glucose tolerance in obese mice [164]. MO leaves enhanced the insulin release, glucose uptake in the liver, and glycogen biosynthesis in alloxan-induced diabetic models [165]. In the same way, the leaf aqueous extract ameliorated the insulin resistance in mice [166]. Methanolic extract of MO leaves improved both glycogen synthesis and glucose tolerance in rats [151]. Moringa aqueous leaf extract showed a remarkable reduction in blood glucose concentration in rats [167]. Systemic and topical applications of MO leaf aqueous extract positively improved wound healing in

TABLE 4: Nutritional significance of *Moringa oleifera*.

Nutritional contents	Members	References
Amino acids	Leucine, aspartate, glutamate, and proline	[89]
Glycosides	Sitogluside	[90]
Minerals	Zinc, iron, potassium, calcium, selenium and phosphorus	[91, 92]
Peptides	LALPVYN, LHIAALVFQ, FHEEDDAKLF, hevein-like peptide FLSeML, LSeMAAL, LSeMMVL, SeMLLAA, LSeMAL, and antimicrobial peptide	[93–96]
Phenolic compounds	Quercetin, phenolic acid, tannins, and saponins	[97–99]
Carbohydrates	Glucosinolate and polysaccharides	[60, 100–102]
Proteins	Protease and amylase	[103, 104]
Vitamins	Pyridoxine, vitamin E, niacin, and ascorbic acid	[89, 105, 106]
Fatty acids	Octadecanoic acids, palmitic acid, and omega-3 fatty acids	[92, 107]
Flavonoids	Flavones, anthocyanins, myricetin, and kaempferol	[99, 108, 109]

FIGURE 2: The hypoglycemic action of *Moringa oleifera*.

diabetic rats [168]. MO enhanced the regulation of circulating glucose through the regulation of Hsp70 and ILP2 (insulin-like peptide 2) [169] and enhanced the uptake of glucose in adipocytes [170]. It has been approved that the flavonoids and polysaccharides isolated from MO leaves had synergistic action to delay glucose diffusion, dialysis, and delaying starch digestion [171]. In the same line, MO polysaccharides improved glucose levels and metabolism in diabetic models [172]. The supplementation of leaves and seeds potentially activated monoamine oxidase (MAO), adenosine triphosphatase (ATPase), adenosine deaminase (ADA), acetylcholinesterase (AChE), arginase, lactate dehydrogenase (LDH), and angiotensin-I converting enzyme (ACE) activities in diabetics [173]. Leaves potentially control both insulin levels and blood glucose levels in rats with polycystic ovaries [144].

3.2.2. *Moringa oleifera* and Lipids. The potential role of MO extracts in lipids metabolism has been illustrated in Figure 3. It has been approved that MO supplementation leads to an improvement in the metabolic status of obese mice [89, 174]. Moreover, it controlled hypertriglyceridemia and hypercholesterolemia in rodent models [175]. Its fermented leaf extracts have the ability to decrease hepatic accumulation of lipids in obese mice [164], and it alleviated the metabolic syndrome in rats inducing an improvement in lipid profile

and adipokines such as adiponectin and leptin [176]. Furthermore, rats that received MO leaf extract showed great resistance against consequences resulting from metabolic syndrome induced by high fructose and fat diets [177]. The leaves reduced lipids absorption in obese rats [42]. Leaf extract lowered levels of TC, HDL-C, LDL-C, and TAGs through an invitro inhibition of pancreatic lipase and α -glucosidase [178]. The same results were obtained when MO was added to ice cream introduced to rats [179]. In adipocytes, MO has the ability to increase glucose uptake, promote lipolysis, and attenuate adipogenesis by controlling the cell cycle genes [170, 180].

Seed oil extracts reduced leptin and resistin indicating antiobesity activity [181]. Serum examinations indicated that dietary polysaccharides extracted from MO leaves reduced TC, TAGs, and LDL-C in rabbits stressed with heat and modulated their lipid metabolism [182]. Similarly, the administration of MO polysaccharides to mice induced protection against high-fat diet-induced obesity [183]. MO prevented the progression of liver damage and inhibited de novo lipogenesis in the liver [184]. In a similar study, it protected liver damage in ethanol-induced liver damage and reduced TAG levels in mice [185]. Seed extracts upregulated the AMPK and PPAR α and downregulated mTOR and SREBP-1 [44]. *Moringa* also downregulated the expression of PPAR γ , C/EBP α , and FAS and upregulated the expression and phosphorylation of ACC1 and AMPK α [186].

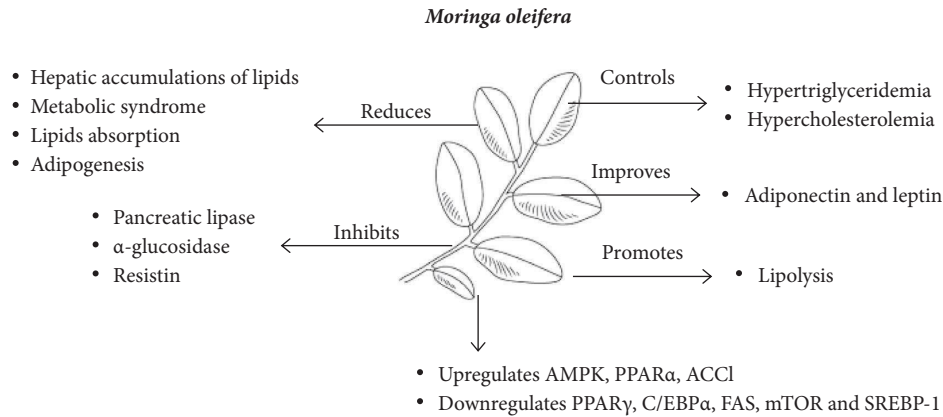


FIGURE 3: The biochemical action of *Moringa oleifera* on lipids.

3.2.3. *Moringa oleifera* and Proteins

(1) *Proteins and Enzymes from Moringa oleifera*. Qualitative analysis of plant parts indicated the presence of many important proteins and enzymes which are included in the synthesis of vitamins, iron transporters, and calcium storage proteins [187]. MO provides proteins for people of low income in consequence, and the plant has been expected to help to reduce kwashiorkor symptoms and its related consequences [188]. The studies approved also that the plant parts could be an alternative resource of proteins used for the preparation of animal diets [189]. It has been approved that MO supplementation enhanced animal reproduction and elevated the protein contents in their milk [124]. Seed oil extracts regulated and enhanced plasma proteins, and reduced the hepatic and renal as well as inflammatory parameters in rats [41]. Polysaccharides isolated from MO also upregulated the expression of enzymes included in protein digestion and absorption in rabbits [118]. Recently, many isolated proteins from MO leaves could improve protein digestibility [190, 191]. In the same way, cysteine peptidase derived from MO seeds is considered a promising alternative to rennet [192]. Milk-clotting peptidase and protease have been isolated and purified from MO seeds and could be used in the dairy industry due to their thermostability [193]. Bioactive compounds like quercetin could target many proteins of high biological functions like p53. Isolated compounds showed great binding affinity to p53 in a molecular docking model the matter which opens the window to apply MO as an active agent in cancer research [194]. Coagulant protein isolated from MO possessed a great antimicrobial action against many isolates of bacteria isolated from water [195]. Leaf extract could modulate the differentiation of brown adipose tissue through the upregulation of bone morphogenetic protein 7 (BMP7) levels in obese mice [196]. The ethanolic extracts were associated with significant normalization of proteins of antioxidant, anti-inflammatory, and antiapoptotic activities [197]. Mass spectrometric results revealed that there was a modulation in the expression of about 125 proteins included in metabolism, signal transduction, transcription, and translation in

models supplemented with pods [198]. MO extracts also have been shown to introduce protection against apoptosis and oxidative stress induced in human neuroblastoma cells through their ability to activate caspase-3 [199]. Leaf extracts protected cells from peroxidative damage and stimulated osteogenic induction in stem cells through the activation of phosphatidylinositol 3-kinase (PI3K/Akt/Foxo1 pathway [200]). MO decreased the inflammatory cytokines as IL-6, IL-1 β , TNF- α , and IFN- β , inhibited the nuclear transfer and expression of the cellular protein transcription factor EB (TFEB), and declined the cellular autophagy [201]. Leaves extracts also alleviated inflammation in colorectal cancer models through the downregulation of TNF- α , IL-2, and IL-6 [202]. Polysaccharides extracted from MO-elevated total proteins and increased protease and amylase activities in early pregnant goats [203]. MO extracts given to laying hens lead to an increase in total proteins and albumin and reduced urea [204]. MO polysaccharides significantly improved the plasma proteins and enhanced protease and amylase activities in goats [103]. MO increased the expression of vascular endothelial growth factor (VEGF) and transforming growth factor beta-1 (TGF- β 1) leading to the contraction of wounds, reduction of epithelization time, increasing antioxidant activities, and reduction of capillary density [205].

(2) *Peptides and Peptide Fractions of Moringa oleifera*. Many peptides isolated from MO are illustrated in Table 5. Furthermore, hydrolysis of MO seeds globulin with trypsin produced many peptides that have potential antioxidant and antihypertensive action [214]. Peptides of hypotensive effect were isolated from MO as angiotensin-converting enzyme (ACE), Gly-Leu-Phe-Phe (GLFF), and renin peptides Leu-Gly-Phe-Phe (LGF) [51]. Many peptides of antimicrobial agents have been purified from MO proteins [215]. Another peptide (<7.5 kDa) has been isolated from MO and possessed an efficient antimicrobial activity [216]. An antimicrobial peptide (AMP) has been also, isolated from MO seeds with an inhibitory action against *S. aureus* through inhibition of DNA gyrase and dihydrofolate reductase [96]. Another peptide of hypoglycemic action has been isolated

from MO and interacted with α -glucosidase through hydrogen bonding and hydrophobic interactions [217].

(3) *Moringa oleifera*, Uric Acid, Urea, and Creatinine. Many studies shed light on the potential influence of MO extracts on protein metabolites such as urea, uric acid, and creatinine. Methanolic extracts of MO decreased both BUN and creatinine in renal ischemic rats' models [218]. Leaves supplementation reduced both serum uric acid and creatinine [219], ameliorated serum uric acid levels [220], improved the hematological indices, and reduced uric acid levels in stressed models [221] in consequence, and MO returned the uric acid to its normal level in diabetic rats and mice [222, 223]. Also, MO seeds reduced the uric acid level in the blood of diabetic rats [224] providing evidence of the hypouricemic power of leaf extracts [225].

3.2.4. *Moringa oleifera* and Minerals. MO is characterized by its huge content of many macro and microelements. Its seeds are rich in iron (Fe), zinc (Zn), phosphorus (P), magnesium (Mg), copper (Cu), and manganese (Mn) [226] while its leaves are full of calcium (Ca), potassium (K), sulphur (S), and Fe [92, 227, 228]. Selenium (Se) and iodide (I) are also present in high concentrations in many parts of edible plants [229]. Its richness of minerals made it a great nutritional supportive plant of high biological value and could be used in the control, management, and treatment of micronutrient malnutrition in both humans and animals, help in bone growth and strength, and competing anemia [126, 134, 188, 230–232].

(1) *Moringa oleifera* and Calcium. MO is approved to contain higher Ca than milk and more potassium (K) than that in bananas [110, 233]. Many methods could be used for the formulation and extraction of MO to increase its bioaccessibility and bioavailability of Ca [234]. Ca contents of leaf extract promoted the growth and development of rats exposed to Ca deficiency, stimulated deposition of Ca in bones, promoted bone growth and strength, and declined resorption of bone [188, 235]. MO has been demonstrated to modulate the Ca signals in tissues, the matter that approved its hypotensive action [236]. Its extracts also were approved to block the entrance and mobilization of Ca^{+2} in sarcolemma [237]. Recently, the plant extracts were used to modify the amorphous/crystallization status of Ca and P the matter which opens the way for its use in drug delivery and nutraceutical applications [238]. Additionally, MO regulates the Ca levels in mitochondria enabling its ability to prevent mitochondrial dysfunction increasing the potentiality of mitochondrial membranes [105].

(2) *Moringa oleifera*, Iron, and Zinc. On a wide scale, MO has been added to formulate diets to compete anemia and bone problems due to its high content of Fe included in its leaves [239]. It is incorporated into the diets of children, pregnant,

and lactating women to overcome iron deficiency and malnutrition [227, 240]. Leaves powder has been used as a fortified food for iron supplementation and to increase its bioaccessibility [241]. The addition of lime juice, ascorbic acid, and citric acid to MO-fortified diets increased iron and zinc bioavailability and bioaccessibility [242]. Fermented leaves also could be another way to increase Fe bioavailability in MO enabling its use to control anemia in children [243]. There are many forms to add MO for animal and human diets; MO leaf flour as a source of Fe and Zn could be added to biscuits to improve pregnant mothers' health who have anemia [244]. For potentiation of children's health and protecting them from the consequences and development of anemia, MO leaf powders have been added to their formulated diets. There is a proportional relationship between the time of using MO in diets and reduction of anemia among children [132, 245].

3.2.5. *Moringa oleifera* and Vitamins. β -Carotenes, α -tocopherols, and L-ascorbic acid, along with vitamins B1, B2, and B3, were identified in many plant parts of MO especially leaves [85, 246–248]. The amount of vitamin A in MO is greater than that in carrots, and the amount of L-ascorbic acid is greater than that in orange [110]. It has been approved also that the recommended daily allowance of vitamins A and C could be covered by the daily use of Moringa seeds and leaves [243, 249]. Clinical studies also approved the ability of MO to improve vitamin A (retinol) kinetics and store with an ameliorated malnutrition status in children [250, 251], improvement of vitamin A levels in postmenopausal women [252], and improved the serum retinol levels in adolescent girls. [253].

