

## **Essential oils in stored product insect pest control**

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## **Supplementary Methods**

**Supplementary Method 1** String used on Scopus database to retrieve the overwide literature for scientometric analyses (Scopus database search: February 2<sup>nd</sup>, 2018)

(TITLE-ABS-KEY(“Essential oil” AND pest OR insect) AND TITLE-ABS-KEY(“stored product” OR food-stuff OR “Internal feeding insect” OR “external feeding insect”) AND TITLE-ABS-KEY(“Pest Management” OR toxicity OR mortality OR repellen\* OR control OR efficacy OR effectiveness OR insecticid\* OR fumigant OR Effect)) AND PUBYEAR > 2003

## **Abbreviations**

AChE= Acetylcholinesterase  
ATPases= Adenosine triphosphatases  
CArE = Carboxylesterase  
EONPs= Essential oil nanoparticles  
FDI= Feeding deterrent index  
GST= Glutathione S-transferase  
IR= Inhibition rate  
KT= Knock-down time  
LC= Lethal concentration  
LD= Lethal dose  
LT= Lethal time  
MFOs= Mixed function oxidases  
PT= Persistence time  
RD= Repellence dose  
RT= Residual activity (time)

## Supplementary Tables

**Table S1** Overview of reviewed studies on EO contact (CT) and ingestion (IT) toxicity towards stored product pests.

Plant Family	Plant species	Insect Species	Tested activity	Main Results	References
Amaryllidaceae	<i>Allium sativum</i>	<i>Sitophilus oryzae</i>	CT	Mortality= 100% after 5 days at 250 mg/kg grain	[1]
		<i>Tribolium castaneum</i>	CT	Mortality= 100% after 4 days at 1.250 mg/kg grain Mortality= 100% after 6 days at 40 mg/kg grain Mortality= 100% after 6 days at 400 mg/kg grain	" " "
Anacardiaceae	<i>Schinus molle</i>	<i>Sitophilus oryzae</i>	CT	$LC_{50}= 0.16 \text{ mg/cm}^2$	[2]
	<i>Schinus terebinthifolius</i>	<i>Sitophilus oryzae</i>	CT	$LC_{50}= 0.42 \text{ mg/cm}^2$	"
Annonaceae	<i>Dennettia tripetala</i>	<i>Sitophilus oryzae</i>	CT	Mortality= 100% after 24h at 2 mg/cm <sup>2</sup>	[3]
	<i>Xylopia aethiopica</i>	<i>Sitophilus oryzae</i>	CT	Mortality= 100% after 24h at 2 mg/cm <sup>2</sup>	"
Apiaceae	<i>Astoma seselifolium</i>	<i>Sitophilus oryzae</i>	CT	$LC_{50}= 0.15 \text{ mg/cm}^2$	[2]
	<i>Carum copticum</i>	<i>Sitophilus granarius</i>	CT	Mortality= 90% in combination with diatomaceous earths (size <37 µm); Synergistic effect	[4]
		<i>Tribolium confusum</i>	CT	Mortality= 36-74% in combination with diatomaceous earths (size <37; < 149 µm); both synergistic and antagonistic effects	"
	<i>Coriandrum sativum</i>	<i>Callosobruchus chinensis</i>	CT	$LC_{50}= 27.26 \mu\text{g}/\text{cm}^2$	[5]
		<i>Corcyra cephalonica</i>	CT	$LC_{50}= 47.93 \mu\text{g}/\text{cm}^2$	"
		<i>Sitophilus oryzae</i>	CT	$LC_{50}= 36.68 \mu\text{g}/\text{cm}^2$	"
	<i>Crithmum maritimum</i>	<i>Oryzaephilus surinamensis</i>	CT	$70.33 \pm 0.70\%$ after 72h with 10% EO solution	[6]
		<i>Rhizopertha dominica</i>	CT	$83.26 \pm 7.50\%$ after 72h with 10% EO solution	"
		<i>Sitophilus granarius</i>	CT	$50.00 \pm 0.58\%$ after 72h with 10% EO solution	"
		<i>Sitophilus oryzae</i>	CT	$93.30 \pm 3.20\%$ after 72h with 10% EO solution	"
Araceae	<i>Pituranthus tortuosus</i>	<i>Tribolium castaneum</i>	CT	$22.16 \pm 1.91\%$ after 72h with 10% EO solution	"
		<i>Tribolium confusum</i>	CT	$33.26 \pm 0.21\%$ after 72h with 10% EO solution	"
		<i>Sitophilus oryzae</i>	CT	$LC_{50}= 0.19 \text{ mg/cm}^2$	[2]
Asparagaceae	<i>Acorus calamus</i>	<i>Callosobruchus chinensis</i>	CT	$LD_{50}= 13.30 \mu\text{g}/\text{cm}^2$ after 24h	[7]
		<i>Sitophilus oryzae</i>	CT	$LD_{50}= 54.46 \mu\text{g}/\text{cm}^2$ after 24h	"
		<i>Tribolium castaneum</i>	CT	$LD_{50}= 166.78 \mu\text{g}/\text{cm}^2$ after 24h	"
Asteraceae	<i>Liriopoe muscari</i>	<i>Lasioderma serricorne</i>	CT	$LD_{50}= 11.28 \mu\text{g}/\text{cm}^2$	[8]
		<i>Liposcelis bostrychophila</i>	CT	$LD_{50}= 21.37 \mu\text{g}/\text{cm}^2$	"
		<i>Tribolium castaneum</i>	CT	$LD_{50}= 13.36 \mu\text{g}/\text{adult}$	"
Asteraceae	<i>Artemisia absinthium</i>	<i>Oryzaephilus surinamensis</i>	CT	$LD_{50}= 0.209$ ; $LD_{95}= 1.963 \mu\text{L}$	[9]
		<i>Tribolium castaneum</i>	CT	$LD_{50}= 2.261$ ; $LD_{95}= 5.921 \mu\text{L}$	"
	<i>Artemisia anethoides</i>	<i>Lasioderma serricorne</i>	CT	$LD_{50}= 24.03 \mu\text{g}/\text{adult}$	[10]
	<i>Artemisia herba-alba</i>	<i>Tribolium castaneum</i>	CT	$LD_{50}= 28.80 \mu\text{g}/\text{adult}$	"
		<i>Callosobruchus maculatus</i>	CT	Mortality= 100% at 20 µL	[11]
		<i>Oryzaephilus surinamensis</i>	CT	$LD_{50}= 2.242$ ; $LD_{95}= 16.864 \mu\text{L}$	[9]
		<i>Tribolium castaneum</i>	CT	$LD_{50}= 7.43$ ; $LD_{95}= 133.32 \mu\text{L}$	"
	<i>Artemisia judaica</i>	<i>Sitophilus oryzae</i>	CT	$LC_{50}= 0,08 \text{ mg/cm}^2$	[2]
	<i>Artemisia monosperma</i>	<i>Sitophilus oryzae</i>	CT	$0,15 LC_{50}\text{mg}/\text{cm}^2$	"
	<i>Artemisia stolonifera</i>	<i>Lasioderma serricorne</i>	CT	$LD_{50}= 12.68 \mu\text{g}/\text{adult}$ , worse than terpinen-4-ol	[12]
Asteraceae	<i>Aster ageratoides</i>	<i>Tribolium castaneum</i>	CT	$LD_{50}= 8.60 \mu\text{g}/\text{adult}$ , better than single compounds	"
		<i>Sitophilus zeamais</i>	CT	$LD_{50}= 27.16 \mu\text{g}/\text{adult}$	[13]
		<i>Tribolium confusum</i>	CT	$LD_{50}= 8.09 \mu\text{g}/\text{adult}$	"
	<i>Calendula officinalis</i>	<i>Sitophilus granarius</i>	CT	Mortality= 99% after 24h with 2.5 mL/kg of EO, 100% after 12h with 5 mL/kg of EO	[14]
		<i>Sitophilus oryzae</i>	CT	$LD_{50}= 163.55 \mu\text{g}/\text{cm}^2$ (flowers EO)	[15]
			CT	$LD_{50}= 424.00 \mu\text{g}/\text{cm}^2$ (roots EO)	"
			CT	$LD_{50}= 485.29 \mu\text{g}/\text{cm}^2$ (leaves EO)	"
			CT	$LD_{50}= 524.31 \mu\text{g}/\text{cm}^2$ (stem EO)	"
		<i>Sitophilus zeamais</i>	CT	$LD_{50}= 308.11 \mu\text{g}/\text{cm}^2$ (flowers EO)	"
			CT	$LD_{50}= 403.05 \mu\text{g}/\text{cm}^2$ (roots EO)	"
Asteraceae	<i>Dahlia pinnata</i>		CT	$LD_{50}= 627.34 \mu\text{g}/\text{cm}^2$ (leaves EO)	"
			CT	$LD_{50}= 828.79 \mu\text{g}/\text{cm}^2$ (leaves EO)	"
	<i>Laggera pterodonta</i>	<i>Lasioderma serricorne</i>	CT	$LD_{50}= 32.97 \mu\text{g}/\text{adult}$	[16]
		<i>Liposcelis bostrychophila</i>	CT	$LD_{50}= 28.53 \mu\text{g}/\text{adult}$	"
	<i>Tagetes erecta</i>	<i>Callosobruchus maculatus</i>	CT	$LD_{50}= 504.5$ ; $LD_{95}= 725.2 \mu\text{g/g adult}$	[17]
Asteraceae		<i>Sitophilus oryzae</i>	CT	$LD_{50}= 392.6$ ; $LD_{95}= 623.2 \mu\text{g/g adult}$	"
		<i>Tribolium castaneum</i>	CT	$LD_{50}= 420.5$ ; $LD_{95}= 679.3 \mu\text{g/g adult}$	"
	<i>Tagetes minuta</i>	<i>Callosobruchus maculatus</i>	CT	$LD_{50}= 310.2$ ; $LD_{95}= 497.8 \mu\text{g/g adult}$	"
		<i>Sitophilus oryzae</i>	CT	$LD_{50}= 192.0$ ; $LD_{95}= 342.5 \mu\text{g/g adult}$	"
Asteraceae		<i>Tribolium castaneum</i>	CT	$LD_{50}= 213.5$ ; $LD_{95}= 372.4 \mu\text{g/g adult}$	"

Plant Family	Plant species	Insect Species	Tested activity	Main Results	References
Atherospermataceae	<i>Tagetes patula</i>	<i>Callosobruchus maculatus</i>	CT	LD <sub>50</sub> = 429.8; LD <sub>95</sub> = 601.7 µg/g adult	"
		<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 222.8; LD <sub>95</sub> = 365.7 µg/g adult	"
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 308.1; LD <sub>95</sub> = 492.4 µg/g adult	"
Chenopodiaceae	<i>Laurelia sempervirens</i>	<i>Sitophilus zeamais</i>	CT	2.3 LC <sub>50</sub> mL/kg grain	[18]
Cupressaceae	<i>Chenopodium ambrosioides</i>	<i>Callosobruchus chinensis</i>	CT	Mortality= 40% with 5-10% of EO contained in the formulations. Protectant action for up to six months of seed storage.	[19]
		<i>Callosobruchus maculatus</i>	CT	LD <sub>50</sub> = 2.8 µL	[19]
		<i>Clausena pentaphylla</i>	CT	Mortality= 40% with 5-10% of EO contained in the formulations. Protectant action for up to six months of seed storage.	[19]
Elaengnaceae	<i>Cupressus lusitanica</i>	<i>Acanthoscelides obtectus</i>	CT	LC <sub>50</sub> = 0.07 % v/w after 168h	[20]
		<i>Sitophilus zeamais</i>	CT	LC <sub>50</sub> = 0.12% v/w after 168h	"
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.79% v/w after 168h	"
Euphorbiaceae	<i>Cupressus macrocarpa</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.32 mg/cm <sup>2</sup>	[2]
		<i>Cupressus sempervirens</i>	CT	LC <sub>50</sub> > 0.6 mg/cm <sup>2</sup>	"
		<i>Juniperus formosana</i>	CT	LD <sub>50</sub> = 81.50 µg/adult	[21]
Geraniaceae	<i>Thuja occidentalis</i>	<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 29.14 µg/adult	"
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 0.3 mg/cm <sup>2</sup>	[2]
		<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 0.3 mg/cm <sup>2</sup>	[2]
Elaengnaceae	<i>Hippophae rhamnoides</i>	<i>Sitophilus granarius</i>	CT	Mortality of 95% after 48h and 100% after 24h at 2.5 mL/kg	[14]
Lamiaceae	<i>Mallotus apelta</i>	<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 211.02 µg/adult	[22]
		<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 46.69 µg/adult	"
		<i>Plodia interpunctella</i>	CT	LD <sub>50</sub> = 37.2; LD <sub>90</sub> = 63.7 µg/cm <sup>2</sup>	[23]
Ocimum	<i>Pelargonium sp</i>	<i>Rhizopertha dominica</i>	CT	LC <sub>50</sub> < 0.006% EO; LC <sub>50</sub> < 0.004 PEG-EO-Nanoparticles. Residual contact toxicity of PEG-EO-nanoparticles for 24 weeks	[24]
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> < 0.2% EO; LC <sub>50</sub> < 0.15 EO-PEG-EO-Nanoparticles. Residual contact toxicity of PEG-EO-nanoparticles for 16 weeks	"
		<i>Pelargonium graveolens</i>	CT	LC <sub>50</sub> = 0.17 mg/cm <sup>2</sup>	[2]
Peraceae	<i>Adhatoda vasica</i>	<i>Callosobruchus chinensis</i>	CT	LD <sub>50</sub> = 6.8 µL	[19]
		<i>Callosobruchus maculatus</i>	CT	LD <sub>50</sub> = 8.4 µL	"
		<i>Calamintha glandulosa</i>	IT	Mortality= 96.67% at 1.14% of EO after 96h.	[25]
Rosaceae	<i>Dracocephalum moldavica</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 22.10 LD <sub>50</sub> µg/adult	[26]
		<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 18.28 µg/adult	"
		<i>Hyptis spicigera</i>	CT	LD <sub>50</sub> = 112.0 ppm	[27]
Solanaceae	<i>Hyptis suaveolens</i>	<i>Tribolium confusum</i>	CT	LD <sub>50</sub> = 57 µg/mg w of adult	[28]
		<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 126 µg/mg w of adult	"
		<i>Callosobruchus maculatus</i>	CT	LD <sub>50</sub> = 101 µg/mg w of adult	"
Sapindaceae	<i>Lavandula angustifolia</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 167 µg/mg w of adult	"
		<i>Plodia interpunctella</i>	CT	LD <sub>50</sub> = 76.3; LD <sub>90</sub> = 129.2 µg/cm <sup>2</sup>	[23]
		<i>Sitophilus granarius</i>	CT	LD <sub>50</sub> = 83.8; LD <sub>90</sub> = 379.7 µg/adult after 24h	[29]
Solanaceae	<i>Lavandula officinalis</i>	<i>Rhizopertha dominica</i>	IT	Mortality= 74.8 and 100% at 2.245 and 4.490 mg/disk (insects fed for 3 days)	"
		<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 0.07 mg/cm <sup>2</sup>	[30]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 0.26 mg/cm <sup>2</sup>	"
Solanaceae	<i>Mentha longifolia subsp. capensis</i>	<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 0.14 µL/g grain	[31]
		<i>Mentha piperita</i>	CT	LD <sub>50</sub> = 53.8; LD <sub>90</sub> = 77.5 µg/cm <sup>2</sup>	[23]
		<i>Mentha sp.</i>	CT	LD <sub>50</sub> = 0.044 µL/mL	[32]
Solanaceae	<i>Mentha viridis</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 239 and 158 ppm after one and two weeks respectively	[33]
		<i>Mosla soochowensis</i>	CT	LD <sub>50</sub> = 25.45 µg/adult	[34]
		<i>Tribolium confusum</i>	CT	LD <sub>50</sub> = 10.23 µg/adult	"
Solanaceae	<i>Ocimum basilicum</i>	<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 0.130 mg/adult	[35]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 0.361 mg/adult	"
		<i>Ocimum canum</i>	CT	LD <sub>50</sub> = 42.9 ppm	[27]
Solanaceae	<i>Ocimum gratissimum</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 37.9; LC <sub>50</sub> = 152.3 µg/mL of EO contained in the powder formulation	[36]
		<i>Sitophilus zeamais</i>	CT	LC <sub>50</sub> = 0.11 mg/cm <sup>2</sup>	[2]
		<i>Tribolium castaneum</i>	IT	Mortality= 10% at 1.14% of EO after 96h.	[25]
Solanaceae	<i>Origanum vulgare</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 1.46 µg/adult. 6 times less toxic than pyretrin	[37]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 1.20 µg/adult, 4.5 times less toxic than pyretrin	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.18 mg/cm <sup>2</sup>	[2]

Plant Family	Plant species	Insect Species	Tested activity	Main Results	References
Lamiaceae	<i>Salvia leiriifolia</i>	<i>Tribolium castaneum</i>	CT	Mortality= 78.3% for non formulated EO and 100% for encapsulated EO at 129.40 µL/30.17 cm <sup>2</sup>	[38]
		<i>Callosobruchus maculatus</i>	CT	Mortality= 100% at 4.66 mg/cm <sup>2</sup> , after 24h	[39]
	<i>Salvia officinalis</i>	<i>Sitophilus oryzae</i>	CT	Mortality= 100% at 4.72 mg/cm <sup>2</sup> , after 24h	"
		<i>Tribolium castaneum</i>	CT	Mortality= 92% at 4.80 mg/cm <sup>2</sup> , after 24h	"
	<i>Salvia sclarea</i>	<i>Tribolium castaneum</i>	IT	No Mortality at the highest tested concentrations (1.14% of EO)	[25]
		<i>Tribolium confusum</i>	IT	No Mortality at the highest tested concentrations (1.14% of EO)	"
	<i>Salvia veneris</i>	<i>Oryzaephilus surinamensis</i>	CT	Mortality= 81.64 ± 2.50% after 72h with 10% EO-acetone solution	[40]
		<i>Rhizopertha dominica</i>	CT	Mortality= 67.09 ± 0.94% after 72h with 10% EO-acetone solution	"
	<i>Satureja hortensis</i>	<i>Sitophilus granarius</i>	CT	Mortality= 70.00 ± 0.00% after 72h with 10% EO-acetone solution	"
		<i>Sitophilus oryzae</i>	CT	Mortality= 71.45 ± 2.50% after 72h with 10% EO-acetone solution	"
Lauraceae	<i>Satureja montana</i>	<i>Tribolium castaneum</i>	CT	Mortality= 32.78 ± 1.52% after 72h with 10% EO-acetone solution	"
		<i>Tribolium confusum</i>	CT	Mortality= 53.50 ± 1.40% after 72h with 10% EO-acetone solution	"
	<i>Schizonepeta multifida</i>	<i>Ephestia kuehniella</i>	CT	LC <sub>50</sub> = 0.27 µL/cm <sup>2</sup>	[41]
		<i>Plodia interpunctella</i>	CT	LC <sub>50</sub> = 0.19 µL/cm <sup>2</sup>	"
	<i>Teucrium chamaedrys</i>	<i>Tribolium castaneum</i>	IT	No Mortality at the tested concentrations (1.14% of EO)	[25]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 30.17 µg/adult	[42]
	<i>Teucrium polium</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 2.75 µg/adult	"
		<i>Tribolium castaneum</i>	IT	Mortality= 10% at the tested concentrations (1.14% of EO) after 96h	[25]
	<i>Thymus marschalianus</i>	<i>Tribolium castaneum</i>	IT	No Mortality at the tested concentrations (1.14% of EO)	"
		<i>Tribolium castaneum</i>	IT	No Mortality at the tested concentrations (1.14% of EO)	"
	<i>Vitex agnus-castus</i>	<i>Sitophilus oryzae</i>	CT	Mortality= 13.33% at the tested concentrations (1.14% of EO) after 96h	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.39 mg/cm <sup>2</sup>	[2]
Monimiaceae	<i>Cinnamomum aromaticum</i>	<i>Callosobruchus maculatus</i>	CT	LD <sub>50</sub> = 27.56 ± 3.78 at 24h; 23.16 ± 3.76 µg/cm <sup>2</sup> at 48h; Concentration-dependent effect on egg hatching and adult emergence	[43]
		<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 7.6-21.3 µg/adult depending on the part of plant	[44]
	<i>Laurus nobilis</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 19 >50 µg/adult depending on the part of plant.	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.44 mg/cm <sup>2</sup>	[30]
	<i>Litsea cubeba</i>	<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.4 mg/cm <sup>2</sup>	"
		<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 27.33 µg/adult	[45]
	<i>Litsea salicifolia</i>	<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 71.56 µg/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 0.079; LD <sub>95</sub> = 0.144 µL/insect	[46]
Myrtaceae	<i>Laurelia sempervirens</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 0.111 and 0.304 µL/insect	"
	<i>Callistemon viminalis</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 38.94, LD <sub>90</sub> = 78.7 µg/mg insect	[47]
		<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 44.05; LD <sub>90</sub> = 102.49 µg/mg insect	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.09 mg/cm <sup>2</sup>	[2]
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.34 mg/cm <sup>2</sup>	[30]
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.46 mg/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 5.1 % (w/v)	[48]
		<i>Plodia interpunctella</i>	CT	No mortality at the tested application rates	[23]
		<i>Callosobruchus chinensis</i>	CT	LC <sub>50</sub> = 59.29 µg/cm <sup>2</sup>	[5]
		<i>Coryca cephalonica</i>	CT	LC <sub>50</sub> = 56. 47 µg/cm <sup>2</sup>	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 52.77 µg/cm <sup>2</sup>	"
Pinaceae	<i>Eucalyptus procera</i>	<i>Callosobruchus maculatus</i>	CT	LC <sub>50</sub> = 0.124 µL/cm <sup>2</sup>	[49]
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.129 µL/cm <sup>2</sup>	"
	<i>Eucalyptus saligna</i>	<i>Acanthoscelides obtectus</i>	CT	LC <sub>50</sub> = 0.003 % v/w after 168h	[20]
		<i>Sitophilus zeamais</i>	CT	LC <sub>50</sub> = 0.005 % v/w after 168h	"
	<i>Myrtus communis</i>	<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.51 % v/w after 168h	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.31 mg/cm <sup>2</sup>	[2]
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.04 mg/cm <sup>2</sup>	[30]
Poaceae	<i>Syzygium cumini</i>	<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 0.13 mg/cm <sup>2</sup>	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> >0.60 mg/cm <sup>2</sup>	[2]
	<i>Pinus longifolia</i>	<i>Callosobruchus chinensis</i>	CT	LC <sub>50</sub> = 74.95 µg/cm <sup>2</sup>	[5]
Cyperaceae	<i>Cymbopogon citratus</i>	<i>Corcyra cephalonica</i>	CT	LC <sub>50</sub> = 64.16 µg/cm <sup>2</sup>	"
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 77.30 µg/cm <sup>2</sup>	"
Poaceae	<i>Cymbopogon citratus</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 0.027 µL/mL	[32]
		<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 20 %(w/v) after 48h	[48]

