

Research Article

Comparison of the Nutritional and Taste Characteristics of 5 Edible Fungus Powders Based on the Composition of Hydrolyzed Amino Acids and Free Amino Acids

Jian Li,^{1,2} Junmei Ma^(b),^{3,4,5} Sufang Fan,^{3,4,5} Shengquan Mi^(b),¹ and Yan Zhang^(b),^{3,4,5}

¹College of Applied Arts and Science, Beijing Union University, Beijing 100191, China

²School of Food and Biological Engineering, Hefei University of Technology, Hefei 230009, China

³Hebei Food Inspection and Research Institute, Hebei Food Safety Key Laboratory, Shijiazhuang 050091, China

⁴Key Laboratory of Special Food Supervision Technology for State Market Regulation, Shijiazhuang 050091, China

⁵Hebei Engineering Research Center for Special Food Safety and Health, Shijiazhuang 050091, China

Correspondence should be addressed to Shengquan Mi; msq65@buu.edu.cn and Yan Zhang; snowwinglv@126.com

Received 28 January 2022; Revised 22 February 2022; Accepted 2 March 2022; Published 7 April 2022

Academic Editor: Tao Feng

Copyright © 2022 Jian Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The nutritional characteristics and taste of some edible fungus powders were scientifically evaluated and compared. Five common edible fungus powders were used as test materials (Agrocybe chaxinggu edible fungus powder, Pleurotus citrinopileatus edible fungus powder, Flammulina velutipes edible fungus powder, Lentinus edodes edible fungus powder, and Hericium erinaceus edible fungus powder). The hydrolyzed amino acid and free amino acid content were measured by an automatic amino acid analyzer, and the ratios of hydrolyzed amino acid and free amino acid components and the taste characteristics of these eatables were systematically compared. The results showed that the total amount of hydrolyzed amino acids contained in the 5 edible fungus powders was between 2.583 and 14.656 g/100 g. The total amount of free amino acids contained in the 5 edible fungus powders was between 0.550 and 2.612 g/100 g. Comparative analysis of the mass fractions and composition of amino acids indicated that Pleurotus citrinopileatus edible fungus powder best met the ideal protein standard. The taste characteristics of protein were evaluated by calculating the taste active value (TAV) of taste-producing free amino acids. The most significant TAV values of the 5 edible fungus powders appeared in glutamic acid, and this amino acid is an umami amino acid. Principal component analysis (PCA) suggested that four principal components could reflect all the information on the free amino acids with a total cumulative variance contribution rate of 100%, and three principal components could reflect most of the information on the hydrolyzed amino acids with a total cumulative variance contribution rate of 99.143%, which could represent the main trends of free amino acids and hydrolyzed acids in edible fungus powder. The comprehensive evaluation model was established, and the comprehensive score indicated that Agrocybe chaxinggu edible fungus powder had the best comprehensive amino acid quality.

1. Introduction

Edible funguses are widely grown all over the world. There are at least 2,000 species of edible funguses in the world, of which about 200 are wild edible funguses [1]. China is the largest producer of edible funguses in the world, accounting for twothirds of the world's production [2, 3]. Edible funguses are widely cultivated for their medicinal and nutritional value. Some edible funguses have been reported as therapeutic foods, useful in preventing diseases such as hypertension, hypercholesterolemia, atherosclerosis, or cancer [4–7]. Edible funguses are valuable health foods, low in calories, lipids, and essential fatty acids, and high in vegetable proteins, minerals, and vitamins [8]. As a delicacy, edible funguses have unique umami, texture, and increasingly high-rated nutritive value. Umami is produced by glutamic acid, ribonucleotide and chemicals that make edible funguses tasty and are widely used in food preparation [2].

An amino acid is a biologically active substance that plays an important role in human metabolism. Its composition and content are important indicators for evaluating the nutritional value of edible funguses [9, 10]. Free amino acids (FAA), also known as nonprotein amino acids, are an important class of taste active ingredients [11], and their content and types are often used as important indicators for food nutritional value and taste and taste evaluation [12]. Some studies have shown that the main taste substances that have a significant impact on the taste of edible funguses are nucleotides, soluble sugars, organic acids, free amino acids, and other taste substances. Among them, free amino acids play an extremely important role in the presentation of the taste and deliciousness of edible funguses [13]. The typical umami taste of edible funguses is attributed to aspartic acid and glutamate [2]. Umami not only alleviates salty, sour or bitter tastes and improves the perception of sweetness, it also reduces the pungent meaty smell and the earthy taste [14, 15]. At present, the analytical methods for amino acids in edible funguses include ninhydrin colorimetry [16], high performance liquid chromatography [14, 17, 18], gas chromatography [19], gas chromatography-mass spectrometry [6, 20], nuclear magnetic resonance [21], amino acid analyzers [22-27], and high performance liquid chromatography-triple quadrupole mass spectrometry [28]. In the present study, the amino acid analyzer was used to analyze the nutritional characteristics of hydrolyzed amino acids and free amino acids of five common edible funguses powder, and their differences in nutritional and taste components were compared. The research results not only provide scientific data for revealing the nutritional value and taste characteristics of edible funguses, but also provide a scientific basis for guiding people to establish a scientific and healthy diet structure.

2. Materials and Methods

2.1. Materials and Reagents. Aspartic acid, threonine, serine, glutamic acid, glycine, alanine, valine, methionine, isoleucine, leucine, tyrosine, phenylalanine, lysine, histidine, arginine, and proline mixed standard solution were obtained from Wako Pure Chemical Industries, Ltd. (2.5 µmol/mL, Tokyo, Japan). Water was purified using a Milli-Q-System (Millipore, Guyancourt, France). Ethanol was HPLC grade and was purchased from Merck (Darmstadt, Germany). Sodium citrate dihydrate, citric acid monohydrate, sodium chloride, sodium hydroxide, sodium borohydride, and hydrochloric acid were premium grade pure and were purchased from Beijing Chemical Reagent Factory (Beijing, China). Anhydrous sodium acetate, ninhydrin, ethylene glycol monomethyl ether, acetic acid, and phenol were analytical grade and were purchased from the Guangfu Fine Chemical Research Institute (Tianjin, China).

