

Research Article

Anti-Inflammatory Effects of Essential Oils of *Amomum aromaticum* Fruits in Lipopolysaccharide-Stimulated RAW264.7 Cells

Nguyen Hai Dang ^{1,2}, Le Thi Van Anh,³ and Nguyen Tien Dat⁴

¹University of Science and Technology of Hanoi, Vietnam Academy of Science and Technology (VAST), 18 Hoang Quoc Viet Cau Giay, Hanoi, Vietnam

²Institute of Marine Biochemistry, VAST, 18 Hoang Quoc Viet Cau Giay, Hanoi, Vietnam

³Graduate University of Science and Technology, VAST, 18 Hoang Quoc Viet Cau Giay, Hanoi, Vietnam

⁴Center for Research and Technology Transfer, VAST, 18 Hoang Quoc Viet Cau Giay, Hanoi, Vietnam

Correspondence should be addressed to Nguyen Hai Dang; nguyenhd@imbc.vast.vn

Received 21 April 2020; Revised 9 May 2020; Accepted 13 May 2020; Published 25 May 2020

Academic Editor: quancai sun

Copyright © 2020 Nguyen Hai Dang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Inflammation is a vital physiologic response of cellular injury, infection, or autoimmune activation. Overproduction of proinflammatory mediators may result in the chronic inflammation that leads to many diseases such as rheumatoid arthritis, asthma, multiple sclerosis, and atherosclerosis. In this study, we assessed for the first time the anti-inflammatory effects of the essential oils of *Amomum aromaticum* fruits (AAE) in RAW264.7 murine macrophage model. As a result, AAE potently inhibited the production of nitric oxide in LPS-induced RAW264.7 cells with the IC_{50} value of $0.45 \pm 0.11 \mu\text{g/ml}$. AAE also dose-dependently reduced the expression of two proinflammatory proteins iNOS and COX-2 in the stimulated cells. Phytochemical analysis revealed that major compositions of the volatile oils including 1,8 cineole (48.22%), geranial (9.24%), neral (6.72%), α -pinene (2.43%), and α -terpineol (2.28%) may contribute greatly to the inhibition effects due to their anti-inflammatory properties. The results suggest for the potential uses of AAE in chronic inflammation prevention.

1. Introduction

Chronic inflammation is an undesirable phenomenon of a prolonged inflammatory response. Overproduction of proinflammatory mediators such as cytokines, interleukins, nitric oxide (NO), inducible nitric oxide synthase (iNOS), and cyclooxygenase (COX-2) may result in various diseases such as rheumatoid arthritis, asthma, multiple sclerosis, and atherosclerosis [1]. Therefore, control of proinflammatory responses is a wise strategy to prevent the development of inflammatory diseases. Since the ancient time, food was determined as an important source for prevention of diseases. There has been accumulation of evidence that increases consumption of certain foods might lower the risk of cardiovascular disease, cancer, and inflammation [2, 3].

Amomum aromaticum Roxb. is a species of the Zingiberaceae family, which is a common spice and food flavoring agent in Vietnam and other Asian countries. The fruits of this plant have been used in traditional medicine for the treatment of cough, abdominal pain, vomiting, diarrhea, and malaria. The oils of seeds have been used in India for benefiting the digestive system, applied to the eyelids to eliminate the inflammation [4, 5]. To date, there has been only few studies about of the phytochemicals as well as the biological activities of this plant. Recently, *A. aromaticum* essential oils are shown as promising antileishmanial agent in a screening program of 37 plants of Vietnam flora [6]. The methanolic extract of *A. aromaticum* exhibited significant antimicrobial activity against *Enterococcus faecalis*, *Staphylococcus aureus*, *Enterobacter aerogenes*, *Proteus mirabilis*,

and *Pseudomonas aeruginosa* with the MIC values ranging from 3.41 to 9.63 mg/ml [7].

In this study, we investigated the phytochemical contents of essential oils of the *A. aromaticum* fruits and its anti-inflammatory properties including NO production inhibition assay and inhibitory effects on the expression of two key enzymes of inflammation process: inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2) in RAW264.7 cells stimulated with LPS.