3.2.6. *Moringa oleifera* and Hormones

(1) *Moringa oleifera* and Reproductive Hormones. Many biochemical studies have demonstrated the possible effect of MO on both males and females. Its effect tended to be dose-dependent. In females, MO reduced serum levels of luteinizing hormone (LH), follicle-stimulating hormone (FSH), and estrogen while in males, there was an increase in serum concentration of FSH, LH, and testosterone [197, 254–256]. This illustrated its effect on male fertility, enhancement of semen quality, and sex hormones. Therefore, MO is more beneficial for male reproduction than that for female reproduction [256–258]. Furthermore, MO extracts improved male fertility through elevation of serum testosterone and gonadotrophin levels in obese rats [259], improved sperm functions, decreased FSH and increased inhibin B levels in mice [260], and normalized the level of testosterone in rats [261]. For its androgenic effect on testosterone chemistry and reserve, MO leaf extracts have been reported to inhibit β -hydroxylation of testosterone by cytochrome P450 3A4 (CYP3A4) [262], increased the testosterone productivity in Leydig TM3 cells [263], and reduced the level of testosterone in prostatic hyperplastic rats [264]. The combination of MO leaves with nanoparticles like zinc oxide nanoparticles

(ZnONPs) could potentiate the biogenic synthesis of LH, FSH, and testosterone through the regulation of the hypothalamus-pituitary-testicular axis [265]. Contrary to the previously mentioned, MO seed extracts have been reported to potentiate male reproductive toxicity as they decreased testosterone, LH, sperm motility, and sperm in rats [266].

(2) *Moringa oleifera* and *Thyroid Hormones*. In related to its effect on thyroid hormones, MO induced a significant decline in serum levels of thyroid-stimulating hormone (TSH) and an elevation in T3 and T4 in patients with primary hypothyroidism [267]. Furthermore, it inhibited the peripheral conversion of T4 to T3 illustrating its potent role in conserving T4. It should be noted that this inhibitory effect was significant in females than in males [268, 269]. On the contrary, the consumption of *Moringa* more than twice a day may be associated with the induction of goiter in human [270].

(3) *Moringa oleifera* and *Metabolic Hormones*. It has been recorded that the MO has a direct effect on β -cells of the pancreas, stimulating insulin secretion [144, 157]. Clinically, high doses of leaf powder in the form of capsules induced a considerable increase in insulin secretion in healthy populations suggesting its role as an agent in the controlling of blood glucose [146]. In vitro studies approved its influence on direct and/or indirect increases in insulin signaling and sensitivity [152]. Accordingly, it provided a protection against the consequences of insulin resistance, hyperinsulinemia, increased low-density lipoprotein (LDL), increased visceral fat, and high liver weight [177]. The isolated agents from MO opened the window for innovative formulation of drugs against diabetes mellitus [148]. On the other hand, MO leaves have been recorded to elevate the plasma insulin such as growth factor I (IGF-I) [271] insulin [144] and leptin hormones [42, 184] leading to inhibiting adipogenesis [180, 272]. On the contrary, others recorded its ability to reduce serum leptin [152, 181, 273, 274] leading to an increase in the energy expenditure and controlling obesity [275].

(4) *Moringa oleifera*, *Prolactin*, and *Progesterone*. Phytohormones such as auxins, cytokinins, and gibberellins have been approved to be contained in significant amounts in MO leaf extracts [276]. Due to the presence of these phytohormones, consumption of MO during pregnancy may be unuseful as the plant could change the hormonal profile of pregnant women. Phytochemical contents and their metabolites may induce uterine contractility leading to abortion, affecting the conception rate, inducing teratogenicity, and producing congenital anomalies [277]. Regardless of its harmful use during pregnancy, the plant could be used as a galactagogue increasing milk letdown in lactating females and improving milk volume and infant weight better than other pharmacological galactagogues [278]. A strange point here has been reported; MO could be used to initiate and establish lactation even in nongestational women [279]. Its leaf extract has been recorded to resist the negative effect of stress on hormonal balance during pregnancy [280] while

consumption of seed extract induced a decrease in serum progesterone in heat-stressed females [281].

(5) *Moringa oleifera*, *Serotonin (5-Hydroxytryptamine)*, and *Melatonin*. It has been recorded that the brain level of serotonin is elevated in aged rats administered aqueous extract of MO [282] while the aqueous extract from its roots significantly elevated the serotonin levels in the cerebral cortex, cerebellum, midbrain, and caudate nucleus [283]. Serotonin release also has been potentiated through the effect of MO 5-hydroxytryptamine 3 (5-HT₃) receptors in experimental ulcer models [284]. The MO seed extract was able to ameliorate the level of serotonin in the cerebellum of titanium oxide nanoparticles induced damaged brain in rats [285]. In depressed rats, it increased serotonin release through the noradrenergic-serotonergic neurotransmission pathway [286] and recovered the serotonin levels in rats affected with Alzheimer's disease [287]. It has a protective effect against gastric ulceration through increasing 5-HT levels and enterochromaffin cell count [288]. It also potentiated the sleep time induced by pentobarbitone and increased serotonin levels in rats [289]. Up to this moment, there were no studies on the effect of MO on melatonin levels in experimental animals or humans. As well as, phytochemical studies revealed the absence of melatonin in MO extracts [290].

3.2.7. *Moringa oleifera* and *Antioxidants*. The ingredients isolated from many plant parts such as leaves, seeds, or buds have been examined for their antioxidant power in many animals' models. Plant extracts have been reported to elevate antioxidant enzyme activities and decrease hydroperoxides (HP) and thiobarbituric acid reactive substances (TBARS) [291].

(1) *Antioxidant Bioactive Ingredients form Moringa oleifera*. Active ingredients such as niazirin and isothiocyanate have been isolated from MO and have been approved for their potent antioxidant activity and could be used to control many oxidant-related stresses [292, 293], while its polysaccharides significantly lead to enhancement of CAT, SOD, and GPx activities and declined malondialdehyde (MDA) and reactive oxygen species (ROS) levels [294]. MO supplementation enhanced the activity of SOD, CAT, GPx, and lower MDA [124, 295]. MO also could compete with the heat stress in birds through its power to modulate the antioxidant system in heat-stressed broilers. The birds fed on MO showed better antioxidant profiles and low lipid peroxide markers [113]. The leaf extract of MO has been approved also to increase TAC and improve tolerance of immunity [154, 166]. Notably, leaf extract also showed an ability to strengthen the intracellular antioxidant defense system (SOD, CAT, and GSH) and lowered MDA in many experimental models [296]. Leaf extracts of MO could be also used to improve the health of muscles through its ability to influence the redox state in myotubes against hydrogen peroxide oxidant stresses. It elevated the activity of thioredoxin (Trx), SOD, CAT, GPx, and GST. It also mitigated lipids and protein peroxidation by lowering the levels of TBARS and protein carbonyls [297]. The methanolic extract of MO

improved the semen quality in rams through its ability to lower the MDA and elevate SOD, GPx, and ascorbic acid in ram frozen semen [298]. Interestingly, supplementation of MO could protect gastric mucosa in ulcerated stomach by increasing the activities of CAT, SOD, and GPx in rats' stomach [33]. In the same prospect, alcoholic extract from MO has been approved to protect the stomach from ulcers induced by bisphenol via its antioxidant activities [32]. Leaves extracts have been approved to improve ulcerative colitis pathological consequences through their antioxidant power, and they elevated CAT and SOD and lowered MDA in rat's serum [299].

(2) *Antioxidant Power of Moringa oleifera at Molecular Level.* At molecular levels, MO tended to reduce the oxidative stress via activation of a basic leucine zipper transcription factor that links to the promoter domain of the antioxidant response element (Nrf2-ARE) signaling, elevating the expression of Nrf2 target genes and declining the expression of transforming growth factor beta-1 (TGF- β 1) signaling [300]. It has been also approved to stimulate the total antioxidant capacity through the suppression of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) translocation, upregulation of the Kelch-like ECH-associated protein 1- (Keap1-) nuclear factor-erythroid 2-related factor 2 (Nrf2) (Nrf2/Keap1) system, reducing the activation of protein kinase C, zeta (PKC ζ), and inhibiting the NADPH oxidase 4 (Nox4) protein expression [301].

4. Conclusion

Due to its huge content of bioactive compounds and ingredients, *Moringa oleifera* has been approved for its medicinal and nutritional value. Further studies are recommended to discover more promising active compounds with therapeutic and synergistic effects for chronic and serious disorders. Furthermore, the molecular effect of MO should be studied on a wide range to explore more molecular mechanisms for understanding its role as a medicinal plant.

Data Availability

The data that support the concept of this review will be available from the corresponding author upon a reasonable request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

All the authors conceptualized the study, provided software, validated the study, provided resources, wrote the original draft, reviewed and edited the manuscript, and visualized the study.

Acknowledgments

The authors are thankful to the Deanship of Scientific Research at the University of Bisha for supporting this work through the Fast-Track Research Support Program. The authors extend their appreciation to the Deanship of Scientific Research at the University of Bisha, Saudi Arabia, for funding this research work through the Promising Program under Grant Number UB-Promising-34-1445.

References

- [1] I. Husain, O. R. Dale, K. Martin et al., "Screening of medicinal plants for possible herb-drug interactions through modulating nuclear receptors, drug-metabolizing enzymes and transporters," *Journal of Ethnopharmacology*, vol. 301, Article ID 115822, 2023.
- [2] M. Malik, R. Sindhu, S. B. Dhull et al., "Nutritional composition, functionality, and processing technologies for amaranth," *Journal of Food Processing and Preservation*, vol. 2023, Article ID 1753029, 24 pages, 2023.
- [3] J. Rani, M. Kapoor, S. B. Dhull, G. Goksen, and S. Jurić, "Identification and assessment of therapeutic phytoconstituents of catharanthus roseus through GC-MS analysis," *Separations*, vol. 10, no. 6, p. 340, 2023.
- [4] E. A. Ayeni, Y. Gong, H. Yuan, Y. Hu, X. Bai, and X. Liao, "Medicinal plants for anti-neurodegenerative diseases in west Africa," *Journal of Ethnopharmacology*, vol. 285, Article ID 114468, 2022.
- [5] S. B. Dhull, P. Kaur, and S. S. Purewal, "Phytochemical analysis, phenolic compounds, condensed tannin content and antioxidant potential in Marwa (*Origanum majorana*) seed extracts," *Resource-Efficient Technologies*, vol. 2, no. 4, pp. 168–174, 2016.
- [6] P. Kaur, S. B. Dhull, K. S. Sandhu, R. K. Salar, and S. S. Purewal, "Tulsi (*Ocimum tenuiflorum*) seeds: in vitro DNA damage protection, bioactive compounds and antioxidant potential," *Food Measure*, vol. 12, no. 3, pp. 1530–1538, 2018.
- [7] J. Rani, S. B. Dhull, P. K. Rose, and M. K. Kidwai, "Drug-induced liver injury and anti-hepatotoxic effect of herbal compounds: a metabolic mechanism perspective," *Phyto-medicine*, vol. 122, Article ID 155142, 2024.
- [8] H. Rouhi-Boroujeni, E. Heidarian, H. Rouhi-Boroujeni, F. Deris, and M. Rafeian-Kopaei, "Medicinal plants with multiple effects on cardiovascular diseases: a systematic review," *Current Pharmaceutical Design*, vol. 23, no. 7, pp. 999–1015, 2017.
- [9] R. A. Bhat, K. R. Hakeem, and M. A. Dervash, *Phytomedicine: A Treasure of Pharmacologically Active Products from Plants*, Academic Press, San Diego, CA, USA, 2021.
- [10] Z. T. Abdel Shakour, R. H. El-Akad, A. I. Elshamy, A. E. N. G. El Gendy, L. A. Wessjohann, and M. A. Farag, "Dissection of *Moringa oleifera* leaf metabolome in context of its different extracts, origin and in relationship to its biological effects as analysed using molecular networking and chemometrics," *Food Chemistry*, vol. 399, Article ID 133948, 2023.