Plant Family	Plant species	Insect Species	Tested activity	Main Results	References
Rutaceae	<i>Cymbopogon giganteus</i>	<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 6.4 % (w/v) after 48h	"
	<i>Cymbopogon martinii</i>	<i>Plodia interpunctella</i>	CT	LD <sub>50</sub> = 22.8; LD <sub>90</sub> = 44.3 µg/cm <sup>2</sup>	[23]
	<i>Cymbopogon schoenanthus</i>	<i>Tribolium castaneum</i>	CT	LC <sub>50</sub> = 8.5 % (w/v) after 48h	[48]
Rutaceae	<i>Atalantia guillauminii</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 24.07 µg/cm <sup>2</sup>	[50]
		<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 55.83 µg/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 17.11 µg/cm <sup>2</sup>	"
	<i>Citrus × bergamia</i>	<i>Rhizopertha dominica</i>	CT	Residual toxicity= 6 weeks at 0.6%; 20 weeks PEG EO <sub>s</sub> NP (w/w wheat)	[24]
		<i>Tribolium castaneum</i>	CT	Residual toxicity= 4 weeks at 0.05%; 12 weeks PEG EO <sub>s</sub> NP (w/w wheat)	"
	<i>Citrus aurantifolia</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.15 mg/cm <sup>2</sup>	[2]
	<i>Citrus bergamia</i>	<i>Plodia interpunctella</i>	CT	LD <sub>50</sub> = 116.2; LD <sub>90</sub> = 188.7 µg/cm <sup>2</sup>	[23]
	<i>Citrus lemon</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.2 mg/cm <sup>2</sup>	[2]
	<i>Citrus paradisi</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.27 mg/cm <sup>2</sup>	"
	<i>Citrus sinensis</i>	<i>Rhizopertha dominica</i>	CT	LD <sub>50</sub> = 351.05 mg/kg; antagonistic effects in combination with diatomaceous earths; synergistic effects in combination with kaolin	[51]
		<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 0.29 mg/cm <sup>2</sup> LC <sub>50</sub> = 0.43 mg/cm <sup>2</sup>	[2] [30]
		<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 0.074 mg/adult	[35]
Celastraceae	<i>Tribolium castaneum</i>		CT	LD <sub>50</sub> = 0.257 mg/adult	"
				LC <sub>50</sub> = 2.42 mg/cm <sup>2</sup>	[30]
	<i>Clausena anisum-olens</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 12.44 µg/adult	[52]
		<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 74.46 µg/adult	"
	<i>Clausena pentaphylla</i>	<i>Callosobruchus maculatus</i>	CT	Mortality= 40% with 10% of EO contained in the formulations. Protectant action for up to six months of seed storage.	[19]
	<i>Dictamnus dasycarpus</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 12.4 µg/adult	[53]
		<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 27.2 µg/cm <sup>2</sup>	"
	<i>Vepris heterophylla</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 349.8 ppm	[27]
	<i>Zanthoxylum armatum</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 18.74 µg/adult	[54]
	<i>Zanthoxylum dissitum</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 32.16 µg/adult	"
Schisandraceae	<i>Attagenus piceus</i>	<i>Attagenus piceus</i>	CT	Mortality= 22% with EO (leaves) and 100% with EO (roots) at 50%(in hexane)	"
		<i>Lasioderma serricorne</i>	CT	Mortality= 33.3% with EO (leaves) and 100% with EO (roots) at 50%(in hexane); LD <sub>50</sub> (EO roots)= 13.8 µg/adult	"
Verbenaceae	<i>Tribolium castaneum</i>		CT	Mortality= 53.3% with EO (leaves) and 100% with EO (roots) at 50%(in hexane); LD <sub>50</sub> (EO roots)= 47.3 µg/adult	"
	<i>Kadsura heterocarpa</i>	<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 25.57 µg/adult	[55]
Vitaceae	<i>Caryopteris incana</i>	<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 122.65 µg/adult	[56]
	<i>Lippia javanica</i> var. <i>javanica</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 6.22 mg/mL	[57]
	<i>Lippia javanica</i> var. <i>whytei</i>	<i>Sitophilus oryzae</i>	CT	LC <sub>50</sub> = 2.96 mg/mL	"
Winteraceae	<i>Cayratia japonica</i>	<i>Sitophilus zeamais</i>	CT	LD <sub>50</sub> = 32.06 mg/adult	[58]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 44.49 mg/adult	"
Zingiberaceae	<i>Drimys winteri</i>	<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> Bark EO= 75.14, LD <sub>90</sub> Bark EO= 132.29 µg/mg insect LD <sub>50</sub> Leaf EO= 84.05, LD <sub>90</sub> Leaf EO= 162.47 µg/mg insect	[47]
	<i>Alpinia blepharocalyx</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 15.02 mg/adult	[59]
	<i>Amomum maximum</i>	<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 67.46 µg/adult	[60]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 29.57 µg/adult	"
	<i>Amomum tsao-ko</i>	<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 6.14 µg/adult	[61]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 16.52 µg/adult	"
	<i>Etlingera yunnanensis</i>	<i>Liposcelis bostrychophila</i>	CT	LD <sub>50</sub> = 47.38 µg/adult	[62]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 23.33 µg/adult	"
	<i>Zingiber officinale</i>	<i>Ephestia kuhniella</i>	CT	LC <sub>50</sub> = 0.61; LC <sub>90</sub> = 1.88 µL/cm <sup>2</sup>	[63]
		<i>Plodia interpunctella</i>	CT	LC <sub>50</sub> = 0.81; LC <sub>90</sub> = 2.68 µL/cm <sup>2</sup>	"
Plant combination	<i>Zingiber purpureum</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 0.136 µL/mL	[32]
		<i>Lasioderma serricorne</i>	CT	LD <sub>50</sub> = 16.3 µg/adult	[64]
		<i>Tribolium castaneum</i>	CT	LD <sub>50</sub> = 39.0 µg/adult	"
Plant combination	<i>Hyptis + Ocimum</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 75.8 ppm	[27]
	<i>Hyptis + Vepris</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 182.1 ppm	"
	<i>Ocimum + Vepris</i>	<i>Sitophilus oryzae</i>	CT	LD <sub>50</sub> = 103.8 ppm	"

**Table S2** Overview of reviewed studies on EO fumigant toxicity towards stored product pests.

Plant Family	Plant species	Insect Species	Main Results	References
Altingiaceae	<i>Liquidambar orientalis</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 22.8 µL/L air; LC <sub>90</sub> = 31.6 µL/L air	[65]
Amaryllidaceae	<i>Allium sativum</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 1.52 µL/L air; LC <sub>95</sub> = 5.49 µL/L air	[66]
Anacardiaceae	<i>Cotinus coggygria</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> > 60 µL/L air	[65]
	<i>Pistacia atlantica</i>	<i>Tribolium castaneum</i>	Gum EO= LC <sub>50</sub> = 29, 57 µL/L air; Fruit EO= LC <sub>50</sub> = 39.66 µL/L air; Leaves EO= LC <sub>50</sub> = 64.84 µL/L air	[67]
	<i>Pistacia lentiscus</i>	<i>Callosobruchus maculatus</i>	Mortality= 90% after 48h at 8 µL/L air	[68]
		<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 28.03 µL/L air; LC <sub>95</sub> = 63.46 µL/L air; LT <sub>50</sub> = 18.58h and LT <sub>95</sub> = 51.46h at 114 µL/L air	[69]
			LC <sub>50</sub> = 8.44 µL/L air; LC <sub>95</sub> = 43.68 µL/L air	[70]
		<i>Sitophilus granarius</i>	Mortality= 33% after 96h at 40 µL	[71]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 8.44 µL/L air; LC <sub>95</sub> = 43.68 µL/L air; LT <sub>50</sub> = 41.05h and LT <sub>95</sub> = 79.89h at 114 µL/L air	[69]
			LC <sub>50</sub> = 28.03 µL/L air; LC <sub>95</sub> = 63.46 µL/L air	[70]
		<i>Tribolium confusum</i>	Mortality= 67% after 96h at 40 µL	[71]
	<i>Pistacia terebinthus</i>	<i>Sitophilus granarius</i>	Mortality= 43% after 96h at 40 µL	"
		<i>Tribolium confusum</i>	Mortality= 40% after 96h at 40 µL	"
	<i>Schinus molle</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 26.89 mg/L	[2]
	<i>Schinus terebinthifolius</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 28.16 mg/L	"
Apiaceae	<i>Apium nodiflorum</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 20.7 µL/L air; LC <sub>90</sub> = 39.3 µL/L air	[65]
	<i>Astoma seselifolium</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 44.43 mg/L	[2]
	<i>Azilia eryngioides</i>	<i>Sitophilus granarius</i>	LC <sub>50</sub> = 20.05 µL/L air; LT <sub>50</sub> = 21.04h at 37.03 µL/L air	[72]
		<i>Tribolium confusum</i>	LC <sub>50</sub> = 46.48 µL/L air; LT <sub>50</sub> = 24.96h at 37.03 µL/L air	"
	<i>Bupleurum fruticosum</i>	<i>Sitophilus oryzae</i>	Crop cultivar LC <sub>50</sub> = 24.9 µL/L air; LC <sub>90</sub> = 33.5 µL/L air	[65]
			Wild plants LC <sub>50</sub> = 35.2 µL/L air; LC <sub>90</sub> = 106.7 µL/L air	"
	<i>Carum copticum</i>	<i>Sitophilus granarius</i>	LD <sub>50</sub> = 24.22 µL/L air; Mortality= 89% at 18 µL/L air of EO and 100% at 12 µL/L of EO-Nanogel after 48h	[73]
			LC <sub>50</sub> = 10.85 µL/L air; LC <sub>90</sub> = 18.77 µL/L air	"
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 0.91 µL/L air	[74]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 33.14 µL/L air	"
		<i>Tribolium confusum</i>	LD <sub>50</sub> = 66.47 µL/L air; Mortality= 72% at 43 µL/L air of EO and 100% at 29 µL/L air of EO-Nanogel after 48h;	[73]
			LC <sub>50</sub> = 26.56 µL/L air; LC <sub>90</sub> = 52.23 µL/L air	"
	<i>Coriandrum sativum</i>	<i>Callosobruchus chinensis</i>	LC <sub>50</sub> = 16.25 µg/cm <sup>2</sup> on 10 cm <sup>2</sup> filter paper	[5]
		<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 72.167 µL/L air; LC <sub>99</sub> = 267.462 µL/L air	[75]
		<i>Corcyra cephalonica</i>	LC <sub>50</sub> = 18.25 µg/cm <sup>2</sup> on 10 cm <sup>2</sup> filter paper	[5]
		<i>Ephestia kuehniella</i>	LC <sub>50</sub> = 62.633 µL/L air; LC <sub>99</sub> = 221.237 µL/L air	[75]
		<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 5.25 µL/L air; LC <sub>95</sub> = 102.31 µL/L air	[76]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 50.956 µL/L air; LC <sub>99</sub> = 221.495 µL/L air	[75]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 18.11 µg/cm <sup>2</sup> on 10 cm <sup>2</sup> filter paper	[5]
			LC <sub>50</sub> = 145.49 µL/L air; LC <sub>95</sub> = 10124.20 µL/L air	[76]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 276.29 µL/L air; LC <sub>95</sub> = 4543.74 µL/L air	"
	<i>Critchmum maritimum</i>	<i>Oryzaephilus surinamensis</i>	Mortality= 90.75 ± 7.00% at 10%(v/v) acetone solution after 48h	[6]
		<i>Rhizopertha dominica</i>	Mortality= 4.53 ± 3.41% at 10%(v/v) acetone solution after 48h	"
		<i>Sitophilus granarius</i>	Mortality= 100.00 ± 0.00% 10%(v/v) acetone solution after 48h	"
		<i>Sitophilus oryzae</i>	Mortality= 100.00 ± 0.00% 10%(v/v) acetone solution after 48h	"
			LC <sub>50</sub> = 17.7 µL/L air; LC <sub>90</sub> = 23.4 µL/L air	[65]
		<i>Tribolium castaneum</i>	Mortality= 0.00% 10%(v/v) acetone solution after 48h	[6]
		<i>Tribolium confusum</i>	Mortality= 0.00% 10%(v/v) acetone solution after 48h	"
			LC <sub>50</sub> = 17.7 µL/L air; LC <sub>90</sub> = 23.4 µL/L air	[65]
	<i>Cuminum cyminum</i>	<i>Sitophilus granarius</i>	Mortality= 18% after 24h, 80% after 48h at 16 µL/L air of EO; Mortality= 97% after 24h, 100% after 48h at 16 µL/L air of EO-Nanogel. PT <sub>50</sub> = 3 days EO and 11 days EO-Nanogel	[77]
			LC <sub>50</sub> = 0.136 mL/mL air	[78]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 0.271 mL/mL air	"
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = larvae= 15.02 µL and adults= 16.26 µL	[79]
		<i>Tribolium confusum</i>	Mortality= 27% after 24h, 94% after 48h at 20 µL/L air of EO; Mortality= 61% after 24h, 100% after 48h at 20 µL/L air of EO-Nanogel; PT <sub>50</sub> = 4 days EO and 21 days EO-Nanogel	[77]
	<i>Ferula gummosa</i>	<i>Sitophilus granarius</i>	LC <sub>50</sub> = 44.25 µL/L air; LC <sub>90</sub> = 82.88 µL/L air	[80]
	<i>Foeniculum vulgare</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> : larvae= 17.48 µL; adults= 18.55 µL	[79]

Plant Family	Plant species	Insect Species	Main Results	References
Asteraceae	<i>Petroselinum crispum</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 71.071 \mu\text{L/L air}$ ; $LC_{99}= 272.487 \mu\text{L/L air}$	[75]
		<i>Epeorus kuehniella</i>	$LC_{50}= 52.412 \mu\text{L/L air}$ ; $LC_{99}= 243.219 \mu\text{L/L air}$	"
	<i>Pimpinella anisum</i>	<i>Plodia interpunctella</i>	$LC_{50}= 55.197 \mu\text{L/L air}$ ; $LC_{99}= 208.107 \mu\text{L/L air}$	"
	<i>Pituranthus tortuosus</i>	<i>Tribolium confusum</i>	Mortality= 75.6% after 14days at 1.5 mL/cm <sup>2</sup>	[81]
		<i>Trogoderma granarium</i>	Mortality= 88.3% after 14days at 1.5 mL/cm <sup>2</sup>	"
		<i>Sitophilus oryzae</i>	$LC_{50}= 41.01 \text{ mg/L}$	[2]
	<i>Achillea wilhelmsii</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 2.65 \mu\text{L/L air}$	[82]
<i>Asteraceae</i>		<i>Tribolium castaneum</i>	$LC_{50}= 10.02 \mu\text{L/L air}$	"
	<i>Aphyllocladus decussatus</i>	<i>Sitophilus zeamais</i>	$LC_{50}= 212.12 \mu\text{L/L air}$	[83]
	<i>Artemisia absinthium</i>	<i>Oryzaephilus surinamensis</i>	$LC_{50}= 5.22 \mu\text{L/L air}$ ; $LC_{95}= 45.18 \mu\text{L/L air}$ ; $LT_{50}= 10.750\text{h}$ and $LT_{95}= 18.258\text{h}$ at 56.82 $\mu\text{L/L air}$	[9]
		<i>Tribolium castaneum</i>	$LC_{50}= 35.18 \mu\text{L/L air}$ ; $LC_{95}= 59.87 \mu\text{L/L air}$ ; $LT_{50}= 75.576\text{h}$ and $LT_{95}= 98.167\text{ h}$ at 56.82 $\mu\text{L/L air}$	"
	<i>Artemisia anethoides</i>	<i>Lasioderma serricorne</i>	$LC_{50}= 8.04 \text{ mg/L air}$	[10]
		<i>Tribolium castaneum</i>	$LC_{50}= 13.05 \text{ mg/L air}$	"
	<i>Artemisia dracunculus</i>	<i>Tribolium castaneum</i>	$LC_{50}= 67.2 \mu\text{L/L air}$	[84]
		<i>Tribolium confusum</i>	$LC_{50}= 178.4 \mu\text{L/L air}$	"
	<i>Artemisia dubia</i>	<i>Liposcelis bostrychophila</i>	$LC_{50}= 0.74 \text{ mg/L air}$	[85]
		<i>Tribolium castaneum</i>	$LC_{50}= 49.54 \text{ mg/L air}$	"
	<i>Artemisia herba-alba</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 99.35 \mu\text{L/L air}$	[86]
		<i>Oryzaephilus surinamensis</i>	$LC_{50}= 3.05 \mu\text{L/L air}$ ; $LC_{95}= 13.41 \mu\text{L/L air}$ ; $LT_{50}= 3.725\text{h}$ and $LT_{95}= 7.119\text{h}$ at 56.82 $\mu\text{L/L air}$	[9]
		<i>Rhizopertha dominica</i>	$LC_{50}= 76.48 \mu\text{L/L air}$	[86]
		<i>Tribolium castaneum</i>	$LC_{50}= 27.76 \mu\text{L/L air}$ ; $LC_{95}= 37.63 \mu\text{L/L air}$ ; $LT_{50}= 17.308\text{h}$ and $LT_{95}= 35.57 \text{ h}$ at 56.82 $\mu\text{L/L air}$	[9]
<i>Asteraceae</i>	<i>Artemisia judaica</i>	<i>Sitophilus oryzae</i>	$LC_{50}= 564.40 \mu\text{L/L air}$	[86]
	<i>Artemisia khorassanica</i>	<i>Plodia interpunctella</i>	$LC_{50}= 29.97 \text{ mg/L}$	[2]
	<i>Artemisia monosperma</i>	<i>Sitophilus oryzae</i>	$LC_{50}= 9.6 \mu\text{L/L air}$ ; $LC_{90}= 24.85 \mu\text{L/L air}$	[87]
	<i>Artemisia scoparia</i>	<i>Callosobruchus maculatus</i>	$LC_{50}> 50 \text{ mg/L}$	[2]
	<i>Artemisia sieberi</i>	<i>Sitophilus oryzae</i>	$LC_{50}= 1.46 \mu\text{L/L air}$ ; $LC_{95}= 7.83 \mu\text{L/L air}$	[88]
		<i>Tribolium castaneum</i>	$LC_{50}= 1.87 \mu\text{L/L air}$ ; $LC_{95}= 7.52 \mu\text{L/L air}$	"
		<i>Callosobruchus maculatus</i>	$LC_{50}= 2.05 \mu\text{L/L air}$ ; $LC_{95}= 11.12 \mu\text{L/L air}$	"
		<i>Callosobruchus maculatus</i>	$LC_{50}= 1.45 \mu\text{L/L air}$ ; $LC_{95}= 7.95 \mu\text{L/L air}$	[89]
		<i>Sitophilus oryzae</i>	$LC_{50}= 1.64 \mu\text{L/L air}$ ; $LC_{95}= 8.84 \mu\text{L/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 3.86 \mu\text{L/L air}$ ; $LC_{95}= 15.55 \mu\text{L/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 4.41 \mu\text{L/L air}$ ; $LC_{95}= 18.06 \mu\text{L/L air}$	"
		<i>Callosobruchus maculatus</i>	$LC_{50}= 16.76 \mu\text{L/L air}$ ; $LC_{95}= 57.32 \mu\text{L/L air}$	"
		<i>Callosobruchus maculatus</i>	$LC_{50}= 20.31 \mu\text{L/L air}$ ; $LC_{95}= 76.03 \mu\text{L/L air}$	"
	<i>Artemisia stolonifera</i>	<i>Lasioderma serricorne</i>	$LC_{50}= 0.99 \text{ mg/L air}$ , better than single components	[12]
<i>Asteraceae</i>		<i>Tribolium castaneum</i>	$LC_{50}= 1.86 \text{ mg/L air}$ , better than single components	"
	<i>Artemisia vulgaris</i>	<i>Tribolium castaneum</i>	Mortality= adults 100%, 12, 14 and 16 days larvae 49%, 53% and 52% at 8.0 $\mu\text{L/ml}$ ; Mortality= eggs 100% at a concentration 20 $\mu\text{L/L}$ air	[90]
	<i>Aster ageratoides</i>	<i>Sitophilus zeamais</i>	$LC_{50}= 13.73 \text{ mg/L}$	[13]
	<i>Dahlia pinnata</i>	<i>Tribolium confusum</i>	$LC_{50}= 12.14 \text{ mg/L}$	"
	<i>Eupatorium glabratum</i>	<i>Sitophilus oryzae</i>	$LC_{50}= 14.10 \text{ mg/L air}$	[15]
		<i>Sitophilus zeamais</i>	$LC_{50}= 18.80 \text{ mg/L air}$	"
		<i>Sitophilus zeamais</i>	$LC_{50}= 18 \mu\text{L/mL air}$ (females 16 $\mu\text{L/mL air}$ and males 20 $\mu\text{L/mL air}$ ); $LC_{95}= 72 \mu\text{L/mL air}$ (females 59.8 $\mu\text{L/mL air}$ and males 73 $\mu\text{L/mL air}$ ); $LT_{50}= 102\text{h}$ at 25- 75 $\mu\text{L/mL air}$	[91]
		<i>Callosobruchus maculatus</i>	$LC_{50}= 1.54 \mu\text{L/L air}$	[82]
		<i>Tribolium castaneum</i>	$LC_{50}= 297.9 \mu\text{L/L air}$	"
	<i>Tagetes erecta</i>	<i>Callosobruchus maculatus</i>	$LD_{50}= 41.7 \text{ ppm}$ and $LD_{95}= 66.2 \text{ ppm}$	[17]
		<i>Sitophilus oryzae</i>	$LD_{50}= 35.2 \text{ ppm}$ and $LD_{95}= 68.8 \text{ ppm}$	"
		<i>Tribolium castaneum</i>	$LD_{50}= 43.9 \text{ ppm}$ and $LD_{95}= 57.9 \text{ ppm}$	"
	<i>Tagetes minuta</i>	<i>Callosobruchus maculatus</i>	$LD_{50}= 20.8 \text{ ppm}$ and $LD_{95}= 38.1 \text{ ppm}$	"
		<i>Sitophilus oryzae</i>	$LD_{50}= 22.5 \text{ ppm}$ and $LD_{95}= 40.3 \text{ ppm}$	"
		<i>Sitophilus zeamais</i>	No mortality	[83]
<i>Asteraceae</i>		<i>Tribolium castaneum</i>	$LD_{50}= 25.2 \text{ ppm}$ and $LD_{95}= 41.9 \text{ ppm}$	[17]
	<i>Tagetes patula</i>	<i>Callosobruchus maculatus</i>	$LD_{50}= 25.7 \text{ ppm}$ and $LD_{95}= 45.7 \text{ ppm}$	"
		<i>Sitophilus oryzae</i>	$LD_{50}= 27.6 \text{ ppm}$ and $LD_{95}= 43.7 \text{ ppm}$	"
		<i>Tribolium castaneum</i>	$LD_{50}= 30.1 \text{ ppm}$ and $LD_{95}= 46.7 \text{ ppm}$	"
	<i>Tanacetum armenum</i>	<i>Acanthoscelides obtectus</i>	$LC_{50}= 272.88 \mu\text{L/L air}$ ; $LC_{99}= 607.14 \mu\text{L/L air}$	[92]
		<i>Tribolium castaneum</i>	No mortality	"
	<i>Tanacetum nubigenum</i>	<i>Tribolium castaneum</i>	$LC_{50}= 8.32 \mu\text{L}$ in 0.25L air after 48h	[93]
<i>Asteraceae</i>	<i>Tanacetum armenum</i>	<i>Epeorus kuehniella</i>	$LC_{50}= 7.93 \mu\text{L/L air}$ after 12h; $LC_{99}= 17.65 \mu\text{L/L air}$ after 12h	[94]
	<i>Laurelia sempervirens</i>	<i>Sitophilus zeamais</i>	$LC_{50}= 177 \mu\text{L/L air}$	[18]
	<i>Brassica rapa</i>	<i>Callosobruchus maculatus</i>	$LC_{50}$ adult= 1.20 ppm; young larvae= 3.94 ppm; old larve= 3.89 ppm; pupae= 3.88 ppm	[95]
<i>Asteraceae</i>		<i>Sitophilus zeamais</i>	$LC_{50}$ adult= 1.21 ppm; young larvae= 4.64 ppm; old larvae= 5.83 ppm; pupae= 6.17 ppm	"