2. Materials and Methods

2.1. Plant Materials and Essential Oil Preparation. The fruits *A. aromaticum* were freshly collected in Ha Giang province in November 2019. The samples were taxonomically identified by Dr. Nguyen the Cuong, Institute of Ecology and Biological Resources (VAST), and voucher specimens were deposited in the Institute of Marine Biochemistry. The samples (500 g) were hydrodistilled in a Clevenger-type apparatus for 4 h, after which the essential oils were separated and dried with anhydrous Na₂SO₄. The obtained oils (AAE) were stored at -5°C until used.

2.2. GC/MS Analysis of Essential Oils. GC/MS analysis was performed using an Agilent GC7890A apparatus coupled to a mass selective detector (Agilent 5976C). A HP-5MS fused silica capillary column (60 m × 0.25 mm id. × 0.25 μm film thickness) was used. Helium was the carrier gas with a flow rate of 1.0 ml/min. The inlet temperature was 240°C, and the oven temperature program was as follows: 60°C to 220°C at 4°C/min and then at 20°C/min to 240°C. The split injection mode was 1:142, the detector temperature was 240°C, and the injection volume was 0.1 μl. The MS interface temperature was 240°C, MS mode, E.I. detector voltage 1300 V, and mass range 40–400 Da at 1.0 scan/s. Identification of components was achieved based on their retention indices and by comparison of their mass spectral fragmentation patterns with those stored on the MS library (NIST08, Wiley09). Component relative contents were calculated based on total ion current without standardization. Data processing was MassFinder4.0.

2.3. Cell Culture. Murine macrophage RAW264.7 cell lines used in this study were obtained from the American Type Culture Collection (Manassas, VA, USA). Cells were maintained in DMEM supplemented with 10% fetal bovine serum (Hyclone, Logan, UT, USA) and penicillin (100 units/ml)-streptomycin (100 μg/ml) (Invitrogen, Carlsbad, CA, USA). Cultures were maintained in a CO₂ incubator-humidified atmosphere 5% CO₂ at 37°C.

2.4. Assay for Inhibition of NO Production. The effects of samples on the NO production in LPS-stimulated RAW264.7 macrophage cells were examined as described previously [8]. The cells were seeded in 96-well plate at 2 × 10⁵ cells/well and incubated for 18 h. The plates were pretreated with AAE (from 0.1 μg/ml to 100 μg/ml) for

30 min and then incubated for another 24 h with or without 1 μg/ml LPS (*Escherichia coli* 0111: B4; Sigma Aldrich, USA). 100 μl of the culture supernatant was transferred to other 96-well plates, and 100 μl of Griess reagent was added. The absorbance of the reaction solution was read at 570 nm with a XMark microplate reader (BioRad, USA). The remaining cell solutions in cultured 96-well plate were used to evaluate cell viability by 3-(4,5-dimethylthiazole-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay [9]. Cardamonin, a known NO production inhibitor, was used as a positive control [10].

2.5. Western Blot Analysis. The RAW264.7 cells were harvested and lysed in a lysis buffer (150 mM NaCl, 50 mM Tris-HCl, pH 7.4, 1 mM EDTA, 1% NP-40, 5 mM sodium orthovanadate, and protease inhibitors cocktail (BD Biosciences)) and then centrifuged for 10 min at 4°C and 15,000 rpm. An equal amount of protein was separated onto SDS-PAGE (sodium dodecyl sulfate polyacrylamide gel electrophoresis) and transferred to a PVDF membrane (Millipore, Germany). The membranes were blocked in 5% nonfat skim milk for 1 h at room temperature, probed with the appropriate primary antibodies, washed, and then incubated with the corresponding secondary antibodies. α-Tubulin was used as the loading control. The signal was developed using the ECL (enhanced chemiluminescence) system (GE Healthcare, UK) and detected in a gel imaging system Azure c300 (Azure Biosciences, UK). The captured images were analyzed and quantified using ImageJ v. 1.53a (NIH, Maryland, USA).

2.6. Statistical Analysis. Data are expressed as the mean ± standard deviation (SD). Statistical significance was assessed by the two-tailed unpaired Student's *t* test, and *P* values less than 0.05 was considered statistically significant.