Agrocybe chaxinggu edible fungus powder, Pleurotus citrinopileatus edible fungus powder, Flammulina velutipes edible fungus powder, Lentinus edodes edible fungus powder, and Hericium erinaceus edible fungus powder were supplied by Henan Longfeng Edible Funguses Industry Research Institute.

2.2. Methods

2.2.1. Determination of Hydrolyzed Amino Acids. Refer to GB 5009.124-2016 [29] for the determination of hydrolyzed amino acids. 0.5 g of each sample was hydrolyzed by an electric heating blast drying oven (Boxun GZX-9240MBE, China) with 10 mL of 6 mol/L hydrogen chloride at 110°C for 22 h under a nitrogen atmosphere, and then filtered through a 0.45 mm membrane filter prior to analysis. The amino acid profiles of each sample were determined by an automatic amino acid analyzer (Hitachi L-8900, Japan). All determinations were carried out in triplicate.

2.2.2. Determination of Free Amino Acids. Refer to the first method in GB/T 30987-2020 [30] for the determination of free amino acids. 0.5 g of the sample was diluted with 100 mL of boiling water. The sample was water bathed by a water bath constant temperature oscillator (Runhua SHA-B, China) at 95°C for 10 min. and then filtered through a 0.45 mm membrane filter prior to analysis. The amino acid profiles of each sample were determined by an automatic amino acid analyzer (Hitachi L-8900, Japan). All determinations were carried out in triplicate.

2.3. Statistical Analysis. All analyses were conducted in triplicate. The results reported were the average of these three replicates. Normal distribution tests of multielements and PCA analysis were performed with SPSS 25.0 software (SPSS, IBM Corp., USA).

3. Results and Discussion

3.1. Hydrolyzed Amino Acid Concentrations in 5 Edible Fungus Powders. The hydrolyzed amino acid is to hydrolyze the protein, polypeptide, and other amino acid chains in the plant to be tested into a single amino acid through acid hydrolysis. Therefore, the hydrolyzed amino acid can fully reflect the kind and content of all single amino acids in the sample. The hydrolyzed amino acid composition and content of 5 kinds of edible fungus powder are shown in Table 1. Results showed that 15 hydrolyzed amino acids were present in all samples tested. The total amount of hydrolyzed amino acids contained in the 5 edible fungus powders was between 2.583 and 14.656 g/100 g. The total hydrolyzed amino acid content of Pleurotus citrinopileatus edible fungus powder was the highest, reaching 14.656 g/100 g, and the total hydrolyzed amino acid content of Hericium erinaceus edible fungus powder was the lowest, at 2.583 g/100 g. Glutamic acid was the most abundant among the 5 kinds of edible fungus powder, accounting for 17.41%, 19.75%, 25.16%, 24.92%, and 16.72% of the total hydrolyzed amino acids, respectively.

3.2. Free Amino Acid Concentrations in 5 Edible Fungus Powders. Free amino acids refer to amino acids that exist in a free state in plants as a single amino acid molecule and can be directly absorbed and utilized. The free amino acid composition and content of 5 kinds of edible fungus powder are shown in Table 2. The total amount of free amino acids

| Compound | <i>Agrocybe chaxinggu</i> edible fungus powder | Pleurotus citrinopileatus edible fungus powder | <i>Flammulina velutipes</i> edible fungus powder | <i>Lentinus edodes</i> edible fungus powder | <i>Hericium erinaceus</i> edible fungus powder |
|------------------------------|--|---|--|---|--|
| Aspartic acid and asparagine | 1.571 ± 0.015 | 0.613 ± 0.001 | 0.531 ± 0.002 | 0.737 ± 0.002 | 0.247 ± 0.005 |
| Threonine | 0.744 ± 0.012 | 0.510 ± 0.009 | 0.147 ± 0.003 | 0.192 ± 0.002 | 0.107 ± 0.005 |
| Serine | 0.768 ± 0.003 | 0.469 ± 0.006 | 0.280 ± 0.006 | 0.399 ± 0.004 | 0.099 ± 0.003 |
| Glutamic acid and glutamine | 2.536 ± 0.012 | 2.894 ± 0.001 | 2.198 ± 0.031 | 2.969 ± 0.030 | 0.432 ± 0.001 |
| Glycine | 0.799 ± 0.008 | 0.867 ± 0.004 | 0.468 ± 0.004 | 0.650 ± 0.002 | 0.165 ± 0.003 |
| Alanine | 1.491 ± 0.004 | 1.427 ± 0.059 | 1.065 ± 0.024 | 1.859 ± 0.014 | 0.414 ± 0.005 |
| Valine | 1.021 ± 0.014 | 1.202 ± 0.023 | 0.585 ± 0.002 | 0.809 ± 0.014 | 0.207 ± 0.012 |
| Methionine | 0.022 ± 0.002 | 0.113 ± 0.011 | ND | ND | ND |
| Isoleucine | 0.583 ± 0.011 | 0.677 ± 0.003 | 0.301 ± 0.004 | 0.423 ± 0.005 | 0.068 ± 0.002 |
| Leucine | 1.379 ± 0.017 | 1.703 ± 0.037 | 0.821 ± 0.001 | 1.135 ± 0.009 | 0.298 ± 0.002 |
| Tyrosine | 0.507 ± 0.004 | 0.636 ± 0.008 | 0.414 ± 0.001 | 0.406 ± 0.015 | 0.111 ± 0.000 |
| Phenylalanine | 0.714 ± 0.010 | 0.849 ± 0.004 | 0.537 ± 0.003 | 0.567 ± 0.025 | 0.112 ± 0.006 |
| Lysine | 0.754 ± 0.002 | 1.082 ± 0.006 | 0.622 ± 0.002 | 0.710 ± 0.005 | 0.102 ± 0.001 |
| Histidine | 0.300 ± 0.004 | 0.410 ± 0.005 | 0.217 ± 0.003 | 0.257 ± 0.003 | 0.057 ± 0.003 |
| Arginine | 0.943 ± 0.009 | 1.204 ± 0.027 | 0.554 ± 0.009 | 0.799 ± 0.012 | 0.165 ± 0.001 |
| Proline | 0.434 ± 0.016 | ND | ND | ND | ND |
| Total | 14.567 ± 0.024 | 14.656 ± 0.287 | 8.739 ± 0.016 | 11.912 ± 0.092 | 2.583 ± 0.010 |

TABLE 1: Contents* of hydrolyzed amino acids in 5 kinds of edible fungus powders.