- [11] K. V. Sashidhara, J. N. Rosaiah, E. Tyagi, R. Shukla, R. Raghbir, and S. M. Rajendran, "Rare dipeptide and urea derivatives from roots of *Moringa oleifera* as potential anti-inflammatory and antinociceptive agents," *European Journal of Medicinal Chemistry*, vol. 44, no. 1, pp. 432–436, 2009.
- [12] N. Z. Abd Rani, E. Kumolosasi, M. Jasamai, J. A. Jamal, K. W. Lam, and K. Husain, "In vitro anti-allergic activity of *Moringa oleifera* Lam. extracts and their isolated compounds," *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, p. 361, 2019.
- [13] P. Chumark, P. Khunawat, Y. Sanvarinda et al., "The in vitro and ex vivo antioxidant properties, hypolipidaemic and antiatherosclerotic activities of water extract of *Moringa oleifera* Lam. leaves," *Journal of Ethnopharmacology*, vol. 116, no. 3, pp. 439–446, 2008.
- [14] C. Li, Z. Li, H. Wu et al., "Therapeutic effect of *Moringa oleifera* leaves on constipation mice based on pharmacodynamics and serum metabolomics," *Journal of Ethnopharmacology*, vol. 282, Article ID 114644, 2022a.
- [15] J. S. Coelho, N. D. Santos, T. H. Napoleao et al., "Effect of *Moringa oleifera* lectin on development and mortality of *Aedes aegypti* larvae," *Chemosphere*, vol. 77, no. 7, pp. 934–938, 2009.
- [16] K. Prabhu, K. Murugan, A. Nareshkumar, N. Ramasubramanian, and S. Bragadeeswaran, "Larvicidal and repellent potential of *Moringa oleifera* against malarial vector, *Anopheles stephensi* Liston (Insecta: Diptera: Culicidae)," *Asian Pacific Journal of Tropical Biomedicine*, vol. 1, no. 2, pp. 124–129, 2011.
- [17] M. E. Sengupta, B. Keraita, A. Olsen et al., "Use of *Moringa oleifera* seed extracts to reduce helminth egg numbers and turbidity in irrigation water," *Water Research*, vol. 46, no. 11, pp. 3646–3656, 2012.
- [18] L. W. Wattenberg, "Chemoprevention of cancer," *Cancer Research*, vol. 45, no. 1, pp. 1–8, 1985.
- [19] M. A. Buabeid, H. S. Yaseen, M. Asif, G. Murtaza, and E. S. A. Arafa, "Anti-inflammatory and anti-angiogenic attributes of *moringa oleifera* Lam. And its nanoclay-based pectin-sericin films," *Frontiers in Pharmacology*, vol. 13, Article ID 890938, 2022.
- [20] E. I. Omodanisi, Y. G. Aboua, and O. O. Oguntibeju, "Assessment of the anti-hyperglycaemic, anti-inflammatory and antioxidant activities of the methanol extract of *moringa oleifera* in diabetes-induced nephrotoxic male wistar rats," *Molecules*, vol. 22, no. 4, p. 439, 2017.
- [21] R. Kumar, S. Varghese, S. Ramamurthy et al., "Assessing the in vitro antioxidant and anti-inflammatory activity of *moringa oleifera* crude extract," *The Journal of Contemporary Dental Practice*, vol. 23, no. 4, pp. 437–442, 2022.
- [22] A. M. E. Sayed, F. A. Omar, M. M. A. Emam, and M. A. Farag, "UPLC-MS/MS and GC-MS based metabolites profiling of *Moringa oleifera* seed with its anti-*Helicobacter pylori* and anti-inflammatory activities," *Natural Product Research*, vol. 36, no. 24, pp. 6433–6438, 2022.
- [23] F. Wang, Y. Bao, C. Zhang et al., "Bioactive components and anti-diabetic properties of *Moringa oleifera* Lam.," *Critical Reviews in Food Science and Nutrition*, vol. 62, no. 14, pp. 3873–3897, 2022a.
- [24] C. Promkum, P. Kupradinun, S. Tuntipopipat, and C. Butryee, "Nutritive evaluation and effect of *Moringa oleifera* pod on clastogenic potential in the mouse," *Asian Pacific Journal of Cancer Prevention*, vol. 11, no. 3, pp. 627–632, 2010.
- [25] S. Shruthi and K. B. Shenoy, "Septilin: a versatile anti-clastogenic, antigenotoxic, antioxidant and histoprotective herbo-mineral formulation on cisplatin-induced toxicity in mice," *Mutation Research, Genetic Toxicology and Environmental Mutagenesis*, vol. 874–875, Article ID 503441, 2022.
- [26] A. G. Bakre, A. O. Aderibigbe, and O. G. Ademowo, "Studies on neuropharmacological profile of ethanol extract of *Moringa oleifera* leaves in mice," *Journal of Ethnopharmacology*, vol. 149, no. 3, pp. 783–789, 2013.
- [27] F. D. Silveira, F. I. F. Gomes, D. R. do Val et al., "Biological and molecular docking evaluation of a benzylisothiocyanate semisynthetic derivative from *moringa oleifera* in a pre-clinical study of temporomandibular joint pain," *Frontiers in Neurosciences*, vol. 16, Article ID 742239, 2022.
- [28] M. K. Segwatibe, S. Cosa, and K. Bassey, "Antioxidant and antimicrobial evaluations of *moringa oleifera* Lam leaves extract and isolated compounds," *Molecules*, vol. 28, no. 2, p. 899, 2023.
- [29] S. Ahmad, S. M. Shah, M. K. Alam, K. Usmanghani, I. Azhar, and M. Akram, "Antipyretic activity of hydro-alcoholic extracts of *Moringa oleifera* in rabbits," *Pakistan journal of pharmaceutical sciences*, vol. 27, no. 4, pp. 931–934, 2014.
- [30] A. Caceres, A. Saravia, S. Rizzo, L. Zabala, E. De Leon, and F. Nave, "Pharmacologie properties of *Moringa oleifera*. 2: screening for antispasmodic, antiinflammatory and diuretic activity," *Journal of Ethnopharmacology*, vol. 36, no. 3, pp. 233–237, 1992.
- [31] L. L. S. Patriota, D. B. M. Ramos, A. Dos Santos et al., "Antitumor activity of *Moringa oleifera* (drumstick tree) flower trypsin inhibitor (MoFTI) in sarcoma 180-bearing mice," *Food and Chemical Toxicology*, vol. 145, Article ID 111691, 2020.
- [32] R. Abo-Elsoud, S. Ahmed Mohamed Abdelaziz, M. Attia Abd Eldaim, and S. M. Hazzaa, "*Moringa oleifera* alcoholic extract protected stomach from bisphenol A-induced gastric ulcer in rats via its anti-oxidant and anti-inflammatory activities," *Environmental Science and Pollution Research*, vol. 29, no. 45, pp. 68830–68841, 2022.
- [33] W. Dalhoumi, F. Guesmi, A. Bouzidi, S. Akermi, N. Hfaiedh, and I. Saidi, "Therapeutic strategies of *Moringa oleifera* Lam. (*Moringaceae*) for stomach and forestomach ulceration induced by HCl/EtOH in rat model," *Saudi Journal of Biological Sciences*, vol. 29, no. 6, Article ID 103284, 2022.
- [34] H. Tian, Z. Wen, Z. Liu, Y. Guo, G. Liu, and B. Sun, "Comprehensive analysis of microbiome, metabolome and transcriptome revealed the mechanisms of *Moringa oleifera* polysaccharide on preventing ulcerative colitis," *International Journal of Biological Macromolecules*, vol. 222, no. Pt A, pp. 573–586, 2022.
- [35] U. Eilert, B. Wolters, and A. Nahrstedt, "The antibiotic principle of seeds of *Moringa oleifera* and *Moringa stenopetala*," *Planta Medica*, vol. 42, no. 05, pp. 55–61, 1981.
- [36] J. R. Peixoto, G. C. Silva, R. A. Costa et al., "In vitro antibacterial effect of aqueous and ethanolic *Moringa* leaf extracts," *Asian Pacific Journal of Tropical Medicine*, vol. 4, no. 3, pp. 201–204, 2011.
- [37] G. H. Viera, J. A. Mourao, A. M. Angelo, R. A. Costa, and R. H. Vieira, "Antibacterial effect (in vitro) of *Moringa oleifera* and *Annona muricata* against Gram positive and Gram negative bacteria," *Revista do Instituto de Medicina Tropical de São Paulo*, vol. 52, no. 3, pp. 129–132, 2010.
- [38] N. Fekadu, H. Basha, A. Meresa, S. Degu, B. Girma, and B. Geleta, "Diuretic activity of the aqueous crude extract and hot tea infusion of *Moringa stenopetala* (Baker f.) Cufod.

- leaves in rats," *Journal of Experimental Pharmacology*, vol. 9, pp. 73–80, 2017.
- [39] P. H. Chuang, C. W. Lee, J. Y. Chou, M. Murugan, B. J. Shieh, and H. M. Chen, "Anti-fungal activity of crude extracts and essential oil of *Moringa oleifera* Lam," *Bioresource Technology*, vol. 98, no. 1, pp. 232–236, 2007.
- [40] P. O. Donli and H. Dauda, "Evaluation of aqueous *Moringa* seed extract as a seed treatment biofungicide for groundnuts," *Pest Management Science*, vol. 59, no. 9, pp. 1060–1062, 2003.
- [41] A. O. Adeoye, J. A. Falode, T. O. Jeje et al., "Modulatory potential of citrus sinensis and moringa oleifera extracts and epiphytes on rat liver mitochondrial permeability transition pore," *Current Drug Discovery Technologies*, vol. 19, no. 3, Article ID e150322202238, 2022a.
- [42] D. A. Alkhudhayri, M. A. Osman, G. M. Alshammari, S. A. Al Maiman, and M. A. Yahya, "Moringa peregrina leaf extracts produce anti-obesity, hypoglycemic, anti-hyperlipidemic, and hepatoprotective effects on high-fat diet fed rats," *Saudi Journal of Biological Sciences*, vol. 28, no. 6, pp. 3333–3342, 2021.
- [43] W. Li, C. Sun, W. Deng et al., "Pharmacokinetic of gastrodigenin rhamnopyranoside from *Moringa* seeds in rodents," *Fitoterapia*, vol. 138, Article ID 104348, 2019.
- [44] M. Liao, C. Sun, R. Li et al., "Amelioration action of gastrodigenin rhamno-pyranoside from *Moringa* seeds on non-alcoholic fatty liver disease," *Food Chemistry*, vol. 379, Article ID 132087, 2022.
- [45] C. Sun, W. Li, Y. Liu et al., "In vitro/in vivo hepatoprotective properties of 1-O-(4-hydroxymethylphenyl)- α -L-rhamnopyranoside from *Moringa oleifera* seeds against carbon tetrachloride-induced hepatic injury," *Food and Chemical Toxicology*, vol. 131, Article ID 110531, 2019.
- [46] S. Ghasi, E. Nwobodo, and J. O. Ofili, "Hypocholesterolemic effects of crude extract of leaf of *Moringa oleifera* Lam in high-fat diet fed wistar rats," *Journal of Ethnopharmacology*, vol. 69, no. 1, pp. 21–25, 2000.
- [47] K. Mehta, R. Balaraman, A. H. Amin, P. A. Bafna, and O. D. Gulati, "Effect of fruits of *Moringa oleifera* on the lipid profile of normal and hypercholesterolaemic rabbits," *Journal of Ethnopharmacology*, vol. 86, no. 2-3, pp. 191–195, 2003.
- [48] P. Siddhuraju and K. Becker, "Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves," *Journal of Agricultural and Food Chemistry*, vol. 51, no. 8, pp. 2144–2155, 2003.
- [49] M. Chan Sun, Z. B. Ruhomally, R. Boojhawon, and V. S. Neergheen-Bhujun, "Consumption of moringa oleifera Lam leaves lowers postprandial blood pressure," *Journal of the American College of Nutrition*, vol. 39, no. 1, pp. 54–62, 2020.
- [50] S. Faizi, B. S. Siddiqui, R. Saleem, K. Aftab, F. Shaheen, and A. H. Gilani, "Hypotensive constituents from the pods of *Moringa oleifera*," *Planta Medica*, vol. 64, no. 03, pp. 225–228, 1998.
- [51] K. Ma, Y. Wang, M. Wang et al., "Antihypertensive activity of the ACE-renin inhibitory peptide derived from *Moringa oleifera* protein," *Food and Function*, vol. 12, no. 19, pp. 8994–9006, 2021.
- [52] M. Mengistu, Y. Abebe, Y. Mekonnen, and T. Tolessa, "In vivo and in vitro hypotensive effect of aqueous extract of *Moringa stenopetala*," *African Health Sciences*, vol. 12, no. 4, pp. 545–551, 2013.
- [53] G. Oboh, O. O. Oluokun, S. I. Oyeleye, and O. B. Ogunsuyi, "Moringa seed-supplemented diets modulate ACE activity but not its gene expression in L-NAME-induced hypertensive rats," *Biomarkers*, vol. 27, no. 7, pp. 684–693, 2022.
- [54] S. J. Stohs and M. J. Hartman, "Review of the safety and efficacy of moringa oleifera," *Phytotherapy Research*, vol. 29, no. 6, pp. 796–804, 2015.
- [55] S. Anudeep, V. K. Prasanna, S. M. Adya, and C. Radha, "Characterization of soluble dietary fiber from *Moringa oleifera* seeds and its immunomodulatory effects," *International Journal of Biological Macromolecules*, vol. 91, pp. 656–662, 2016.
- [56] M. C. Coriolano, J. de Santana Brito, L. L. de Siqueira Patriota et al., "Immunomodulatory effects of the water-soluble lectin from moringa oleifera seeds (WSMoL) on human peripheral blood mononuclear cells (PBMC)," *Protein and Peptide Letters*, vol. 25, no. 3, pp. 295–301, 2018.
- [57] L. L. de Siqueira Patriota, D. K. D. do Nascimento Santos, B. R. da Silva Barros et al., "Evaluation of the in vivo acute toxicity and in vitro hemolytic and immunomodulatory activities of the moringa oleifera flower trypsin inhibitor (MoFTI)," *Protein and Peptide Letters*, vol. 28, no. 6, pp. 665–674, 2021.
- [58] E. E. El Shanawany, E. A. Fouad, H. G. Keshta, S. E. Hassan, A. G. Hegazi, and E. H. Abdel-Rahman, "Immunomodulatory effects of *Moringa oleifera* leaves aqueous extract in sheep naturally co-infected with *Fasciola gigantica* and *Clostridium novyi*," *Journal of Parasitic Diseases*, vol. 43, no. 4, pp. 583–591, 2019.
- [59] A. Gupta, M. K. Gautam, R. K. Singh et al., "Immunomodulatory effect of *Moringa oleifera* Lam. extract on cyclophosphamide induced toxicity in mice," *Indian Journal of Experimental Biology*, vol. 48, no. 11, pp. 1157–1160, 2010.
- [60] K. Sharma, M. Kumar, R. Waghmare et al., "Moringa (*Moringa oleifera* Lam.) polysaccharides: extraction, characterization, bioactivities, and industrial application," *International Journal of Biological Macromolecules*, vol. 209, no. Pt A, pp. 763–778, 2022.
- [61] M. Yang, L. Tao, X. R. Kang et al., "Recent developments in *Moringa oleifera* Lam. polysaccharides: a review of the relationship between extraction methods, structural characteristics and functional activities," *Food Chemistry X*, vol. 14, Article ID 100322, 2022a.
- [62] S. Afrin, A. Hossain, and S. Begum, "Effects of *Moringa oleifera* on working memory: an experimental study with memory-impaired Wistar rats tested in radial arm maze," *BMC Research Notes*, vol. 15, no. 1, p. 314, 2022.
- [63] M. M. Bin-Meferij and A. F. El-Kott, "The radioprotective effects of *Moringa oleifera* against mobile phone electromagnetic radiation-induced infertility in rats," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 8, pp. 12487–12497, 2015.
- [64] T. Ramabulana, R. D. Mavunda, P. A. Steenkamp, L. A. Piater, I. A. Dubery, and N. E. Madala, "Perturbation of pharmacologically relevant polyphenolic compounds in *Moringa oleifera* against photo-oxidative damages imposed by gamma radiation," *Journal of Photochemistry and Photobiology B: Biology*, vol. 156, pp. 79–86, 2016.
- [65] A. V. Rao, P. U. Devi, and R. Kamath, "In vivo radioprotective effect of *Moringa oleifera* leaves," *Indian Journal of Experimental Biology*, vol. 39, no. 9, pp. 858–863, 2001.
- [66] M. Sinha, D. K. Das, S. Bhattacharjee, S. Majumdar, and S. Dey, "Leaf extract of *Moringa oleifera* prevents ionizing