3. Results and Discussion

3.1. Phytochemicals of AAE Analyzed by GC/MS. The essential oils of *A. aromaticum* fruits (AAE) obtained by hydrodistillation yields 1.49% based on a calculation with the dry weight of fruits. A total of 25 compounds were identified by using GC/MS data in combination with the MS library analyses (Table 1). The major chemical group of AAE is monoterpene with more than 81% of the total contents. Of which, 1,8-cineole (or eucalyptol, 48.22%), geranial (9.24%), neral (6.72%), α-pinene (2.43%), α-terpineol (2.28%), and β-pinene (2.18%) are among the most abundance monoterpenes of the fruit oils. Four aliphatic aldehydes was found including *n*-octanal, 2-octenal, (*E,E*)-decenal, and (*E,E*)-dodecanal which comprise about 8.26% of the AAE content. Meanwhile, only one sesquiterpene *E*-nerolidol (1.69%) was found. Our results were in good agreement with the previous report which shows monoterpenes as the major contents together with the presence of aliphatic aldehyde and sesquiterpene groups [6]. The fruit essential oils contained 55.2% of 1,8-cineole which was slightly higher than that of our findings. The distribution of relative quantities of major

TABLE 1: Chemical compositions of essential oils of *A. aromaticum* fruits.

No.	Compounds	RI	Relative percentage (%)
1	α -Thujene	930	0.16
2	α -Pinene	939	2.43
3	Sabinene	978	0.54
4	β -Pinene	984	2.18
5	Myrcene	991	0.46
6	<i>n</i> -Octanal	1003	0.47
7	α -Phellandrene	1010	1.43
8	<i>O</i> -Cymene	1029	0.51
9	Limonene	1034	2.9
10	1,8-Cineole	1038	48.22
11	(<i>E</i>)- β -cymene	1048	0.83
12	2-Octenal	1058	0.95
13	γ -Terpinene	1063	0.3
14	Linalool	1101	0.37
15	Isoneral	1166	0.19
16	δ -Terpineol	1174	0.21
17	Isogeranial	1184	0.3
18	Terpinen-4-ol	1185	0.92
19	α -Terpineol	1198	2.28
20	Neral	1246	6.72
21	Geraniol	1256	1.33
22	(<i>E,E</i>)-Decenal	1264	4.9
23	Geranial	1275	9.24
24	(<i>E,E</i>)-Dodecanal	1470	1.94
25	<i>E</i> -Nerolidol	1570	1.69
Total identified (%)			91.47
Yield^a (%)			1.49

^aYield calculated based on the fresh materials; RI: retention index.

monoterpenes in both studies was found to be similar. The difference of quantities of individual compounds in both samples may be due to the variation of the origin of samples, seasons of collection, or the environmental factors. The presence of high content of 1,8-cineole was not only found in *A. aromaticum* but also in some other *Amomum* species including *A. tsao-ko* (23.87%–45.24%) [11–13] and *A. subulatum* Roxb (20%–89%) [14–16] depending on parts of the plant used for analysis.

3.2. AAE Reduced the NO Production in LPS-Induced RAW264.7 Cells by Inhibiting the Expressions of iNOS and COX-2. NO is an important signaling molecule in various physiological and pathophysiological responses [17]. Searching for inhibitors of NO production in LPS-stimulated macrophages has been a worldwide effort for the development of anti-inflammatory agents. The in vitro anti-inflammatory activity of AAE was investigated by determining its NO production inhibitory effect in LPS-stimulated RAW264.7 cells. The primary screening results showed that AAE inhibited potently the NO production (about 100%) in the stimulated cells at concentration of 100 μ g/ml. We further evaluated the potency of the inhibitory activity of AAE by determining its IC₅₀ value. As the results, the IC₅₀ value of AAE was determined as 0.45 ± 0.11 μ g/ml which was slightly higher than the positive control, cardamomin (0.59 ± 0.18 μ g/ml). Treatment of AAE at the screening

concentration after 24 h had no impact on the cell viability (data not shown). Next, we investigated the effects of AAE on the two key enzymes of inflammation process: iNOS (inducible nitric oxide synthase), mainly responsible for the production of NO and COX-2 (cyclooxygenase-2), in charge of production inflammatory mediators such as PGE₂ (prostaglandin E₂). The western blot analysis revealed that AAE dose-dependently inhibited the expression of both enzymes. Remarkably, at a concentration of 0.3 μ g/ml, the inhibitory effects of AAE against iNOS and COX-2 expressions were still observed significantly (Figure 1(b)). To our knowledge, this is the first report of this potent anti-inflammatory activity of the *A. aromaticum* essential oils.