Plant Family	Plant species	Insect Species	Main Results	References
Chenopodiaceae	<i>Chenopodium ambrosioides</i>	<i>Sitophilus zeamais</i>	Mortality= 100% after 24h at 500 µL/L air	[96]
		<i>Tribolium confusum</i>	Mortality= 100% after 14days at 1.5 mL/cm <sup>2</sup>	[81]
		<i>Trogoderma granarium</i>	Mortality= 92% after 14days at 1.5 mL/cm <sup>2</sup>	"
		<i>Acanthoscelides obtectus</i>	LC <sub>50</sub> = 302.08 µL/L air; LC <sub>99</sub> = 628.90 µL/L air	[92]
	<i>Chenopodium botrys</i>	<i>Ephestia kuehniella</i>	LC <sub>50</sub> = 32.74 µL/L air after 12h; LC <sub>99</sub> = 63.53 µL/L air after 12h	[94]
		<i>Tribolium castaneum</i>	No mortality	"
Cupressaceae	<i>Cupressus macrocarpa</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 30.34 mg/L	[2]
	<i>Cupressus sempervirens</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 17.16 mg/L	"
	<i>Juniperus polycarpus</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 368 µL/L air after 24h	[97]
	<i>Juniperus sabina</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 302 µL/L air after 24h	"
	<i>Platycladus orientalis</i>	<i>Callosobruchus maculatus</i>	Levaes EO LC <sub>50</sub> = 2.99 µL/L air after 48h; LC <sub>95</sub> = 18.06 µL/L air after 48h; Fruit EO LC <sub>50</sub> = 3.98 µL/L air after 48h; LC <sub>95</sub> = 27.19 µL/L air after 48h	[98]
		<i>Sitophilus oryzae</i>	Leaves EO LC <sub>50</sub> = 11.08 µL/L air after 48h; LC <sub>95</sub> = 43.44 µL/L air after 48h; Fruit EO LC <sub>50</sub> = 14.27 µL/L air after 48h; LC <sub>95</sub> = 54.15 µL/L air after 48h	"
		<i>Tribolium castaneum</i>	Leaves EO LC <sub>50</sub> = 26.31 µL/L air after 48h; LC <sub>95</sub> = 114.69 µL/L air after 48h; Fruit EO LC <sub>50</sub> = 29.23 µL/L air after 48h; LC <sub>95</sub> = 197.57 µL/L air after 48h	"
	<i>Thuja occidentalis</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 43.11 mg/L	[2]
	<i>Mallotus apelta</i>	<i>Liposcelis bostrychophila</i>	LC <sub>50</sub> = 3.21 mg/L air	[22]
		<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 48.42 mg/L air	"
		<i>Lasioderma serricorne</i>	No mortality	[99]
Euphorbiaceae	<i>Ricinus communis</i>	<i>Tribolium castaneum</i>	No mortality	"
	<i>Geranium maculatum</i>	<i>Plodia interpunctella</i>	KT <sub>50</sub> = 32.6 min and KT <sub>90</sub> = 43.6 min at 40µL; RT <sub>50</sub> = 2.532 at 0.05%(w/w) in 20 g of whole grain wheat	[23]
	<i>Pelargonium</i>	<i>Rhizopertha dominica</i>	Mortality= higher for PEG EONP than for pure EO at 3%(w/w wheat)	[24]
		<i>Tribolium castaneum</i>	Mortality= higher for PEG EONP than for EOs at 0.25%(w/w wheat)	"
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> > 50 mg/L	[2]
Hypericaceae	<i>Hypericum hyssopifolium</i>	<i>Sitophilus granarius</i>	Mortality= 43% after 96h at 40 µL	[71]
		<i>Tribolium confusum</i>	Mortality= 50% after 96h at 40 µL	"
	<i>Hypericum scabrum</i>	<i>Ephestia kuehniella</i>	Mortality= 72% at 10 µL in 4L air	[100]
		<i>Sitophilus granarius</i>	Mortality= 73% at 10 µL in 4L air	"
Lamiaceae	<i>Agastache foeniculum</i>	<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 21.565 µL/L air	[101]
		<i>Oryzaephilus surinamensis</i>	LC <sub>50</sub> = 18.781 µL/L air	"
		<i>Rhizopertha dominica</i>	LC <sub>50</sub> = 14.17 µL/L air; LC <sub>95</sub> = 166.01 µL/L air; LT <sub>50</sub> = 10.05h LT <sub>95</sub> = 62.22h at 42 µL/L air	[102]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 22.24 µL/L air; LC <sub>95</sub> = 129.93 µL/L air; LT <sub>50</sub> = 12.47h and LT <sub>95</sub> = 48.66h at 42 µL/L air	"
	<i>Dracocephalum moldavica</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 2.65 mg/L air	[26]
		<i>Tribolium confusum</i>	LC <sub>50</sub> = 0.88 mg/L air	"
	<i>Hyptis spicigera</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 1.30 µL/L air; LC <sub>90</sub> = 2.84 µL/L air	[103]
		<i>Sitophilus zeamais</i>	Mortality= 68% after 2days at 2 µL/L air	[104]
	<i>Hyptis suaveolens</i>	<i>Tribolium castaneum</i>	Mortality= 3% after 2days at 2 µL/L air	"
		<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 5.53 µL/L air; LC <sub>90</sub> = 7.80 µL/L air	[103]
	<i>Hyssopus officinalis</i>	<i>Rhizopertha dominica</i>	LC <sub>50</sub> = 4.7 mg/L air	[28]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 12.0 mg/L air	"
	<i>Lavandula angustifolia</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 10.6 mg/L air	"
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 23.2 mg/L air	"
	<i>Lavandula officinalis</i>	<i>Ephestia kuehniella</i>	Mortality= 14% at 10 µL in 4L air	[100]
		<i>Sitophilus granarius</i>	Mortality= 14% at 10 µL in 4L air	"
	<i>Lavandula stoechas</i>	<i>Plodia interpunctella</i>	KT <sub>50</sub> = 35.2 min and KT <sub>90</sub> = 48.3 min at 40µL; RT <sub>50</sub> = 1.309 at 0.05%(w/w) in 20 g of whole grain wheat	[23]
		<i>Sitophilus granarius</i>	LC <sub>50</sub> = 1.6 mg/L air and LC <sub>90</sub> = 4.1 mg/L air in absence of grains; LC <sub>50</sub> = 10.9 mg/L air and LD <sub>90</sub> = 47.6 mg/L air in presence of grains	[29]
	<i>Tribolium castaneum</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 36.57 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
		<i>Lasioderma serricorne</i>	LC <sub>50</sub> >100 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	"
	<i>Rhizopertha dominica</i>	<i>Rhizopertha dominica</i>	LC <sub>50</sub> = 3.835 µL/L air	[105]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 5.66 µL/L air	"
	<i>Mentha × piperita</i>	<i>Corcyra cephalonica</i>	LC <sub>50</sub> = 39.685 µL/L air	"
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = larvae= 343.96 µL/L air	[106]
	<i>Mentha longifolia</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = adults= 85.04 µL/L air	"
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 2.05 µL/L air	[82]
	<i>Mentha longifolia subsp. capensis</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 13.05 µL/L air	"
			Mortality= 60-80% after 3days at 24 -32 µL/L air	[31]

Plant Family	Plant species	Insect Species	Main Results	References
	<i>Mentha piperita</i>	<i>Plodia interpunctella</i>	$KT_{50}= 27.1$ min and $KT_{90}= 47.2$ min at $40\mu\text{L}$ ; $RT_{50}= 3.081$ at 0.05% (w/w) in 20 g of whole grain wheat $LC_{50}= 0.135 \text{ mL/mL air}$	[23]
		<i>Sitophilus oryzae</i>	$LC_{50}= 0.421 \text{ mL/mL air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 8.46 \mu\text{L/L air}$	[99]
	<i>Mentha pulegium</i>	<i>Lasioderma serricorne</i>	$LC_{50}= 0.038 \mu\text{L/L air}; LC_{90}= 0.137 \mu\text{L/L air}$	[107]
		<i>Sitophilus granarius</i>	$LC_{50}= 11.57 \mu\text{L/L air}$	[99]
		<i>Tribolium castaneum</i>	$LC_{50}= 3.51 \mu\text{L/cm}^2$	[32]
	<i>Mentha sp.</i>	<i>Sitophilus oryzae</i>	$LD_{50}= 0.5 \mu\text{L/L}; LD_{99}= 2.4 \mu\text{L/L}$	[108]
	<i>Mentha spicata</i>	<i>Ephestia kuehniella</i>	$LD_{50}= 0.4 \mu\text{L/L}; LD_{99}= 2.1 \mu\text{L/L}$	"
	<i>Micromeria fruticosa</i>	<i>Plodia interpunctella</i>	Mortality= 24% at 10 $\mu\text{l}$ in 4L air	[100]
		<i>Ephestia kuehniella</i>	Mortality= 12% at 10 $\mu\text{l}$ in 4L air	"
	<i>Minthostachys verticillata</i>	<i>Sitophilus granarius</i>	$LC_{50}= 116.61 \mu\text{L/L air}$	[83]
	<i>Mosla soochowensis</i>	<i>Sitophilus zeamais</i>	$LC_{50}= 12.19 \text{ mg/L air}$	[34]
		<i>Sitophilus zeamais</i>	$LC_{50}= 10.26 \text{ mg/L air}$	"
	<i>Ocimum americanum</i>	<i>Tribolium confusum</i>	$LC_{50}= 0.23 \mu\text{L/L air}; LC_{90}= 0.71 \mu\text{L/L air}$	[103]
	<i>Ocimum basilicum</i>	<i>Callosobruchus maculatus</i>	Mortality= 0-100% at 5 $\mu\text{l}$ depending on genotype (i.e. 18 genotypes tested)"	[109]
		<i>Callosobruchus maculatus</i>	$LD_{50}= 1.4 \mu\text{L/L}; LD_{99}= 4.1 \mu\text{L/L}$	[108]
		<i>Ephestia kuehniella</i>	$LD_{50}= 1.2 \mu\text{L/L}; LD_{99}= 3.8 \mu\text{L/L}$	"
		<i>Plodia interpunctella</i>	Mortality=>95% at 3mL of acetone solution (10% v/v)	[110]
		<i>Sitophilus oryzae</i>	$LC_{50}= 0.014 \text{ mg/cm}^3 \text{ air}$	[35]
		<i>Sitophilus zeamais</i>	$LC_{50}= 0.106 \text{ mg/cm}^3 \text{ air}$	"
		<i>Tribolium castaneum</i>	Mortality= 86.3% after 14days at 1.5 $\text{mL/cm}^2$	[81]
		<i>Tribolium confusum</i>	Mortality= 100% after 14days at 1.5 $\text{mL/cm}^2$	"
	<i>Ocimum gratissimum</i>	<i>Trogoderma granarium</i>	$LC_{50}= 0.20$ after 24h $\mu\text{L/L air}$	[111]
		<i>Callosobruchus chinensis</i>	$LC_{50}= 0.19$ after 72h $\mu\text{L/L air}$	"
		<i>Oryzaephilus surinamensis</i>	$LC_{50}= 0.20$ after 72h $\mu\text{L/L air}$	"
		<i>Rhizopertha dominica</i>	$LC_{50}= 0.50$ after 72h $\mu\text{L/L air}$	"
		<i>Sitophilus oryzae</i>	$LC_{50}= 24.9$ after 72 $\mu\text{L/L air}$	"
	<i>Origanum acutidens</i>	<i>Tribolium castaneum</i>	Mortality= 79% at 10 $\mu\text{l}$ in 4L air	[100]
		<i>Ephestia kuehniella</i>	Mortality= 88.3 $\pm$ 1.9 third instar larvae after 72h at 2 $\mu\text{L/L air}$	[112]
		<i>Lasioderma serricorne</i>	Mortality= 100.0 $\pm$ 3.1 adults after 72h at 2 $\mu\text{L/L air}$	"
		<i>Sitophilus granarius</i>	Mortality= 93.3 $\pm$ 1.6 adults after 72h at 2 $\mu\text{L/L air}$	"
	<i>Origanum majorana</i>	<i>Acanthoscelides obtectus</i>	Mortality= 74% at 10 $\mu\text{l}$ in 4L air	[100]
		<i>Ephestia kuehniella</i>	$LC_{50}= 45.31 \mu\text{L/L air}; LC_{99}= 70.75 \mu\text{L/L air}$	[92]
		<i>Plodia interpunctella</i>	Egg Mortality: $LT_{50}= 57.98\text{h}$ and $LT_{99}= 127.82\text{h}$ at 200 $\mu\text{L/L air}$	[113]
		<i>Tribolium castaneum</i>	$LC_{50}= 3.27 \mu\text{L/L air}$ after 12h; $LC_{99}= 5.13 \mu\text{L/L air}$ after 12h	[94]
			Egg Mortality: $LT_{50}= 35.66\text{h}$ and $LT_{99}= 90.19\text{h}$ at 200 $\mu\text{L/L air}$	[113]
	<i>Origanum minutiflorum</i>	<i>Tribolium castaneum</i>	$LC_{50}= 3.32 \mu\text{g/mL}$	[114]
	<i>Origanum onites</i>	<i>Acanthoscelides obtectus</i>	$LC_{50}= 92.80 \mu\text{L/L air}; LC_{99}= 159.03 \mu\text{L/L air}$	[92]
		<i>Ephestia kuehniella</i>	$LC_{50}= 34.571 \mu\text{g/mL}$	[114]
		<i>Plodia interpunctella</i>	$LC_{50}= 209.29 \mu\text{L/L air}; LC_{99}= 360.08 \mu\text{L/L air}$	[92]
		<i>Tribolium castaneum</i>	$LC_{50}= 236.42 \mu\text{L/L air}; LC_{99}= 444.68 \mu\text{L/L air}$	[115]
			$LC_{50}= 7.52 \mu\text{L/L air}; LC_{99}= 12.72 \mu\text{L/L air}$	"
		<i>Ephestia kuehniella</i>	$LC_{50}= 9.81 \mu\text{L/L air}$ after 12h; $LC_{99}= 25.03 \mu\text{L/L air}$ after 12h	[94]
	<i>Origanum syriacum</i>	<i>Plodia interpunctella</i>	$LC_{50}= 4.06 \mu\text{L/L air}; LC_{99}= 5.77 \mu\text{L/L air}$	[115]
	<i>Origanum vulgare</i>	<i>Tribolium castaneum</i>	$LC_{50}= 52.91 \mu\text{g/mL}$	[114]
	<i>Perilla frutescens</i>	<i>Tribolium castaneum</i>	$LC_{50}= 270.71 \mu\text{L/L air}; LC_{99}= 479.09 \mu\text{L/L air}$	[92]
		<i>Dermestes maculatus</i>	$LC_{50}= 10.93 \mu\text{g/mL}$	[114]
		<i>Lasioderma serricorne</i>	$LC_{50}= 1.64 \text{ mg/L}$	[2]
		<i>Tribolium castaneum</i>	$LC_{50}= 72.75 \mu\text{g/mL}$	[114]
			$LC_{50}= 0.06 \mu\text{L/L}$ ; Larvae= 0.09 $\mu\text{L/L}$ ; Pupae= 0.16 $\mu\text{L/L}$ ; Eggs= 0.10 $\mu\text{L/L}$ . With $\text{CO}_2$ (25% or 60%) $LC_{50}$ decreased 3 and 6 times respectively	[116]
		<i>Tribolium castaneum</i>	$LC_{50}= 4.16 \text{ mg/L air}$	[117]
		<i>Sitophilus oryzae</i>	$LC_{50}= 1.21 \text{ mg/L air}$	"
	<i>Perovskia abrotanoides</i>	<i>Tribolium castaneum</i>	$LC_{50}= 4.10 \text{ mg/L air}$	"
			$LC_{50}= 18.75 \mu\text{L/L air}; LC_{95}= 53.00 \mu\text{L/L air}; LT_{50}= 11.54\text{h}$ and $LT_{95}= 17.55\text{h}$ at 32 $\mu\text{L/L air}$	[118]
		<i>Tribolium castaneum</i>	$LC_{50}= 11.39 \mu\text{L/L air}; LC_{95}= 45.46 \mu\text{L/L air}; LT_{50}= 4.53\text{h}$ and $LT_{95}= 9.21$ at 32 $\mu\text{L/L air}$	"
	<i>Perovskia atriplicifolia</i>	<i>Tribolium castaneum</i>	Mortality= 5% with 7.65 $\mu\text{L/L EO}$ and 25.8 % with 7.65 $\mu\text{L/L EO}$ combined with 230 Gy irradiation	[119]

Plant Family	Plant species	Insect Species	Main Results	References
			LD <sub>5</sub> , LD <sub>25</sub> and LD <sub>50</sub> were increased 8.5, 13.0 and 16.0 times, respectively, in combination with irradiation (900 Gy)	[120]
Rosmarinus officinalis		<i>Epeorus kuehniella</i>	LC <sub>50</sub> = 100.52 µL/L air	[80]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 0.93 µL/L air; LC <sub>90</sub> = 6.33 µL/L air	[107]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 26.71 mg/L	[2]
		<i>Tribolium castaneum</i>	Mortality= 71.6% after 24h at 27.76 µL/L of EO and 96.6% at 27.76 µL/L of polycaprolactone nanocapsules loading EO	[38]
			LC <sub>50</sub> = 1.17 µg/mL	[114]
			LC <sub>50</sub> = 103.28 µL/L air	[80]
			Mortality= 5% with 4.84 µL/L EO and 50% with 4.84 µL/L EO combined with 230 Gy irradiation	[119]
Salvia cryptantha		<i>Acanthoscelides obtectus</i>	LC <sub>50</sub> = 120.27 µL/L air; LC <sub>99</sub> = 253.34 µL/L air	[92]
Salvia fruticosa		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 372.550 µL/L air; LC <sub>99</sub> = 654.27 µL/L air	"
Salvia leriiifolia		<i>Sitophilus oryzae</i>	Cultivated crop: LC <sub>50</sub> = 15.5 µL/L air; LC <sub>90</sub> = 27.0 µL/L air	[65]
		<i>Callosobruchus maculatus</i>	Wild plant: LC <sub>50</sub> = 7.4 µL/L air; LC <sub>90</sub> = 10.1 µL/L air	[121]
		<i>Rhizopertha dominica</i>	Mortality= 100% after 24h at 5.21mg/cm <sup>2</sup> for EO from vegetative and flowering plants	[122]
		<i>Sitophilus granarius</i>	LC <sub>50</sub> = 25.87 µL/L air after 48h; LC <sub>95</sub> = 333.65 µL/L air after 48 ; LT <sub>50</sub> = 29.26h and LT <sub>95</sub> = 89.02h at 40 µL/L air	
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 56.12µL/L air after 48h; LC <sub>95</sub> = 744.76 µL/L air after 48h; LT <sub>50</sub> = 30.46h and LT <sub>95</sub> = 107.99h at 65 µL/L air	[121]
		<i>Tribolium castaneum</i>	Mortality= 100% after 24h at 5.21 mg/cm <sup>2</sup> for EO from vegetative and flowering plants	"
Salvia limbata		<i>Epeorus kuehniella</i>	Mortality= 88% and 100% after 24h at 5.21 mg/cm <sup>2</sup> for EO from vegetative and flowering plants, respectively	
Salvia microphylla		<i>Sitophilus granarius</i>	Mortality= 6% at 10 µl in 4L air	[100]
Salvia nemerosa		<i>Sitophilus oryzae</i>	Mortality= 7% at 10 µl in 4L air	"
Salvia officinalis		<i>Epeorus kuehniella</i>	LC <sub>50</sub> > 60 µL/L air	[65]
		<i>Sitophilus granarius</i>	Mortality= 0% at 10 µl in 4L air	"
		<i>Callosobruchus maculatus</i>	Mortality= 10% at 10 µl in 4L air	"
		<i>Sitophilus granarius</i>	Mortality= 100% after 48h at 4 µL/L air	[68]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 342.97 µL/L air; LC <sub>90</sub> = 614.05 µL/L air	[80]
Salvia pomifera		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 9.9 µL/L air; LC <sub>90</sub> = 17.4 µL/L air	[65]
Salvia veneris		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 311.09 µL/L air; LC <sub>90</sub> = 508.09 µL/L air	[80]
			LC <sub>50</sub> = 4.74 µg/mL	[114]
		<i>Rhizopertha dominica</i>	LC <sub>50</sub> = 4.4 µL/L air; LC <sub>90</sub> = 6.5 µL/L air	[65]
		<i>Sitophilus granarius</i>	Mortality= 90.75 ± 7.00% at 1 µL/mL air of 10% acetone solution (v/v) after 48 h	[40]
		<i>Sitophilus oryzae</i>	Mortality= 0.00% at 1 µL/mL air of 10% acetone solution (v/v) after 48 h	"
		<i>Tribolium castaneum</i>	Mortality= 97.63 ± 7.00% at 1 µL/mL air of 10% acetone solution (v/v) after 48 h	"
		<i>Tribolium confusum</i>	Mortality= 100.00 ± 0.00% at 1 µL/mL air of 10% acetone solution (v/v) after 48 h	"
Satureja hortensis		<i>Epeorus kuehniella</i>	Mortality= 0.00% at 1 µL/mL air of 10% acetone solution (v/v) after 48 h	"
		<i>Plodia interpunctella</i>	Mortality= 0.00% at 1 µL/mL air of 10% acetone solution (v/v) after 48 h	"
		<i>Sitophilus granarius</i>	Mortality= 62% at 10 µl in 4L air	[100]
Satureja thymbra		<i>Tribolium castaneum</i>	LC <sub>50</sub> = larvae= 80.9 µL/L air after 9h	[41]
		<i>Acanthoscelides obtectus</i>	LC <sub>50</sub> = larvae= 139.8 µL/L air after 9h	"
		<i>Epeorus kuehniella</i>	Mortality= 66% at 10 µl in 4L air	[100]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 192.35 µL/L air	[41]
Schizonepeta multifida		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 104.99 µL/L air; LC <sub>99</sub> = 215.41 µL/L air	[92]
			LC <sub>50</sub> = 124.75 µL/L air; LC <sub>99</sub> = 336.64 µL/L air	[115]
Sideritis perfoliata			Egg Mortality= LT <sub>50</sub> = 58.58h and LT <sub>99</sub> = 158.50h at 50 µL/L air	[113]
			LC <sub>50</sub> = 10.34 µL/L air; LC <sub>99</sub> = 21.27 µL/L air	[115]
			LC <sub>50</sub> = 13.92 µL/L air after 12h; LC <sub>99</sub> = 33.11 µL/L air after 12h	[94]
Teucrium capitatum		<i>Sitophilus oryzae</i>	Egg Mortality= LT <sub>50</sub> = 34.64h and LT <sub>99</sub> = 81.88h at 50 µL/L air	[115]
			LC <sub>50</sub> = 3.43 µL/L air; LC <sub>99</sub> = 7.72 µL/L air	[115]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> =133.85 µL/L air; LC <sub>99</sub> = 221.21 µL/L air	[92]
		<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 8.33 mg/cm <sup>3</sup>	[42]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 26.41 mg/cm <sup>3</sup>	"
		<i>Acanthoscelides obtectus</i>	Mortality= 100% at 70 and 140 µL/L air	[123]
		<i>Tribolium castaneum</i>	Mortality= 76.7% and 100% at 70 and 140 µL/L air, respectively	"
			LC <sub>50</sub> = 37.9 µL/L air; LC <sub>90</sub> = 90.9 µL/L air	[65]

Plant Family	Plant species	Insect Species	Main Results	References
Lamiaceae	<i>Teucrium polium</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 148.96 µL/L air	[124]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 360.25 µL/L air	"
	<i>Thymbra capitata</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 3.4 µL/L air; LC <sub>90</sub> = 5.8 µL/L air	[65]
	<i>Thymus daenensis</i>	<i>Ephestia kuehniella</i>	LC <sub>50</sub> = 1instar larvae= 16.30 µL/L air; 3instar larvae= 4483 µL/L air; adults= 0.191 µL/L air	[125]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 1instar larvae= 25.32 µL/L air; 3instar larvae= 3480 µL/L air; adults= 0.274 µL/L air	"
	<i>Thymus persicus</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 3.34 µL/L air	[126]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 236.9 µL/L air	"
	<i>Thymus sipyleus</i>	<i>Acanthoscelides obtectus</i>	LC <sub>50</sub> = 27.36 µL/L air; LC <sub>99</sub> = 43.05 µL/L air	[92]
		<i>Tribolium castaneum</i>	No mortality	"
	<i>Thymus vulgaris</i>	<i>Ephestia kuehniella</i>	Mortality= 24% at 10 µL in 4L air	[100]
Lauraceae		<i>Sitophilus granarius</i>	Mortality= 4% at 10 µL in 4L air	"
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 8.14 µg/mL	[114]
	<i>Vitex agnus-castus</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 39.85 mg/L	[2]
	<i>Vitex pseudo-negundo</i>	<i>Plodia interpunctella</i>	LC <sub>50</sub> = 23.05 µL/L air; LC <sub>90</sub> = 56.84 µL/L air	[87]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 31.96 µL/L air	[127]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 47.27 µL/L air	"
	<i>Zataria multiflora</i>	<i>Plodia interpunctella</i>	LC <sub>50</sub> = 1.75 µL/L air; LC <sub>90</sub> = 24.26 µL/L air	[107]
	<i>Ziziphora clinopodioides</i>	<i>Ephestia kuehniella</i>	LC <sub>50</sub> = larvae= 54.61 µL/L air; adults= 1.39 µL/L air	[128]
	<i>Cinnamomum aromaticum</i>	<i>Callosobruchus maculatus</i>	LD <sub>50</sub> = 434.69 ± 32.56 µg/cm <sup>2</sup> after 24h	[43]
	<i>Cinnamomum camphora</i>	<i>Lasioderma serricorne</i>	LC <sub>50</sub> : Flowers<3.3 mg/L air; Leaves= 2.5 mg/L air; Barks= 3.0 mg/L air	[44]
Laurales		<i>Tribolium castaneum</i>	LC <sub>50</sub> : Flowers= 5 mg/L air; Leaves<3.1 mg/L air; Barks<3.2 mg/L air	"
		<i>Trogoderma granarium</i>	Mortality= 100% after 14days at 1.5 mL/cm <sup>2</sup>	[81]
		<i>Tribolium confusum</i>	Mortality= 89% after 14days at 1.5 mL/cm <sup>2</sup>	"
	<i>Laurus azorica</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 33.7 µL/L air; LC <sub>90</sub> = 56.2 µL/L air	[65]
	<i>Laurus nobilis</i>	<i>Acanthoscelides obtectus</i>	LC <sub>50</sub> = 108.09 µL/L air; LC <sub>99</sub> = 208.55 µL/L air	[92]
		<i>Callosobruchus maculatus</i>	Mortality= 90% after 48h at 4 µL/L air	[68]
		<i>Ephestia kuehniella</i>	Egg Mortality: LT <sub>50</sub> = 46.26h and LT <sub>99</sub> = 120.23h at 200 µL/L air	[113]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 1.6 µL/L air; LC <sub>95</sub> = 13.01 µL/L air	[129]
		<i>Rhizopertha dominica</i>	LC <sub>50</sub> = 14.16 µL/L air after 12h; LC <sub>99</sub> = 31.38 µL/L air after 12h	[94]
		<i>Sitophilus oryzae</i>	Egg Mortality: LT <sub>50</sub> = 38.98h and LT <sub>99</sub> = 108.87h at 200 µL/L air	[113]
Laurales		<i>Tribolium castaneum</i>	LC <sub>50</sub> and LT <sub>50</sub> = Tunisia= 113.42 µL/L air and 19.58h; Algeria= 98.95 µL/L air and 17.58h; Marocco= 67.9 µL/L air and 14.25h	[130]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 41.7 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
			LC <sub>50</sub> = 8.0 µL/L air; LC <sub>90</sub> = 11.0 µL/L air	[65]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 47.02 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
			LC <sub>50</sub> : adults= 243.78 µL/L air and larvae= 69.63 µL/L air; LC <sub>90</sub> : adults= 685.85 µL/L air and larvae= 211.64 µL/L air	[131]
			LC <sub>50</sub> = 192.78 µL/L air; LC <sub>99</sub> = 300.13 µL/L air	[92]
			LC <sub>50</sub> and LT <sub>50</sub> = Tunisia= 217.10 µL/L air and 55.67h; Algeria= 193.95 µL/L air and 51.53h; Marocco= 172.37 µL/L air and 43.05h	[130]
	<i>Litsea cubeba</i>	<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 22.97 mg/L air	[45]
		<i>Liposcelis bostrychophila</i>	LC <sub>50</sub> = 0.73 mg/L air	"
	<i>Litsea salicifolia</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 4.4 µL/L air; LC <sub>95</sub> = 174.6 µL/L air	[46]
Monimiaceae		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 845.2 µL/L air; LC <sub>95</sub> = 1345 µL/L air	"
	<i>Laurelia sempervirens</i>	<i>Tribolium castaneum</i>	Bark EO: LC <sub>50</sub> = 1.63 µL/L air after 24h; LC <sub>90</sub> = 8.61 µL/L air after 24h; LT <sub>50</sub> = 9-3days and LT <sub>90</sub> = 17-7days at 25-100 µL/L air	[47]
Moringaceae			Leaves EO (LC <sub>50</sub> )= 1.66 µL/L air after 24h; LC <sub>90</sub> = 8.44 µL/L air after 24h; LT <sub>50</sub> = 9-4days and LT <sub>90</sub> = 19-9days at 25-100 µL/L air	"
	<i>Moringa oleifera</i>	<i>Cryptolestes ferrugineus</i>	Average Mortality= 20.44% µL/L air	[132]
		<i>Tribolium castaneum</i>	Average Mortality= 12.03% µL/L air	"
Myrtaceae		<i>Trogoderma granarium</i>	Average Mortality= 9.41% µL/L air	"
	<i>Callistemon viminalis</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 16.17 mg/L	[2]
	<i>Eucalyptus camaldulensis</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 3.97 µL/L air; LC <sub>95</sub> = 10.86 µL/L air	[133]
		<i>Cryptolestes ferrugineus</i>	Average Mortality= 23.31% µL/L air	[132]
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 22.22 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 12.06 µL/L air; LC <sub>95</sub> = 16.94 µL/L air	[133]
		<i>Trogoderma granarium</i>	Average Mortality= 17.43% µL/L air	[132]
			LC <sub>50</sub> = 57.7 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
			LC <sub>50</sub> = 33.50 µL/L air; LC <sub>95</sub> = 92.59 µL/L air	[133]
	<i>Eucalyptus citriodora</i>	<i>Tribolium castaneum</i>	Average Mortality= 14.27% µL/L air	[132]
<i>Eucalyptus floribundi</i>		<i>Oryzaephilus surinamensis</i>	LC <sub>50</sub> = 2.0 mL/L air	[48]
		<i>Rhizopertha dominica</i>	LC <sub>50</sub> = 43.54 µL/L air; LC <sub>95</sub> = 174.47 µL/L air	[134]
		<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 34.39 µL/L air; LC <sub>95</sub> = 77.07 µL/L air	"
	<i>Eucalyptus globulus</i>		Mortality= 92% after 48h at 4 µL/L air	[68]