- radiation-induced oxidative stress in mice,” *Journal of Medicinal Food*, vol. 14, no. 10, pp. 1167–1172, 2011.
- [67] M. Elghandour, A. Maggiolino, P. Vazquez-Mendoza et al., “Moringa oleifera as a natural alternative for the control of gastrointestinal parasites in equines: a review,” *Plants*, vol. 12, no. 9, p. 1921, 2023.
- [68] U. K. Azlan, N. A. Khairul Annuar, A. Mediani et al., “An insight into the neuroprotective and anti-neuroinflammatory effects and mechanisms of Moringa oleifera,” *Frontiers in Pharmacology*, vol. 13, Article ID 1035220, 2022.
- [69] A. Cuschieri, E. Camilleri, and R. Blundell, “Cerebroprotective effects of Moringa oleifera derivatives extracts against MCAO ischemic stroke: a systematic review and meta-analysis,” *Heliyon*, vol. 9, no. 6, Article ID e16622, 2023.
- [70] M. M. Moremane, B. Abrahams, and C. Tiloke, “Moringa oleifera: a review on the antiproliferative potential in breast cancer cells,” *Current Issues in Molecular Biology*, vol. 45, no. 8, pp. 6880–6902, 2023.
- [71] N. E. Shafiq and A. F. Mahdee, “Moringa oleifera use in maintaining oral health and its potential use in regenerative dentistry,” *The Scientific World Journal*, vol. 2023, Article ID 8876189, 8 pages, 2023.
- [72] X. H. Hu, X. Y. Yang, J. Lian et al., “Moringa oleifera leaf attenuate osteoporosis in ovariectomized rats by modulating gut microbiota composition and MAPK signaling pathway,” *Biomedicine and Pharmacotherapy*, vol. 161, Article ID 114434, 2023.
- [73] Z. Li, X. Li, S. Tang et al., “Moringa oleifera Lam. Leaf improves constipation of rats induced by low-fiber-diet: a proteomics study,” *Journal of Ethnopharmacology*, vol. 318, no. Pt A, Article ID 116936, 2024.
- [74] S. Zhang, Y. Cao, Y. Huang et al., “Aqueous M. oleifera leaf extract alleviates DSS-induced colitis in mice through suppression of inflammation,” *Journal of Ethnopharmacology*, vol. 318, no. Pt B, Article ID 116929, 2024.
- [75] G. Barzan, A. Sacco, A. M. Giovannozzi et al., “Development of innovative antioxidant food packaging systems based on natural extracts from food industry waste and Moringa oleifera leaves,” *Food Chemistry*, vol. 432, Article ID 137088, 2024.
- [76] A. Bangar, H. Khan, A. Kaur, K. Dua, and T. G. Singh, “Understanding mechanistic aspect of the therapeutic role of herbal agents on neuroplasticity in cerebral ischemic-reperfusion injury,” *Journal of Ethnopharmacology*, vol. 319, no. Pt 2, Article ID 117153, 2024.
- [77] X. Xiao, J. Wang, C. Meng et al., “Moringa oleifera Lam and its therapeutic effects in immune disorders,” *Frontiers in Pharmacology*, vol. 11, Article ID 566783, 2020.
- [78] Z. J. Sha, Y. H. Liu, B. Yang et al., “[Textual research on traditional application of Moringa],” *Zhongguo Zhongyao Zazhi*, vol. 45, no. 12, pp. 2800–2807, 2020.
- [79] Moringamix, “The one which never dies,” in *The Beautiful Original Name the Moringa Tree, that Is Used in many African Languages*, Moringamix, Philippines, 2020.
- [80] A. Pareek, M. Pant, M. M. Gupta et al., “Moringa oleifera: an updated comprehensive review of its pharmacological activities, ethnomedicinal, phytopharmaceutical formulation, clinical, phytochemical, and toxicological aspects,” *International Journal of Molecular Sciences*, vol. 24, no. 3, p. 2098, 2023.
- [81] A. Senthilkumar, N. Karuvantevida, L. Rastrelli, S. S. Kurup, and A. J. Cheruth, “Traditional uses, pharmacological efficacy, and phytochemistry of moringa peregrina (forssk.) fiori-A review,” *Frontiers in Pharmacology*, vol. 9, p. 465, 2018.
- [82] S. Kumar, P. K. Verma, A. Shukla et al., “Moringa oleifera L. leaf extract induces cell cycle arrest and mitochondrial apoptosis in Dalton’s Lymphoma: an in vitro and in vivo study,” *Journal of Ethnopharmacology*, vol. 302, no. Pt A, Article ID 115849, 2023.
- [83] M. Potesta, A. Minutolo, A. Gismondi et al., “Cytotoxic and apoptotic effects of different extracts of Moringa oleifera Lam on lymphoid and monocytoid cells,” *Experimental and Therapeutic Medicine*, vol. 18, no. 1, pp. 5–17, 2019.
- [84] R. Gupta, G. M. Kannan, M. Sharma, and S. J. S. Flora, “Therapeutic effects of Moringa oleifera on arsenic-induced toxicity in rats,” *Environmental Toxicology and Pharmacology*, vol. 20, no. 3, pp. 456–464, 2005.
- [85] H. K. J. Dhongade, B. K. Paikra, and B. Gidwani, “Phytochemistry and pharmacology of moringa oleifera Lam,” *Journal of Pharmacopuncture*, vol. 20, no. 3, pp. 194–200, 2017.
- [86] Z. Islam, S. M. R. Islam, F. Hossen, K. Mahtab-Ul-Islam, M. R. Hasan, and R. Karim, “Moringa oleifera is a prominent source of nutrients with potential health benefits,” *International Journal of Food Science*, vol. 2021, Article ID 6627265, 11 pages, 2021.
- [87] S. Grosshagauer, P. Pirkwieser, K. Kraemer, and V. Somoza, “The future of moringa foods: a food chemistry perspective,” *Frontiers in Nutrition*, vol. 8, Article ID 751076, 2021.
- [88] K. Ercan, O. F. Geceseffa, M. E. Taysi, O. A. Ali Ali, and S. Taysi, “Moringa oleifera: a review of its occurrence, pharmacological importance and oxidative stress,” *Mini-Reviews in Medicinal Chemistry*, vol. 21, no. 3, pp. 380–396, 2021.
- [89] Y. J. Kim and H. S. Kim, “Screening Moringa species focused on development of locally available sustainable nutritional supplements,” *Nutrition Research and Practice*, vol. 13, no. 6, pp. 529–534, 2019.
- [90] B. Zainab, Z. Ayaz, M. S. Alwahibi et al., “In-silico elucidation of Moringa oleifera phytochemicals against diabetes mellitus,” *Saudi Journal of Biological Sciences*, vol. 27, no. 9, pp. 2299–2307, 2020.
- [91] S. V. Patil, B. V. Mohite, K. R. Marathe, N. S. Salunkhe, V. Marathe, and V. S. Patil, “Moringa tree, gift of nature: a review on nutritional and industrial potential,” *Current Pharmacology Reports*, vol. 8, no. 4, pp. 262–280, 2022.
- [92] C. Trigo, M. L. Castello, M. D. Ortola, F. J. Garcia-Mares, and M. Desamparados Soriano, “Moringa oleifera: an unknown crop in developed countries with great potential for industry and adapted to climate change,” *Foods*, vol. 10, no. 1, p. 31, 2020.
- [93] B. Chen, J. Miao, H. Ye et al., “Purification, identification, and mechanistic investigation of novel selenium-enriched antioxidant peptides from moringa oleifera seeds,” *Journal of Agricultural and Food Chemistry*, vol. 71, no. 11, pp. 4625–4637, 2023.
- [94] S. G. Kini, K. H. Wong, W. L. Tan, T. Xiao, and J. P. Tam, “Morintides: cargo-free chitin-binding peptides from Moringa oleifera,” *BMC Plant Biology*, vol. 17, no. 1, p. 68, 2017.
- [95] L. Tao, F. Gu, Y. Liu et al., “Preparation of antioxidant peptides from Moringa oleifera leaves and their protection against oxidative damage in HepG2 cells,” *Frontiers in Nutrition*, vol. 9, Article ID 1062671, 2022.
- [96] X. Wang, L. He, Z. Huang et al., “Isolation, identification and characterization of a novel antimicrobial peptide from

- Moringa oleifera seeds based on affinity adsorption,” *Food Chemistry*, vol. 398, Article ID 133923, 2023.
- [97] Q. Gao, Z. Wei, Y. Liu et al., “Characterization, large-scale HSCCC separation and neuroprotective effects of polyphenols from moringa oleifera leaves,” *Molecules*, vol. 27, no. 3, p. 678, 2022.
- [98] S. S. Ndlovu, T. Ghazi, and A. A. Chaturgoon, “The potential of moringa oleifera to ameliorate HAART-induced pathophysiological complications,” *Cells*, vol. 11, no. 19, p. 2981, 2022.
- [99] S. B. Rode, A. Dadmal, and H. V. Salankar, “Nature’s gold (moringa oleifera): miracle properties,” *Cureus*, vol. 14, no. 7, Article ID e26640, 2022.
- [100] K. E. Coello, J. Frias, C. Martinez-Villaluenga, M. E. Cartea, R. Abilleira, and E. Penas, “Potential of germination in selected conditions to improve the nutritional and bioactive properties of moringa (moringa oleifera L.),” *Foods*, vol. 9, no. 11, p. 1639, 2020.
- [101] N. A. Lopez-Rodriguez, M. Gaytan-Martinez, M. de la Luz Reyes-Vega, and G. Loarca-Pina, “Glucosinolates and isothiocyanates from moringa oleifera: chemical and biological approaches,” *Plant Foods for Human Nutrition*, vol. 75, no. 4, pp. 447–457, 2020.
- [102] T. Tshabalala, B. Ncube, N. E. Madala et al., “Scribbling the cat: a case of the “Miracle” Plant, moringa oleifera,” *Plants*, vol. 8, no. 11, p. 510, 2019.
- [103] A. Afzal, T. Hussain, and A. Hameed, “Moringa oleifera supplementation improves antioxidant status and biochemical indices by attenuating early pregnancy stress in beetal goats,” *Frontiers in Nutrition*, vol. 8, Article ID 700957, 2021a.
- [104] S. Sultana, “Nutritional and functional properties of Moringa oleifera,” *Metabolism Open*, vol. 8, Article ID 100061, 2020.
- [105] E. Gonzalez-Burgos, I. Urena-Vacas, M. Sanchez, and M. P. Gomez-Serranillos, “Nutritional value of moringa oleifera Lam. Leaf powder extracts and their neuroprotective effects via antioxidative and mitochondrial regulation,” *Nutrients*, vol. 13, no. 7, p. 2203, 2021.
- [106] N. Qi, X. Gong, C. Feng, X. Wang, Y. Xu, and L. Lin, “Simultaneous analysis of eight vitamin E isomers in Moringa oleifera Lam. leaves by ultra performance convergence chromatography,” *Food Chemistry*, vol. 207, pp. 157–161, 2016.
- [107] D. Barhoi, P. Upadhaya, S. N. Barbhuiya, A. Giri, and S. Giri, “Aqueous extract of moringa oleifera exhibit potential anticancer activity and can be used as a possible cancer therapeutic agent: a study involving in vitro and in vivo approach,” *Journal of the American College of Nutrition*, vol. 40, no. 1, pp. 70–85, 2021.
- [108] S. W. A. Adeoye, O. S. Adeshina, M. G. Yusuf, and A. Omole, “Hepatoprotective and renoprotective effect of moringa oleifera seed oil on dichlorvos-induced toxicity in male wistar rats,” *Nigerian Journal of Physiological Sciences*, vol. 37, no. 1, pp. 119–126, 2022b.
- [109] A. K. Dhakad, M. Ikram, S. Sharma, S. Khan, V. V. Pandey, and A. Singh, “Biological, nutritional, and therapeutic significance of Moringa oleifera Lam,” *Phytotherapy Research*, vol. 33, no. 11, pp. 2870–2903, 2019.
- [110] B. S. Mushtaq, M. B. Hussain, R. Omer et al., “Moringa oleifera in malnutrition: a comprehensive review,” *Current Drug Discovery Technologies*, vol. 18, no. 2, pp. 235–243, 2021.
- [111] A. T. Oyeyinka, O. A. Adebo, M. Siwela, and K. Pillay, “Dataset on effect of decolourisation on metabolomic profile of Moringa oleifera leaf powder,” *Data in Brief*, vol. 44, Article ID 108508, 2022.
- [112] S. H. Abu Hafsa, S. A. Ibrahim, Y. Z. Eid, and A. A. Hassan, “Effect of dietary Moringa oleifera leaves on the performance, ileal microbiota and antioxidative status of broiler chickens,” *Journal of Animal Physiology and Animal Nutrition*, vol. 104, no. 2, pp. 529–538, 2020.
- [113] O. A. Jimoh, O. T. Daramola, H. O. Okin-Aminu, and O. A. Ojo, “Performance, hemato-biochemical indices and oxidative stress markers of broiler chicken fed phytogenic during heat stress condition,” *Journal of Animal Science and Technology*, vol. 64, no. 5, pp. 970–984, 2022.
- [114] W. Monir, M. A. Abdel-Rahman, S. El-Din Hassan, E. S. Mansour, and S. M. M. Awad, “Pomegranate peel and moringa-based diets enhanced biochemical and immune parameters of Nile tilapia against bacterial infection by *Aeromonas hydrophila*,” *Microbial Pathogenesis*, vol. 145, Article ID 104202, 2020.
- [115] S. Tabassum, S. M. Hussain, S. Ali et al., “Partial replacement of fish meal with Moringa oleifera leaf meal in practical diets of *Cirrhinus mrigala* fingerlings,” *Brazilian Journal of Biology*, vol. 83, Article ID e246333, 2021.
- [116] E. E. Babiker, F. Al Juhaimi, K. Ghafoor, H. E. Mohamed, and K. A. Abdoun, “Effect of partial replacement of alfalfa hay with Moringa species leaves on milk yield and composition of Najdi ewes,” *Tropical Animal Health and Production*, vol. 48, no. 7, pp. 1427–1433, 2016.
- [117] T. W. Kekana, U. Marume, and F. V. Nherera-Chokuda, “Prepartum supplementation of Moringa oleifera leaf meal: effects on health of the dam, colostrum quality, and acquisition of immunity in the calf,” *Journal of Dairy Science*, vol. 105, no. 7, pp. 5813–5821, 2022.
- [118] A. R. Khalid, T. B. Yasoob, Z. Zhang, X. Zhu, and S. Hang, “Dietary Moringa oleifera leaf powder improves jejunal permeability and digestive function by modulating the microbiota composition and mucosal immunity in heat stressed rabbits,” *Environmental Science and Pollution Research*, vol. 29, no. 53, pp. 80952–80967, 2022.
- [119] E. Faustin Evaris, L. Sarmiento Franco, C. Sandoval Castro, J. Segura Correa, and J. A. Caamal Maldonado, “Male layer chicken’s response to dietary moringa oleifera meal in a tropical climate,” *Animals*, vol. 12, no. 14, p. 1843, 2022.
- [120] O. N’Nanle, A. Tete-Benissan, D. Nideou, O. M. Onagbesan, and K. Tona, “Use of Moringa oleifera leaves in broiler production chain. 1- Effect on Sasso breeder hens performances, internal quality of hatching eggs and serum lipids,” *Veterinary Medicine and Science*, vol. 6, no. 3, pp. 485–490, 2020.
- [121] H. M. N. Tolba, A. A. Elmaaty, G. K. Farag, D. A. Mansour, and H. A. Elakkad, “Immunological effect of Moringa Oleifera leaf extract on vaccinated and non-vaccinated Hubbard chickens experimentally infected with Newcastle virus,” *Saudi Journal of Biological Sciences*, vol. 29, no. 1, pp. 420–426, 2022.
- [122] J. Liu, G. Ma, Y. Wang, and Y. Zhang, “Moringa oleifera leaf flavonoids protect bovine mammary epithelial cells from hydrogen peroxide-induced oxidative stress in vitro,” *Reproduction in Domestic Animals*, vol. 55, no. 6, pp. 711–719, 2020.
- [123] M. K. Bashir, K. S. Huque, N. R. Sarker, and N. Sultana, “Quality assessment and feeding impact of Moringa feed on intake, digestibility, enteric CH₄ emission, rumen fermentation, and milk yield,” *Journal of Advanced Veterinary and Animal Research*, vol. 7, no. 3, pp. 521–529, 2020.