*Value $(g/100 g) = mean \pm SD (n = 3)$. ND: not detectable.

TABLE 2: Contents* of free amino acids in 5 kinds of edible fungus powders.

| Compound | <i>Agrocybe chaxinggu</i> edible fungus powder | <i>Pleurotus citrinopileatus</i> edible fungus powder | <i>Flammulina velutipes</i> edible fungus powder | <i>Lentinus edodes</i> edible fungus powder | <i>Hericium erinaceus</i> edible fungus powder |
|---------------|--|---|---|---|--|
| Aspartic acid | 0.070 ± 0.001 | 0.016 ± 0.000 | 0.018 ± 0.000 | 0.022 ± 0.000 | 0.016 ± 0.001 |
| Threonine | 0.139 ± 0.003 | 0.286 ± 0.000 | 0.373 ± 0.001 | 0.327 ± 0.004 | 0.037 ± 0.001 |
| Serine | 0.079 ± 0.002 | 0.107 ± 0.000 | 0.068 ± 0.002 | 0.015 ± 0.000 | 0.004 ± 0.000 |
| Glutamic acid | 0.505 ± 0.003 | 0.396 ± 0.000 | 0.739 ± 0.003 | 0.448 ± 0.007 | 0.100 ± 0.002 |
| Glycine | 0.047 ± 0.000 | 0.032 ± 0.000 | 0.049 ± 0.000 | 0.022 ± 0.000 | 0.019 ± 0.001 |
| Alanine | 0.326 ± 0.002 | 0.207 ± 0.000 | 0.409 ± 0.001 | 0.135 ± 0.002 | 0.085 ± 0.003 |
| Valine | 0.164 ± 0.002 | 0.117 ± 0.000 | 0.118 ± 0.000 | 0.053 ± 0.001 | 0.047 ± 0.002 |
| Methionine | ND | 0.007 ± 0.000 | ND | ND | ND |
| Isoleucine | 0.101 ± 0.001 | 0.033 ± 0.000 | 0.060 ± 0.000 | 0.012 ± 0.000 | 0.022 ± 0.000 |
| Leucine | 0.168 ± 0.002 | 0.063 ± 0.000 | 0.103 ± 0.000 | 0.017 ± 0.000 | 0.064 ± 0.002 |
| Tyrosine | 0.064 ± 0.001 | 0.086 ± 0.001 | 0.159 ± 0.000 | 0.032 ± 0.001 | 0.038 ± 0.001 |
| Phenylalanine | 0.115 ± 0.002 | 0.131 ± 0.000 | 0.216 ± 0.001 | 0.046 ± 0.000 | 0.046 ± 0.003 |
| Lysine | 0.066 ± 0.002 | 0.079 ± 0.000 | 0.151 ± 0.001 | 0.035 ± 0.000 | 0.007 ± 0.000 |
| Histidine | 0.027 ± 0.000 | 0.028 ± 0.000 | 0.052 ± 0.000 | 0.009 ± 0.000 | 0.008 ± 0.000 |
| Arginine | 0.106 ± 0.002 | 0.062 ± 0.000 | 0.057 ± 0.001 | 0.055 ± 0.001 | 0.024 ± 0.001 |
| Proline | 0.026 ± 0.002 | ND | 0.039 ± 0.002 | ND | 0.034 ± 0.002 |
| Total | 2.003 ± 0.018 | 1.651 ± 0.001 | 2.612 ± 0.005 | 1.228 ± 0.017 | 0.550 ± 0.018 |

*Value $(g/100 g) = mean \pm SD (n = 3)$. ND: not detectable.

contained in the 5 edible fungus powders was between 0.550 and 2.612 g/100 g. The total free amino acid content of *Flammulina velutipes* edible fungus powder was the highest, reaching 2.612 g/100 g, and the total free amino acid content of *Hericium erinaceus* edible fungus powder was the lowest, at 0.550 g/100 g. Glutamic acid was the most abundant among the 5 kinds of edible fungus powders, accounting for 25.21%, 23.99%, 28.29%, 36.48%, and 18.18% of the total free amino acids, respectively.

3.3. The Difference in the Ratio of Essential Amino Acids and Nonessential Amino Acids. In order to scientifically evaluate the component structure of hydrolyzed amino acids, indicators such as the ratio of essential amino acids (EAAs) to nonessential amino acids (NEAAs) have been introduced. According to the essential amino acid model of protein nutritional value proposed by the World Health Organization (WHO) and the United Nations Food and Agriculture Organization (FAO) in 1973, the EAA/(EAA + NEAA)

TABLE 3: Comparative analysis of the mass fractions and composition of amino acids in 5 edible fungus powders.

| Sample | EAA/NEAA | EAA/(EAA + NEAA) |
|--|----------|------------------|
| Agrocybe chaxinggu edible fungus powder | 55.81 | 35.82 |
| Pleurotus citrinopileatus edible fungus powder | 72.03 | 41.87 |
| Flammulina velutipes edible fungus powder | 52.62 | 34.48 |
| Lentinus edodes edible fungus powder | 47.48 | 32.20 |
| Hericium erinaceus edible fungus powder | 52.85 | 34.58 |

TABLE 4: Flavored amino acid content (g/100 g) in 5 kinds of edible fungus powders.

| Sample | Umami amino acids | Sweet amino acids | Bitterness amino acids | Tasteless amino acids |
|--|-------------------|-------------------|------------------------|-----------------------|
| Agrocybe chaxinggu edible fungus powder | 0.574 | 0.616 | 0.682 | 0.131 |
| Pleurotus citrinopileatus edible fungus powder | 0.412 | 0.633 | 0.441 | 0.163 |
| Flammulina velutipes edible fungus powder | 0.757 | 0.969 | 0.606 | 0.310 |
| Lentinus edodes edible fungus powder | 0.469 | 0.499 | 0.193 | 0.067 |
| Hericium erinaceus edible fungus powder | 0.115 | 0.179 | 0.211 | 0.045 |