Plant Family	Plant species	Insect Species	Main Results	References
		<i>Lasioderma serricorne</i>	$LC_{50}= 11.222 \mu\text{L/L air}$	[135]
		<i>Plodia interpunctella</i>	$KT_{50}= 8.34 \text{ min and } KT_{90}= 17.4 \text{ min at } 40\mu\text{L}$ ; $RT_{50}= 0.444 \text{ at } 0.05\%(\text{w/w}) \text{ in } 20 \text{ g of whole grain wheat}$	[23]
		<i>Rhizopertha dominica</i>	$LC_{50}= 3.529 \mu\text{L/L air}$	[135]
	<i>Eucalyptus intortexta</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 2.55 \mu\text{L/L air}; LC_{95}= 8.46 \mu\text{L/L air}$	[133]
		<i>Sitophilus oryzae</i>	$LC_{50}= 6.93 \mu\text{L/L air}; LC_{95}= 13.40 \mu\text{L/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 11.59 \mu\text{L/L air}; LC_{95}= 16.10 \mu\text{L/L air}$	"
	<i>Eucalyptus leucoxylon</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 2.76 \mu\text{L/L air}$	[136]
		<i>Sitophilus oryzae</i>	$LC_{50}= 8.48 \mu\text{L/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 13.15 \mu\text{L/L air}$	"
	<i>Eucalyptus obliqua</i>	<i>Callosobruchus chinensis</i>	$LC_{50}= 21.70 \mu\text{g/cm}^2 \text{ on } 10 \text{ cm}^2 \text{ filter paper}$	[5]
		<i>Coryca cephalonica</i>	$LC_{50}= 22.37 \mu\text{g/cm}^2 \text{ on } 10 \text{ cm}^2 \text{ filter paper}$	"
		<i>Sitophilus oryzae</i>	$LC_{50}= 30.29 \mu\text{g/cm}^2 \text{ on } 10 \text{ cm}^2 \text{ filter paper}$	"
	<i>Eucalyptus procera</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 11.722 \mu\text{L/L air after } 24\text{h}$	[49]
		<i>Tribolium castaneum</i>	$LC_{50}= 13.9 \mu\text{L/L air after } 24\text{h}$	"
	<i>Eucalyptus radiata</i>	<i>Callosobruchus maculatus</i>	Mortality= 100% after 48h 8 $\mu\text{L/L air}$	[68]
	<i>Eucalyptus sargentii</i>	<i>Callosobruchus maculatus</i>	$LC_{50}= 3.87 \mu\text{L/L air}; LC_{95}= 11.18 \mu\text{L/L air}$	[133]
		<i>Sitophilus oryzae</i>	$LC_{50}= 12.91 \mu\text{L/L air}; LC_{95}= 19.17 \mu\text{L/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 18.38 \mu\text{L/L air} LC_{95}= 27.93 \mu\text{L/L air}$	"
	<i>Melaleuca alternifolia</i>	<i>Sitophilus zeamais</i>	$LC_{50}= 8.42 \text{ mg/L air after } 24\text{h}, 7.70 \text{ mg/L air after } 48\text{h}, 6.78 \text{ mg/L air after } 72\text{h}$	[137]
	<i>Myrtus communis</i>	<i>Acanthoscelides obtectus</i>	$LC_{50}= 27.36 \mu\text{L/L air}; LC_{99}= 43.05 \mu\text{L/L air}$	[92]
		<i>Callosobruchus maculatus</i>	$LC_{50}= 49.58 \mu\text{L/L air}; LC_{99}= 76.07 \mu\text{L/L air}$	[115]
		<i>Ephestia kuhniella</i>	Mortality= 100% after 48h at 8 $\mu\text{L/L air}$	[68]
		<i>Plodia interpunctella</i>	Egg Mortality: $LT_{50}= 22.23\text{h}$ and $LT_{99}= 53.36\text{h}$ at 200 $\mu\text{L/L air}$	[113]
		<i>Sitophilus oryzae</i>	$LC_{50}= 0.6 \mu\text{L/L air}; LC_{95}= 4.00 \mu\text{L/L air}$	[129]
		<i>Tribolium castaneum</i>	$LC_{50}= 12.74 \mu\text{L/L air}; LC_{99}= 29.43 \mu\text{L/L air}$	[115]
			$LC_{50}= 15.15 \mu\text{L/L air after } 12\text{h}; LC_{99}= 29.85 \mu\text{L/L air after } 12\text{h}$	[94]
			Egg Mortality: $LT_{50}= 50.42\text{h}$ and $LT_{99}= 179.33\text{h}$ at 200 $\mu\text{L/L air}$	[113]
			$LC_{50}= 22.61 \mu\text{L/L air}; LC_{99}= 41.74 \mu\text{L/L air}$	[115]
			$LC_{50}= 27.4 \text{ mg/L}$	[2]
			$LC_{50}= 20.3 \mu\text{L/L air}; LC_{90}= 49.4 \mu\text{L/L air}$	[65]
			$LC_{50}= \text{adults}= 56.11 \mu\text{L/L air and larvae}= 183.659 \mu\text{L/L air}; LC_{90}= \text{adults}= 144.01 \mu\text{L/L air and larvae}= 656.84 \mu\text{L/L air}$	[131]
	<i>Syzygium aromaticum</i>	<i>Sitophilus oryzae</i>	$LC_{50}= 56.98 \mu\text{L/L air}; LC_{99}= 89.73 \mu\text{L/L air}$	[92]
		<i>Tribolium castaneum</i>	$LC_{50}= 41.76 \text{ mg/cm}^2 \text{ on } 6 \text{ cm}^2 \text{ filter paper}$	[30]
			$LC_{50}>100 \text{ mg/cm}^2 \text{ on } 6 \text{ cm}^2 \text{ filter paper}$	"
	<i>Syzygium cumini</i>	<i>Sitophilus oryzae</i>	$LC_{50}= 6.63 \text{ ppm at } 72\text{h}$	[138]
			$LC_{50}> 50 \text{ mg/L}$	[2]
Pinaceae	<i>Pinus longifolia</i>	<i>Callosobruchus chinensis</i>	$LC_{50}= 33.11 \mu\text{g/cm}^2 \text{ on } 10 \text{ cm}^2 \text{ filter paper}$	[5]
		<i>Coryca cephalonica</i>	$LC_{50}= 33.75 \mu\text{g/cm}^2 \text{ on } 10 \text{ cm}^2 \text{ filter paper}$	"
		<i>Sitophilus oryzae</i>	$LC_{50}= 47.88 \mu\text{g/cm}^2 \text{ on } 10 \text{ cm}^2 \text{ filter paper}$	"
Piperaceae	<i>Piper nigrum</i>	<i>Coryca cephalonica</i>	$LC_{50}= \text{larvae}= 530.53 \mu\text{L/L air}$	[106]
		<i>Sitophilus oryzae</i>	$LC_{50}= \text{adults}= 287.70 \mu\text{L/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= \text{larvae}= 14.02 \mu\text{L and adults}= 15.26 \mu\text{L}$	[79]
Poaceae	<i>Cymbopogon citratus</i>	<i>Oryzaephilus surinamensis</i>	$LC_{50}= 33.1 \mu\text{L/L air}$	[139]
		<i>Sitophilus oryzae</i>	$LC_{50}= 4.15 \mu\text{l/cm}^2$	[32]
		<i>Sitophilus zeamais</i>	$LC_{50}> 604 \mu\text{L/L air}$	[139]
		<i>Tribolium castaneum</i>	$LC_{50}= 4.2 \text{ mL/L air}$	[48]
	<i>Cymbopogon giganteus</i>	<i>Tribolium castaneum</i>	$LC_{50}= 2.1 \text{ mL/L air}$	"
	<i>Cymbopogon martinii</i>	<i>Oryzaephilus surinamensis</i>	$LC_{50}= 37.2 \mu\text{L/L air}$	[139]
		<i>Plodia interpunctella</i>	$KT_{50}= 92.6 \text{ min and } KT_{90}= 119.3 \text{ min at } 40\mu\text{L}$ ; $RT_{50}= 4.855 \text{ at } 0.05\%(\text{w/w}) \text{ in } 20 \text{ g of whole grain wheat}$	[23]
	<i>Cymbopogon nardus</i>	<i>Sitophilus zeamais</i>	$LC_{50}= 159 \mu\text{L/L air}$	[139]
		<i>Oryzaephilus surinamensis</i>	$LC_{50}= 46.9 \mu\text{L/L air}$	"
		<i>Sitophilus zeamais</i>	$LC_{50}> 604 \mu\text{L/L air}$	"
	<i>Cymbopogon schoenanthus</i>	<i>Tribolium castaneum</i>	$LC_{50}= 2.3 \text{ mL/L air}$	[48]
Ranuncolaceae	<i>Nigella sativa</i>	<i>Cryptolestes ferrugineus</i>	Average Mortality= 22.95% $\mu\text{L/L air}$	[132]
		<i>Tribolium castaneum</i>	Average Mortality= 20.06% $\mu\text{L/L air}$	"
		<i>Trogoderma granarium</i>	$LC_{50}= 4.64 \text{ at } 72\text{h ppm}$	[138]
			Average Mortality= 10.94% $\mu\text{L/L air}$	[132]
Rutaceae	<i>Atalantia guillauminii</i>	<i>Lasioderma serricorne</i>	$LC_{50}= 12.06 \text{ mg/L air}$	[50]
		<i>Liposcelis bostrychophila</i>	$LC_{50}> 16.75 \text{ mg/L air}$	"
		<i>Tribolium castaneum</i>	$LC_{50}= 17.60 \text{ mg/L air}$	"
		<i>Sitophilus oryzae</i>	$LC_{50}= 101.69 \mu\text{L/L air}; LC_{90}= 244.43 \mu\text{L/L air}$	[140]
	<i>Citrus × bergamia</i>	<i>Rhizopertha dominica</i>	$LC_{50}= 113.67 \mu\text{L/L air}; LC_{90}= 261.98 \mu\text{L/L air}$	"
			Mortality= higher for PEG EONP than for EO at 0.6% (w/w wheat)	[24]

Plant Family	Plant species	Insect Species	Main Results	References
		<i>Tribolium castaneum</i>	Mortality= higher for PEG EONP than for EO at 0.05%(w/w wheat) LC <sub>50</sub> = 29.37 mg/L LC <sub>50</sub> = 4.08*10 <sup>-3</sup> %(v/v EO/air)	" [2] [141]
	<i>Citrus aurantifolia</i>	<i>Sitophilus oryzae</i>	KT <sub>50</sub> = 68.7 min and KT <sub>90</sub> = 96.9 min at 40µL; RT <sub>50</sub> = 0.631 at 0.05%(w/w) in 20 g of whole grain wheat	[23]
	<i>Citrus aurantium</i>	<i>Tribolium confusum</i>	LC <sub>54</sub> = 33.02*10 <sup>-3</sup> %(v/v EO/air)	[141]
	<i>Citrus bergamia</i>	<i>Plodia interpunctella</i>	LC <sub>50</sub> = 9.89 mg/L LC <sub>50</sub> = 121.23 µL/L air; LC <sub>99</sub> = 260.72 µL/L air	[2] [92]
	<i>Citrus lemon</i>	<i>Tribolium confusum</i>	Egg Mortality: LT <sub>50</sub> = 52.92 h and LT <sub>99</sub> = 185.65h at 200 µL/L air	[113]
	<i>Citrus limon</i>	<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 4.05 µL/L air after 12h; LC <sub>99</sub> = 5.57 µL/L air after 12h	[92]
		<i>Acanthoscelides obtectus</i>	Egg Mortality: LT <sub>50</sub> = 50.23h and LT <sub>99</sub> = 193.26h at 200 µL/L air	[113]
		<i>Ephestia kuehniella</i>	LC <sub>50</sub> = 664.87 µL/L air; LC <sub>99</sub> = 1311.75 µL/L air	[92]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 5.09*10 <sup>-3</sup> %(v/v EO/air)	[141]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 24.13 mg/L	[2]
	<i>Citrus paradisi</i>	<i>Sitophilus oryzae</i>	Mortality: 78.8% and 81% after 24h and 48h at 28 µL/L air	[142]
	<i>Citrus reticulata</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 33.8 at 24h µL/L air and 28.2 µL/L air at 48h for pure EO; LC <sub>50</sub> = 18.1 µL/L air at 24h and 12.2 µL/L air at 48h for EO in combination with diethyl maleate	[143]
		<i>Tribolium confusum</i>	LC <sub>50</sub> = 3.49*10 <sup>-3</sup> %(v/v EO/air)	[141]
	<i>Citrus sinensis</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 201.17 µL/L air; LC <sub>99</sub> = 275.83 µL/L air	[107]
		<i>Sitophilus granarius</i>	LC <sub>50</sub> = 367.75 µL/L air; LC <sub>99</sub> = 508.91 µL/L air	"
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 19.67 mg/L	[2]
		<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 63.89 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 0.020 mg/cm <sup>3</sup> air	[35]
		<i>Tribolium confusum</i>	LC <sub>50</sub> = 35 mg/cm <sup>2</sup> on 6 cm <sup>2</sup> filter paper	[30]
	<i>Zanthoxylum armatum</i>	<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 0.130 mg/cm <sup>3</sup> air	[35]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 362.40 µL/L air; LC <sub>99</sub> = 442.69 µL/L air	[107]
		<i>Stegobium paniceum</i>	LC <sub>50</sub> = 4.03*10 <sup>-3</sup> %(v/v EO/air)	[141]
	<i>Zanthoxylum bungeanum</i>		LC <sub>50</sub> = 12.54 mg/L air	[54]
			LC <sub>50</sub> = 4.28 mg/L air	"
	<i>Zanthoxylum dissitum</i>	<i>Attagenus piceus</i>	Adults were more vulnerable to fumigant treatment than the larvae	[144]
		<i>Lasioderma serricorne</i>	Mortality= 13.3% and 3.3% for EOs from leaves and roots, respectively, at 50% hexane solution (v/v)	[145]
		<i>Tribolium castaneum</i>	Mortality= 40% and 10% for EOs from leaves and roots, respectively, at 50% hexane solution (v/v)	"
			Mortality= 13.3% for both EOs from leaves and roots at 50% hexane solution (v/v)	"
Schisandraceae	<i>Kadsura heterocarpa</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 14.04 mg/L air	[55]
Solanaceae	<i>Datura stramonium</i>	<i>Cryptolestes ferrugineus</i>	Average Mortality= 28.49% µL/L air	[132]
		<i>Tribolium castaneum</i>	Average Mortality= 18.90% µL/L air	[132]
		<i>Trogoderma granarium</i>	Average Mortality= 14.46% µL/L air	"
Verbenaceae	<i>Aloysia polystachya</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 218.65 µL/L air and 230.74 µL/L air for EOs with different origin	[83]
	<i>Caryopteris incana</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 10.05 mg/L	[56]
	<i>Lippia javanica</i> var. <i>javanica</i>	<i>Sitophilus oryzae</i>	LD <sub>50</sub> = 254 µg/cm <sup>3</sup> at 72h, 216 µg/cm <sup>3</sup> air at 120h	[57]
	<i>Lippia multiflora</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 0.47 µL/L air; LC <sub>99</sub> = 6.44 µL/L air	[103]
Vitaceae	<i>Cayratia japonica</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = 10.05 mg/L air	[58]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 15.67 mg/L air	"
Winteraceae	<i>Drimys winteri</i>	<i>Tribolium castaneum</i>	Bark EO: LC <sub>50</sub> = 10.45 µL/L air after 24; LC <sub>99</sub> = 24.86 µL/L air after 24h; LT <sub>50</sub> = 13-3days and LT <sub>90</sub> = 24-5days at 25-100 µL/L air Leaves EO: LC <sub>50</sub> = 8.96 µL/L air after 24h; LC <sub>99</sub> = 31.54 µL/L air after 24h; LT <sub>50</sub> = 11-4days and LT <sub>90</sub> = 23-9days at 25-100 µL/L air	[47]
Zingiberaceae	<i>Alpinia blepharocalyx</i>	<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 3.83 mg/L air	[59]
	<i>Alpinia conchigera</i>	<i>Sitophilus zeamais</i>	LC <sub>50</sub> = adults= 85 µL/L, eggs> 593 µL/L, larvae= 437 µL/L, pupae= 278 µL/L	[146]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = adults= 73 µL/L, eggs> 593 µL/L, larvae= 196 µL/L, pupae= 414 µL/L	"
	<i>Amomum maximum</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 23.09 mg/L air	[60]
	<i>Amomum tsao-ko</i>	<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 8.70 mg/L air	"
	<i>Curcuma zedoaria</i>	<i>Tribolium castaneum</i>	LC <sub>50</sub> = 5.85 mg/L air	"
		<i>Sitophilus zeamais</i>	Mortality= 6% at 593 µL/L	[146]
		<i>Tribolium castaneum</i>	No mortality	"

Plant Family	Plant species	Insect Species	Main Results	References
	<i>Elettaria cardamomum</i>	<i>Callosobruchus maculatus</i>	LC <sub>50</sub> = 78.79 mg/cm <sup>3</sup> air after 24h and 55.27 mg/cm <sup>3</sup> air after 48 h; LC <sub>90</sub> = 175.24 mg/cm <sup>3</sup> air after 24h and 95.56 mg/cm <sup>3</sup> air after 48h	[147]
		<i>Ephestia kuehniella</i>	LC <sub>50</sub> = 1.57 mg/cm <sup>3</sup> air after 24h; LC <sub>90</sub> = 3.87 mg/cm <sup>3</sup> air after 24h; LT <sub>50</sub> 11.78-12.96h and LT <sub>90</sub> = 19.02-17.77h at 3.32-4 mg/cm <sup>3</sup>	"
		<i>Sitophilus granarius</i>	LC <sub>50</sub> = 220.76 µL/L air; LC <sub>90</sub> = 353.94 µL/L air	[80]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 482.70 mg/cm <sup>3</sup> after 24h and 402.93 mg/cm <sup>3</sup> after 48h; LC <sub>90</sub> = 891.23 mg/cm <sup>3</sup> after 24h and 600 mg/cm <sup>3</sup> after 48h	"
<i>Zingiber officinale</i>		<i>Ephestia kuehniella</i>	LC <sub>50</sub> = 258.95 µL/L air and LC <sub>90</sub> = 2831.65 µL/L air after 9h	[63]
		<i>Plodia interpunctella</i>	LC <sub>50</sub> = 69.05 µL/L air and LC <sub>90</sub> = 182.15 µL/L air after 9h	"
		<i>Sitophilus oryzae</i>	LC <sub>50</sub> = 1.18 µl/cm <sup>2</sup>	[32]
		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 374.95 µL/L air and LC <sub>90</sub> = 1124.2 µL/L air at 48h	[63]
<i>Zingiber purpureum</i>		<i>Lasioderma serricorne</i>	LC <sub>50</sub> = 9.3 mg/L air	[64]
<i>Zingiber zerumbet</i>		<i>Tribolium castaneum</i>	LC <sub>50</sub> = 13.6 mg/L air	"
		<i>Sitophilus zeamais</i>	Mortality= 4% at 593 µL/L	[146]
		<i>Tribolium castaneum</i>	No mortality	"