- [124] J. J. Sun, P. Wang, G. P. Chen et al., "Effect of Moringa oleifera supplementation on productive performance, colostrum composition and serum biochemical indexes of sow," *Journal of Animal Physiology and Animal Nutrition*, vol. 104, no. 1, pp. 291–299, 2020.
- [125] S. Seshadri and V. S. Nambiar, "Kanjero (*Digera arvensis*) and drumstick leaves (*Moringa oleifera*): nutrient profile and potential for human consumption," *World Review of Nutrition and Dietetics*, vol. 91, pp. 41–59, 2003.
- [126] S. Vishwakarma, C. Genu Dalbhagat, S. Mandliya, and H. Niwas Mishra, "Investigation of natural food fortificants for improving various properties of fortified foods: a review," *Food Research International*, vol. 156, Article ID 111186, 2022.
- [127] C. T. Lockett Christopher C. Calvert, C. C. Calvert, and L. E. Grivetti, "Energy and micronutrient composition of dietary and medicinal wild plants consumed during drought. Study of rural Fulani, northeastern Nigeria," *International Journal of Food Sciences and Nutrition*, vol. 51, no. 3, pp. 195–208, 2000.
- [128] Suhartini, V. Hadju, S. R. A. Unde, Nurjaqin, and B. Bahar, "Moringa oleifera capsule and diet in young women with dropout school," *Gaceta Sanitaria*, vol. 35, no. Suppl 2, pp. S211–S215, 2021.
- [129] J. Ahmad, I. Khan, S. K. Johnson, I. Alam, and Z. U. Din, "Effect of incorporating stevia and moringa in cookies on postprandial glycemia, appetite, palatability, and gastrointestinal well-being," *Journal of the American College of Nutrition*, vol. 37, no. 2, pp. 133–139, 2018.
- [130] A. Leone, S. Bertoli, S. Di Lello et al., "Effect of moringa oleifera leaf powder on postprandial blood glucose response: in vivo study on saharawi people living in refugee camps," *Nutrients*, vol. 10, no. 10, p. 1494, 2018.
- [131] M. Glover-Amengor, R. Aryeetey, E. Afari, and A. Nyarko, "Micronutrient composition and acceptability of Moringa oleifera leaf-fortified dishes by children in Ada-East district, Ghana," *Food Science and Nutrition*, vol. 5, no. 2, pp. 317–323, 2017.
- [132] L. Menasria, S. Blaney, B. Main et al., "Mitigated impact of provision of local foods combined with nutrition education and counseling on young child nutritional status in Cambodia," *Nutrients*, vol. 10, no. 10, p. 1450, 2018.
- [133] V. Ramarason Rakotosamimanana, D. Valentin, and G. Arvisenet, "How to use local resources to fight malnutrition in Madagascar? A study combining a survey and a consumer test," *Appetite*, vol. 95, pp. 533–543, 2015.
- [134] M. Barichella, G. Pezzoli, S. A. Faierman et al., "Nutritional characterisation of Zambian Moringa oleifera: acceptability and safety of short-term daily supplementation in a group of malnourished girls," *International Journal of Food Sciences and Nutrition*, vol. 70, no. 1, pp. 107–115, 2019.
- [135] L. Boateng, I. Ashley, A. Ohemeng, M. Asante, and M. Steiner-Asiedu, "Improving blood retinol concentrations with complementary foods fortified with moringa oleifera leaf powder- A pilot study," *Yale Journal of Biology and Medicine*, vol. 91, no. 2, pp. 83–94, 2018a.
- [136] L. Boateng, R. Nyarko, M. Asante, and M. Steiner-Asiedu, "Acceptability of complementary foods that incorporate moringa oleifera leaf powder among infants and their caregivers," *Food and Nutrition Bulletin*, vol. 39, no. 1, pp. 137–148, 2018b.
- [137] K. Dixit, D. V. Kamath, K. V. Alluri, and B. A. Davis, "Efficacy of a novel herbal formulation for weight loss demonstrated in a 16-week randomized, double-blind, placebo-controlled clinical trial with healthy overweight adults," *Diabetes, Obesity and Metabolism*, vol. 20, no. 11, pp. 2633–2641, 2018.
- [138] A. O. Ademosun, G. Oboh, and O. F. Ajeigbe, "Influence of Moringa (*Moringa oleifera*) enriched ice creams on rats' brain: exploring the redox and cholinergic systems," *Current Research in Food Science*, vol. 5, pp. 366–373, 2022a.
- [139] G. Oboh, O. M. Agunloye, S. A. Adefegha, A. J. Akinyemi, and A. O. Ademiluyi, "Caffeic and chlorogenic acids inhibit key enzymes linked to type 2 diabetes (in vitro): a comparative study," *Journal of Basic and Clinical Physiology and Pharmacology*, vol. 26, no. 2, pp. 165–170, 2015.
- [140] K. Tadera, Y. Minami, K. Takamatsu, and T. Matsuoka, "Inhibition of ALPHA-glucosidase and ALPHA-amylase by flavonoids," *Journal of Nutritional Science and Vitaminology*, vol. 52, no. 2, pp. 149–153, 2006.
- [141] P. Ader, M. Block, S. Pietzsch, and S. Wolfram, "Interaction of quercetin glucosides with the intestinal sodium/glucose co-transporter (SGLT-1)," *Cancer Letters*, vol. 162, no. 2, pp. 175–180, 2001.
- [142] M. Ndong, M. Uehara, S. Katsumata, and K. Suzuki, "Effects of oral administration of moringa oleifera Lam on glucose tolerance in goto-kakizaki and wistar rats," *Journal of Clinical Biochemistry and Nutrition*, vol. 40, no. 3, pp. 229–233, 2007.
- [143] W. Sangkitikomol, A. Rocejanasaroj, and T. Tencomnao, "Effect of Moringa oleifera on advanced glycation end-product formation and lipid metabolism gene expression in HepG2 cells," *Genetics and Molecular Research*, vol. 13, no. 1, pp. 723–735, 2014.
- [144] S. Siahaan, B. Santoso, and Widjiati, "Effectiveness of moringa oleifera leaves on TNF- α expression, insulin levels, glucose levels and follicle count in *Rattus norvegicus* PCOS model," *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 15, pp. 3255–3270, 2022.
- [145] C. Egbuna, C. G. Awuchi, G. Kushwaha et al., "Bioactive compounds effective against type 2 diabetes mellitus: a systematic review," *Current Topics in Medicinal Chemistry*, vol. 21, no. 12, pp. 1067–1095, 2021.
- [146] P. Anthanont, N. Lumlerdkij, P. Akarasereenont, S. Vannasaeng, and A. Sriwijitkamol, "Moringa oleifera leaf increases insulin secretion after single dose administration: a preliminary study in healthy subjects," *Medical Journal of the Medical Association of Thailand*, vol. 99, no. 3, pp. 308–313, 2016.
- [147] S. Kushwaha, P. Chawla, and A. Kochhar, "Effect of supplementation of drumstick (*Moringa oleifera*) and amaranth (*Amaranthus tricolor*) leaves powder on antioxidant profile and oxidative status among postmenopausal women," *Journal of Food Science and Technology*, vol. 51, no. 11, pp. 3464–3469, 2014.
- [148] P. C. Paula, J. T. A. Oliveira, D. O. B. Sousa et al., "Insulin-like plant proteins as potential innovative drugs to treat diabetes-The Moringa oleifera case study," *New Biotech*, vol. 39, no. Pt A, pp. 99–109, 2017a.
- [149] R. Taweerutchana, N. Lumlerdkij, S. Vannasaeng, P. Akarasereenont, and A. Sriwijitkamol, "Effect of moringa oleifera leaf capsules on glycemic control in therapy-naive type 2 diabetes patients: a randomized placebo controlled study," *Evidence-based Complementary and Alternative Medicine*, vol. 2017, Article ID 6581390, 6 pages, 2017.
- [150] M. A. Abd Eldaim, A. Shaban Abd Elrasoul, and S. A. Abd Elaziz, "An aqueous extract from Moringa oleifera leaves ameliorates hepatotoxicity in alloxan-induced diabetic rats,"