The phytochemicals are considered as the major contributors to the biological activity of a plant samples. In our study, we found that the fruit essential oils showed remarkable anti-inflammatory effects. The major composition of AAE, as indicated, is 1,8-cineole which comprises about 48% of the total oil content. Interestingly, 1,8-cineole was demonstrated as a very promising anti-inflammatory agent. Molecular mechanism studies indicated that 1,8-cineole effectively reduced the expression of proinflammatory cytokines such as TNF-IL-1 β and IL-6 with the IC₅₀ values ranging from 0.2 to 7.0 μ M. It was found to be a potent inhibitor of NF- κ B activation [18]. 1,8-cineole also displayed its anti-inflammatory properties in various animal models. This compound was advanced to clinical trials for bronchial asthma. When administered as an adjunct therapy with prednisolone, 1,8-cineole showed a significant improvement in respiratory volume and quality of asthma. The effect was still maintained when the dosage of prednisolone was decreased by 36% [19]. Other major compositions of AAE such as α -pinene [20], α -terpineol [21], geraniol [22], neral, and geranial [23] also exhibited their effects of anti-inflammation. It is demonstrated that the chief monoterpenes of AAE seem to greatly contribute to the anti-inflammatory activity of the fruit oils.

The anti-inflammatory properties of essential oils of some other *Amomum* species were reported. The fruit extract of *A. tsao-ko* displayed potent anti-inflammatory effects in RAW264.7 cells stimulated with LPS [24]. Further studies showed that the effects were achieved because this extract induced the expression of heme oxygenase-1 which consequently increased the Nrf-2 activation. The similar effects were also obtained from different extracts and isolated compounds from *A. tsao-ko* [25–27]. Agnihotri et al. investigated the topical anti-inflammatory effect of the fruit essential oils of *A. subulatum*. The results showed that the volatile oils exhibited moderate activities compared with standard drug, diclofenac [14]. The extracts of *A. compactum*, *A. xanthoides*, and *A. vilosum* also demonstrated their anti-inflammatory activities in vitro and in vivo [28–30]. Interestingly, there have been very few studies on the *Amomum* essential oils with anti-inflammation. In our study, we have reported that AAE is a promising anti-inflammatory agent by potently inhibiting the production of nitric oxide, the expressions of iNOS and COX-2 in LPS-induced RAW264.7 murine macrophages. Notably, *A. aromaticum* has been traditionally used as a common spice suggesting its safety effects in therapeutic use.