**Table S3** Overview of reviewed studies on EO repellence towards stored product pests.

Plant Family	Plant species	Insect Species	Main results	References
Anacardiaceae	<i>Pistacia lentiscus</i>	<i>Callosobruchus maculatus</i>	Repellency rate= 71.87% at 12 µL; 100% of Repellency at 16 µL	[68]
		<i>Rhizopertha dominica</i>	$RD_{50} = 0.01 \mu\text{L}/\text{cm}^2$	[148]
		<i>Sitophilus zeamais</i>	$RD_{50} = 0.037 \mu\text{L}/\text{cm}^2$	"
		<i>Tribolium castaneum</i>	$RD_{50} = 0.025 \mu\text{L}/\text{cm}^2$	"
Annonaceae	<i>Cananga odorata</i>	<i>Tribolium castaneum</i>	Better repellent activity than the commercial repellent IR3535 at the highest tested concentration (5 mL/g)	[149]
		<i>Callosobruchus maculatus</i>	Repellent at 10 µL EO solution (1 mg/mL)	[150]
		<i>Sitophilus oryzae</i>	Repellent at 10 µL EO solution (1 mg/mL)	"
		<i>Sitophilus oryzae</i>	Repellent at 10 µL EO solution (1 mg/mL)	"
Apiaceae	<i>Anethum graveolens</i>	<i>Plodia interpunctella</i>	100% of Repellency at 2µL of EO.	[151]
		<i>Plodia interpunctella</i>	88% of Repellency at 2µL of EO.	"
		<i>Lasioderma serricorne</i>	$RD_{50} = 0.049 \mu\text{L}/\text{cm}^2$ ; $RD_{95} = 20.948 \mu\text{L}/\text{cm}^2$	[76]
	<i>Carum carvi</i>	<i>Sitophilus oryzae</i>	$RD_{50} = 0.084 \mu\text{L}/\text{cm}^2$ ; $RD_{95} = 4.592 \mu\text{L}/\text{cm}^2$	[76]
		<i>Tribolium castaneum</i>	$RD_{50} = 0.136 \mu\text{L}/\text{cm}^2$ ; $RD_{95} = 7.684 \mu\text{L}/\text{cm}^2$	[78]
		<i>Sitophilus oryzae</i>	60.4% of Repellency at 5 µL	"
	<i>Coriandrum sativum</i>	<i>Tribolium castaneum</i>	66.4% of Repellency at 5 µL	"
		<i>Cuminum cyminum</i>	Repellent at 0.2% concentration (v/v)	[79]
		<i>Tribolium castaneum</i>	70% of repelled insects after 5h at 0.1% EO-methanol solution (v/v)	[152]
	<i>Foeniculum vulgare</i>	<i>Cryptolestes ferrugineus</i>	32% of Repellency at 2µL of EO	[153]
		<i>Plodia interpunctella</i>	41.2% of repelled insects after 5h at 0.1% EO-methanol solution (v/v)	[152]
		<i>Tenebrio molitor</i>	68% of repelled insects after 1h at 0.1% EO-methanol solution (v/v)	"
	<i>Petroselinum sativum</i>	<i>Tribolium castaneum</i>	Repellent at 0.2% concentration (v/v)	"
		<i>Plodia interpunctella</i>	9.48% of Repellency at 2µL of EO	[151]
		<i>Callosobruchus maculatus</i>	71.60% of Repellency at 2µL of EO	[153]
	<i>Prangos acaulis</i>	<i>Sitophilus oryzae</i>	83.60% of Repellency at 2µL of EO	"
		<i>Tribolium castaneum</i>	63.60% of Repellency at 2µL of EO	"
Asparagaceae	<i>Liriope muscari</i>	<i>Lasioderma serricorne</i>	Repellency= 86% at 78.63 nL/cm <sup>2</sup> of EO after 2h	[8]
		<i>Liposcelis bostrychophila</i>	Repellency= 96% at 6.32 nL/cm <sup>2</sup> of EO after 4h	"
		<i>Tribolium castaneum</i>	Repellency= 92% at 15.73 nL/cm <sup>2</sup> of EO after 2h	"
Asteraceae	<i>Achillea millefolium</i>	<i>Plodia interpunctella</i>	32% of Repellency at 2µL of EO	[151]
		<i>Plodia interpunctella</i>	60% of Repellency at 2µL of EO	"
		<i>Artemisia absinthium</i>	64% of Repellency at 2µL of EO	[153]
	<i>Artemisia anethoides</i>	<i>Plodia interpunctella</i>	Percentage Repellency= 60 after 4 h at 15.73nL/cm <sup>2</sup>	[10]
		<i>Tribolium castaneum</i>	Percentage Repellency= 100 after 2 and 4h at 15.73nL/cm <sup>2</sup>	"
		<i>Artemisia dracunculus</i>	40% of Repellency at 2µL of EO.	[151]
	<i>Artemisia dubia</i>	<i>Plodia interpunctella</i>	Repellent activity similar to DEET at the highest tested concentration of 63.16 nL/cm <sup>2</sup>	[85]
		<i>Liposcelis bostrychophila</i>	Repellent activity similar to DEET at five tested concentrations after 2 h exposure	"
		<i>Tribolium castaneum</i>		
	<i>Artemisia scoparia</i>	<i>Callosobruchus maculatus</i>	Mean repelled insects= 48.57%	[88]
		<i>Sitophilus oryzae</i>	Mean repelled insects= 62.01%	"
		<i>Tribolium castaneum</i>	Mean repelled insects= 63.80%	"
	<i>Artemisia stolonifera</i>	<i>Lasioderma serricorne</i>	Repellency > 80% at 39.32 nL/cm <sup>2</sup>	[12]
		<i>Tribolium castaneum</i>	Repellency > 80% at 39.32 nL/cm <sup>2</sup>	"
		<i>Artemisia vulgaris</i>	75% of repelled insects at 0.6 µL/mL after 3h	[90]
	<i>Eupatorium glabratum</i>	<i>Tribolium castaneum</i>	Repellence index= 0.72 (females) at 0.4 µL oil/cm <sup>2</sup> ; 0.56 (males) at 0.4 µL oil/cm <sup>2</sup>	[91]
		<i>Sitophilus zeamais</i>	Repellency >90% at 78.63 nL/cm <sup>2</sup>	[16]
		<i>Laggera pterodonta</i>	Repellency >90% at 63.17 nL/cm <sup>2</sup>	"
	<i>Tagetes erecta</i>	<i>Lasioderma serricorne</i>	45% of oviposition reduction at 70,000 ppm	[17]
		<i>Liposcelis bostrychophila</i>	Repellency= 95% at 5 µL/g grain	[149]
		<i>Tribolium castaneum</i>	88% of oviposition reduction at 70,000 ppm	[17]
		<i>Tagetes lucida</i>	45% of oviposition reduction at 70,000 ppm	"
		<i>Tagetes minuta</i>	Repellency= 74-87% 20 µL/plate; FDI= 83-94% at 0.25L air	[93]
	<i>Tagetes patula</i>	<i>Tribolium castaneum</i>		
		<i>Tanacetum nubigenum</i>		
Atherospermataceae	<i>Laurelia sempervirens</i>	<i>Sitophilus zeamais</i>	Repellence index (RI)= 0.46 mL/kg grain (RI < 1 repellent)	[18]
Chenopodiaceae	<i>Chenopodium ambrosioides</i>	<i>Callosobruchus chinensis</i>	Strong oviposition repellence	[154]
		<i>Callosobruchus maculatus</i>	84.21% FDI at 5% concentration	[19]
	<i>Clausena pentaphylla</i>	<i>Callosobruchus chinensis</i>	Strong oviposition repellence	[154]
			100% FDI at 5% EO concentration	[19]
Cucurbitaceae	<i>Citrullus colocynthis</i>	<i>Rhizopertha dominica</i>	FDI= 27.17-52.59% with 2 mL of 2-6% EO-acetone solution	[155]
		<i>Tribolium castaneum</i>	FDI= 37.22-71.74% with 2 mL of 2-6% EO-acetone solution	"

Plant Family	Plant species	Insect Species	Main results	References
		<i>Trogoderma granarium</i>	FDI= 32.54-62.83% with 2 mL of 2-6% EO-acetone solution	"
Cupressaceae	<i>Cupressus lusitanica</i>	<i>Acanthoscelides obtectus</i>	Repellency= 32.2% at 0.2% v/w after 30 days	[20]
		<i>Sitophilus zeamais</i>	Repellency= 49.30% at 0.2% v/w after 30 days	"
		<i>Tribolium castaneum</i>	Repellency= 13.2% at 0.2% v/w after 30 days	"
	<i>Juniperus formosana</i>	<i>Liposcelis bostrychophila</i>	Repellency > 70% at 63.17 nL/cm <sup>2</sup> after 4 h	[21]
	<i>Juniperus polycarpus</i>	<i>Tribolium castaneum</i>	Repellency > 90% at 63.17 nL/cm <sup>2</sup> after 4 h	"
	<i>Juniperus sabina</i>	<i>Tribolium castaneum</i>	96% of repelled insects at 15 µL/mL EO-acetone solution after 24h	[97]
			82.67% of repelled insects at 15 µL/mL EO-acetone solution after 24h	"
Euphorbiaceae	<i>Croton malambo</i>	<i>Tribolium castaneum</i>	92 and 86% of repelled insects at 0.2µL/cm <sup>2</sup> after 2 and 4 hours respectively	[156]
	<i>Ricinus communis</i>	<i>Lasioderma serricorne</i>	50% of repelled insects after 5h at 0.214 µL/cm <sup>2</sup>	[99]
		<i>Tribolium castaneum</i>	Repellency of 60-80% during 1-24h of exposure at 0.314 µL/cm <sup>2</sup>	"
Geraniaceae	<i>Pelargonium</i>	<i>Rhizopertha dominica</i>	FDI= 39-83% depending on formulation (EO vs EO-nanoparticles), the EOs alone produced the higher FDI	[24]
		<i>Tribolium castaneum</i>	FDI= 3-60% depending on formulation (EO vs EO-nanoparticles), the EOs alone produced the higher FDI	"
Lamiaceae	<i>Adhatoda vasica</i>	<i>Callosobruchus chinensis</i>	Repellency= 36% at 1 µL/mL	[154]
		<i>Callosobruchus maculatus</i>	Repellency= 36% at 1 µL/mL	"
	<i>Hypericum hemsleyanum</i>	<i>Tribolium castaneum</i>	Repellency of 70% after 72h at 31.5 µg/cm <sup>2</sup>	[157]
	<i>Hyptis spicigera</i>	<i>Sitophilus oryzae</i>	Repellency= 62.5% after 2h (0.031, 0.062, 0.125, and 0.251 µL/cm <sup>2</sup> )	[27]
	<i>Hyptis suaveolens</i>	<i>Callosobruchus maculatus</i>	Repellence of 85% after 5h at 9.2 mg/cm <sup>2</sup>	[28]
		<i>Rhizopertha dominica</i>	Repellence of 70% after 5h at 9.2 mg/cm <sup>2</sup>	"
		<i>Sitophilus oryzae</i>	Repellence of 69% after 5h at 9.2 mg/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	Repellence of 44% after 5h at 9.2 mg/cm <sup>2</sup>	"
	<i>Hyssopus officinalis</i>	<i>Plodia interpunctella</i>	Repellency= 7.69% at 2 µL	[151]
	<i>Laurus nobilis</i>	<i>Cryptolestes ferrugineus</i>	Repellence= 60% after 5h at 0.1% v/v	[152]
		<i>Sitophilus zeamais</i>	Repellence= 13.7% after 5h at 0.1% v/v	"
		<i>Tenebrio molitor</i>	Repellence= 75% after 1h at 0.1% v/v	"
	<i>Lavandula angustifolia</i>	<i>Plodia interpunctella</i>	62% of Repellency at 2µL of EO.	[151]
		<i>Sitophilus granarius</i>	FDI= 8.9% at 1.125 mg/disk	[29]
	<i>Lavandula hybrida</i>	<i>Cryptolestes ferrugineus</i>	no Repellency at 0.1 % = v/v	[152]
		<i>Sitophilus zeamais</i>	Repellency= 50% after 5h at 0.1% v/v	"
		<i>Tenebrio molitor</i>	no Repellency at 0.1% v/v	"
	<i>Melissa officinalis</i>	<i>Plodia interpunctella</i>	Repellency= 40% at 2 µL	[151]
	<i>Menta piperita</i>	<i>Tenebrio molitor</i>	Luring effect in Y-tube and wind tunnel	[158]
	<i>Mentha haplocalyx</i>	<i>Tribolium castaneum</i>	Repellency of 83% after 72h at 31.5µg/cm <sup>2</sup>	[157]
	<i>Mentha longifolia subsp. <i>capensis</i></i>	<i>Sitophilus zeamais</i>	Re repellency= 90–100% at 24–32µL/cm <sup>2</sup>	[31]
	<i>Mentha piperita</i>	<i>Plodia interpunctella</i>	Repellency= 56% at 5 µL	[151]
		<i>Sitophilus oryzae</i>	Repellency= 55.2% at 5µL	[78]
		<i>Tribolium castaneum</i>	Repellency= 61.2% at 5 µL	"
	<i>Mentha pulegium</i>	<i>Lasioderma serricorne</i>	RD <sub>50</sub> = 0.010 µL/cm <sup>2</sup> ; RD <sub>95</sub> = 0.526 µL/cm <sup>2</sup>	[99]
		<i>Tribolium castaneum</i>	RD <sub>50</sub> = 0.015 µL/cm <sup>2</sup> ; RD <sub>95</sub> = 0.370 µL/cm <sup>2</sup>	"
	<i>Mentha viridis</i>	<i>Rhizopertha dominica</i>	Repellency of 94.6% at 1.75% w/w (treated packaging)	[159]
		<i>Sitophilus granarius</i>	Repellency of 100% at 1.75% w/w (treated packaging)	"
		<i>Stegobium paniceum</i>	Repellency of 96.5% at 1.75% w/w (treated packaging)	"
		<i>Tribolium castaneum</i>	Repellency of 99.7% at 1.75% w/w (treated packaging)	"
	<i>Ocimum canum</i>	<i>Sitophilus oryzae</i>	Repellency= 33.7% after 2h (0.031, 0.062, 0.125, and 0.251 µL/cm <sup>2</sup> )	[27]
	<i>Ocimum gratissimum</i>	<i>Callosobruchus chinensis</i>	Repellency= 93% after 24h at 0.2 µL/g grain	[111]
		<i>Rhizopertha dominica</i>	Repellency= 79% after 24h at 0.2 µL/g grain	"
		<i>Sitophilus oryzae</i>	Repellency= 100% after 24h at 0.2 µL/g grain	"
		<i>Tribolium castaneum</i>	Repellency= 78% after 24h at 0.2 µL/g grain	"
	<i>Perilla frutescens</i>	<i>Lasioderma serricorne</i>	Repellency > 70% after 2 and 4h at 7.86nL/cm <sup>2</sup>	[37]
		<i>Tribolium castaneum</i>	Repellency > 70% after 2 and 4h at 7.86nL/cm <sup>2</sup>	[117]
		<i>Plodia interpunctella</i>	Repellency > 90% after 2 and 4h at 7.86nL/cm <sup>2</sup>	[37]
	<i>Rosmarinus officinalis</i>	<i>Plodia interpunctella</i>	Repellency= 100% at 5 µL	[151]
	<i>Salvia multicaulis</i>	<i>Plodia interpunctella</i>	Repellency= 80% at 5 µL	"
	<i>Salvia officinalis</i>	<i>Callosobruchus maculatus</i>	Repellency rate= 82.18%(4-16 µL tested)	[68]
	<i>Satureja hortensis</i>	<i>Plodia interpunctella</i>	Repellency= 32% at 2µL	[151]
		<i>Ephesia kuehniella</i>	Repellency= 80% at 3.2µL/L air	[41]
		<i>Plodia interpunctella</i>	Repellency= 55% at 3.2µL/L air	"
		<i>Tribolium castaneum</i>	Repellency= 82.5% at 3.2µL/L air	"
	<i>Teucrium polium</i>	<i>Callosobruchus maculatus</i>	Repellency= 52% at 3 µL/mL acetone	[124]
		<i>Tribolium castaneum</i>	Repellency= 60% at 3 µL/mL acetone	"

Plant Family	Plant species	Insect Species	Main results	References
Lauraceae	<i>Thymus daenensis</i>	<i>Ephestia kuehniella</i>	Repellency= 64.4% at 22.22 µL/L air	[125]
		<i>Plodia interpunctella</i>	Repellency= 85.8% at 2.77 µL/L air	"
	<i>Thymus vulgaris</i>	<i>Plodia interpunctella</i>	Repellency= 93.33% at 2µL	[151]
	<i>Ziziphora clinopodioides</i>	<i>Ephestia kuehniella</i>	Repellency of 89.89% at 8,000 ppm	[128]
		<i>Plodia interpunctella</i>	Repellency= 68% at 2µL	[151]
	<i>Cinnamomum aromaticum</i>	<i>Callosobruchus maculatus</i>	Dose-dependent effect on oviposition deterrence. Egg laying reduction of 60-11.7% at 62.5-7.86 µg/cm <sup>2</sup>	[43]
	<i>Cinnamomum camphora</i>	<i>Ephestia kuehniella</i>	Treated packaging penetration reduced of 64%	[160]
Lauraceae	<i>Laurus nobilis</i>	<i>Sitotroga cerealella</i>	Treated packaging penetration reduced of 63%	"
		<i>Callosobruchus maculatus</i>	Repellency rate= 72.75%(4-16 µL tested)	[68]
		<i>Ephestia kuehniella</i>	Repellency= 84.2% at 2 µL/L air	[129]
	<i>Litsea cubeba</i>	<i>Lasioderma serricorne</i>	RD <sub>50</sub> = 37.84 µL/cm <sup>2</sup>	[161]
		<i>Rhizopertha dominica</i>	RD <sub>50</sub> : Tunisia= 0.036; Algeria= 0.033; Morocco= 0.013 µL/cm <sup>2</sup>	[130]
	<i>Litsea salicifolia</i>	<i>Tribolium castaneum</i>	RD <sub>50</sub> : Tunisia= 0.139; Algeria= 0.096; Morocco= 0.045 µL/cm <sup>2</sup>	"
		<i>Lasioderma serricorne</i>	Repellencies of 78% and 82%(after 2-4h) at 78.63 nL/cm <sup>2</sup>	[45]
		<i>Liposcelis bostrychophila</i>	Repellencies of 84% and 78%(after 2-4h) at 31.58 nL/cm <sup>2</sup>	"
Meliaceae	<i>Azadiracta indica</i>	<i>Sitophilus zeamais</i>	Repellency of 72-100% after 5h at 0.16-0.63 µg/cm <sup>2</sup>	[46]
		<i>Tribolium castaneum</i>	FDI= 29.63% at 10% EO concentration	"
		<i>Tribolium castaneum</i>	Repellency of 76-100% after 5h at 0.16-0.63 µg/cm <sup>2</sup>	"
	<i>Melia azadiracta</i>	<i>Tribolium castaneum</i>	FDI= 84.62% at 10% EO concentration	"
		<i>Rhizopertha dominica</i>	FDI= 60% at 2 mL of 2-6% EO-acetone solution	[155]
		<i>Tribolium castaneum</i>	FDI= 80% at 2 mL of 2-6% EO-acetone solution	"
		<i>Trogoderma granarium</i>	FDI= 40% at 2 mL of 2-6% EO-acetone solution	"
Monimiaceae	<i>Laurelia sempervirens</i>	<i>Rhizopertha dominica</i>	FDI= 55% at 2 mL of 2-6% EO-acetone solution	"
		<i>Tribolium castaneum</i>	FDI= 70% at 2 mL of 2-6% EO-acetone solution	"
Moringaceae	<i>Moringa oleifera</i>	<i>Tribolium castaneum</i>	FDI= 35% at 2 mL of 2-6% EO-acetone solution	"
		<i>Cryptolestes ferrugineus</i>	Repellency= 93% after 4h at 0.032µL/cm <sup>2</sup>	[47]
		<i>Tribolium castaneum</i>	Repellency= 97% after 4h at 0.032µL/cm <sup>2</sup>	"
Myrtaceae	<i>Eucalyptus</i>	<i>Tribolium castaneum</i>	FDI= 45.72%	[162]
		<i>Oryzaephilus surinamensis</i>	FDI= 33.8%	"
		<i>Rhizopertha dominica</i>	FDI= 15.4%	"
	<i>Eucalyptus camaldulensis</i>	<i>Tenebrio molitor</i>	Repellent in Y-tube and wind tunnel	[158]
		<i>Orizaephilus surinamensis</i>	55% of repelled insects at 0.08 µL/cm <sup>2</sup> after 24h	[163]
		<i>Rhizopertha dominica</i>	58.75% of repelled insects at 0.08 µL/cm <sup>2</sup> after 24h	"
	<i>Eucalyptus citriodora</i>	<i>Cryptolestes ferrugineus</i>	FDI= 72.44%	[162]
		<i>Tribolium castaneum</i>	FDI= 37.5%	"
		<i>Trogoderma granarium</i>	FDI= 18%	"
	<i>Eucalyptus floribundi</i>	<i>Tribolium castaneum</i>	Repellency= 90% at 5 µL/g grain	[149]
		<i>Oryzaephilus surinamensis</i>	FDI= 79.25% at 75 µL/L air	[134]
		<i>Rhizopertha dominica</i>	FDI= 88.89% at 140 µL/L air	"
Myrtaceae	<i>Eucalyptus globulus</i>	<i>Tribolium castaneum</i>	FDI= 81.11% at 75 µL/L air	"
		<i>Callosobruchus maculatus</i>	FDI= 74.61% at 140 µL/L air	"
		<i>Callosobruchus maculatus</i>	Repellency rate= 81.31%(4-16 µL tested)	[68]
	<i>Eucalyptus radiata</i>	<i>Callosobruchus maculatus</i>	Repellency rate= 81.31%(4-16 µL tested)	"
		<i>Acanthoscelides obtectus</i>	Repellency= 49.3% at 0.2% v/w after 30 days	[20]
		<i>Sitophilus zeamais</i>	Repellency= 17.8% at 0.2% v/w after 30 days	"
	<i>Eucalyptus saligna</i>	<i>Tribolium castaneum</i>	Repellency= 33.6% at 0.2% v/w after 30 days	"
		<i>Callosobruchus maculatus</i>	Repellency rate= 74.75%(4-16 µL tested)	[68]
		<i>Ephestia kuehniella</i>	Repellency= 61.3% at 2 µL/L air	[129]
Myrtaceae	<i>Myrtus communis</i>	<i>Ephestia kuehniella</i>	Treated packaging penetration reduced of 76%	[160]
		<i>Lepinotus reticulatus</i>	No Repellency (used as negative control)	[164]
		<i>Liposcelis bostrychophila</i>	Repellent (used as negative control)	"
	<i>Syzygium aromaticum</i>	<i>Liposcelis brunnea</i>	Repellent (used as negative control)	"
		<i>Liposcelis corrodens</i>	No Repellency (used as negative control)	"
		<i>Liposcelis decolor</i>	No Repellency (used as negative control)	"
	<i>Eucalyptus citriodora</i>	<i>Liposcelis entomophila</i>	Repellent (used as negative control)	"
		<i>Liposcelis paeta</i>	Repellent (used as negative control)	"
		<i>Sitotroga cerealella</i>	Treated packaging penetration reduced of 75%	[160]
Piperaceae	<i>Piper nigrum</i>	<i>Tribolium castaneum</i>	Repellent (used as negative control)	[138]
		<i>Tribolium castaneum</i>	Repellent at 0.2% v/v	[79]
Poaceae	<i>Cymbopogon citratus</i>	<i>Oryzaephilus surinamensis</i>	RD <sub>50</sub> = 0.05 µL/cm <sup>2</sup>	[139]
		<i>Sitophilus zeamais</i>	RD <sub>50</sub> = 0.06 µL/cm <sup>2</sup>	"
	<i>Cymbopogon martinii</i>	<i>Oryzaephilus surinamensis</i>	RD <sub>50</sub> = 0.07 µL/cm <sup>2</sup>	"
		<i>Sitophilus zeamais</i>	RD <sub>50</sub> = 0.03 µL/cm <sup>2</sup>	"
	<i>Cymbopogon nardus</i>	<i>Oryzaephilus surinamensis</i>	RD <sub>50</sub> = 0.03 µL/cm <sup>2</sup>	"
Poaceae	<i>Cymbopogon citratus</i>	<i>Sitophilus zeamais</i>	RD <sub>50</sub> = 0.04 µL/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	Repellency= 95% at 5 µL/g grain	[149]