- Biochemistry and Cell Biology*, vol. 95, no. 4, pp. 524–530, 2017.
- [151] L. A. Olayaki, J. E. Irekpita, M. T. Yakubu, and O. O. Ojo, “Methanolic extract of *Moringa oleifera* leaves improves glucose tolerance, glycogen synthesis and lipid metabolism in alloxan-induced diabetic rats,” *Journal of Basic and Clinical Physiology and Pharmacology*, vol. 26, no. 6, pp. 585–593, 2015.
- [152] C. Waterman, P. Rojas-Silva, T. B. Tumer et al., “Isothiocyanate-rich *Moringa oleifera* extract reduces weight gain, insulin resistance, and hepatic gluconeogenesis in mice,” *Molecular Nutrition and Food Research*, vol. 59, no. 6, pp. 1013–1024, 2015.
- [153] Y. Bao, J. Xiao, Z. Weng, X. Lu, X. Shen, and F. Wang, “A phenolic glycoside from *Moringa oleifera* Lam. improves the carbohydrate and lipid metabolisms through AMPK in db/db mice,” *Food Chemistry*, vol. 311, Article ID 125948, 2020.
- [154] Y. Wen, Y. Liu, Q. Huang et al., “*Moringa oleifera* Lam. seed extract protects kidney function in rats with diabetic nephropathy by increasing GSK-3 β activity and activating the Nrf2/HO-1 pathway,” *Phytomedicine*, vol. 95, Article ID 153856, 2022.
- [155] W. Khan, R. Parveen, K. Chester, S. Parveen, and S. Ahmad, “Hypoglycemic potential of aqueous extract of moringa oleifera leaf and in vivo GC-MS metabolomics,” *Frontiers in Pharmacology*, vol. 8, p. 577, 2017.
- [156] P. C. Paula, D. O. Sousa, J. T. Oliveira et al., “A protein isolate from moringa oleifera leaves has hypoglycemic and antioxidant effects in alloxan-induced diabetic mice,” *Molecules*, vol. 22, no. 2, p. 271, 2017b.
- [157] E. S. Attakpa, G. A. Bertin, N. W. Chabi, J. M. Ategbro, B. Seri, and N. A. Khan, “*Moringa oleifera*-rich diet and T cell calcium signaling in spontaneously hypertensive rats,” *Physiological Research*, vol. 66, no. 5, pp. 753–767, 2017a.
- [158] J. Ahmad, I. Khan, and R. Blundell, “*Moringa oleifera* and glycemic control: a review of current evidence and possible mechanisms,” *Phytotherapy Research*, vol. 33, no. 11, pp. 2841–2848, 2019.
- [159] K. Vargas-Sanchez, E. Garay-Jaramillo, and R. E. Gonzalez-Reyes, “Effects of moringa oleifera on glycaemia and insulin levels: a review of animal and human studies,” *Nutrients*, vol. 11, no. 12, p. 2907, 2019.
- [160] F. S. Owens 3rd, O. Dada, J. W. Cyrus, O. O. Adedoyin, and G. Adunlin, “The effects of *Moringa oleifera* on blood glucose levels: a scoping review of the literature,” *Complementary Therapies in Medicine*, vol. 50, Article ID 102362, 2020.
- [161] S. L. Haber, R. P. McMahon, J. Barajas, A. R. Hayes, and H. Hussein, “Effects of *Moringa oleifera* in patients with type 2 diabetes,” *American Journal of Health-System Pharmacy*, vol. 77, no. 22, pp. 1834–1837, 2020.
- [162] S. Gomez-Martinez, L. E. Diaz-Prieto, I. Vicente Castro et al., “*Moringa oleifera* leaf supplementation as a glycemic control strategy in subjects with prediabetes,” *Nutrients*, vol. 14, no. 1, p. 57, 2021.
- [163] C. Waterman, J. L. Graham, C. D. Arnold et al., “*Moringa* isothiocyanate-rich seed extract delays the onset of diabetes in UC davis type-2 diabetes mellitus rats,” *Scientific Reports*, vol. 10, no. 1, p. 8861, 2020.
- [164] H. Joung, B. Kim, H. Park et al., “Fermented moringa oleifera decreases hepatic adiposity and ameliorates glucose intolerance in high-fat diet-induced obese mice,” *Journal of Medicinal Food*, vol. 20, no. 5, pp. 439–447, 2017.
- [165] M. A. Hassan, T. Xu, Y. Tian et al., “Health benefits and phenolic compounds of *Moringa oleifera* leaves: a comprehensive review,” *Phytomedicine*, vol. 93, Article ID 153771, 2021.
- [166] M. J. Tuorkey, “Effects of *Moringa oleifera* aqueous leaf extract in alloxan induced diabetic mice,” *Interventional Medicine and Applied Science*, vol. 8, no. 3, pp. 109–117, 2016.
- [167] S. Woldekidan, A. Mulu, W. Ergetie et al., “Evaluation of antihyperglycemic effect of extract of moringa stenopetala (baker f.) aqueous leaves on alloxan-induced diabetic rats,” *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 14, pp. 185–192, 2021.
- [168] I. M. Azevedo, I. Araujo-Filho, M. M. A. Teixeira, M. Moreira, and A. C. Medeiros, “Wound healing of diabetic rats treated with *Moringa oleifera* extract,” *Acta Cirurgica Brasileira*, vol. 33, no. 9, pp. 799–805, 2018.
- [169] O. H. Oyeniran, G. Oboh, A. O. Ademiluyi, and H. I. Umar, “Mistletoe infested *Moringa oleifera* and *Terminalia catappa* leaves supplemented diet enhances antioxidant and insulin-like peptide mRNA levels in *Drosophila melanogaster*,” *Food Chemistry: Molecular Sciences*, vol. 5, Article ID 100124, 2022.
- [170] K. Vasanth, G. C. Minakshi, K. Velu et al., “Anti-adipogenic β -sitosterol and lupeol from *Moringa oleifera* suppress adipocyte differentiation through regulation of cell cycle progression,” *Journal of Food Biochemistry*, vol. 46, no. 8, Article ID e14170, 2022.
- [171] Y. Yang, L. Lin, M. Zhao, and X. Yang, “The hypoglycemic and hypolipemic potentials of *Moringa oleifera* leaf polysaccharide and polysaccharide-flavonoid complex,” *International Journal of Biological Macromolecules*, vol. 210, pp. 518–529, 2022.
- [172] N. A. Lopez-Rodriguez, L. K. Sanchez-Ortiz, R. Reynoso-Camacho, J. R. Riesgo-Escovar, and G. Loarca-Pina, “Chronic consumption of moringa leaf powder (*Moringa oleifera*) concentration-dependent effects in a *Drosophila melanogaster* type 2 diabetes model,” *Journal of the American Nutraceutical Association*, vol. 42, no. 3, pp. 285–294, 2023.
- [173] S. I. Oyeleye, A. O. Ademiluyi, O. O. Raymond, and G. Oboh, “Synergistic cardioprotective ability of co-administration of *Moringa* supplemented diets and acarbose in diabetic cardiomyopathy involves attenuation of cholinergic, purinergic, monoaminergic, renin-angiotensin system, and antioxidant pathways,” *Journal of Food Biochemistry*, vol. 46, no. 11, Article ID e14475, 2022.
- [174] E. R. M. Zunica, S. Yang, A. Coulter, C. White, J. P. Kirwan, and L. A. Gilmore, “*Moringa oleifera* seed extract concomitantly supplemented with chemotherapy worsens tumor progression in mice with triple negative breast cancer and obesity,” *Nutrients*, vol. 13, no. 9, p. 2923, 2021.
- [175] S. Watanabe, H. Okoshi, S. Yamabe, and M. Shimada, “*Moringa oleifera* Lam. In diabetes mellitus: a systematic review and meta-analysis,” *Molecules*, vol. 26, no. 12, p. 3513, 2021.
- [176] S. M. Awad, N. M. El-Shei, H. Abdel-Sabo, and H. M. Ismail Abo, “*Moringa*, rosemary and purslane leaves extracts alleviate metabolic syndrome in rats induced by high fat-high fructose diet,” *Pakistan Journal of Biological Sciences*, vol. 24, no. 10, pp. 1022–1033, 2021.
- [177] H. M. Irfan, N. A. K. Khan, and M. Z. Asmawi, “*Moringa oleifera* Lam. leaf extracts reverse metabolic syndrome in Sprague Dawley rats fed high-fructose high fat diet for 60-days,” *Archives of Physiology and Biochemistry*, vol. 128, no. 5, pp. 1202–1208, 2022.

- [178] G. L. Chen, Y. B. Xu, J. L. Wu, N. Li, and M. Q. Guo, "Hypoglycemic and hypolipidemic effects of *Moringa oleifera* leaves and their functional chemical constituents," *Food Chemistry*, vol. 333, Article ID 127478, 2020a.
- [179] A. O. Ademosun, A. R. Olaniyan, O. F. Ajeigbe, and G. Oboh, "Functional cereals' anti-diabetic property, phenolic composition, and role on glycemic indices in-vitro," *Journal of Food Biochemistry*, vol. 46, no. 11, Article ID e14150, 2022b.
- [180] G. Muni Swamy, G. Ramesh, R. Devi Prasad, and B. Meriga, "Astragalol, (3-O-glucoside of kaempferol), isolated from *Moringa oleifera* leaves modulates leptin, adiponectin secretion and inhibits adipogenesis in 3T3-L1 adipocytes," *Archives of Physiology and Biochemistry*, vol. 128, no. 4, pp. 938–944, 2022.
- [181] O. E. Kilany, H. M. A. Abdelrazek, T. S. Aldayel, S. Abdo, and M. M. A. Mahmoud, "Anti-obesity potential of *Moringa oleifera* seed extract and lycopene on high fat diet induced obesity in male Sprague Dawley rats," *Saudi Journal of Biological Sciences*, vol. 27, no. 10, pp. 2733–2746, 2020.
- [182] T. B. Yasoob, A. R. Khalid, Z. Zhang, X. Zhu, and S. Hang, "Liver transcriptome of rabbits supplemented with oral *Moringa oleifera* leaf powder under heat stress is associated with modulation of lipid metabolism and up-regulation of genes for thermo-tolerance, antioxidation, and immunity," *Nutrition Research*, vol. 99, pp. 25–39, 2022.
- [183] L. Li, L. Ma, Y. Wen et al., "Crude polysaccharide extracted from *moringa oleifera* leaves prevents obesity in association with modulating gut microbiota in high-fat diet-fed mice," *Frontiers in Nutrition*, vol. 9, Article ID 861588, 2022b.
- [184] C. A. Monraz-Mendez, R. Escutia-Gutierrez, J. S. Rodriguez-Sanabria et al., "*Moringa oleifera* improves MAFLD by inducing epigenetic modifications," *Nutrients*, vol. 14, no. 20, p. 4225, 2022.
- [185] C. G. Kim, S. N. Chang, S. M. Park et al., "*Moringa oleifera* mitigates ethanol-induced oxidative stress, fatty degeneration and hepatic steatosis by promoting Nrf2 in mice," *Phytomedicine*, vol. 100, Article ID 154037, 2022.
- [186] N. Elarabany, A. Hamad, and S. M. AlSobeai, "Evaluating anti-obesity potential, active components, and antioxidant mechanisms of *Moringa peregrina* seeds extract on high-fat diet-induced obesity," *Journal of Food Biochemistry*, vol. 46, no. 10, Article ID e14265, 2022.
- [187] S. N. Pasha, K. M. Shafi, A. G. Joshi et al., "The transcriptome enables the identification of candidate genes behind medicinal value of Drumstick tree (*Moringa oleifera*)," *Genomics*, vol. 112, no. 1, pp. 621–628, 2020.
- [188] R. Ponka, P. M. Zhung, G. Zomegni, C. G. Tchouape, and E. Fokou, "Organoleptic and physicochemical properties of soy-milk yoghurt enriched with *moringa oleifera* root powder," *Global Challenges*, vol. 6, no. 5, Article ID 2100097, 2022.
- [189] B. Su and X. Chen, "Current status and potential of *moringa oleifera* leaf as an alternative protein source for animal feeds," *Frontiers in Veterinary Science*, vol. 7, p. 53, 2020.
- [190] J. Agrawal, K. A. Kumar, D. Indrani, and C. Radha, "Effect of *Moringa oleifera* seed flour on the rheological, physico-sensory, protein digestibility and fatty acid profile of cookies," *Journal of Food Science and Technology*, vol. 59, no. 12, pp. 4731–4739, 2022.
- [191] T. Benhammouche, A. Melo, Z. Martins et al., "Nutritional quality of protein concentrates from *Moringa Oleifera* leaves and in vitro digestibility," *Food Chemistry*, vol. 348, Article ID 128858, 2021.
- [192] X. Wang, Q. Zhao, L. He et al., "Milk-clotting properties on bovine caseins of a novel cysteine peptidase from germinated *Moringa oleifera* seeds," *Journal of Dairy Science*, vol. 105, no. 5, pp. 3770–3781, 2022b.
- [193] F. Wang, Y. Bao, X. Shen et al., "Niazirin from *Moringa oleifera* Lam. attenuates high glucose-induced oxidative stress through PKC ζ /Nox4 pathway," *Phytomedicine*, vol. 86, Article ID 153066, 2021a.
- [194] S. Rath, M. Jagadeb, and R. Bhuyan, "Molecular docking of bioactive compounds derived from *Moringa oleifera* with p53 protein in the apoptosis pathway of oral squamous cell carcinoma," *Genomics Inform*, vol. 19, no. 4, p. e46, 2021.
- [195] A. S. Taiwo, K. Adenike, and O. Aderonke, "Efficacy of a natural coagulant protein from *Moringa oleifera* (Lam) seeds in treatment of Opa reservoir water, Ile-Ife, Nigeria," *Heliyon*, vol. 6, no. 1, Article ID e03335, 2020.
- [196] M. Syamsunarno, F. Alia, N. Anggraeni, V. A. Sumirat, S. Praptama, and N. Atik, "Ethanol extract from *Moringa oleifera* leaves modulates brown adipose tissue and bone morphogenetic protein 7 in high-fat diet mice," *Veterinary World*, vol. 14, no. 5, pp. 1234–1240, 2021.
- [197] G. Albasher, R. Alrajhi, E. Alshammry, and R. Almeer, "*Moringa oleifera* leaf extract attenuates Pb acetate-induced testicular damage in rats," *Combinatorial Chemistry & High Throughput Screening*, vol. 24, no. 10, pp. 1593–1602, 2021.
- [198] P. Phannasil, S. Roytrakul, N. Phaonakrop et al., "Protein expression profiles that underpin the preventive and therapeutic potential of *Moringa oleifera* Lam against azoxymethane and dextran sodium sulfate-induced mouse colon carcinogenesis," *Oncology Letters*, vol. 20, no. 2, pp. 1792–1802, 2020.
- [199] I. Amara, M. L. Ontario, M. Scuto et al., "*Moringa oleifera* protects SH-SY5Y Cells from DEHP-induced endoplasmic reticulum stress and apoptosis," *Antioxidants*, vol. 10, no. 4, p. 532, 2021.
- [200] M. Liu, H. Ding, H. Wang et al., "*Moringa oleifera* leaf extracts protect BMSC osteogenic induction following peroxidative damage by activating the PI3K/Akt/Foxo1 pathway," *Journal of Orthopaedic Surgery and Research*, vol. 16, no. 1, p. 150, 2021.
- [201] Y. Xiong, M. S. R. Rajoka, H. M. Mehwish et al., "Virucidal activity of *moringa A* from *moringa oleifera* seeds against influenza A viruses by regulating TFEB," *International Immunopharmacology*, vol. 95, Article ID 107561, 2021.
- [202] M. L. Cuellar-Nunez, E. Gonzalez de Mejia, and G. Loarca-Pina, "*Moringa oleifera* leaves alleviated inflammation through downregulation of IL-2, IL-6, and TNF- α in a colitis-associated colorectal cancer model," *Food Research International*, vol. 144, Article ID 110318, 2021.
- [203] A. Afzal, T. Hussain, A. Hameed, M. Shahzad, M. U. Mazhar, and G. Yang, "Dietary *moringa oleifera* alters periparturient plasma and milk biochemical indicators and promotes productive performance in goats," *Frontiers in Veterinary Science*, vol. 8, Article ID 787719, 2021.
- [204] Z. M. Chen, W. H. Chang, A. J. Zheng, H. Y. Cai, and G. H. Liu, "Tolerance evaluation of *Moringa oleifera* extract to Hailan brown laying hens," *Journal of Animal Physiology and Animal Nutrition*, vol. 104, no. 5, pp. 1375–1383, 2020b.
- [205] A. A. Al-Ghanayem, M. S. Alhussaini, M. Asad, and B. Joseph, "Effect of *moringa oleifera* leaf extract on excision wound infections in rats: antioxidant, antimicrobial, and gene expression analysis," *Molecules*, vol. 27, no. 14, p. 4481, 2022.