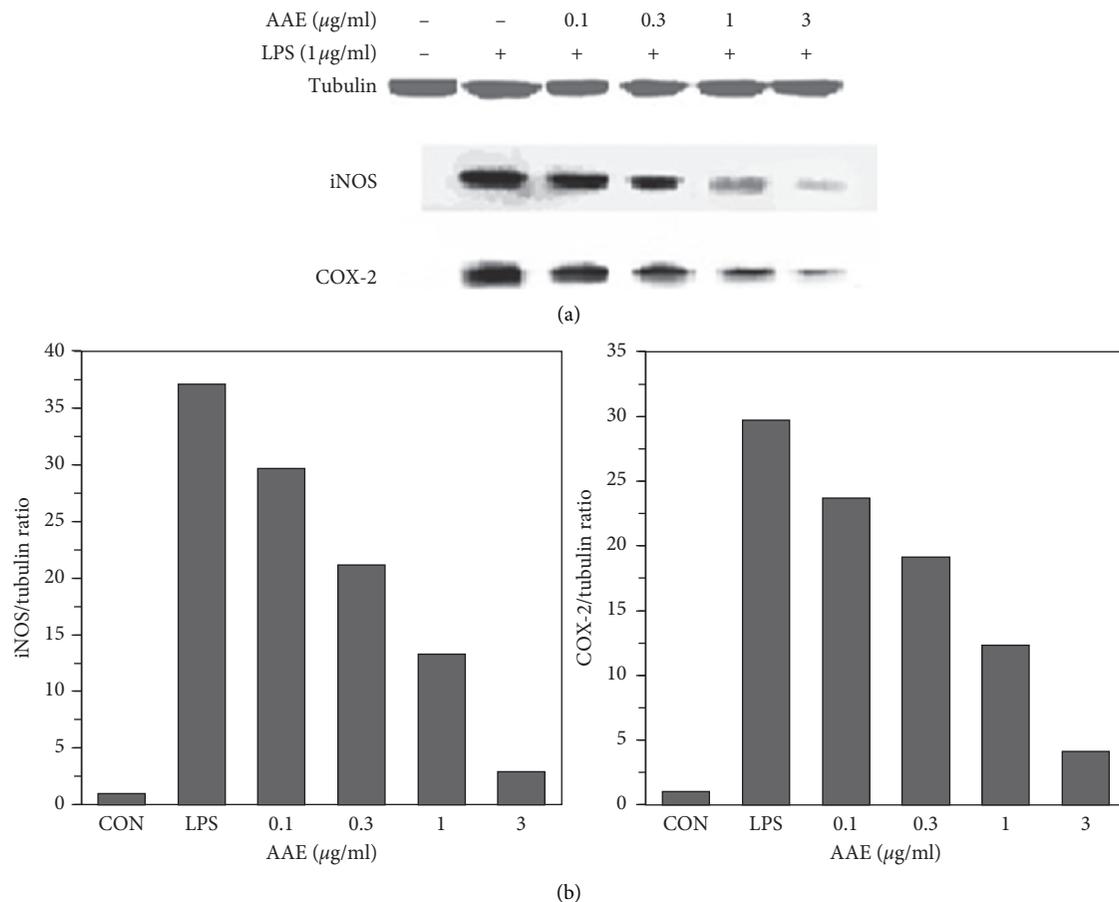


FIGURE 1: Inhibitory effect of AAE on LPS-induced iNOS and COX-2 expression in RAW264.7 murine macrophages. (a) Western blot analysis of iNOS and COX-2 expression in LPS-induced RAW264.7 cells after 24 h of treatment with AAE in different concentrations; (b) quantitation was analyzed by ImageJ 1.53a (NIH, USA). The ratio of the relative intensity of iNOS or COX-2 to tubulin is expressed; CON: control, LPS: lipopolysaccharide.

4. Conclusions

For the first time, the anti-inflammatory properties of the fruit essential oils of *Amomum aromaticum* Roxb. were investigated. The volatile oils displayed potent inhibitory effects against the production of nitric oxide; the expression of two proinflammatory enzymes iNOS and COX-2 in RAW264.7 macrophages was stimulated with LPS. Phytochemical investigation revealed that the essential oils contain various anti-inflammatory compositions including 1,8 cineole (48.22%), geranial (9.24%), neral (6.72%), α -pinene (2.43%), and α -terpineol (2.28%). These findings suggest that essential oils of *A. aromaticum* fruits can be an alternative natural source for prevention of chronic inflammation. Further studies are necessary for evaluation of anti-inflammation mechanisms of action and in vivo assessments of the very promising essential oils.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

All authors declare that they have no conflicts of interest.

Acknowledgments

This research was funded by the Vietnam National Foundation for Science and Technology Development (NAFOSTED) under grant no. 104.01-2017.307.

References

- [1] J. Kanterman, M. Sade-Feldman, and M. Baniyash, "New insights into chronic inflammation-induced immunosuppression," *Seminars in Cancer Biology*, vol. 22, no. 4, pp. 307–318, 2012.
- [2] H. Boeing, A. Bechthold, A. Bub et al., "Critical review: vegetables and fruit in the prevention of chronic diseases," *European Journal of Nutrition*, vol. 51, no. 6, pp. 637–663, 2012.
- [3] Y. Peng, R. Gan, H. Li et al., "Absorption, metabolism, and bioactivity of vitexin: recent advances in understanding the efficacy of an important nutraceutical," *Critical Reviews in Food Science and Nutrition*, pp. 1–16, 2020.