TABLE 5: The TAV of free amino acid in 5 kinds of edible fungus powders.

| | Teste | Tests threads ald | | TAV of 5 kind | s of edible fungus | powders | |
|---------------|--------------------------|------------------------------------|-----------------------|------------------------------|-------------------------|--------------------|-----------------------|
| Compound | Taste characteristics | Taste threshold (mg/100 g) [34] | Agrocybe chaxinggu | Pleurotus citrinopileatus | Flammulina velutipes | Lentinus edodes | Hericium erinaceus |
| Aspartic acid | Umami | 3.00 | 23.22 | 5.30 | 6.03 | 7.29 | 5.24 |
| Threonine | Sweet | 260.00 | 0.53 | 1.10 | 1.44 | 1.26 | 0.14 |
| Serine | Sweet | 150.00 | 0.52 | 0.72 | 0.45 | 0.10 | 0.02 |
| Glutamic acid | Umami | 5.00 | 100.93 | 79.30 | 147.84 | 89.50 | 19.95 |
| Glycine | Sweet | 110.00 | 0.42 | 0.29 | 0.45 | 0.20 | 0.17 |
| Alanine | Sweet | 60.00 | 5.44 | 3.45 | 6.82 | 2.25 | 1.42 |
| Valine | Bitterness | 150.00 | 1.09 | 0.78 | 0.78 | 0.35 | 0.32 |
| Methionine | Bitterness | 30.00 | 0.00 | 0.24 | 0.00 | 0.00 | 0.00 |
| Isoleucine | Bitterness | 90.00 | 1.13 | 0.37 | 0.67 | 0.14 | 0.25 |
| Leucine | Bitterness | 380.00 | 0.44 | 0.17 | 0.27 | 0.04 | 0.17 |
| Tyrosine | Tasteless | 260.00 | 0.25 | 0.33 | 0.61 | 0.12 | 0.15 |
| Phenylalanine | Bitterness | 150.00 | 0.77 | 0.87 | 1.44 | 0.31 | 0.31 |
| Lysine | Tasteless | 50.00 | 1.33 | 1.59 | 3.02 | 0.69 | 0.14 |
| Histidine | Bitterness | 20.00 | 1.34 | 1.39 | 2.60 | 0.44 | 0.38 |
| Arginine | Bitterness | 10.00 | 10.64 | 6.23 | 5.73 | 5.53 | 2.38 |
| Proline | Sweet | 300.00 | 0.09 | 0.00 | 0.13 | 0.00 | 0.11 |

value in an ideal protein should reach about 40%, and the EAA/NEAA value should be above 60% [31]. The results (Table 3) showed that the value of EAA/(EAA + NEAA) was between 32.20% and 41.87%, and the value of EAA/NEAA was 47.48%–72.03%. In contrast, among the 5 kinds of edible fungus powders, *Lentinus edodes* edible fungus powder had the lowest EAA/(EAA + NEAA) and EAA/NEAA values, and *Pleurotus citrinopileatus* edible fungus powder best met the ideal protein standard.

3.4. The Difference in Taste Characteristics and Taste Activity Values. The determination of hydrolyzed amino acids was mainly to study their nutritional properties. The flavored amino acids in structural proteins are mostly in a combined state and have little effect on flavor, while free amino acids are mainly used to participate in the formation of taste substances. Therefore, the taste characteristics of free amino acids in the five test samples were compared, and they were divided into 4 categories: umami amino acids, sweet amino acids, bitterness amino acids, and tasteless

amino acids [32]. The results (Table 4) showed that *Pleurotus citrinopileatus* edible fungus powder, *Flammulina velutipes* edible fungus powder, and *Lentinus edodes* edible fungus powder had the highest content of sweet amino acids. *Agrocybe chaxinggu* edible fungus powder and *Hericium erinaceus* edible fungus powder had the highest content of bitterness amino acids.

The absolute content and relative ratio of flavored amino acids may be closely related to the taste of the food. Therefore, the taste active value (TAV) of 16 amino acids in 5 kinds of test samples was analyzed and compared. TAV is the ratio of the content of each taste amino acid in the sample to its corresponding taste threshold [33]. In general, when TAV > 1, the taste-producing substance is considered to have a significant impact on the taste-producing effect of the sample; when TAV < 1, it means that the substance has no significant taste-producing effect [34]. The results (Table 5) showed that among the 5 edible fungus powders, the TAV values of aspartic acid, glutamic acid, alanine, and arginine were also greater than 1. The TAV values of serine, glycine, methionine, leucine, tyrosine, and proline were also