- [206] P. Kashyap, S. Kumar, C. S. Riar et al., "Recent advances in drumstick (*moringa oleifera*) leaves bioactive compounds: composition, health benefits, bioaccessibility, and dietary applications," *Antioxidants*, vol. 11, no. 2, p. 402, 2022.
- [207] S. Patil, S. Mushtaq, A. T. Raj et al., "Moringa oleifera: antioxidant, anticancer, anti-inflammatory, and related properties of extracts in cell lines: a review of medicinal effects, phytochemistry, and applications," *The Journal of Contemporary Dental Practice*, vol. 22, no. 12, pp. 1483–1492, 2022.
- [208] Y. Jiang, R. Liu, L. Huang et al., "Spiroleiferthione A and oleiferthione A: two unusual isothiocyanate-derived thio- ketone alkaloids from moringa oleifera Lam. Seeds," *Phar- maceuticals*, vol. 16, no. 3, p. 452, 2023.
- [209] R. Banerji, A. Bajpai, and S. C. Verma, "Oil and fatty acid diversity in genetically variable clones of *Moringa oleifera* from India," *Journal of Oleo Science*, vol. 58, no. 1, pp. 9–16, 2009.
- [210] A. A. Karim and A. Azlan, "Fruit pod extracts as a source of nutraceuticals and pharmaceuticals," *Molecules*, vol. 17, no. 10, pp. 11931–11946, 2012.
- [211] Y. Kc, R. Rai, N. Katuwal et al., "Phytochemicals, nutritional, antioxidant activity, and sensory analyses of *Moringa oleifera* Lam. collected from mid-hill region of Nepal," *Natural Product Research*, vol. 36, no. 1, pp. 470–473, 2022.
- [212] A. Razzak, K. R. Roy, U. Sadia, and W. Zzaman, "Effect of thermal processing on physicochemical and antioxidant properties of raw and cooked moringa oleifera Lam. Pods," *International Journal of Food Science*, vol. 2022, Article ID 1502857, 5 pages, 2022.
- [213] M. Z. M. Salem, H. M. Ali, and M. Akrami, "Moringa oleifera seeds-removed ripened pods as alternative for papersheet production: antimicrobial activity and their phytoconstituents profile using HPLC," *Scientific Reports*, vol. 11, no. 1, Article ID 19027, 2021.
- [214] T. A. Aderinola, T. N. Fagbemi, V. N. Enujiugha, A. M. Alashi, and R. E. Aluko, "In vitro antihypertensive and antioxidative properties of trypsin-derived *Moringa oleifera* seed globulin hydrolyzate and its membrane fractions," *Food Science and Nutrition*, vol. 7, no. 1, pp. 132–138, 2019.
- [215] S. Chandrashekar, R. Vijayakumar, R. Chelliah et al., "In vitro and in silico screening and characterization of anti- microbial napin bioactive protein in Brassica juncea and moringa oleifera," *Molecules*, vol. 26, no. 7, p. 2080, 2021.
- [216] S. Chandrashekar, R. Vijayakumar, R. Chelliah, and D. H. Oh, "Identification and purification of potential bioactive peptide of moringa oleifera seed extracts," *Plants*, vol. 9, no. 11, p. 1445, 2020.
- [217] J. Chen, G. Liu, Y. Hong et al., "Regulation of atherosclerosis by toll-like receptor 4 induced by serum amyloid 1: a sys- tematic in vitro study," *BioMed Research International*, vol. 2022, Article ID 4887593, 14 pages, 2022.
- [218] A. Stephen Akinrinde, O. Oduwole, F. Joseph Akinrinmade, and F. Bolawaye Bolaji-Alabi, "Nephroprotective effect of methanol extract of *Moringa oleifera* leaves on acute kidney injury induced by ischemia-reperfusion in rats," *African Health Sciences*, vol. 20, no. 3, pp. 1382–1396, 2020.
- [219] A. A. A. Abdel-Wareth and J. Lohakare, "Moringa oleifera leaves as eco-friendly feed additive in diets of hy-line Brown hens during the late laying period," *Animals*, vol. 11, no. 4, p. 1116, 2021.
- [220] A. E. El-Hadary and M. F. Ramadan, "Antioxidant traits and protective impact of *Moringa oleifera* leaf extract against diclofenac sodium-induced liver toxicity in rats," *Journal of Food Biochemistry*, vol. 43, no. 2, Article ID e12704, 2019.
- [221] H. S. Hamed and Y. S. El-Sayed, "Antioxidant activities of *Moringa oleifera* leaf extract against pendimethalin-induced oxidative stress and genotoxicity in Nile tilapia, *Oreochromis niloticus* (L.)," *Fish Physiology and Biochemistry*, vol. 45, no. 1, pp. 71–82, 2019.
- [222] B. B. Balakrishnan, K. Krishnasamy, V. Mayakrishnan, and A. Selvaraj, "Moringa concanensis Nimmo extracts ame- liorates hyperglycemia-mediated oxidative stress and upre- gulates PPAR γ and GLUT4 gene expression in liver and pancreas of streptozotocin-nicotinamide induced diabetic rats," *Biomedicine and Pharmacotherapy*, vol. 112, Article ID 108688, 2019.
- [223] O. H. Elhamalawy, F. S. Al-Anany, and A. I. El Makawy, "Thiamethoxam-induced hematological, biochemical, and genetic alterations and the ameliorated effect of *Moringa oleifera* in male mice," *Toxicology Reports*, vol. 9, pp. 94–101, 2022.
- [224] A. L. Al-Malki and H. A. El Rabey, "The antidiabetic effect of low doses of *Moringa oleifera* Lam. seeds on streptozotocin induced diabetes and diabetic nephropathy in male rats," *BioMed Research International*, vol. 2015, Article ID 381040, 13 pages, 2015.
- [225] H. Tian, Y. Liang, G. Liu et al., "Moringa oleifera poly- saccharides regulates caecal microbiota and small intestinal metabolic profile in C57BL/6 mice," *International Journal of Biological Macromolecules*, vol. 182, pp. 595–611, 2021.
- [226] A. Gautier, C. M. Duarte, and I. Sousa, "Moringa oleifera seeds characterization and potential uses as food," *Foods*, vol. 11, no. 11, p. 1629, 2022.
- [227] V. V. Matabura and L. M. P. Rweyemamu, "Formulation of plant-based food and characterisation of the nutritional composition: a case study on soy-moringa beverage," *Journal of Food Science and Technology*, vol. 59, no. 10, pp. 3794–3805, 2022.
- [228] R. Penalver, L. Martinez-Zamora, J. M. Lorenzo, G. Ros, and G. Nieto, "Nutritional and antioxidant properties of moringa oleifera leaves in functional foods," *Foods*, vol. 11, no. 8, p. 1107, 2022.
- [229] D. B. Kumssa, E. J. Joy, S. D. Young, D. W. Odee, E. L. Ander, and M. R. Broadley, "Variation in the mineral element concentration of *Moringa oleifera* Lam. and *M. stenopetala* (Bak. f.) Cuf.: role in human nutrition," *PLoS One*, vol. 12, no. 4, Article ID e0175503, 2017.
- [230] A. C. Adi, Q. Rachmah, and A. N. Arimbi, "The acceptance and nutritional value of crispy noodles supplemented with moringa oleifera as a functional snack for children in a food insecure area," *Preventive Nutrition and Food Science*, vol. 24, no. 4, pp. 387–392, 2019.
- [231] K. D. Mandal, M. R. Das, M. Pati et al., "Effect of *Moringa oleifera* on hematological parameters of calves reared in industrial fluorotic area," *Veterinary World*, vol. 8, no. 11, pp. 1364–1369, 2015.
- [232] B. S. Ogunsina, C. Radha, and D. Indrani, "Quality char- acteristics of bread and cookies enriched with debittered *Moringa oleifera* seed flour," *International Journal of Food Sciences and Nutrition*, vol. 62, no. 2, pp. 185–194, 2011.
- [233] M. E. Olson, R. P. Sankaran, J. W. Fahey, M. A. Grusak, D. Odee, and W. Nouman, "Leaf protein and mineral concentrations across the "Miracle Tree" Genus moringa," *PLoS One*, vol. 11, no. 7, Article ID e0159782, 2016.
- [234] Y. R. Yun, S. J. Oh, M. J. Lee et al., "Antioxidant activity and calcium bioaccessibility of *Moringa oleifera* leaf hydrolysate,

- as a potential calcium supplement in food,” *Food Science and Biotechnology*, vol. 29, no. 11, pp. 1563–1571, 2020.
- [235] J. Dai, L. Tao, C. Shi et al., “Fermentation improves calcium bioavailability in moringa oleifera leaves and prevents bone loss in calcium-deficient rats,” *Food Science and Nutrition*, vol. 8, no. 7, pp. 3692–3703, 2020.
- [236] E. S. Attakpa, M. M. Sangare, G. J. Behanzin, J. M. Ategbro, B. Seri, and N. A. Khan, “Moringa olifera Lam. Stimulates activation of the insulin-dependent Akt pathway. Antidiabetic effect in a diet-induced obesity (DIO) mouse model,” *Folia Biologica (Praha)*, vol. 63, no. 2, pp. 42–51, 2017.
- [237] D. Aekthammarat, P. Pannangpetch, and P. Tangsucharit, “Moringa oleifera leaf extract induces vasorelaxation via endothelium-dependent hyperpolarization and calcium channel blockade in mesenteric arterial beds isolated from L-NAME hypertensive rats,” *Clinical and Experimental Hypertension*, vol. 42, no. 6, pp. 490–501, 2020.
- [238] R. Gelli, M. Tonelli, F. Ridi et al., “Modifying the crystallization of amorphous magnesium-calcium phosphate nanoparticles with proteins from Moringa oleifera seeds,” *Journal of Colloid and Interface Science*, vol. 589, pp. 367–377, 2021.
- [239] K. K. Khoja, M. F. Aslam, P. A. Sharp, and G. O. Latunde-Dada, “In vitro bioaccessibility and bioavailability of iron from fenugreek, baobab and moringa,” *Food Chemistry*, vol. 335, Article ID 127671, 2021.
- [240] S. Arora and S. Arora, “Nutritional significance and therapeutic potential of Moringa oleifera: the wonder plant,” *Journal of Food Biochemistry*, vol. 45, no. 10, Article ID e13933, 2021.
- [241] R. van der Merwe, J. Kruger, M. G. Ferruzzi, K. G. Duodu, and J. R. N. Taylor, “Improving iron and zinc bioaccessibility through food-to-food fortification of pearl millet with tropical plant foodstuffs (moringa leaf powder, roselle calyces and baobab fruit pulp),” *Journal of Food Science and Technology*, vol. 56, no. 4, pp. 2244–2256, 2019.
- [242] S. Mawouma, R. Ponka, and C. M. Mbofung, “Acceptability and solubility of iron and zinc contents of modified Moringa oleifera sauces consumed in the Far-north region of Cameroon,” *Food Science and Nutrition*, vol. 5, no. 2, pp. 344–348, 2017.
- [243] S. Alphonse, L. D. Kaale, F. Millinga, and L. M. Rweyemamu, “Enrichment of fermented cassava meal “mchuchume” with micronutrient ingredients from soya bean flour and Moringa oleifera leaves powder,” *Journal of the Science of Food and Agriculture*, vol. 101, no. 9, pp. 3575–3581, 2021.
- [244] M. S. Manggul, H. Hidayanty, S. Arifuddin, M. Ahmad, V. Hadju, and A. N. Usman, “Biscuits containing Moringa oleifera leaves flour improve conditions of anemia in pregnant women,” *Gaceta Sanitaria*, vol. 35, no. Suppl 2, pp. S191–S195, 2021.
- [245] A. E. Shija, S. F. Rumisha, N. M. Oriyo, S. P. Kilima, and J. J. Massaga, “Effect of Moringa Oleifera leaf powder supplementation on reducing anemia in children below two years in Kisarawe District, Tanzania,” *Food Science and Nutrition*, vol. 7, no. 8, pp. 2584–2594, 2019.
- [246] F. Hodas, M. R. T. Zorzenon, and P. G. Milani, “Moringa oleifera potential as a functional food and a natural food additive: a biochemical approach,” *Anais da Academia Brasileira de Ciencias*, vol. 93, no. suppl 4, Article ID e20210571, 2021.
- [247] X. Kou, B. Li, J. B. Olayanju, J. M. Drake, and N. Chen, “Nutraceutical or pharmacological potential of moringa oleifera Lam,” *Nutrients*, vol. 10, no. 3, p. 343, 2018.
- [248] H. Wen Lee, X. Bi, and C. Jeyakumar Henry, “Carotenoids, tocopherols and phyloquinone content of 26 green leafy vegetables commonly consumed in Southeast Asia,” *Food Chemistry*, vol. 385, Article ID 132729, 2022.
- [249] A. Palizban, B. Bakhshaei, and G. Asghari, “Quantitative analysis of the nutritional components in leaves and seeds of the Persian Moringa peregrina (Forssk.) Fiori,” *Pharmacognosy Research*, vol. 7, no. 3, pp. 242–248, 2015.
- [250] M. A. Arif, M. Inam-Ur-Raheem, W. Khalid et al., “Effect of antioxidant-rich moringa leaves on quality and functional properties of strawberry juice,” *Evidence-based Complementary and Alternative Medicine*, vol. 2022, Article ID 8563982, 11 pages, 2022.
- [251] V. Lopez-Teros, J. L. Ford, M. H. Green et al., “Use of a “Super-child” Approach to assess the vitamin A equivalence of moringa oleifera leaves, develop a compartmental model for vitamin A kinetics, and estimate vitamin A total body stores in young Mexican children,” *The Journal of Nutrition*, vol. 147, no. 12, pp. 2356–2363, 2017.
- [252] S. Brar, C. Haugh, N. Robertson et al., “The impact of Moringa oleifera leaf supplementation on human and animal nutrition, growth, and milk production: a systematic review,” *Phytotherapy Research*, vol. 36, no. 4, pp. 1600–1615, 2022.
- [253] M. Khanam, K. I. Sanin, G. Ara et al., “Effects of Moringa oleifera leaves on hemoglobin and serum retinol levels and underweight status among adolescent girls in rural Bangladesh,” *Frontiers in Nutrition*, vol. 9, Article ID 959890, 2022.
- [254] P. K. Ajuogu, O. O. Mgbere, D. S. Bila, and J. R. McFarlane, “Hormonal changes, semen quality and variance in reproductive activity outcomes of post pubertal rabbits fed Moringa oleifera Lam. leaf powder,” *Journal of Ethnopharmacology*, vol. 233, pp. 80–86, 2019.
- [255] N. I. El-Desoky, N. M. Hashem, A. Elkomy, and Z. R. Abo-Elezz, “Physiological response and semen quality of rabbit bucks supplemented with Moringa leaves ethanolic extract during summer season,” *Animal*, vol. 11, no. 9, pp. 1549–1557, 2017.
- [256] Y. M. El-Gindy, S. M. Zahran, M. H. Ahmed, M. J. Adegbeye, A. Z. M. Salem, and M. Y. Salam, “Enhancing semen quality, antioxidant status and sex hormones of V-line rabbit bucks fed on supplemented diets with dried moringa leaves,” *Animal Biotechnology*, vol. 34, no. 7, pp. 2626–2635, 2022.
- [257] H. H. Abd, H. A. Ahmed, and T. F. Mutar, “Moringa oleifera leaves extract modulates toxicity, sperms alterations, oxidative stress, and testicular damage induced by tramadol in male rats,” *Toxicology Research*, vol. 9, no. 2, pp. 101–106, 2020.
- [258] B. Ogunlade, S. O. Jeje, S. A. Adelakun, and G. T. Akingbade, “Moringa oleifera restored semen quality, hormonal profile, and testicular morphology against Highly Active Antiretroviral Therapy-induced toxicity in adult male Wistar rats,” *JBRA Assisted Reproduction*, vol. 26, no. 1, pp. 3–12, 2022.
- [259] S. M. Greish, G. S. Abdel Kader, E. Z. Abdelaziz, D. A. Eltamany, H. S. Sallam, and N. M. Abogresha, “Lycopene is superior to moringa in improving fertility markers in diet-induced obesity male rats,” *Saudi Journal of Biological Sciences*, vol. 28, no. 5, pp. 2956–2963, 2021.
- [260] G. Nayak, A. Rao, P. Mullick et al., “Ethanolic extract of Moringa oleifera leaves alleviate cyclophosphamide-induced testicular toxicity by improving endocrine function and modulating cell specific gene expression in mouse testis,”