- [4] P. K. Sharma, N. S. Chauhan, and B. Lal, "Observations on the traditional phytotherapy among the inhabitants of Parvati valley in western Himalaya, India," *Journal of Ethnopharmacology*, vol. 92, no. 2-3, pp. 167-176, 2004.
- [5] C. P. Kala, "Ethnomedicinal botany of the apatani in the eastern Himalayan region of India," *Journal of Ethnobiology and Ethnomedicine*, vol. 1, no. 1, p. 11, 2005.
- [6] T. B. Le, C. Beaufay, D. T. Nghiem, M.-P. Mingeot-Leclercq, and J. Quetin-Leclercq, "In vitro anti-leishmanial activity of essential oils extracted from Vietnamese plants," *Molecules*, vol. 22, no. 7, p. 1071, 2017.
- [7] S. Rath and R. N. Padhy, "Monitoring in vitro antibacterial efficacy of 26 Indian spices against multidrug resistant urinary tract infecting bacteria," *Integrative Medicine Research*, vol. 3, no. 3, pp. 133-141, 2014.
- [8] L. T. Vien, T. T. H. Hanh, P. T. T. Huong et al., "Pyrrole oligoglycosides from the starfish *Acanthaster planci* suppress lipopolysaccharide-induced nitric oxide production in RAW264.7 macrophages," *Chemical and Pharmaceutical Bulletin*, vol. 64, no. 11, pp. 1654-1657, 2016.
- [9] T. Mosmann, "Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays," *Journal of Immunological Methods*, vol. 65, no. 1-2, pp. 55-63, 1983.
- [10] S. Hatziieremia, A. I. Gray, V. A. Ferro, A. Paul, and R. Plevin, "The effects of cardamomin on lipopolysaccharide-induced inflammatory protein production and MAP kinase and NF κ B signalling pathways in monocytes/macrophages," *British Journal of Pharmacology*, vol. 149, no. 2, pp. 188-198, 2009.
- [11] Q. Cui, L.-T. Wang, J.-Z. Liu et al., "Rapid extraction of *Amomum tsao-ko* essential oil and determination of its chemical composition, antioxidant and antimicrobial activities," *Journal of Chromatography B*, vol. 1061-1062, pp. 364-371, 2017.
- [12] Y. Yang, Y. Yue, Y. Runwei, and Z. Guolin, "Cytotoxic, apoptotic and antioxidant activity of the essential oil of *Amomum tsao-ko*," *Bioresource Technology*, vol. 101, no. 11, pp. 4205-4211, 2010.
- [13] Y. Wang, C.-X. You, C.-F. Wang et al., "Chemical constituents and insecticidal activities of the essential oil from *Amomum tsaoko* against two stored-product insects," *Journal of Oleo Science*, vol. 63, no. 10, pp. 1019-1026, 2014.
- [14] S. A. Agnihotri, S. R. Wakode, and M. Ali, "Chemical composition, antimicrobial and topical anti-inflammatory activity of essential oil of *Amomum subulatum* fruits," *Acta Poloniae Pharmaceutica*, vol. 69, no. 6, pp. 1177-1181, 2012.
- [15] R. Joshi, P. Sharma, V. Sharma, R. Prasad, R. K. Sud, and A. Gulati, "Analysis of the essential oil of large cardamom (*Amomum subulatum*Roxb.) growing in different agro-climatic zones of himachal pradesh, India," *Journal of the Science of Food and Agriculture*, vol. 93, no. 6, pp. 1303-1309, 2013.
- [16] P. Satyal, N. S. Dosoky, B. L. Kincer, and W. N. Setzer, "Chemical compositions and biological activities of *Amomum subulatum* essential oils from Nepal," *Natural product communications*, vol. 7, no. 9, pp. 1233-1236, 2012.
- [17] S. Moncada, R. M. Palmer, and E. A. Higgs, "Nitric oxide: physiology, pathophysiology, and pharmacology," *Pharmacological Reviews*, vol. 43, no. 2, pp. 109-142, 1991.