| | | | | TABLE 6: | Correlatio | n analysis | s of free a | TABLE 6: Correlation analysis of free amino acids in 5 kinds of edible fungus powders. | in 5 kinds o | of edible f | wod sngur | lers. | | | | |
|--|------------------|------------------|-------------|-------------------|--------------|-------------|-------------|--|--------------|-------------|--------------|---|--------------|-----------|------------|--------|
| Amino acids | Aspartic acid | Threonine Serine | Serine | Glutamic acid | Glycine | Alanine | Valine N | Aethionine | Isoleucine | Leucine | l'yrosine P | Glycine Alanine Valine Methionine Isoleucine Leucine Tyrosine Phenylalanine Lysine Histidine Arginine Proline | Lysine 1 | Histidine | Arginine F | roline |
| Aspartic acid | 1 | | | | | | | | | | | | | | | |
| Threonine | -0.320 | 1 | | | | | | | | | | | | | | |
| Serine | 0.271 | 0.349 | 1 | | | | | | | | | | | | | |
| Glutamic acid | 0.200 | 0.783 | 0.502 | 1 | | | | | | | | | | | | |
| Glycine | 0.519 | 0.333 | 0.680 | 0.819 | 1 | | | | | | | | | | | |
| Alanine | 0.385 | 0.457 | 0.624 | 0.888^{*} | 0.983^{**} | 1 | | | | | | | | | | |
| Valine | 0.699 | 0.122 | 0.839 | 0.583 | 0.889^{*} | 0.799 | 1 | | | | | | | | | |
| Methionine | -0.296 | 0.214 | 0.670 | -0.101 | -0.073 | -0.106 | 0.195 | 1 | | | | | | | | |
| Isoleucine | 0.842 | -0.119 | 0.545 | 0.493 | 0.867 | 0.772 | 0.910^{*} | -0.197 | 1 | | | | | | | |
| Leucine | 0.802 | -0.276 | 0.493 | 0.353 | 0.807 | 0.702 | 0.860 | -0.198 | 0.980^{**} | 1 | | | | | | |
| Tyrosine | -0.153 | 0.577 | 0.559 | 0.767 | 0.763 | 0.843 | 0.492 | 0.111 | 0.379 | 0.351 | 1 | | | | | |
| Phenylalanine | 0.007 | 0.557 | 0.682 | 0.808 | 0.856 | 0.908^{*} | 0.646 | 0.160 | 0.515 | 0.475 | 0.982^{**} | 1 | | | | |
| Lysine | -0.023 | 0.697 | 0.634 | 0.894^{*} | 0.833 | 0.906^{*} | 0.591 | 0.117 | 0.449 | 0.375 | 0.968^{**} | 0.981^{**} | 1 | | | |
| Histidine | 0.047 | 0.555 | 0.657 | 0.832 | 0.878 | 0.930^{*} | 0.655 | 0.100 | 0.546 | 0.501 | 0.978^{**} | 0.998** | 0.983^{**} | 1 | | |
| Arginine | 0.874 | 0.119 | 0.624 | 0.527 | 0.707 | 0.605 | 0.875 | 0.023 | 0.832 | 0.728 | 0.134 | 0.311 | 0.320 | 0.337 | 1 | |
| Proline | 0.146 | -0.343 | -0.173 | 0.118 | 0.419 | 0.431 | 0.159 | -0.593 | 0.449 | 0.556 | 0.432 | 0.382 | 0.284 | 0.408 | -0.102 | 1 |
| Note: relevance is Pearson's type; *significant correlation ($p < 0.05$); ** extremely significant correlation ($p < 0.01$). | ; Pearson's | type; *signific | ant correls | ation $(p < 0)$. | 05); **extre | mely signi | ficant corr | elation $(p < 0$. | 01). | | | | | | | |

| Component | | Initial eigenva | lue |] | Rotate the sum of squa | ares loading |
|-----------|-------|-----------------|----------------|-------|------------------------|----------------|
| Component | Total | Variance (%) | Accumulate (%) | Total | Variance (%) | Accumulate (%) |
| 1 | 9.241 | 57.759 | 57.759 | 7.125 | 44.532 | 44.532 |
| 2 | 3.455 | 21.592 | 79.351 | 5.259 | 32.869 | 77.402 |
| 3 | 2.183 | 13.645 | 92.996 | 1.916 | 11.977 | 89.378 |
| 4 | 1.121 | 7.004 | 100 | 1.699 | 10.622 | 100 |

TABLE 7: Results of principal component analysis.

TABLE 8: Contribution value of element principal component.

| A | | Com | ponent | |
|---------------|-------|--------|--------|--------|
| Amino acids | 1 | 2 | 3 | 4 |
| Aspartic acid | 0.432 | 0.854 | 0.178 | -0.228 |
| Threonine | 0.417 | -0.750 | 0.176 | -0.482 |
| Serine | 0.745 | -0.103 | 0.589 | 0.296 |
| Glutamic acid | 0.843 | -0.301 | -0.045 | -0.443 |
| Glycine | 0.991 | 0.112 | -0.064 | -0.031 |
| Alanine | 0.984 | -0.040 | -0.148 | -0.089 |
| Valine | 0.882 | 0.334 | 0.315 | 0.104 |
| Methionine | 0.037 | -0.397 | 0.776 | 0.488 |
| Isoleucine | 0.809 | 0.587 | -0.011 | 0.029 |
| Leucine | 0.743 | 0.634 | -0.089 | 0.195 |
| Tyrosine | 0.814 | -0.490 | -0.256 | 0.176 |
| Phenylalanine | 0.902 | -0.377 | -0.131 | 0.163 |
| Lysine | 0.884 | -0.454 | -0.112 | -0.017 |
| Histidine | 0.917 | -0.344 | -0.164 | 0.117 |
| Arginine | 0.675 | 0.510 | 0.443 | -0.295 |
| Proline | 0.349 | 0.253 | -0.847 | 0.311 |

less than 1. The TAV of free amino acid in Agrocybe chaxinggu edible fungus powder was between 0.00 and 100.93. The TAV of free amino acid in Pleurotus citrinopileatus edible fungus powder was between 0.00 and 79.30. The TAV of free amino acid in Flammulina velutipes edible fungus powder was between 0.00 and 147.84. The TAV of free amino acid in Lentinus edodes edible fungus powder was between 0.00 and 89.50. The TAV of free amino acid in Hericium erinaceus edible fungus powder was between 0.00 and 19.95. According to the TAV value, the most significant free amino acid that affects the taste of the 5 edible fungus powders that can be screened out was glutamic acid in the umami amino acid, followed by aspartic acid in umami amino acids, arginine in bitterness amino acids, and alanine in sweet amino acids. And Flammulina velutipes edible fungus powder has the most outstanding umami.

3.5. Correlation Analysis and PCA of Free Amino Acids. A correlation analysis was performed on the 16 free amino acid components of 5 edible fungus powders, and the results are shown in Table 6. There were positive correlations and negative correlations between amino acids, and most of them were positive. The results showed that there was a strong correlation between the free amino acids of the five edible fungus powders, which could be comprehensively evaluated by PCA.

PCA is a multivariate statistical analysis method that analyses a few variables which can reveal the internal structure sufficiently by studying the relationship between multiple original variables [35]. According to the rule that the characteristic value is greater than 1 and the cumulative variance contribution rate is greater than 80%, four principal component factors were obtained through rotation and extraction factors, and the total contribution rate was 100%, indicating that the experimental data can fully reflect the original information (Table 7).