Plant Family	Plant species	Insect Species	Main results	References
Ranuncolaceae	<i>Cymbopogon distans</i>	<i>Liposcelis bostrychophila</i>	Repellency of 88% and 64% at 26 and 13 nL/cm <sup>2</sup>	[165]
		<i>Tribolium castaneum</i>	Repellency of 96% and 90% at 26 and 13 nL/cm <sup>2</sup>	"
	<i>Cymbopogon nardus</i>	<i>Tribolium castaneum</i>	50% of infestation reduction at 0.2 g/m <sup>2</sup> carton board	[166]
Rosaceae	<i>Nigella sativa</i>	<i>Cryptolestes ferrugineus</i>	FDI= 42.98%	[162]
		<i>Tribolium castaneum</i>	87.33% of repelled insects at 0.08% EO concentration	[138]
		<i>Trogoderma granarium</i>	FDI= 39.2%	[162]
			FDI= 16.8%	"
Rutaceae	<i>Prunus amygdalus</i>	<i>Rhizopertha dominica</i>	Repellency of 99.2% at 1.75% w/w (treated packaging)	[159]
		<i>Sitophilus granarius</i>	Repellency of 100% at 1.75% w/w (treated packaging)	"
		<i>Stegobium paniceum</i>	Repellency of 97.8% at 1.75% w/w (treated packaging)	"
		<i>Tribolium castaneum</i>	Repellency of 100% at 1.75% w/w (treated packaging)	"
	<i>Atalantia guillauminii</i>	<i>Lasioderma serricorne</i>	Repellency= 82-64% after 4h at 78.63 and 15.73 nL/cm <sup>2</sup>	[50]
		<i>Liposcelis bostrychophila</i>	Repellency= 92-88% after 4h at 31.58 and 6.32 nL/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	Repellency=> 82% at 3.15-78.63 nL /cm <sup>2</sup>	"
<i>Atalantia monophylla</i>	<i>Callosobruchus maculatus</i>	Repellency= 85.24% after 3h at 25µL	[140]	
	<i>Citrus × bergamia</i>	<i>Sitophilus oryzae</i>	Repellency= 75.24% after 3h at 25µL	"
		<i>Rhizopertha dominica</i>	FDI= 2 mg/disc, the EOs alone produced the higher FDI mg/flour disc	[24]
		<i>Tribolium castaneum</i>	FDI= 2 mg/disc PEG EO&NP more feeding deterrent than EOs mg/flour disc	"
	<i>Citrus bergamia</i>	<i>Cryptolestes ferrugineus</i>	Repellency 85% after 5h at 0.1% v/v	[152]
	<i>Sitophilus zeamais</i>	Repellency 56.3% after 5h at 0.1% v/v	"	
	<i>Tenebrio molitor</i>	no Repellency at 0.1 =% v/v	"	
	<i>Clausena anisum-olens</i>	<i>Lasioderma serricorne</i>	Repellency= 86-96% after 2-4h at 39.2 nL/cm <sup>2</sup>	[52]
	<i>Liposcelis bostrychophila</i>	Repellency= 94-92% after 2-4h at 39.2 nL/cm <sup>2</sup>	"	
	<i>Clausena pentaphylla</i>	<i>Callosobruchus maculatus</i>	FDI= 88.52% at 5% EO concentration	[19]
	<i>Dictamnus dasycarpus</i>	<i>Lasioderma serricorne</i>	Repellency= 66-67% after 2-4h at 7.86 nL/cm <sup>2</sup>	[53]
	<i>Liposcelis bostrychophila</i>	Repellency= 92-98% after 2-4h at 6.32 nL/cm <sup>2</sup>	"	
	<i>Evodia calcicola</i>	<i>Lasioderma serricorne</i>	Repellency= 34% after 4h at 39.32 nL/cm <sup>2</sup>	[167]
		<i>Liposcelis bostrychophila</i>	Repellency= 86% after 4h at 31.58 nL/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	Repellency= 90% after 4h at 7.86 nL/cm <sup>2</sup>	"
<i>Evodia trichotoma</i>	<i>Lasioderma serricorne</i>	Repellency= 76% after 4h at 39.32 nL/cm <sup>2</sup>	"	
		<i>Liposcelis bostrychophila</i>	Repellency= 94% after 4h at 31.58 nL/cm <sup>2</sup>	"
		<i>Tribolium castaneum</i>	Repellency= 86% after 4h at 7.86 nL/cm <sup>2</sup>	"
<i>Glycosmis lucida</i>	<i>Liposcelis bostrychophila</i>	Repellency= 100% at 31.58 nL/cm <sup>2</sup>	[168]	
		<i>Tribolium castaneum</i>	Repellency= 100% at 78.63 nL/cm <sup>2</sup>	"
<i>Murraya alata</i>	<i>Tribolium castaneum</i>	Repellency= 96% after 2h and 100% after 4h at 15.73 nL/cm <sup>2</sup>	[169]	
<i>Murraya euchrestifolia</i>	<i>Tribolium castaneum</i>	Repellency= 92% after 2h and 89% after 4h at 15.73 nL/cm <sup>2</sup>	"	
	<i>Murraya exotica</i>	<i>Tribolium castaneum</i>	Repellency= 66% after 2h and 76% after 4h at 15.73 nL/cm <sup>2</sup>	"
	<i>Murraya koenigii</i>	<i>Tribolium castaneum</i>	Repellency= 59% after 2h and 44% after 4h at 15.73 nL/cm <sup>2</sup>	"
	<i>Murraya kwangsiensis</i>	<i>Tribolium castaneum</i>	Repellency= 85% after 2h and 98% after 4h at 15.73 nL/cm <sup>2</sup>	"
	<i>Murraya microphylla</i>	<i>Lasioderma serricorne</i>	Repellent as the positive control DEET after 2h exposure	"
	<i>Murraya tetramera</i>	<i>Tribolium castaneum</i>	Repellency= 94% after 2h and 100% after 4h at 15.73 nL/cm <sup>2</sup>	"
	<i>Vepris heterophylla</i>	<i>Sitophilus oryzae</i>	Repellency= 42.5% after 2h (0.031, 0.062, 0.125, and 0.251 µL/cm <sup>2</sup> )	[27]
	<i>Zanthoxylum armatum</i>	<i>Lasioderma serricorne</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	[170]
	<i>Tribolium castaneum</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	"	
	<i>Zanthoxylum bungeanum</i>	<i>Stegobium paniceum</i>	Oviposition deterrent	[144]
	<i>Zanthoxylum dimorphophyllum</i>	<i>Lasioderma serricorne</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	[170]
	<i>Tribolium castaneum</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	"	
	<i>Zanthoxylum dimorphophyllum var. spinifolium</i>	<i>Lasioderma serricorne</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	"
	<i>Zanthoxylum dissitum</i>	<i>Tribolium castaneum</i>	Repellency > 90% after 4 h at 78.63 nL/ cm <sup>2</sup>	"
	<i>Lasioderma serricorne</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	"	
	<i>Zanthoxylum piasezkii</i>	<i>Tribolium castaneum</i>	Repellency > 90% after 4 h at 78.63 nL/ cm <sup>2</sup>	"
	<i>Zanthoxylum stenophyllum</i>	<i>Lasioderma serricorne</i>	Repellency > 90% after 4 h at 78.63 nL/ cm <sup>2</sup>	"
	<i>Tribolium castaneum</i>	Repellency > 90% after 4 h at 15.73 nL/ cm <sup>2</sup>	"	

Plant Family	Plant species	Insect Species	Main results	References
Solanaceae	<i>Datura stramonium</i>	<i>Cryptolestes ferrugineus</i>	FDI= 78.79%	[162]
		<i>Tribolium castaneum</i>	FDI= 38.9%	"
		<i>Trogoderma granarium</i>	FDI= 20%	"
	<i>Eucalyptus camaldulensis</i>	<i>Rhizopertha dominica</i>	FDI= 30% with 2 mL of 2-6% EO-acetone solution	[155]
		<i>Tribolium castaneum</i>	FDI= 40% with 2 mL of 2-6% EO-acetone solution	"
		<i>Trogoderma granarium</i>	FDI= 25% with 2 mL of 2-6% EO-acetone solution	"
	<i>Nicotiana tabacum</i>	<i>Rhizopertha dominica</i>	FDI= 40% with 2 mL of 2-6% EO-acetone solution	"
		<i>Tribolium castaneum</i>	FDI= 60% with 2 mL of 2-6% EO-acetone solution	"
		<i>Trogoderma granarium</i>	FDI= 30% with 2 mL of 2-6% EO-acetone solution	"
Stemonaceae	<i>Stemona japonica</i>	<i>Tribolium castaneum</i>	no Repellency at all tested concentrations	[157]
Verbenaceae	<i>Lippia alba</i>	<i>Tribolium castaneum</i>	Repellency= 95% at 5 $\mu\text{L}/\text{g}$ grain	[149]
Winteraceae	<i>Drimys winteri</i>	<i>Tribolium castaneum</i>	Repellency= 100% after 4h at 0.2 $\mu\text{L}/\text{cm}^2$	[47]
			Repellency= 97% after 4h at 0.2 $\mu\text{L}/\text{cm}^2$	"
Zingiberaceae	<i>Amomum maximum</i>	<i>Liposcelis bostrychophila</i>	Repellency= 84% at 63.17 nL/cm <sup>2</sup>	[60]
		<i>Tribolium castaneum</i>	Repellency= 100% at 78.63 nL/cm <sup>2</sup>	"
	<i>Elettaria cardamomum</i>	<i>Callosobruchus maculatus</i>	Oviposition deterrent at highest doses	[147]
		<i>Liposcelis bostrychophila</i>	Repellency= 70% at 15.73 nL/cm <sup>2</sup>	[62]
	<i>Etlingera yunnanensis</i>	<i>Tribolium castaneum</i>	Repellency= 80% at 15.73 nL/cm <sup>2</sup>	"
		<i>Ephestia kuehniella</i>	Repellency= 80% at 3.2 $\mu\text{L}/\text{L}$ air	[63]
	<i>Zingiber officinale</i>	<i>Plodia interpunctella</i>	Repellency= 90% at 3.2 $\mu\text{L}/\text{L}$ air	"
		<i>Tribolium castaneum</i>	Repellency= 77.5% at 3.2 $\mu\text{L}/\text{L}$ air	"
Combined plant family	<i>Hyptis + Ocimum</i>	<i>Lasioderma serricorne</i>	88% of repelled insects after 4h at 39.32 nL/cm <sup>2</sup>	[64]
		<i>Tribolium castaneum</i>	86% of repelled insects after 4h at 39.32 nL/cm <sup>2</sup>	"
		<i>Sitophilus oryzae</i>	Repellency= 77.5% after 2h (0.031, 0.062, 0.125, and 0.251 $\mu\text{L}/\text{cm}^2$ )	[27]
	<i>Hyptis + Vepris</i>	<i>Sitophilus oryzae</i>	Repellency= 41.2% after 2h (0.031, 0.062, 0.125, and 0.251 $\mu\text{L}/\text{cm}^2$ )	"
	<i>Ocimum + Vepris</i>	<i>Sitophilus oryzae</i>	Repellency= 62.5% after 2h (0.031, 0.062, 0.125, and 0.251 $\mu\text{L}/\text{cm}^2$ )	"

**Table S4** Overview of reviewed studies on EO sublethal physiological effects towards stored product pests.

Plant Family	Plant species	Incesct Species	Main results	References
Amaryllidaceae	<i>Allium sativum</i>	<i>Sitophilus oryzae</i>	Reduction in number of adult developing from eggs, larvae or adults treated with EO alone or in combination with diatomaceous earth (DE).	[1]
		<i>Tribolium castaneum</i>	Reduction in number of adult developing from eggs, larvae or adults treated with EO alone or in combination with diatomaceous earth (DE).	"
Anacardiaceae	<i>Pistacia lentiscus</i>	<i>Callosobruchus maculatus</i>	Hatching inhibition at 12-16 µl of EO/50g seeds	[68]
Apiaceae	<i>Carum copticum</i>	<i>Sitophilus granarius</i>	The movement pattern of insects increases when exposed to the EO.	[4]
		<i>Tribolium confusum</i>	The movement pattern of insects increases when exposed to the EO."	"
	<i>Heracleum persium</i>	<i>Callosobruchus maculatus</i>	EO reduces the female longevity (26.98%), the fecundity rate (39.58%) and the fertility rate (29.6%).	[171]
	<i>Pimpinella anisum</i>	<i>Tribolium confusum</i>	IR= 80.4% after 14 days at 78.78 mL/cm <sup>2</sup>	[81]
		<i>Trogoderma granarium</i>	IR= 92.1% after 14 days at 78.78 mL/cm <sup>2</sup>	"
Asteraceae	<i>Artemisia herba-alba</i>	<i>Callosobruchus maculatus</i>	90% egg hatching reduction 100% egg laying reduction	[11] "
	<i>Artemisia khorassanica</i>	<i>Plodia interpunctella</i>	Biological parameters were affected by EO treatment= Larval period, body weight, pupal period, developmental time, Female longevity, fecundity and fertility. Reduction of protein, lipids and glycogen	[87]
	<i>Tagetes erecta</i>	<i>Tribolium castaneum</i>	Egg hatchability reduction of 64.4% at 70,000 ppm	[17]
	<i>Tagetes minuta</i>	<i>Tribolium castaneum</i>	Egg hatchability reduction of 91.4% at 70,000 ppm	"
	<i>Tagetes patula</i>	<i>Tribolium castaneum</i>	Egg hatchability reduction of 84.3% at 70,000 ppm	"
Atherospermataceae	<i>Laurelia sempervirens</i>	<i>Sitophilus zeamais</i>	Dose dependent reduction of adult emergence.	[18]
Chenopodiaceae	<i>Chenopodium ambrosioides</i>	<i>Tribolium confusum</i>	IR= 100% at 1.5 mL/cm <sup>2</sup>	[81]
		<i>Trogoderma granarium</i>	IR= 92.1% at 1.5 mL/cm <sup>2</sup>	"
Geraniaceae	<i>Geranium maculatum</i>	<i>Plodia interpunctella</i>	Significant decrease in egg laying	[23]
Lamiaceae	<i>Lavandula angustifolia</i>	<i>Plodia interpunctella</i>	Significant decrease in egg laying	[23]
		<i>Sitophilus granarius</i>	In the range of sublethal doses between 1.125 and 0.281 mg/disk, RGR values did not vary significantly and were similar to those of control.	[29]
	<i>Mentha piperita</i>	<i>Plodia interpunctella</i>	Significant decrease in egg laying	[23]
	<i>Mentha viridis</i>	<i>Sitophilus oryzae</i>	Dose dependent reduction of adult emergence.	"
	<i>Ocimum basilicum</i>	<i>Tribolium confusum</i>	IR= 88.7% at 1.5 mL/cm <sup>2</sup>	[81]
	<i>Salvia officinalis</i>	<i>Trogoderma granarium</i>	IR= 100% at 1.5 mL/cm <sup>2</sup>	"
	<i>Salvia verbenaca</i>	<i>Callosobruchus maculatus</i>	Hatching inhibition and total fecundity reduction at 4-16 µl of EO/50g seeds."	[68]
	<i>Vitex pseudo-negundo</i>	<i>Callosobruchus maculatus</i>	Dose dependent reduction of egg hatching, egg laying and longevity	[11]
		<i>Plodia interpunctella</i>	Biological parameters were affected by EO treatment= Larval period, Body weight, pupal period, Female longevity, Fecundity and Fertility. Reduction of protein, lipids and glycogen	[87]
	<i>Ziziphora clinopodioides</i>	<i>Ephestia kuehniella</i>	Dose dependent reduction of egg hatching and egg laying.	[128]
Lauraceae	<i>Cinnamomum aromaticum</i>	<i>Callosobruchus maculatus</i>		[43]
	<i>Cinnamomum camphora</i>	<i>Trogoderma granarium</i>	IR= 100% at 1.5 mL/cm <sup>2</sup>	[81]
	<i>Cinnamomum camphora,</i> <i>Laurus nobilis</i>	<i>Tribolium confusum</i>	IR= 82.7% at 1.5 mL/cm <sup>2</sup>	"
		<i>Callosobruchus maculatus</i>	Dose dependent reduction of egg hatching and adult emergence	[68]
Liliaceae	<i>Scilla maritima</i>	<i>Callosobruchus maculatus</i>	egg hactching, egg laying and longevity	[11]
Myrtaceae	<i>Eucalyptus camaldulensis</i>	<i>Callosobruchus maculatus</i>	EO reduces the female longevity (28.44%), the fecundity rate (27.58%) and the fertility rate (14.71%)."	[171]
	<i>Eucalyptus floribundi</i>	<i>Oryzaephilus surinamensis</i>	RGR decline at 25-75 µl/l air	[134]

<i>Eucalyptus globulus</i>	<i>Rhizopertha dominica</i> <i>Callosobruchus maculatus</i>	RGR decline at 25-75 µl/l air Hatching inhibition and total fecundity reduction at 4(8)-16 µl of EO/50g seeds."	"
<i>Eucalyptus radiata</i>	<i>Plodia interpunctella</i> <i>Callosobruchus maculatus</i>	Significant decrease in egg laying Hatching inhibition and total fecundity reduction at 4(8)-16 µl of EO/50g seeds."	[23]
<i>Myrtus communis</i>	<i>Callosobruchus maculatus</i>	Hatching inhibition and total fecundity reduction at 4(8)-16 µl of EO/50g seeds."	[68]
<i>Cymbopogon martinii</i>	<i>Plodia interpunctella</i>	Significant decrease in egg laying	"
<i>Citrus sinensis</i>	<i>Rhizopertha dominica</i>	Dose dependent reduction of progeny production; sinergism EO-inert dusts	[23] [51]

**Table S5** Overview of reviewed studies on EO mode of action towards stored product pests.

Plant Family	Plant species	Insect Species	Main Results	References
Apiaceae	<i>Carum carvi</i>	<i>Trogoderma granarium</i>	Severe effects found in mid-gut of the EO treated larvae  Ovarioles affected, after the treatment with 0.50 mL diluted EO. The germarium and the follicular epithelium of developing oocytes of the ovarioles showed faint nuclei	[172] “
Asteraceae	<i>Artemisia dracunculus</i>	<i>Tribolium castaneum</i>	Esterases activity equal to the control; GST activity positively correlated to the different EO concentrations. MFOs decrease with increase of EO concentration	[84]
		<i>Tribolium confusum</i>	Esterases activity equal to the control; GST activity negatively correlated to the different EO concentrations. MFOs increase with increase of EO concentration	“
	<i>Artemisia judaica</i>	<i>Sitophilus oryzae</i>	AChE $I_{50}$ = 16.1 mg/L; ATPases $I_{50}$ = 21.4 mg/L	[2]
	<i>Artemisia khorassanica</i>	<i>Plodia interpunctella</i>	Reduction of body protein, lipids and glycogen	[87]
	<i>Artemisia monosperma</i>	<i>Sitophilus oryzae</i>	AChE $I_{50}$ = 120 mg/L; ATPases $I_{50}$ = 24.6mg/L	[2]
Lamiaceae	<i>Origanum vulgare</i>	<i>Sitophilus oryzae</i>	AChE $I_{50}$ = 61.3 mg/L; ATPases $I_{50}$ = 6.07 mg/L	“
	<i>Vitex pseudo-negundo</i>	<i>Plodia interpunctella</i>	Reduction of body protein, lipids and glycogen	[87]
Myrtaceae	<i>Callistemon viminalis</i>	<i>Sitophilus oryzae</i>	AChE $I_{50}$ = 28.5 mg/L; ATPases $I_{50}$ = 4.69 mg/L	[2]
	<i>Melaleuca alternifolia</i>	<i>Sitophilus zeamais</i>	EO significantly inhibits the activity af GTS, CarE and AChE. RNA-Seq identified a total of 3,562 differentially expressed genes (DEGs), of which 2,836 and 726 were up-regulated and down-regulated.	[137]
Rutaceae	<i>Atalantia monophylla</i>	<i>Callosobruchus maculatus</i>	Total Protein, total esterase, AChE and GST affected at LC <sub>10</sub> -LC <sub>30</sub> EO application rates.	[140]
		<i>Sitophilus oryzae</i>	Total Protein, total esterase, AChE and GST affected at LC <sub>10</sub> -LC <sub>30</sub> EO application rates.	“
	<i>Citrus aurantifolia</i>	<i>Sitophilus oryzae</i>	AChE $I_{50}$ = 29.4 mg/L; ATPases $I_{50}$ = 11.4 mg/L	[2]
	<i>Citrus lemon</i>	<i>Sitophilus oryzae</i>	AChE $I_{50}$ = 20.2 mg/L; ATPases $I_{50}$ = 9.69 mg/L	“

## Reference list

1. Yang, F.-L., Liang, G.-W., Xu, Y.-J., Lu, Y.-Y., & Zeng, L. (2010). Diatomaceous earth enhances the toxicity of garlic, Allium sativum, essential oil against stored-product pests. *Journal of Stored Products Research*, 46(2), 118–123. doi:10.1016/j.jspr.2010.01.001
2. Abdelgaleil, S. A. M., Mohamed, M. I. E., Shawir, M. S., & Abou-Taleb, H. K. (2016). Chemical composition, insecticidal and biochemical effects of essential oils of different plant species from Northern Egypt on the rice weevil, Sitophilus oryzae L. *Journal of Pest Science*, 89(1), 219–229. doi:10.1007/s10340-015-0665-z
3. Ukeh, D. A., Oku, E. E., Udo, I. A., Nta, A. I., & Ukeh, J. A. (2012). Insecticidal effect of fruit extracts from Xylopia aethiopica and Dennettia tripetala (Annonaceae) against Sitophilus oryzae (Coleoptera: Curculionidae) | Efecto insecticida de extractos de fruta de Xylopia aethiopica y Dennettia tripetala (Annonaceae) co. *Chilean Journal of Agricultural Research*, 72(2), 195–200. doi:10.4067/S0718-58392012000200005
4. Ziae, M., Moharramipour, S., & Francikowski, J. (2014). The synergistic effects of Carum copticum essential oil on diatomaceous earth against Sitophilus granarius and Tribolium confusum. *Journal of Asia-Pacific Entomology*, 17(4), 817–822. doi:10.1016/j.aspen.2014.08.001
5. Usha Rani, P. (2012). Fumigant and contact toxic potential of essential oils from plant extracts against stored product pests. *Journal of Biopesticides*, 5(2), 120–128.
6. Polatoğlu, K., Karakoç, T. C., Yücel Yücel, Y., Güçel, S., Demirci, B., Başer, K. H. C., & Demirci, F. (2016). Insecticidal activity of edible Crithmum maritimum L. essential oil against Coleopteran and Lepidopteran insects. *Industrial Crops and Products*, 89, 383–389. doi:10.1016/j.indcrop.2016.05.032
7. Talukder, D., & Muslma Khanam, L. A. (2009). Toxicity of four plant based products against three stored product pests. *Journal of Bio-Science*, 17(1), 149–153.
8. Wu, Y., Zhang, W.-J., Wang, P.-J., Yang, K., Huang, D.-Y., Wei, J.-Y., ... Du, S.-S. (2015). Contact toxicity and repellency of the essential oil of Liriope muscari (Decn.) bailey against three insect tobacco storage pests. *Molecules*, 20(1), 1676–1685. doi:10.3390/molecules20011676
9. Bachrouch, O., Ferjani, N., Haouel, S., & Jemâa, J. M. B. (2015). Major compounds and insecticidal activities of two Tunisian Artemisia essential oils toward two major coleopteran pests. *Industrial Crops and Products*, 65, 127–133. doi:10.1016/j.indcrop.2014.12.007
10. Liang, J.-Y., Wang, W.-T., Zheng, Y.-F., Zhang, D., Wang, J.-L., Guo, S.-S., ... Zhang, J. (2017). Bioactivities and chemical constituents of essential oil extracted from Artemisia anethoides against two stored product insects. *Journal of Oleo Science*, 66(1), 71–76. doi:10.5650/jos.ess16080
11. Fatiha, R. A., Kada, R., Khelil, A., & Pujade-Villar, J. (2014). Biological control against the cowpea weevil (*Callosobruchus chinensis* L., Coleoptera: Bruchidae) using essential oils of some medicinal plants. *Journal of Plant Protection Research*, 54(3), 211–217. doi:10.2478/jppr-2014-0032
12. Zhang, W.-J., Yang, K., You, C.-X., Wang, Y., Wang, C.-F., Wu, Y., ... Deng, Z.-W. (2015). Bioactivity of essential oil from artemisia stolonifera (Maxim.) komar. and its main compounds against two stored-product insects. *Journal of Oleo Science*, 64(3), 299–307. doi:10.5650/jos.ess14187
13. Chu, S. S., Liu, S. L., Liu, Q. Z., Jiang, G. H., & Liu, Z. L. (2013). Chemical composition and insecticidal activities of the essential oil of the flowering aerial parts of Aster ageratoides. *Journal of the Serbian Chemical Society*, 78(2), 209–216. doi:10.2298/JSC120130043C
14. Keszthelyi, S., Hoffmann, R., Pónya, Z., & Pál-Fám, F. (2017). Acute and persistence effects of oil of Hippophae rhamnoides and Calendula officinalis on Sitophilus granarius (Coleoptera: Curculionidae) in stored maize. *Acta Phytopathologica et Entomologica Hungarica*, 52(2), 255–264. doi:10.1556/038.52.2017.025
15. Wang, D.-C., Qiu, D.-R., Shi, L.-N., Pan, H.-Y., Li, Y.-W., Sun, J.-Z., ... Qin, J.-C. (2015). Identification of insecticidal constituents of the essential oils of Dahlia pinnata Cav. against Sitophilus zeamais and Sitophilus oryzae. *Natural Product Research*, 29(18), 1748–1751. doi:10.1080/14786419.2014.998218
16. Guo, S.-S., Zhang, W.-J., You, C.-X., Liang, J.-Y., Yang, K., Geng, Z.-F., ... Wang, C.-F. (2017). Chemical Composition of Essential Oil Extracted from Laggera pterodonta and its Bioactivities Against Two