- [18] K. Y. Kim, H. S. Lee, and G. H. Seol, "Eucalyptol suppresses matrix metalloproteinase-9 expression through an extracellular signal-regulated kinase-dependent nuclear factor-kappa B pathway to exert anti-inflammatory effects in an acute lung inflammation model," *Journal of Pharmacy and Pharmacology*, vol. 67, no. 8, pp. 1066-1074, 2015.
- [19] H. Worth, C. Schacher, and U. Dethlefsen, "Concomitant therapy with cineole (eucalyptole) reduces exacerbations in COPD: a placebo-controlled double-blind trial," *Respiratory Research*, vol. 10, no. 1, 2009.
- [20] A. T. Rufino, M. Ribeiro, F. Judas et al., "Anti-inflammatory and chondroprotective activity of (+)- α -Pinene: structural and enantiomeric selectivity," *Journal of Natural Products*, vol. 77, no. 2, pp. 264-269, 2014.
- [21] S. Held, P. Schieberle, and V. Somoza, "Characterization of α -terpineol as an anti-inflammatory component of orange juice by in vitro studies using oral buccal cells," *Journal of Agricultural and Food Chemistry*, vol. 55, no. 20, pp. 8040-8046, 2007.
- [22] J. Wang, B. Su, H. Zhu, C. Chen, and G. Zhao, "Protective effect of geraniol inhibits inflammatory response, oxidative stress and apoptosis in traumatic injury of the spinal cord through modulation of NF- κ B and p38 MAPK," *Experimental and Therapeutic Medicine*, vol. 12, no. 6, pp. 3607-3613, 2016.
- [23] P.-C. Liao, T.-S. Yang, J.-C. Chou et al., "Anti-inflammatory activity of neral and geraniol isolated from fruits of *Litsea cubeba* Lour," *Journal of Functional Foods*, vol. 19, pp. 248-258, 2015.
- [24] J.-S. Shin, S. Ryu, D. S. Jang, Y.-W. Cho, E. K. Chung, and K.-T. Lee, "*Amomum tsao-ko* fruit extract suppresses lipopolysaccharide-induced inducible nitric oxide synthase by inducing heme oxygenase-1 in macrophages and in septic mice," *International Journal of Experimental Pathology*, vol. 96, no. 6, pp. 395-405, 2015.
- [25] K. Lee, S. Kim, S. Sung, and Y. Kim, "Inhibitory constituents of lipopolysaccharide-induced nitric oxide production in BV2 microglia isolated from *Amomum tsao-ko*," *Planta Medica*, vol. 74, no. 8, pp. 867-869, 2008.
- [26] B. Li, H.-J. Choi, D.-S. Lee et al., "*Amomum tsao-ko* suppresses lipopolysaccharide-induced inflammatory responses in RAW264.7 macrophages via Nrf2-dependent heme oxygenase-1 expression," *The American Journal of Chinese Medicine*, vol. 42, no. 5, pp. 1229-1244, 2014.
- [27] T.-T. Zhang, C.-L. Lu, and J.-G. Jiang, "Neuroprotective and anti-inflammatory effects of diphenylheptanes from the fruits of *Amomum tsaoko*, a Chinese spice," *Plant Foods for Human Nutrition*, vol. 71, no. 4, pp. 450-453, 2016.
- [28] Z. Chen, W. Ni, C. Yang et al., "Therapeutic effect of *Amomum villosum* on inflammatory bowel disease in rats," *Frontiers in Pharmacology*, vol. 9, p. 639, 2018.
- [29] Y.-A. Choi, J. K. Choi, Y. H. Jang et al., "Anti-inflammatory effect of *Amomum xanthioides* in a mouse atopic dermatitis model," *Molecular Medicine Reports*, vol. 16, no. 6, pp. 8964-8972, 2017.
- [30] J.-A. Lee, M.-Y. Lee, I.-S. Shin, C.-S. Seo, H. Ha, and H. K. Shin, "Anti-inflammatory Effects of *Amomum compactum* on RAW 264.7 cells via induction of heme oxygenase-1," *Archives of Pharmacal Research*, vol. 35, no. 4, pp. 739-746, 2012.