The first principal component was mainly composed of glycine, alanine, histidine, phenylalanine, lysine, valine, glutamic acid, tyrosine, isoleucine, serine, leucine, and arginine. The second principal component was mainly composed of aspartic acid, leucine, isoleucine, and arginine. And the third principal component was mainly composed of methionine and serine (Table 8).

3.6. Correlation Analysis and PCA of Hydrolyzed Amino Acids. A correlation analysis was performed on the 16 hydrolyzed amino acid components of 5 edible fungus powders, and the results are shown in Table 9. The results showed that the correlation coefficients between most hydrolyzed amino acids were greater than 0.7, indicating that there were strong correlations among the 16 hydrolyzed amino acids in 5 kinds of edible fungus powders, so they could be further studied by PCA.

PCA was performed on the hydrolyzed amino acid content of the 5 samples, and the results are shown in Tables 10 and 11. It can be seen from Tables 10 and 11 that the cumulative contribution rate of the three principal components reaches 99.143%, indicating that three

| | | | | | CONTRACTOR | in anna 1 | | innual configuration and the animal and a second and as | | AT ATATA T | ind man | | | | | |
|---|------------------|------------------|------------|-------------------|---------------|-------------|---------------|--|--------------|--------------|--------------|---|--------------|-------------|------------|--------|
| Amino acids | Aspartic acid | Threonine Serine | Serine | Glutamic acid | Glycine | Alanine | Valine 1 | Methionine | Isoleucine | Leucine | Tyrosine H | Glycine Alanine Valine Methionine Isoleucine Leucine Tyrosine Phenylalanine Lysine Histidine Arginine Proline | Lysine | Histidine / | Arginine P | roline |
| Aspartic acid | 1 | | | | | | | | | | | | | | | |
| Threonine | 0.853 | 1 | | | | | | | | | | | | | | |
| Serine | 0.953^{*} | 0.927^{*} | 1 | | | | | | | | | | | | | |
| Glutamic acid | 0.513 | 0.481 | 0.684 | 1 | | | | | | | | | | | | |
| Glycine | 0.663 | 0.785 | 0.852 | 0.903^{*} | 1 | | | | | | | | | | | |
| Alanine | 0.576 | 0.444 | 0.692 | 0.948^{*} | 0.832 | 1 | | | | | | | | | | |
| Valine | 0.607 | 0.783 | 0.817 | 0.875 | 0.995^{**} | 0.785 | 1 | | | | | | | | | |
| Methionine | 0.041 | 0.519 | 0.319 | 0.418 | 0.648 | 0.235 | 0.724 | 1 | | | | | | | | |
| Isoleucine | 0.630 | 0.807 | 0.834 | 0.861 | 0.994^{**} | 0.769 | 0.999^{**} | 0.721 | 1 | | | | | | | |
| Leucine | 0.568 | 0.758 | 0.787 | 0.876 | 0.990^{**} | 0.780 | 0.999^{**} | 0.746 | 0.996^{**} | 1 | | | | | | |
| Tyrosine | 0.515 | 0.688 | 0.734 | 0.889^{*} | 0.956^{*} | 0.734 | 0.964^{**} | 0.712 | 0.963^{**} | 0.967** | 1 | | | | | |
| Phenylalanine | 0.573 | 0.714 | 0.779 | 0.910^{*} | 0.974^{**} | 0.777 | 0.975** | 0.673 | 0.974^{**} | 0.975** | 0.996^{**} | 1 | | | | |
| Lysine | 0.435 | 0.612 | 0.673 | 0.917^{*} | 0.952^{*} | 0.777 | 0.962^{**} | 0.728 | 0.955^{*} | 0.970^{**} | 0.989^{**} | 0.985^{**} | 1 | | | |
| Histidine | 0.480 | 0.686 | 0.717 | 0.886^{*} | 0.969^{**} | 0.753 | 0.983^{**} | 0.769 | 0.979^{**} | 0.989^{**} | 0.988^{**} | 0.986^{**} | 0.993^{**} | 1 | | |
| Arginine | 0.548 | 0.737 | 0.772 | 0.885^{*} | 0.988^{**} | 0.790 | 0.997** | 0.746 | 0.993^{**} | 0.999^{**} | 0.966^{**} | 0.974^{**} | 0.974^{**} | 0.990** | 1 | |
| Proline | 0.932^{*} | 0.817 | 0.824 | 0.178 | 0.414 | 0.245 | 0.369 | -0.057 | 0.402 | 0.324 | 0.266 | 0.318 | 0.158 | 0.225 | 0.297 | 1 |
| Note: relevance is Pearson's type; * significant correlation ($p < 0.05$); ** extremely significant correlation ($p < 0.01$). | Pearson's | type; *signific | ant correl | ation $(p < 0)$. | .05); ** extr | emely sign. | ificant corre | elation $(p < 0)$ | 01). | | | | | | | |

TABLE 9: Correlation analysis of free amino acids in 5 kinds of edible funguses powder.

| Common ont | | Initial eigenval | lue |] | Rotate the sum of squ | ares loading |
|------------|--------|------------------|----------------|-------|-----------------------|----------------|
| Component | Total | Variance (%) | Accumulate (%) | Total | Variance (%) | Accumulate (%) |
| 1 | 12.568 | 78.547 | 78.547 | 6.213 | 38.834 | 38.834 |
| 2 | 2.294 | 14.335 | 92.882 | 5.491 | 34.318 | 73.152 |
| 3 | 1.002 | 6.261 | 99.143 | 4.158 | 25.99 | 99.143 |

TABLE 10: Results of principal component analysis.