- Stored Product Insects. *Journal of Food Processing and Preservation*, 41(2). doi:10.1111/jfpp.12941
17. Krishna, A., Prajapati, V., Bhasney, S., Tripathi, A. K., & Kumar, S. (2005). Potential toxicity of new genotypes of Tagetes (Asteraceae) species against stored grain insect pests. *International Journal of Tropical Insect Science*, 25(2), 122–128. doi:10.1079/IJT200560
18. Torres, C., Silva, G., Tapia, M., Rodríguez, J. C., Figueroa, I., Lagunes, A., ... Tucuch, I. (2014). Insecticidal activity of laurelia sempervirens (Ruiz & pav.) tul. essential oil against sitophilus zeamais motschulsky. *Chilean Journal of Agricultural Research*, 74(4), 421–426. doi:10.4067/S0718-58392014000400007
19. Pandey, A. K., Singh, P., Palni, U. T., & Tripathi, N. N. (2014). Invivo evaluation of two essential oil based botanical formulations (EOBBFs) for the use against stored product pathogens and pests, Aspergillus species and Callosobruchus species (Coleoptera: Bruchidae). *Journal of Stored Products Research*, 59, 285–291. doi:10.1016/j.jspr.2014.09.001
20. Bett, P. K., Deng, A. L., Ogendo, J. O., Kariuki, S. T., Kamatenesi-Mugisha, M., Mihale, J. M., & Torto, B. (2017). Residual contact toxicity and repellence of Cupressus lusitanica Miller and Eucalyptus saligna Smith essential oils against major stored product insect pests. *Industrial Crops and Products*, 110, 65–74. doi:10.1016/j.indcrop.2017.09.046
21. Guo, S., Zhang, W., Liang, J., You, C., Geng, Z., Wang, C., & Du, S. (2016). Contact and Repellent Activities of the Essential Oil from Juniperus formosana against Two Stored Product Insects. *Molecules*, 21(4). doi:10.3390/molecules21040504
22. Liu, X. C., Chen, X. B., & Liu, Z. L. (2014). Gas chromatography-mass spectrometric analysis and insecticidal activity of essential oil of aerial parts of Mallotus apelta (Lour.) Muell.-Arg. (Euphorbiaceae). *Tropical Journal of Pharmaceutical Research*, 13(9), 1515–1520. doi:10.4314/tjpr.v13i9.19
23. Jesser, E. N., Werdin-González, J. O., Murray, A. P., & Ferrero, A. A. (2017). Efficacy of essential oils to control the Indian meal moth, Plodia interpunctella (Hübner) (Lepidoptera: Pyralidae). *Journal of Asia-Pacific Entomology*, 20(4), 1122–1129. doi:10.1016/j.aspen.2017.08.004
24. Werdin González, J. O., Gutiérrez, M. M., Ferrero, A. A., & Fernández Band, B. (2014). Essential oils nanoformulations for stored-product pest control - Characterization and biological properties. *Chemosphere*, 100, 130–138. doi:10.1016/j.chemosphere.2013.11.056
25. Popović, A., Šućur, J., Orčić, D., & Štrbac, P. (2013). Effects of essential oil formulations on the adult insect Tribolium castaneum (Herbst) (Col., Tenebrionidae) | Efikasnost etarskih ulja na odrasle insekte Tribolium castaneum (Herbst) (Col., Tenebrionidae). *Journal of Central European Agriculture*, 14(2), 181–193. doi:10.5513/JCEA01/14.2.1246
26. Chu, S. S., Liu, S. L., Liu, Q. Z., Liu, Z. L., & Du, S. S. (2011). Composition and toxicity of Chinese Dracocephalum moldavica (Labiatae) essential oil against two grain storage insects. *Journal of Medicinal Plant Research*, 5(21), 5262–5267.
27. Ngassoum, M. B., Ngamo Tinkeu, L. S., Ngatanko, L., Tapondjou, L. A., Lognay, G., Malaisse, F., & Hance, T. (2007). Chemical composition, insecticidal effect and repellent activity of essential oils of three aromatic plants, alone and in combination, towards sitophilus oryzae l. (coleoptera: Curculionidae). *Natural Product Communications*, 2(12), 1229–1232.
28. Tripathi, A. K., & Upadhyay, S. (2009). Repellent and insecticidal activities of Hyptis suaveolens (Lamiaceae) leaf essential oil against four stored-grain coleopteran pests. *International Journal of Tropical Insect Science*, 29(4), 219–228. doi:10.1017/S1742758409990282
29. Germinara, G. S., Di Stefano, M. G., De Acutis, L., Pati, S., Delfine, S., De Cristofaro, A., & Rotundo, G. (2017). Bioactivities of Lavandula angustifolia essential oil against the stored grain pest Sitophilus granarius. *Bulletin of Insectology*, 70(1), 129–138.
30. El-Bakry, A. M., Abdel-Aziz, N. F., Sammour, E. A., & Abdelgaleil, S. A. M. (2016). Insecticidal activity of natural plant essential oils against some stored product insects and their side effects on wheat seed germination. *Egyptian Journal of Biological Pest Control*, 26(1), 83–88.
31. Odeyemi, O. O., Masika, P., & Afolayan, A. J. (2008). Insecticidal activities of essential oil from the leaves of Mentha longifolia L. subsp. capensis against Sitophilus zeamais (Motschulsky) (Coleoptera: Curculionidae). *African Entomology*, 16(2), 220–225. doi:10.4001/1021-3589-16.2.220

32. Franz, A. R., Knaak, N., & Fiúza, L. M. (2011). Toxic effects of essential plant oils in adult *Sitophilus oryzae* (Linnaeus) (Coleoptera, Curculionidae). *Revista Brasileira de Entomologia*, 55(1), 116–120. doi:10.1590/S0085-56262011000100018
33. Derbalah, A., & Ahmed, S. (2011). Oil and powder of spearmint as an alternative to *sitophilus oryzae* chemical control of wheat grains. *Journal of Plant Protection Research*, 51(2), 145–150. doi:10.2478/v10045-011-0025-9
34. Chen, X.-B., Chen, R., & Luo, Z.-R. (2017). Chemical composition and insecticidal properties of essential oil from aerial parts of *mosla soochowensis* against two grain storage insects. *Tropical Journal of Pharmaceutical Research*, 16(4), 905–910. doi:10.4314/tjpr.v16i4.23
35. Kim, S.-I., & Lee, D.-W. (2014). Toxicity of basil and orange essential oils and their components against two coleopteran stored products insect pests. *Journal of Asia-Pacific Entomology*, 17(1), 13–17. doi:10.1016/j.aspen.2013.09.002
36. Nguemtchouin, M. G. M., Ngassoum, M. B., Chalier, P., Kamga, R., Ngamo, L. S. T., & Cretin, M. (2013). *Ocimum gratissimum* essential oil and modified montmorillonite clay, a means of controlling insect pests in stored products. *Journal of Stored Products Research*, 52, 57–62. doi:10.1016/j.jspr.2012.09.006
37. You, C. X., Yang, K., Wu, Y., Zhang, W. J., Wang, Y., Geng, Z. F., ... Liu, Z. L. (2014). Chemical composition and insecticidal activities of the essential oil of *Perilla frutescens* (L.) Britt. aerial parts against two stored product insects. *European Food Research and Technology*, 239(3), 481–490. doi:10.1007/s00217-014-2242-8
38. Khoobdel, M., Ahsaei, S. M., & Farzaneh, M. (2017). Insecticidal activity of polycaprolactone nanocapsules loaded with *Rosmarinus officinalis* essential oil in *Tribolium castaneum* (Herbst). *Entomological Research*, 47(3), 175–184. doi:10.1111/1748-5967.12212
39. Hashemi, S. M., Hosseini, B., Estaji, A., Hashemi, S. M., Hosseini, B., & Estaji, A. (2013). Chemical Composition and Insecticidal Properties of the Essential Oil of *Salvia leiriifolia* Benth (Lamiaceae) at Two Developmental Stages. *Journal of Essential Oil-Bearing Plants*, 16(6), 806–816. doi:10.1080/0972060X.2013.854493
40. Polatoğlu, K., Karakoç, Ö. C., Yücel Yücel, Y., Güçel, S., Demirci, B., Demirci, F., & Başer, K. H. C. (2017). Insecticidal activity of *Salvia veneris* Hedge. Essential oil against coleopteran stored product insects and *Spodoptera exigua* (Lepidoptera). *Industrial Crops and Products*, 97, 93–100. doi:10.1016/j.indcrop.2016.12.012
41. Maedeh, M., Hamzeh, I., Hossein, D., Majid, A., & Reza, R. K. (2011). Bioactivity of essential oil from *Satureja hortensis* (Laminaceae) against three stored-product insect species. *African Journal of Biotechnology*, 10(34), 6620–6627.
42. Liu, Z. L., Chu, S. S., & Jiang, G. H. (2011). Toxicity of *Schizonpetra multifida* essential oil and its constituent compounds towards two grain storage insects. *Journal of the Science of Food and Agriculture*, 91(5), 905–909. doi:10.1002/jsfa.4263
43. Islam, R., Khan, R. I., Al-Reza, S. M., Jeong, Y. T., Song, C. H., & Khalequzzaman, M. (2009). Chemical composition and insecticidal properties of *Cinnamomum aromaticum* (Nees) essential oil against the stored product beetle *Callosobruchusmaculatus* (F.). *Journal of the Science of Food and Agriculture*, 89(7), 1241–1246. doi:10.1002/jsfa.3582
44. Guo, S., Geng, Z., Zhang, W., Liang, J., Wang, C., Deng, Z., & Du, S. (2016). The chemical composition of essential oils from *Cinnamomum camphora* and their insecticidal activity against the stored product pests. *International Journal of Molecular Sciences*, 17(11). doi:10.3390/ijms17111836
45. Yang, K., Wang, C. F., You, C. X., Geng, Z. F., Sun, R. Q., Guo, S. S., ... Deng, Z. W. (2014). Bioactivity of essential oil of *Litsea cubeba* from China and its main compounds against two stored product insects. *Journal of Asia-Pacific Entomology*, 17(3), 459–466. doi:10.1016/j.aspen.2014.03.011
46. Ko, K., Juntarajumnong, W., & Chandrapatya, A. (2010). Insecticidal activities of essential oils from fruits of *Litsea salicifolia* Roxb. ex Wall. Against *Sitophilus zeamais* motschulsky and *tribolium castaneum* (Herbst). *Pakistan Journal of Zoology*, 42(5), 551–557.
47. Zapata, N., & Smagghe, G. (2010). Repellency and toxicity of essential oils from the leaves and bark of *Laurelia sempervirens* and *Drimys winteri* against *Tribolium castaneum*. *Industrial Crops and*

- Products*, 32(3), 405–410. doi:10.1016/j.indcrop.2010.06.005
48. Bossou, A. D., Ahoussi, E., Ruysbergh, E., Adams, A., Smagghe, G., De Kimpe, N., ... Mangelinckx, S. (2015). Characterization of volatile compounds from three *Cymbopogon* species and *Eucalyptus citriodora* from Benin and their insecticidal activities against *Tribolium castaneum*. *Industrial Crops and Products*, 76, 306–317. doi:10.1016/j.indcrop.2015.06.031
49. Nouri-Ganbalani, G., Ebadollahi, A., & Nouri, A. (2016). Chemical Composition of the Essential Oil of *Eucalyptus procera* Dehnh. and Its Insecticidal Effects Against Two Stored Product Insects. *Journal of Essential Oil-Bearing Plants*, 19(5), 1234–1242. doi:10.1080/0972060X.2016.1178606
50. Yang, K., You, C.-X., Wang, C.-F., Lei, N., Guo, S.-S., Geng, Z.-F., ... Deng, Z.-W. (2015). Chemical composition and bioactivity of essential oil of *Atalantia guillauminii* against three species stored product insects. *Journal of Oleo Science*, 64(10), 1101–1109. doi:10.5650/jos.ess15135
51. Campolo, O., Romeo, F. V., Malacrinò, A., Laudani, F., Carpinteri, G., Fabroni, S., ... Palmeri, V. (2014). Effects of inert dusts applied alone and in combination with sweet orange essential oil against *Rhyzopertha dominica* (Coleoptera: Bostrichidae) and wheat microbial population. *Industrial Crops and Products*, 61, 361–369. doi:10.1016/j.indcrop.2014.07.028
52. You, C. X., Jiang, H. Y., Zhang, W. J., Guo, S. S., Yang, K., Lei, N., ... Athanassiou, C. (2015). Contact toxicity and repellency of the main components from the essential oil of *Clausena anisum-olens* against two stored product insects. *Journal of Insect Science*, 15(1). doi:10.1093/jisesa/iev071
53. Yang, K., Guo, S.-S., Geng, Z.-F., Du, S.-S., Wang, C.-F., & Deng, Z.-W. (2015). Contact toxicity and repellency of the essential oil of *dictamnus dasycarpus* roots from China against two stored-product insects. *Chemistry and Biodiversity*, 12(6), 980–986. doi:10.1002/cbdv.201400262
54. Wang, C.-F., Zhang, W.-J., You, C.-X., Guo, S.-S., Geng, Z.-F., Fan, L., ... Wang, Y.-Y. (2015). Insecticidal constituents of essential oil derived from *Zanthoxylum armatum* against two stored-product insects. *Journal of Oleo Science*, 64(8), 861–868. doi:10.5650/jos.ess15068
55. Li, H. Q., Bai, C. Q., Chu, S. S., Zhou, L., Du, S. S., Liu, Z. L., & Liu, Q. Z. (2011). Chemical composition and toxicities of the essential oil derived from *Kadsura heteroclita* stems against *Sitophilus zeamais* and *Meloidogyne incognita*. *Journal of Medicinal Plant Research*, 5(19), 4943–4948.
56. Chu, S. S., Liu, Q. Z., Zhou, L., Du, S. S., & Liu, Z. L. (2011). Chemical composition and toxic activity of essential oil of *Caryopteris incana* against *Sitophilus zeamais*. *African Journal of Biotechnology*, 10(42), 8476–8480.
57. Kamanula, J. F., Belmain, S. R., Hall, D. R., Farman, D. I., Goyder, D. J., Mvumi, B. M., ... Stevenson, P. C. (2017). Chemical variation and insecticidal activity of *Lippia javanica* (Burm. f.) Spreng essential oil against *Sitophilus zeamais* Motschulsky. *Industrial Crops and Products*, 110, 75–82. doi:10.1016/j.indcrop.2017.06.036
58. Liu, Z. L., Yang, K., Huang, F., Liu, Q. Z., Zhou, L., & Du, S. S. (2012). Chemical composition and toxicity of the essential oil of *Cayratia japonica* against two grain storage insects. *Journal of Essential Oil Research*, 24(3), 237–240. doi:10.1080/10412905.2012.676765
59. Wang, Y., You, C. X., Yang, K., Chen, R., Zhang, W. J., Wu, Y., ... Deng, Z. W. (2015). Chemical constituents and insecticidal activities of the essential oil from *Alpinia blepharocalyx* rhizomes against *Lasioderma serricorne*. *Journal of the Serbian Chemical Society*, 80(2), 171–178. doi:10.2298/JSC140422068W
60. Guo, S.-S., You, C.-X., Liang, J.-Y., Zhang, W.-J., Yang, K., Geng, Z.-F., ... Lei, N. (2015). Essential oil of *Amomum maximum* Roxb. and its bioactivities against two stored-product insects. *Journal of Oleo Science*, 64(12), 1307–1314. doi:10.5650/jos.ess15160
61. Wang, Y., You, C.-X., Wang, C.-F., Yang, K., Chen, R., Zhang, W.-J., ... Deng, Z.-W. (2014). Chemical constituents and insecticidal activities of the essential oil from *Amomum tsaoko* against two stored-product insects. *Journal of Oleo Science*, 63(10), 1019–1026. doi:10.5650/jos.ess14087
62. Guo, S.-S., You, C.-X., Liang, J.-Y., Zhang, W.-J., Geng, Z.-F., Wang, C.-F., ... Lei, N. (2015). Chemical composition and bioactivities of the essential oil from *Etlingera yunnanensis* against two stored product insects. *Molecules*, 20(9), 15735–15747. doi:10.3390/molecules200915735
63. Maedeh, M., Hamzeh, I., Hossein, D., Majid, A., & Reza, R. K. (2012). Bioactivity of essential oil from *zingiber officinale* (zingiberaceae) against three stored-product insect species. *Journal of Essential*

- Oil-Bearing Plants*, 15(1), 122–133. doi:10.1080/0972060X.2012.10644028
64. Wang, Y., You, C. X., Yang, K., Wu, Y., Chen, R., Zhang, W. J., ... Han, J. (2015). Bioactivity of Essential Oil of Zingiber purpureum Rhizomes and Its Main Compounds against Two Stored Product Insects. *Journal of Economic Entomology*, 108(3), 925–932. doi:10.1093/jee/tov030
65. Koutsaviti, A., Antonopoulou, V., Vlassi, A., Antonatos, S., Michaelakis, A., Papachristos, D. P., & Tzakou, O. (2017). Chemical composition and fumigant activity of essential oils from six plant families against Sitophilus oryzae (Col: Curculionidae). *Journal of Pest Science*, 1–14. doi:10.1007/s10340-017-0934-0
66. Yang, F.-L., Zhu, F., & Lei, C.-L. (2010). Garlic essential oil and its major component as fumigants for controlling Tribolium castaneum (Herbst) in chambers filled with stored grain. *Journal of Pest Science*, 83(3), 311–317. doi:10.1007/s10340-010-0300-y
67. Sadeghi, A., Pourya, M., & Smagghe, G. (2016). Insecticidal activity and composition of essential oils from Pistacia atlantica subsp. kurdica against the model and stored product pest beetle Tribolium castaneum. *Phytoparasitica*, 44(5), 601–607. doi:10.1007/s12600-016-0551-0
68. Toudert-Taleb, K., Hedjal-Chebheb, M., Hami, H., Debras, J.-F., & Kellouche, A. (2014). Composition of essential oils extracted from six aromatic plants of kabylian origin (Algeria) and evaluation of their bioactivity on callosobruchus maculatus (Fabricius, 1775) (Coleoptera: Bruchidae). *African Entomology*, 22(2), 417–427. doi:10.4001/003.022.0220
69. Bachrouch, O., Jemâa, J. M. B., Talou, T., Marzouk, B., & Abderrabba, M. (2010). Fumigant toxicity of Pistacia lentiscus essential oil against Tribolium castaneum and Lasioderma serricorne. *Bulletin of Insectology*, 63(1), 129–135.
70. Mediouni-Ben Jemâa, J., Bachrouch, O., Marzouk, B., & Abderrabba, M. (2010). *Fumigant toxicity of essential oil from pistacia lentiscus L. (Anacardiaceae) against stored-product insects*. Acta Horticulturae (Vol. 853).
71. Yildirim, E., Kesdek, M., & Kordali, S. (2005). Effects of essential oils of three plant species on Tribolium confusum du Val and Sitophilus granarius (L.) (Coleoptera: Tenebrionidae and Curculionidae). *Fresenius Environmental Bulletin*, 14(7), 574–578.
72. Ebadollahi, A., & Mahboubi, M. (2011). Insecticidal activity of the essential oil isolated from Azilia eryngioides (PAU) hedge et lamond against two beetle pests. *Chilean Journal of Agricultural Research*, 71(3), 406–411.
73. Ziae, M., Moharramipour, S., & Mohsenifar, A. (2014). Toxicity of Carum copticum essential oil-loaded nanogel against Sitophilus granarius and Tribolium confusum. *Journal of Applied Entomology*, 138(10), 763–771. doi:10.1111/jen.12133
74. Sahaf, B. Z., Moharramipour, S., & Meshkatalasadat, M. H. (2007). Chemical constituents and fumigant toxicity of essential oil from Carum copticum against two stored product beetles. *Insect Science*, 14(3), 213–218. doi:10.1111/j.1744-7917.2007.00146.x
75. Maroufpoor, M., Ebadollahi, A., Vafaee, Y., & Badiee, E. (2016). Chemical Composition and Toxicity of the Essential Oil of Coriandrum sativum L. and Petroselinum crispum L. Against Three Stored-Product Insect Pests. *Journal of Essential Oil-Bearing Plants*, 19(8), 1993–2002. doi:10.1080/0972060X.2016.1256234
76. Sriti Eljazi, J., Bachrouch, O., Salem, N., Msada, K., Aouini, J., Hammami, M., ... Mediouni Ben Jemaa, J. (2018). Chemical composition and insecticidal activity of essential oil from coriander fruit against Tribolium castaneum, Sitophilus oryzae, and Lasioderma serricorne. *International Journal of Food Properties*, 1–13. doi:10.1080/10942912.2017.1381112
77. Ziae, M., Moharramipour, S., & Mohsenifar, A. (2014). MA-chitosan nanogel loaded with Cuminum cyminum essential oil for efficient management of two stored product beetle pests. *Journal of Pest Science*, 87(4), 691–699. doi:10.1007/s10340-014-0590-6
78. Lashgari, A., Mashayekhi, S., Javadzadeh, M., & Marzban, R. (2014). Effect of Mentha piperita and Cuminum cyminum essential oil on Tribolium castaneum and Sitophilus oryzae. *Archives of Phytopathology and Plant Protection*, 47(3), 324–329. doi:10.1080/03235408.2013.809230
79. Chaubey, M. K. (2007). Toxicity of essential oils from Cuminum cyminum (Umbelliferae), piper nigrum (Piperaceae) and foeniculum vulgare (Umbelliferae) against stored-product beetle Tribolium

- castaneum herbst (Coleopetera: Tenebrionidae). *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 6(1), 1719–1727.
80. Mahmoudvand, M., Abbasipour, H., Rastegar, F., Hosseinpour, M. H., & Basij, M. (2012). Efficacy of some plants as a post-harvest protectant against some major stored pests. *Archives of Phytopathology and Plant Protection*, 45(7), 806–811. doi:10.1080/03235408.2011.597151
81. Nenaah, G. E., & Ibrahim, S. I. A. (2011). Chemical composition and the insecticidal activity of certain plants applied as powders and essential oils against two stored-products coleopteran beetles. *Journal of Pest Science*, 84(3), 393–402. doi:10.1007/s10340-011-0354-5
82. Khani, A., & Asghari, J. (2012). Insecticide activity of essential oils of mentha longifolia, pulicaria gnaphalodes and Achillea wilhelmsii against two stored product pests, the flour beetle, *Tribolium castaneum*, and the cowpea weevil, *callosobruchus maculatus*. *Journal of Insect Science*, 12. doi:10.1673/031.012.7301
83. Herrera, J. M., Zunino, M. P., Massuh, Y., Pizzollito, R. P., Dambolena, J. S., Gañan, N. A., & Zygaldo, J. A. (2014). Fumigant toxicity of five essential oils rich in ketones against *Sitophilus zeamais* (Motschulsky) | Toxicidad fumigante de cinco aceites esenciales rico en cetonas contra *Sitophilus zeamais* (Motschulsky). *AgriScientia*, 31(1), 35–41.
84. Shojaei, A., Talebi, K., Sharifian, I., & Ahsaei, S. M. (2017). Evaluation of detoxifying enzymes of *Tribolium castaneum* and *Tribolium confusum* (Col.: Tenebrionidae) exposed to essential oil of *Artemisia dracunculus* L. *Tenebrionidae*. *Biharean Biologist*, 11(1), 5–9.
85. Liang, J.-Y., Guo, S.-S., Zhang, W.-J., Geng, Z.-F., Deng, Z.-W., Du, S.-S., & Zhang, J. (2017). Fumigant and repellent activities of essential oil extracted from *Artemisia dubia* and its main compounds against two stored product pests. *Natural Product Research*, 1–5. doi:10.1080/14786419.2017.1331227
86. Sharifian, I., Hashemi, S. M., Aghaei, M., & Alizadeh, M. (2012). Insecticidal activity of essential oil of *Artemisia herba-alba* Asso against three stored product beetles. *Biharean Biologist*, 6(2), 90–93.
87. Borzoui, E., Naseri, B., Abedi, Z., & Karimi-Pormehr, M. S. (2016). Lethal and Sublethal Effects of Essential Oils from *Artemisia khorassanica* and *Vitex pseudo-negundo* Against *Plodia interpunctella* (Lepidoptera: Pyralidae). *Environmental Entomology*, 45(5), 1220–1226. doi:10.1093/ee/nvw100
88. Negahban, M., Moharrampour, S., & Sefidkon, F. (2006). Chemical Composition and Insecticidal Activity of *Artemisia scoparia* Essential Oil against Three Coleopteran Stored-Product Insects. *Journal of Asia-Pacific Entomology*, 9(4), 381–388. doi:10.1016/S1226-8615(08)60318-0
89. Negahban, M., Moharrampour, S., & Sefidkon, F. (2007). Fumigant toxicity of essential oil from *Artemisia sieberi* Besser against three stored-product insects. *Journal of Stored Products Research*, 43(2), 123–128. doi:10.1016/j.jspr.2006.02.002
90. Wang, J., Zhu, F., Zhou, X. M., Niu, C. Y., & Lei, C. L. (2006). Repellent and fumigant activity of essential oil from *Artemisia vulgaris* to *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Journal of Stored Products Research*, 42(3), 339–347. doi:10.1016/j.jphotochem.2005.09.007
91. Pimienta-Ramírez, L., García-Rodríguez, Y. M., Ríos-Ramírez, E. M., Lindig-Cisneros, R., & Espinosa-García, F. J. (2016). Chemical composition and evaluation of the essential oil from *Eupatorium glabratum* as biopesticide against *Sitophilus zeamais* and several stored maize fungi. *Journal of Essential Oil Research*, 28(2), 113–120. doi:10.1080/10412905.2015.1093969
92. Karabörklü, S., Ayvaz, A., & Yilmaz, S. (2010). Bioactivities of different essential oils against the adults of two stored product insects. *Pakistan Journal of Zoology*, 42(6), 679–686.
93. Haider, S. Z., Mohan, M., Pandey, A. K., & Singh, P. (2015). Repellent and fumigant activities of *Tanacetum nubigenum* Wallich. ex DC essential oils against *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Journal of Oleo Science*, 64(8), 895–903. doi:10.5650/jos.ess15094
94. Karabörkü, S., Ayvaz, A., Yilmaz, S., & Akbulut, M. (2011). Chemical composition and fumigant toxicity of some essential oils against *ephestia kuehniella*. *Journal of Economic Entomology*, 104(4), 1212–1219. doi:10.1603/EC10284
95. Sousa, A. H., Faroni, L. R. D., & Freitas, R. S. (2014). Relative toxicity of mustard essential oil to insect-pests of stored products | Toxicidade relativa do óleo essencial de mostarda para insetos-praga de produtos armazenados. *Revista Caatinga*, 27(2), 222–226.