- Journal of Ethnopharmacology*, vol. 259, Article ID 112922, 2020.
- [261] S. S. Elblehi, O. I. El Euony, and A. F. El-Nahas, "Partial ameliorative effect of Moringa leaf ethanolic extract on the reproductive toxicity and the expression of steroidogenic genes induced by subchronic cadmium in male rats," *Environmental Science and Pollution Research*, vol. 26, no. 23, pp. 23306–23318, 2019.
- [262] T. G. Monera, A. R. Wolfe, C. C. Maponga, L. Z. Benet, and J. Guglielmo, "Moringa oleifera leaf extracts inhibit 6 β -hydroxylation of testosterone by CYP3A4," *Journal of Infection in Developing Countries*, vol. 2, no. 5, pp. 379–383, 2008.
- [263] C. S. Opuwari, M. N. Matshipi, M. K. Phaahla et al., "Androgenic effect of aqueous leaf extract of Moringa oleifera on Leydig TM3 cells in vitro," *Andrologia*, vol. 52, no. 11, Article ID e13825, 2020.
- [264] I. O. Ishola, K. O. Yemitan, O. O. Afolayan, C. C. Anunobi, and T. E. Durojaiye, "Potential of Moringa oleifera in the Treatment of Benign Prostate Hyperplasia: role of Antioxidant Defence Systems," *Medical Principles and Practice*, vol. 27, no. 1, pp. 15–22, 2018.
- [265] J. K. Akintunde, T. I. Farai, M. R. Arogundade, and J. T. Adeleke, "Biogenic zinc-oxide nanoparticles of Moringa oleifera leaves abrogates rotenone induced neuroendocrine toxicity by regulation of oxidative stress and acetylcholinesterase activity," *Biochemistry and Biophysics Reports*, vol. 26, Article ID 100999, 2021.
- [266] O. O. Obembe, "Sex hormones and oxidative stress biomarkers of male Wistar rats treated with Moringa oleifera seed fractions," *JBRA Assisted Reproduction*, vol. 23, no. 4, pp. 408–413, 2019.
- [267] M. A. Alam, M. A. Quamri, and N. Haider, "Efficacy and safety of Barg-e-Sahajna (Moringa olifera Lam.) in primary hypothyroidism," *Drug Metabolism and Personalized Therapy*, vol. 0, no. 0, 2021.
- [268] A. R. Khalid, T. B. Yasooob, Z. Zhang et al., "Supplementation of Moringa oleifera leaf powder orally improved productive performance by enhancing the intestinal health in rabbits under chronic heat stress," *Journal of Thermal Biology*, vol. 93, Article ID 102680, 2020.
- [269] P. Tahiliani and A. Kar, "Role of Moringa oleifera leaf extract in the regulation of thyroid hormone status in adult male and female rats," *Pharmacological Research*, vol. 41, no. 3, pp. 319–323, 2000.
- [270] C. Abuye, A. M. Omwega, and J. K. Imungi, "Familial tendency and dietary association of goitre in Gamo-Gofa, Ethiopia," *East African Medical Journal*, vol. 76, no. 8, pp. 447–451, 1999.
- [271] S. M. Abdel-Raheem and E. H. Hassan, "Effects of dietary inclusion of Moringa oleifera leaf meal on nutrient digestibility, rumen fermentation, ruminal enzyme activities and growth performance of buffalo calves," *Saudi Journal of Biological Sciences*, vol. 28, no. 8, pp. 4430–4436, 2021.
- [272] I. Nallamuthu, A. Jain, and T. Anand, "Comparative evaluation of *Brassica oleracea*, *Ocimum basilicum*, and Moringa oleifera leaf extracts on lipase inhibition and adipogenesis in 3T3-L1 adipocytes," *Journal of Food Biochemistry*, vol. 46, no. 7, Article ID e14158, 2022.
- [273] M. Abdel-Latif, T. Sakran, Y. K. Badawi, and D. S. Abdel-Hady, "Influence of Moringa oleifera extract, vitamin C, and sodium bicarbonate on heat stress-induced HSP70 expression and cellular immune response in rabbits," *Cell Stress and Chaperones*, vol. 23, no. 5, pp. 975–984, 2018.
- [274] S. M. Ezzat, M. H. El Bishbishy, N. M. Aborehab et al., "Upregulation of MC4R and PPAR- α expression mediates the anti-obesity activity of Moringa oleifera Lam. in high-fat diet-induced obesity in rats," *Journal of Ethnopharmacology*, vol. 251, Article ID 112541, 2020.
- [275] S. Kundimi, K. C. Kavungala, S. Sinha et al., "Combined extracts of Moringa oleifera, Murraya koeingii leaves, and Curcuma longa rhizome increases energy expenditure and controls obesity in high-fat diet-fed rats," *Lipids in Health and Disease*, vol. 19, no. 1, p. 198, 2020.
- [276] C. V. Mashamaite, B. L. Ngcobo, A. Manyevere, I. Bertling, and O. A. Fawole, "Assessing the usefulness of moringa oleifera leaf extract as a biostimulant to supplement synthetic fertilizers: a review," *Plants*, vol. 11, no. 17, p. 2214, 2022.
- [277] N. Bernstein, M. Akram, Z. Yaniv-Bachrach, and M. Daniyal, "Is it safe to consume traditional medicinal plants during pregnancy?" *Phytotherapy Research*, vol. 35, no. 4, pp. 1908–1924, 2021.
- [278] S. C. Foong, M. L. Tan, W. C. Foong, L. A. Marasco, J. J. Ho, and J. H. Ong, "Oral galactagogues (natural therapies or drugs) for increasing breast milk production in mothers of non-hospitalised term infants," *Cochrane Database of Systematic Reviews*, vol. 5, no. 5, Article ID CD011505, 2020.
- [279] A. Schnell, "Successful Co-lactation by a queer couple: a case study," *Journal of Human Lactation*, vol. 38, no. 4, pp. 644–650, 2022.
- [280] N. I. El-Desoky, N. M. Hashem, A. Gonzalez-Bulnes, A. G. Elkomy, and Z. R. Abo-Elezz, "Effects of a nano-encapsulated moringa leaf ethanolic extract on the physiology, metabolism and reproductive performance of rabbit does during summer," *Antioxidants*, vol. 10, no. 8, p. 1326, 2021.
- [281] V. B. Mutwedu, A. W. Nyongesa, J. M. Kitaa, R. B. B. Ayagirwe, C. Baharanyi, and J. M. Mbaria, "Effects of Moringa oleifera aqueous seed extracts on reproductive traits of heat-stressed New Zealand white female rabbits," *Frontiers in Veterinary Science*, vol. 9, Article ID 883976, 2022.
- [282] A. Nair D, T. J. James, S. L. Sreelatha, B. J. Kariyil, and S. N. Nair, "Moringa oleifera (Lam.): a natural remedy for ageing?" *Natural Product Research*, vol. 35, no. 24, pp. 6216–6222, 2021.
- [283] K. Ray, R. Hazra, and D. Guha, "Central inhibitory effect of Moringa oleifera root extract: possible role of neurotransmitters," *Indian Journal of Experimental Biology*, vol. 41, no. 11, pp. 1279–1284, 2003.
- [284] S. Debnath, D. Biswas, K. Ray, and D. Guha, "Moringa oleifera induced potentiation of serotonin release by 5-HT(3) receptors in experimental ulcer model," *Phytomedicine*, vol. 18, no. 2-3, pp. 91–95, 2011.
- [285] M. A. Kandeil, E. T. Mohammed, K. S. Hashem, L. Aleya, and M. M. Abdel-Daim, "Moringa seed extract alleviates titanium oxide nanoparticles (TiO₂-NPs)-induced cerebral oxidative damage, and increases cerebral mitochondrial viability," *Environmental Science and Pollution Research*, vol. 27, no. 16, pp. 19169–19184, 2020.
- [286] G. Kaur, M. Invally, R. Sanzagiri, and H. S. Buttar, "Evaluation of the antidepressant activity of Moringa oleifera alone and in combination with fluoxetine," *Journal of Ayurveda and Integrative Medicine*, vol. 6, no. 4, pp. 273–279, 2015.
- [287] R. Ganguly and D. Guha, "Alteration of brain monoamines & EEG wave pattern in rat model of Alzheimer's disease & protection by Moringa oleifera," *Indian Journal of Medical Research*, vol. 128, no. 6, pp. 744–751, 2008.

- [288] S. Debnath and D. Guha, "Role of *Moringa oleifera* on enterochromaffin cell count and serotonin content of experimental ulcer model," *Indian Journal of Experimental Biology*, vol. 45, no. 8, pp. 726–731, 2007.
- [289] K. Ray, R. Hazra, P. K. Debnath, and D. Guha, "Role of 5-hydroxytryptamine in *Moringa oleifera* induced potentiation of pentobarbitone hypnosis in albino rats," *Indian Journal of Experimental Biology*, vol. 42, no. 6, pp. 632–635, 2004.
- [290] T. Padumanonda, J. Johns, A. Sangkasat, and S. Tiyaoranant, "Determination of melatonin content in traditional Thai herbal remedies used as sleeping aids," *Daru Journal of Pharmaceutical Sciences*, vol. 22, no. 1, p. 6, 2014.
- [291] B. Y. Aju, R. Rajalakshmi, and S. Mini, "Protective role of *Moringa oleifera* leaf extract on cardiac antioxidant status and lipid peroxidation in streptozotocin induced diabetic rats," *Heliyon*, vol. 5, no. 12, Article ID e02935, 2019.
- [292] A. Jaja-Chimedza, L. Zhang, K. Wolff et al., "A dietary isothiocyanate-enriched moringa (*Moringa oleifera*) seed extract improves glucose tolerance in a high-fat-diet mouse model and modulates the gut microbiome," *Journal of Functional Foods*, vol. 47, pp. 376–385, 2018.
- [293] X. Wang, L. He, Q. Zhao et al., "Protein function analysis of germinated *Moringa oleifera* seeds, and purification and characterization of their milk-clotting peptidase," *International Journal of Biological Macromolecules*, vol. 171, pp. 539–549, 2021b.
- [294] F. Gu, L. Tao, R. Chen et al., "Ultrasonic-cellulase synergistic extraction of crude polysaccharides from moringa *oleifera* leaves and alleviation of insulin resistance in HepG2 cells," *International Journal of Molecular Sciences*, vol. 23, no. 20, p. 12405, 2022.
- [295] Y. Tian, L. Lin, M. Zhao, A. Peng, and K. Zhao, "Xanthine oxidase inhibitory activity and antihyperuricemic effect of *Moringa oleifera* Lam. leaf hydrolysate rich in phenolics and peptides," *Journal of Ethnopharmacology*, vol. 270, Article ID 113808, 2021b.
- [296] F. T. Mthiyane, P. V. Dlodla, K. Ziqubu et al., "A review on the antidiabetic properties of moringa *oleifera* extracts: focusing on oxidative stress and inflammation as main therapeutic targets," *Frontiers in Pharmacology*, vol. 13, Article ID 940572, 2022.
- [297] R. Ceci, M. Maldini, M. E. Olson et al., "Moringa *oleifera* leaf extract protects C2C12 myotubes against H₂O₂-induced oxidative stress," *Antioxidants*, vol. 11, no. 8, p. 1435, 2022.
- [298] I. E. El-Seadawy, M. S. Kotp, A. M. A. El-Maaty, A. M. Fadl, H. R. El-Sherbiny, and E. A. Abdelnaby, "The impact of varying doses of moringa leaf methanolic extract supplementation in the cryopreservation media on sperm quality, oxidants, and antioxidant capacity of frozen-thawed ram sperm," *Tropical Animal Health and Production*, vol. 54, no. 6, p. 344, 2022.
- [299] P. A. Noubissi, Q. Njilifac, M. A. Fokam Tagne et al., "Anxiolytic and anti-colitis effects of *Moringa oleifera* leaf-aqueous extract on acetic acid-induced colon inflammation in rat," *Biomedicine and Pharmacotherapy*, vol. 154, Article ID 113652, 2022.
- [300] D. Cheng, L. Gao, S. Su et al., "Moringa isothiocyanate activates Nrf2: potential role in diabetic nephropathy," *The AAPS Journal*, vol. 21, no. 2, p. 31, 2019.
- [301] M. Louisa, C. G. H. Patintingan, and B. W. K. Wardhani, "Moringa *oleifera* Lam. In cardiometabolic disorders: a systematic review of recent studies and possible mechanism of actions," *Frontiers in Pharmacology*, vol. 13, Article ID 792794, 2022.