TABLE 11: Contribution value of element principal component.

| | | Component | |
|---------------|-------|-----------|--------|
| Amino acids | 1 | 2 | 3 |
| Aspartic acid | 0.674 | 0.732 | -0.100 |
| Threonine | 0.805 | 0.474 | 0.355 |
| Serine | 0.861 | 0.508 | -0.018 |
| Glutamic acid | 0.889 | -0.176 | -0.419 |
| Glycine | 0.998 | -0.017 | -0.035 |
| Alanine | 0.805 | -0.027 | -0.562 |
| Valine | 0.994 | -0.077 | 0.049 |
| Methionine | 0.657 | -0.467 | 0.582 |
| Isoleucine | 0.996 | -0.045 | 0.073 |
| Leucine | 0.989 | -0.126 | 0.054 |
| Tyrosine | 0.964 | -0.185 | 0.022 |
| Phenylalanine | 0.980 | -0.126 | -0.023 |
| Lysine | 0.952 | -0.288 | -0.037 |
| Histidine | 0.971 | -0.232 | 0.047 |
| Arginine | 0.985 | -0.152 | 0.036 |
| Proline | 0.436 | 0.889 | 0.131 |

TABLE 12: Comprehensive scores of 5 edible funguses powder.

| Samula | Free amino acid | | Hydrolyzed amino a | acid |
|--|---------------------|------|---------------------|------|
| Sample | Comprehensive score | Rank | Comprehensive score | Rank |
| Agrocybe chaxinggu edible fungus powder | 2.040 | 1 | 2.027 | 1 |
| Pleurotus citrinopileatus edible fungus powder | 0.065 | 3 | 1.180 | 2 |
| Flammulina velutipes edible fungus powder | 0.733 | 2 | -0.844 | 4 |
| Lentinus edodes edible fungus powder | -1.424 | 5 | -0.381 | 3 |
| Hericium erinaceus edible fungus powder | -1.413 | 4 | -1.979 | 5 |

components can represent the information of all hydrolyzed amino acids in 5 kinds of edible fungus powders and can better reflect the relationship of free amino acids in 5 kinds of edible fungus powders.

The first principal component was mainly composed of glycine, isoleucine, valine, leucine, arginine, phenylalanine, histidine, tyrosine, lysine, glutamic acid, serine, alanine, threonine, aspartic acid, and methionine. The second principal component was mainly composed of proline, aspartic acid, and serine. And the third principal component was mainly composed of methionine (Table 11).

3.7. Comprehensive Evaluation. A comprehensive evaluation model was established based on the contribution rates of the eigenvalues corresponding to the three principal components of hydrolyzed amino acids and the four principal components of free amino acids. The free amino acid model was $F = 0.445F_1 + 0.329F_2 + 0.120F_3 + 0.106F_4$. The hydrolyzed amino acid model was $F' = 0.392F'_1 + 0.346F'_2 + 0.262F'_3$. A comprehensive score was calculated for each sample, followed

by ranking and evaluating the amino acid content of each sample. It can be seen from Table 12 that the comprehensive scores of hydrolyzed amino acids and free amino acids of *Agrocybe chaxinggu* edible fungus powder were greater than those of other varieties, indicating that the comprehensive quality of hydrolyzed amino acids and free amino acids of this variety was higher, and it is a variety with better amino acid quality.

4. Conclusions and Discussion

In this study, the composition and content of free amino acids and hydrolyzed amino acids of *Agrocybe chaxinggu* edible fungus powder, *Pleurotus citrinopileatus* edible fungus powder, *Flammulina velutipes* edible fungus powder, *Lentinus edodes* edible fungus powder, and *Hericium erinaceus* edible fungus powder were analyzed. The results showed that the total amount of hydrolyzed amino acids contained in the 5 edible fungus powders was between 2.583 and 14.656 g/100 g. The total amount of free amino acids contained in the 5 edible fungus powders was between 0.550 and 2.612 g/100 g. The total amount of hydrolyzed amino acids was relatively large, which may be caused by the simultaneous thermal degradation and Maillard reaction, which hydrolyzed macromolecular proteins or polypeptides into small molecular amino acids. Comparative analysis of the mass fractions and composition of amino acids indicated that Pleurotus citrinopileatus edible fungus powder best met the ideal protein standard. The delicious taste of edible fungus powder is determined by the balance and mutual influence of different free amino acids, which play an important role in the taste of edible fungus powder. In this study, the TAV of free amino acids of five edible fungus powders was evaluated. The TAV of amino acids is related to water solubility. Water-soluble amino acids may be partly due to hydrolysis, or may be due to water-solubility in nature, which was related to the structure and properties of the amino acids themselves [36]. The most significant free amino acid that affects the taste of the 5 edible fungus powders which can be screened out was the glutamic acid in the umami amino acid. And Flammulina velutipes edible fungus powder has the most outstanding umami. However, the amino acids alone might not be good enough to be responsible for the taste. Fatty acids might also contribute a lot in forming the taste. The taste of edible fungus powder was systematically evaluated, and the results of fatty acids need to be determined later [37]. PCA extracted four principal components from 16 free amino acids with a cumulative variance contribution rate of 100%, and extracted three principal components from 16 hydrolyzed amino acids with a cumulative variance contribution rate of 99.143%, which can better reflect the comprehensive information on the quality of the 5 edible fungus powders. A comprehensive evaluation model was established, and the comprehensive quality of amino acids in Agrocybe chaxinggu edible fungus powder was the best. This paper only studies the amino acid compositions and contents of 5 edible fungus powders at present. Further research will collect more edible fungus powder samples to determine their amino acid composition and content in order to establish a more detailed edible fungus powder quality evaluation system, laying a theoretical foundation for the development of edible fungus resources.

Data Availability

The data used to support the findings of this study are included within the article.

Disclosure

Jian Li and Junmei Ma are the co-first authors.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Jian Li and Junmei Ma contributed equally to this paper. YZ and SQM conceived and designed the study. JL and JMM

performed the experiments. JL and JMM wrote the paper. YZ, SQM, and SSF reviewed and edited the manuscript. All authors read and approved the manuscript.

Acknowledgments

This work was supported by National Key Research and Development Program of China (Project no. 2018YFC1311400).