96. Jaramillo C, B. E., Duarte R, E., & Delgado, W. (2012). Bioactivity of essential oil from Colombian Chenopodium ambrosioides | Bioactividad del aceite esencial de Chenopodium ambrosioides Colombiano. *Revista Cubana de Plantas Medicinales*, 17(1), 54–64.
97. Khani, A., Rashid, B., & Mirshekar, A. (2017). Chemical composition and insecticidal efficacy of Juniperus polycarpus and Juniperus sabina essential oils against Tribolium confusum (Coleoptera: Tenebrionidae). *International Journal of Food Properties*, 1–9. doi:10.1080/10942912.2017.1338726
98. Hashemi, S. M., & Safavi, S. A. (2012). Chemical constituents and toxicity of essential oils of oriental arborvitae, Platycladus orientalis (L.) Franco, against three stored-product beetles | Componentes químicos y toxicidad de aceites esenciales de tuya oriental, Platycladus orientalis (L.) Fr. *Chilean Journal of Agricultural Research*, 72(2), 188–194.
99. Salem, N., Bachrouch, O., Sriti, J., Msaada, K., Khammassi, S., Hammami, M., ... Mediouni Ben Jemaa, J. (2018). Fumigant and repellent potentials of Ricinus communis and Mentha pulegium essential oils against Tribolium castaneum and Lasioderma serricorne. *International Journal of Food Properties*, 1–15. doi:10.1080/10942912.2017.1382508
100. Yildirim, E., Kesdek, M., Aslan, I., Calmasur, O., & Sahin, F. (2005). The effects of essential oils from eight plant species on two pests of stored product insects. *Fresenius Environmental Bulletin*, 14(1), 23–27.
101. Ebadollahi, A., Safaralizadeh, M., Pourmirza, A., & Gheibi, S. (2010). Toxicity of essential oil of Agastache foeniculum (Pursh) Kuntze to Oryzaephilus surinamensis L. and Lasioderma serricorne F. *Journal of Plant Protection Research*, 50(2), 215–219. doi:10.2478/v10045-010-0037-x
102. Ebadollahi, A. (2011). Chemical constituents and toxicity of Agastache foeniculum (Pursh) kuntze essential oil against two stored-product insect pests | Componentes químicos y toxicidad del aceite esencial de Agastache foeniculum (Pursh) Kuntze contra dos plagas de insectos de . *Chilean Journal of Agricultural Research*, 71(2), 212–217. doi:10.4067/S0718-58392011000200005
103. Ilboudo, Z., Dabiré, L. C. B., Nébié, R. C. H., Dicko, I. O., Dugravot, S., Cortesero, A. M., & Sanon, A. (2010). Biological activity and persistence of four essential oils towards the main pest of stored cowpeas, Callosobruchus maculatus (F.) (Coleoptera: Bruchidae). *Journal of Stored Products Research*, 46(2), 124–128. doi:10.1016/j.jspr.2009.12.002
104. Othira, J. O., Onek, L. A., Deng, L. A., & Omolo, E. O. (2009). Insecticidal potency of Hyptis spicigera preparations against Sitophilus zeamais (L.) and Tribolium castaneum (herbst) on stored maize grains. *African Journal of Agricultural Research*, 4(3), 187–192.
105. Ebadollahi, A., Safaralizadeh, M., & Pourmirza, A. (2010). Fumigant toxicity of lavandula stoechas L. Oil against three insect pests attacking stored products. *Journal of Plant Protection Research*, 50(1), 56–60. doi:10.2478/v10045-010-0010-8
106. Khani, M., Muhamad Awang, R., & Omar, D. (2012). Insecticidal effects of peppermint and black pepper essential oils against rice weevil, Sitophilus oryzae L. and rice moth, Corcyra cephalonica (St.). *Journal of Medicinal Plants*, 11(43), 97–110.
107. Mahmoudvand, M., Abbasipour, H., Basij, M., Hosseinpour, M. H., Rastegar, F., & Nasiri, M. B. (2011). Fumigant toxicity of some essential oils on adults of some stored-product pests | Toxicidad fumigante de algunos aceites esenciales sobre adultos de algunas plagas de productos almacenados. *Chilean Journal of Agricultural Research*, 71(1), 83–89. doi:10.4067/S0718-58392011000100010
108. Eliopoulos, P. A., Hassiotis, C. N., Andreadis, S. S., & Porichi, A.-E. E. (2015). Fumigant toxicity of essential oils from basil and spearmint against two major pyralid pests of stored products. *Journal of Economic Entomology*, 108(2), 805–810. doi:10.1093/jee/tov029
109. Pascual-Villalobos, M. J., & Ballesta-Acosta, M. C. (2003). Chemical variation in an Ocimum basilicum germplasm collection and activity of the essential oils on Callosobruchus maculatus. *Biochemical Systematics and Ecology*. doi:10.1016/S0305-1978(02)00183-7
110. Follett, P. A., Rivera-Leong, K., & Myers, R. (2014). Rice weevil response to basil oil fumigation. *Journal of Asia-Pacific Entomology*, 17(2), 119–121. doi:10.1016/j.aspen.2013.11.008
111. Ogendo, J. O., Kostyukovsky, M., Ravid, U., Matasyoh, J. C., Deng, A. L., Omolo, E. O., ... Shaaya, E. (2008). Bioactivity of Ocimum gratissimum L. oil and two of its constituents against five insect pests attacking stored food products. *Journal of Stored Products Research*, 44(4), 328–334.

- doi:10.1016/j.jspr.2008.02.009
- 112. Çaglar, Ö., Çalmaşur, Ö., Aslan, I., & Kaya, O. (2007). Insecticidal effect of essential oil of *Origanum acutidens* against several stored product pests. *Fresenius Environmental Bulletin*, 16(11 A), 1395–1400.
  - 113. Ayvaz, A., Karaborklu, S., & Sagdic, O. (2009). Fumigant toxicity of five essential oils against the eggs of *Ephestia kuehniella* zeller and *Plodia interpunctella* (Hübner) (Lepidoptera: Pyralidae). *Asian Journal of Chemistry*, 21(1), 596–604.
  - 114. Demirel, N., Sener, O., Arslan, M., Uremis, I., Uluc, F. T., & Cabuk, F. (2009). Toxicological responses of confused flour beetle, *tribolium confusum* du val (Coleoptera: Tenebrinoidea) to various plant essential oils. *Asian Journal of Chemistry*, 21(8), 6403–6410.
  - 115. Ayvaz, A., Sagdic, O., Karaborklu, S., & Ozturk, I. (2010). Insecticidal activity of the essential oils from different plants against three stored-product insects. *Journal of insect science (Online)*, 10, 21.
  - 116. Ye, Y., Zhang, B., Zhang, R., & Wang, K. (2015). The toxic effects of *Perilla frutescens* essential oils in combination with CO<sub>2</sub>-enriched modified atmospheres on the life stages of *Dermestes maculatus* Degeer (Coleoptera: Dermestidae). *Turkiye Entomoloji Dergisi*, 39(3), 261–270.  
doi:10.16970/ted.47667
  - 117. You, C.-X., Wang, Y., Zhang, W.-J., Yang, K., Wu, Y., Geng, Z.-F., ... Liu, Z.-L. (2014). Chemical constituents and biological activities of the Purple *Perilla* essential oil against *Lasioderma serricorne*. *Industrial Crops and Products*, 61, 331–337. doi:10.1016/j.indcrop.2014.07.021
  - 118. Arabi, F., Moharramipour, S., & Sefidkon, F. (2008). Chemical composition and insecticidal activity of essential oil from *Perovskia abrotanoides* (Lamiaceae) against *Sitophilus oryzae* (Coleoptera: Curculionidae) and *Tribolium castaneum* (Coleoptera: Tenebrionidae). *International Journal of Tropical Insect Science*, 28(3), 144–150. doi:10.1017/S1742758408079861
  - 119. Ahmadi, M., Abd-allah, A. M. M., & Moharramipour, S. (2013). Combination of gamma radiation and essential oils from medicinal plants in managing *Tribolium castaneum* contamination of stored products. *Applied Radiation and Isotopes*, 78, 16–20. doi:10.1016/j.apradiso.2013.03.012
  - 120. Ahmadi, M., Moharramipour, S., Mozdarani, H., & Negahban, M. (2008). Combined effect of gamma radiation and *Perovskia atriplicifolia* for the control of red flour beetle, *Tribolium castaneum*. *Communications in agricultural and applied biological sciences*, 73(3), 643–650.
  - 121. Hashemi, S. M., Hosseini, B., Estaji, A., Hashemi, S. M., Hosseini, B., & Estaji, A. (2013). Chemical Composition and Insecticidal Properties of the Essential Oil of *Salvia leiriifolia* Benth (Lamiaceae) at Two Developmental Stages. *Journal of Essential Oil-Bearing Plants*, 16(6), 806–816.  
doi:10.1080/0972060X.2013.854493
  - 122. Hosseini, B., Estaji, A., & Hashemi, S. M. (2013). Fumigant toxicity of essential oil from *Salvia leiriifolia* (Benth) against two stored product insect pests. *Australian Journal of Crop Science*, 7(6), 855–860.
  - 123. Karabörklü, S. (2014). Chemical characterization of *Sideritis perfoliata* L. essential oil and its fumigant toxicity against two pest insects. *Journal of Food, Agriculture and Environment*, 12(2), 434–437.
  - 124. Khani, A., & Heydarian, M. (2014). Fumigant and repellent properties of sesquiterpene-rich essential oil from *Teucrium polium* subsp. *capitatum* (L.). *Asian Pacific Journal of Tropical Medicine*, 7(12), 956–961. doi:10.1016/S1995-7645(14)60169-3
  - 125. Moazeni, N., Khajeali, J., Izadi, H., & Mahdian, K. (2014). Chemical composition and bioactivity of *Thymus daenensis* Celak (Lamiaceae) essential oil against two lepidopteran stored-product insects. *Journal of Essential Oil Research*, 26(2), 118–124. doi:10.1080/10412905.2013.860412
  - 126. Saroukolai, A. T., Moharramipour, S., & Meshkatalasadat, M. H. (2010). Insecticidal properties of *Thymus persicus* essential oil against *Tribolium castaneum* and *Sitophilus oryzae*. *Journal of Pest Science*, 83(1), 3–8. doi:10.1007/s10340-009-0261-1
  - 127. Sahaf, B. Z., Moharramipour, S., & Meshkatalasadat, M. H. (2008). Fumigant toxicity of essential oil from *Vitex pseudo-negundo* against *Tribolium castaneum* (Herbst) and *Sitophilus oryzae* (L.). *Journal of Asia-Pacific Entomology*, 11(4), 175–179. doi:10.1016/j.aspen.2008.09.001
  - 128. Kheirkhah, M., Ghasemi, V., Yazdi, A. K., & Rahban, S. (2015). Chemical composition and insecticidal activity of essential oil from *Ziziphora clinopodioides* Lam. used against the Mediterranean flour

- moth, *Ephestia kuehniella* Zeller. *Journal of Plant Protection Research*, 55(3), 260–265. doi:10.1515/jppr-2015-0037
129. Salehi, T., Karimi, J., Hasanshahi, G., Askarianzadeh, A., Abbasipour, H., & Garjan, A. S. (2014). The Effect of Essential Oils from *Laurus nobilis* and *Myrtus communis* on the Adults of Mediterranean Flour Moth, *Ephestia kuehniella* Zeller (Lep.: Pyralidae). *Journal of Essential Oil-Bearing Plants*, 17(4), 553–561. doi:10.1080/0972060X.2014.935059
130. Mediouni Ben Jemâa, J., Tersim, N., Toudert, K. T., & Khouja, M. L. (2012). Insecticidal activities of essential oils from leaves of *Laurus nobilis* L. from Tunisia, Algeria and Morocco, and comparative chemical composition. *Journal of Stored Products Research*, 48, 97–104. doi:10.1016/j.jspr.2011.10.003
131. Senfi, F., Safaralizadeh, M. H., Safavi, S. A., & Aramideh, S. (2014). Fumigant toxicity of *Laurus nobilis* and *Myrtus communis* essential oils on larvae and adults of the Red flour beetle, *Tribolium castaneum* Herbst (Col.: Tenebrionidae). *Archives of Phytopathology and Plant Protection*, 47(4), 472–476. doi:10.1080/03235408.2013.812819
132. Saleem, S., Ul Hasan, M., Sagheer, M., & Talib Sahi, S. (2014). Insecticidal activity of essential oils of four medicinal plants against different stored grain insect pests. *Pakistan Journal of Zoology*, 46(5), 1407–1414.
133. Negahban, M., & Moharramipour, S. (2007). Fumigant toxicity of *Eucalyptus intertexta*, *Eucalyptus sargentii* and *Eucalyptus camaldulensis* against stored-product beetles. *Journal of Applied Entomology*, 131(4), 256–261. doi:10.1111/j.1439-0418.2007.01152.x
134. Parsia Aref, S., Valizadegan, O., & Farashiani, M. E. (2016). The Insecticidal Effect of Essential Oil of *Eucalyptus floribundi* Against Two Major Stored Product Insect Pests; *Rhyzopertha dominica* (F.) and *Oryzaephilus surinamensis* (L.). *Journal of Essential Oil-Bearing Plants*, 19(4), 820–831. doi:10.1080/0972060X.2014.958569
135. Ebadollahi, A., Safaralizadeh, M. H., & Pourmirza, A. A. (2010). Fumigant toxicity of essential oils of *Eucalyptus globulus* labill and *Lavandula stoechas* L., grown in Iran, against the two coleopteran insect pests; *Lasioderma serricorne* F. and *Rhyzopertha dominica* F. *Egyptian Journal of Biological Pest Control*, 20(1), 1–5.
136. Kambouzia, J., Negahban, M., & Moharramipour, S. (2009). Fumigant toxicity of *Eucalyptus leucoxylon* against stored product insects. *American-Eurasian Journal of Sustainable Agriculture*, 3(2), 229–233.
137. Liao, M., Xiao, J.-J., Zhou, L.-J., Liu, Y., Wu, X.-W., Hua, R.-M., ... Cao, H.-Q. (2016). Insecticidal activity of *Melaleuca alternifolia* essential oil and RNA-Seq analysis of *Sitophilus zeamais* transcriptome in response to oil fumigation. *PLoS ONE*, 11(12). doi:10.1371/journal.pone.0167748
138. Kamran, H. M., Mansoor-Ul-Hasan, Sagheer, M., Khan, A. A., Aatif, H. M., Ijaz, M., ... Abbas, S. K. (2017). Bioactivity of Three Plant Essential Oils against Red Flour Beetle (*Tribolium castaneum*) (Coleoptera: Tenebrionidae). *Zeitschrift fur Arznei- und Gewurzpflanzen*, 22(1), 14–19.
139. Hernandez-Lambrano, R., Pajaro-Castro, N., Caballero-Gallardo, K., Stashenko, E., & Olivero-Verbel, J. (2015). Essential oils from plants of the genus *Cymbopogon* as natural insecticides to control stored product pests. *Journal of Stored Products Research*, 62, 81–83. doi:10.1016/j.jspr.2015.04.004
140. Nattudurai, G., Baskar, K., Paulraj, M. G., Islam, V. I. H., Ignacimuthu, S., & Duraipandiyan, V. (2017). Toxic effect of *Atalantia monophylla* essential oil on *Callosobruchus maculatus* and *Sitophilus oryzae*. *Environmental Science and Pollution Research*, 24(2), 1619–1629. doi:10.1007/s11356-016-7857-9
141. Campolo, O., Malacrinò, A., Zappalà, L., Laudani, F., Chiera, E., Serra, D., ... Palmeri, V. (2014). Fumigant bioactivity of five *Citrus* essential oils against *Tribolium confusum*. *Phytoparasitica*, 42(2), 223–233. doi:10.1007/s12600-013-0355-4
142. Safavi, S. A., & Mobki, M. (2016). Susceptibility of *Tribolium castaneum* (Herbst, 1797) larvae to essential oils of *Citrus reticulata* Blanco fruit peels and the synergist, diethyl maleate. *Biharean Biologist*, 10(2), 82–85.
143. Mobki, M., Safavi, S. A., Safaralizadeh, M. H., Panahi, O., & Afshar, A. T. (2014). Effect of diethyl

- maleate on the toxicity of essential oil from *Citrus reticulata* fruit peels to *Tribolium castaneum* Herbst under laboratory conditions. *Archives of Phytopathology and Plant Protection*, 47(9), 1023–1029. doi:10.1080/03235408.2013.817069
144. Li, C. (2011). Effect of essential oil from *zanthoxylum bungeanum maxim* on drugstore beetle *stegobium paniceum* (L.) (Coleoptera, Anobiidae). In *ITME 2011 - Proceedings: 2011 IEEE International Symposium on IT in Medicine and Education* (Vol. 2, pp. 588–592). doi:10.1109/ITiME.2011.6132180
145. Wang, C.-F., Yang, K., You, C.-X., Zhang, W.-J., Guo, S.-S., Geng, Z.-F., ... Wang, Y.-Y. (2015). Chemical composition and insecticidal activity of essential oils from *Zanthoxylum dissitum* leaves and roots against three species of storage pests. *Molecules*, 20(5), 7990–7999. doi:10.3390/molecules20057990
146. Suthisut, D., Fields, P. G., & Chandrapatya, A. (2011). Fumigant toxicity of essential oils from three Thai plants (Zingiberaceae) and their major compounds against *Sitophilus zeamais*, *Tribolium castaneum* and two parasitoids. *Journal of Stored Products Research*, 47(3), 222–230. doi:10.1016/j.jspr.2011.03.002
147. Abbasipour, H., Mahmoudvand, M., Rastegar, F., & Hosseinpour, M. H. (2011). Fumigant toxicity and oviposition deterrence of the essential oil from cardamom, *Elettaria cardamomum*, against three stored - Product insects. *Journal of Insect Science*, 11. doi:10.1673/031.011.16501
148. Bougherra, H. H., Bedini, S., Flamini, G., Cosci, F., Belhamel, K., & Conti, B. (2015). *Pistacia lentiscus* essential oil has repellent effect against three major insect pests of pasta. *Industrial Crops and Products*, 63, 249–255. doi:10.1016/j.indcrop.2014.09.048
149. Olivero-Verbel, J., Tirado-Ballestas, I., Caballero-Gallardo, K., & Stashenko, E. E. (2013). Essential oils applied to the food act as repellents toward *Tribolium castaneum*. *Journal of Stored Products Research*, 55, 145–147. doi:10.1016/j.jspr.2013.09.003
150. Ukeh, D. A., Adie, E. B., & Ukeh, J. A. (2011). Insecticidal and repellent activities of pepper fruit, *Dennettia tripetala* (G. Baker) against the cowpea Beetle, *Callosobruchus maculatus* (Fabricius). *Biopesticides International*, 7(1), 15–23.
151. Karahroodi, Z. R., Moharrampour, S., & Rahbarpour, A. (2009). Investigated repellency effect of some essential oils of 17 native medicinal plants on adults *Plodia interpunctella*. *American-Eurasian Journal of Sustainable Agriculture*.
152. Cosimi, S., Rossi, E., Cioni, P. L., & Canale, A. (2009). Bioactivity and qualitative analysis of some essential oils from Mediterranean plants against stored-product pests: Evaluation of repellency against *Sitophilus zeamais* Motschulsky, *Cryptolestes ferrugineus* (Stephens) and *Tenebrio molitor* (L.). *Journal of Stored Products Research*, 45(2), 125–132. doi:10.1016/j.jspr.2008.10.002
153. Moharrampour, S., Taghizadeh, A., Meshkatalasadat, M. H., Fathipour, Y., & Talebi, A. A. (2009). Repellent activity and persistence of essential oil extracted from *Prangos acaulis* to three stored-product beetles. *American-Eurasian Journal of Sustainable Agriculture*, 3(2), 202–204.
154. Pandey, A. K., Palni, U. T., & Tripathi, N. N. (2014). Repellent activity of some essential oils against two stored product beetles *Callosobruchus chinensis* L. and *C. maculatus* F. (Coleoptera: Bruchidae) with reference to *Chenopodium ambrosioides* L. oil for the safety of pigeon pea seeds. *Journal of Food Science and Technology*, 51(12), 4066–4071. doi:10.1007/s13197-012-0896-4
155. Akhtar, S., Mansoor-Ul-Hasan, Sagheer, M., & Javed, N. (2015). Antifeedant effect of essential oils of five indigenous medicinal plants against stored grain insect pests. *Pakistan Journal of Zoology*, 47(4), 1045–1050.
156. Jaramillo-Colorado, B. E., Karen, M. C., Edisson, D. R., Elena, S., & Jesus, O. V. (2014). Volatile Secondary Metabolites from Colombian *Croton malambo* (Karst) by Different Extraction Methods and Repellent Activity of its Essential Oil. *Journal of Essential Oil-Bearing Plants*, 17(5), 992–1001. doi:10.1080/0972060X.2014.895185
157. Wagan, T. A., Hu, D., He, Y., Nawaz, M., Nazir, T., Mabubu, J. I., & Hua, H. (2016). Repellency of three plant essential oils against red flour beetle *Tribolium castaneum* (Herbst, 1797) (Coleoptera: Tenebrionidae). *Turkiye Entomoloji Dergisi*, 40(4), 347–354. doi:10.16970/ted.67713
158. Wang, Y.-C., Li, P., & Chi, D.-F. (2016). Electrophysiological and behavioral responses of *Tenebrio*

- molitor L. to fourteen kinds of plant volatiles. *Journal of Asia-Pacific Entomology*, 19(2), 261–267. doi:10.1016/j.aspen.2016.03.002
159. Allahvaisi, S. (2010). Reducing insects contaminations through stored foodstuffs by use of packaging and repellency essential oils. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 38(3), 21–24.
160. Allahvaisi, S., Maroufpoor, M., Abdolmaleki, A., Hoseini, S.-A., & Ghasemzadeh, S. (2011). The effect of plant oils for reducing contamination of stored packaged-foodstuffs | Wpływ olejków roślinnych na ograniczenie występowania szkodników magazynowych w przechowywanych produktach spożywczych. *Journal of Plant Protection Research*, 51(1), 82–86. doi:10.2478/v10045-011-0015-y
161. Jemaa, J. M.-B., Tersim, N., & Khouja, M. L. (2013). *Repellent action of laurus nobilis essential oil against the cigarette beetle lasioderma serricorne fabricius (Coleoptera: Anobiidae)*. *Acta Horticulturae* (Vol. 997).
162. Saleem, S., ul Hasan, M., Ali, Q., Hanif, C. M. S., Sajid, M. W., Akhtar, S., ... Mahmood, A. (2017). Effectiveness of four medicinal plant essential oils as feeding deterrent towards different strains of stored grain insect pests. *Pakistan Journal of Agricultural Sciences*, 54(4), 769–774. doi:10.21162/PAKJAS/17.4534
163. Khemira, S., Jemaa, J. M.-B., Haouel, S., & Khouja, M. L. (2013). *Repellent activity of essential oil of eucalyptus astringens against rhizopertha dominica and oryzaephilus surinamensis*. *Acta Horticulturae* (Vol. 997).
164. Diaz-Montano, J., Campbell, J. F., Phillips, T. W., & Throne, J. E. (2015). Evaluation of Potential Attractants for Six Species of Stored-Product Psocids (Psocoptera: Liposcelididae, Trogidae). *Journal of Economic Entomology*, 108(3), 1398–1407. doi:10.1093/jee/tov028
165. Zhang, J. S., Zhao, N. N., Liu, Q. Z., Liu, Z. L., Du, S. S., Zhou, L., & Deng, Z. W. (2011). Repellent constituents of essential oil of Cymbopogon distans aerial parts against two stored-product insects. *Journal of Agricultural and Food Chemistry*, 59(18), 9910–9915. doi:10.1021/jf202266n
166. Wong, K. K. Y., Signal, F. A., Campion, S. H., & Motion, R. L. (2005). Citronella as an insect repellent in food packaging. *Journal of Agricultural and Food Chemistry*, 53(11), 4633–4636. doi:10.1021/jf050096m
167. Yang, K., You, C.-X., Wang, C.-F., Guo, S.-S., Li, Y.-P., Wu, Y., ... Du, S.-S. (2014). Composition and repellency of the essential oils of evodia calcicola chun ex huang and evodia trichotoma (lour.) pierre against three stored product insects. *Journal of Oleo Science*, 63(11), 1169–1176. doi:10.5650/jos.ess14140
168. Guo, S.-S., Zhang, W.-J., Yang, K., Liang, J.-Y., You, C.-X., Wang, C.-F., ... Du, S.-S. (2017). Repellence of the main components from the essential oil of Glycosmis lucida Wall. ex Huang against two stored product insects. *Natural Product Research*, 31(10), 1201–1204. doi:10.1080/14786419.2016.1226825
169. You, C.-X., Zhang, W.-J., Guo, S.-S., Wang, C.-F., Yang, K., Liang, J.-Y., ... Deng, Z.-W. (2015). Chemical composition of essential oils extracted from six Murraya species and their repellent activity against Tribolium castaneum. *Industrial Crops and Products*, 76, 681–687. doi:10.1016/j.indcrop.2015.07.044
170. Zhang, W.-J., Zhang, Z., Chen, Z.-Y., Liang, J.-Y., Geng, Z.-F., Guo, S.-S., ... Deng, Z.-W. (2017). Chemical Composition of Essential Oils from Six Zanthoxylum Species and Their Repellent Activities against Two Stored-Product Insects. *Journal of Chemistry*, 2017. doi:10.1155/2017/1287362
171. Izakmehri, K., Saber, M., Mehrvar, A., Hassanpouraghdam, M. B., & Vojoudi, S. (2013). Lethal and sublethal effects of essential oils from Eucalyptus camaldulensis and Heracleum persicum against the adults of callosobruchus maculatus. *Journal of Insect Science*, 13. doi:10.1673/031.013.15201
172. Osman, S. E. I., Swidan, M. H., Kheirallah, D. A., & Nour, F. E. (2016). Histological effects of essential oils, their monoterpenoids and insect growth regulators on midgut, integument of larvae and ovaries of khapra beetle, Trogoderma granarium everts. *Journal of Biological Sciences*, 16(3), 93–101. doi:10.3923/jbs.2016.93.101