References

- P Kalač, "A review of chemical composition and nutritional value of wild-growing and cultivated mushrooms," *Journal of the Science of Food and Agriculture*, vol. 93, no. 2, pp. 209–218, 2013.
- [2] Y. Zhang, C. Venkitasamy, Z. Pan, and W. Wang, "Recent developments on umami ingredients of edible mushrooms—a review," *Trends in Food Science & Technology*, vol. 33, no. 2, pp. 78–92, 2013.
- [3] S. T. Chang, "Overview of mushroom cultivation and utilization as functional foods," *Mushrooms as functional foods*, vol. 260, 2008.
- [4] T. Girish and M. K. Rai, "Biotechnological potential of mushrooms: drugs and dye production," *International Journal of Medicinal Mushrooms*, vol. 8, no. 4, 2006.
- [5] H. Kawagishi, "Function of mushrooms and their active principles," *Food Style*, vol. 7, no. 9, pp. 70–73, 2003.
- [6] H. Kawagishi, M. Ando, T. Mizuno, H. Yokota, and S. Konishi, "A novel fatty acid from the mushroom Hericium erinaceum," *Agricultural & Biological Chemistry*, vol. 54, no. 5, pp. 1329–1331, 1990.
- [7] V. M. Dembitsky, A. O. Terent'ev, and D. O. Levitsky, "Amino and fatty acids of wild edible mushrooms of the genus boletus," *Records of Natural Products*, vol. 4, no. 4, pp. 218–223, 2010.
- [8] R. Ana, B. Sunčica, and Š. Zdenko, "Analysis of nucleosides and monophosphate nucleotides from mushrooms with reversed-phase HPLC," *Journal of Separation Science*, vol. 33, no. 8, pp. 1024–1033, 2010.
- [9] S. J. M. Mdachi, M. H. H. Nkunya, V. A. Nyigo, and I. T. Urasa, "Amino acid composition of some Tanzanian wild mushrooms," *Food Chemistry*, vol. 86, no. 2, pp. 179–182, 2004.
- [10] R. Bonku and J. Yu, "Health aspects of peanuts as an outcome of its chemical composition," *Food Science and Human Wellness*, vol. 9, no. 1, pp. 21–30, 2020.
- [11] D. Tagkouli, A. Kaliora, G. Bekiaris et al., "Free amino acids in three pleurotus species cultivated on agricultural and agroindustrial by-products," *Molecules*, vol. 25, no. 17, p. 4015, 2020.
- [12] J. W. Xia, Y. R. Ma, Z. Li, and X. G. Zhang, "Acrodictys-like wood decay fungi from southern China, with two new families Acrodictyaceae and Junewangiaceae," *Scientific Reports*, vol. 7888, pp. 1–21, 2017.
- [13] S.-Y. Tsai, T.-P. Wu, S.-J. Huang, and J.-L. Mau, "Nonvolatile taste components of Agaricus bisporus harvested at different stages of maturity," *Food Chemistry*, vol. 103, no. 4, pp. 1457–1464, 2007.
- [14] J.-L. Mau, Y.-L. Chen, R.-C. Chien, Y.-C. Lo, and S.-D. Lin, "Taste quality of the hot water extract from Flammulina velutipes and its application in umami seasoning," *Food Science and Technology Research*, vol. 24, no. 2, pp. 201–208, 2018.

- [15] J.-L. Mau, "The umami taste of edible and medicinal mushrooms," *International Journal of Medicinal Mushrooms*, vol. 7, no. 1-2, pp. 119–126, 2005.
- [16] Z. Chen, H. Gao, W. Wu et al., "Effects of fermentation with different microbial species on the umami taste of Shiitake mushroom (Lentinus edodes)," *Lebensmittel-Wissenschaft & Technologie*, vol. 141, Article ID 110889, 2021.
- [17] D. Kumari, M. S. Reddy, and R. C. Upadhyay, "Nutritional composition and antioxidant activities of 18 different wild Cantharellus mushrooms of Northwestern Himalayas," *Food Science and Technology International*, vol. 17, no. 6, pp. 557–567, 2010.
- [18] S.-Y. Chen, K.-J. Ho, Y.-J. Hsieh, L.-T. Wang, and J.-L. Mau, "Contents of lovastatin, γ-aminobutyric acid and ergothioneine in mushroom fruiting bodies and mycelia," *Lebensmittel-Wissenschaft* & *Technologie*, vol. 47, no. 2, pp. 274–278, 2012.
- [19] M. M. Poojary, V. Orlien, P. Passamonti, and K. Olsen, "Enzyme-assisted extraction enhancing the umami taste amino acids recovery from several cultivated mushrooms," *Food Chemistry*, vol. 234, pp. 236–244, 2017.
- [20] D. Luo, J. Wu, Z. Ma, P. Tang, X. Liao, and F. Lao, "Production of high sensory quality Shiitake mushroom (Lentinus edodes) by pulsed air-impingement jet drying (AID) technique," *Food Chemistry*, vol. 341, Article ID 128290, 2021.
- [21] D. Liu, Y.-Q. Chen, X.-W. Xiao et al., "Nutrient properties and nuclear magnetic resonance-based metabonomic analysis of macrofungi," *Foods*, vol. 8, no. 9, p. 397, 2019.
- [22] L. Y. Zhou, W. Li, and W. J. Pan, "Effects of thermal processing on nutritional characteristics and non-volatile flavor components from Tricholoma lobayense," *Emirates Journal of Food and Agriculture*, vol. 29, no. 4, pp. 285–292, 2017.
- [23] W. Tang, C. Liu, J. Liu et al., "Purification of polysaccharide from Lentinus edodes water extract by membrane separation and its chemical composition and structure characterization," *Food Hydrocolloids*, vol. 105, Article ID 105851, 2020.
- [24] X. Li, Y. Guo, Y. Zhuang, Y. Qin, and L. Sun, "Nonvolatile taste components, nutritional values, bioactive compounds and antioxidant activities of three wild Chanterelle mushrooms," *International Journal of Food Science and Technology*, vol. 53, no. 8, pp. 1855–1864, 2018.
- [25] Z. Qing, J. Cheng, X. Wang, D. Tang, X. Liu, and M. Zhu, "The effects of four edible mushrooms (Volvariella volvacea, Hypsizygus marmoreus, Pleurotus ostreatus and Agaricus bisporus) on physicochemical properties of beef paste," *Lebensmittel-Wissenschaft & Technologie*, vol. 135, Article ID 110063, 2021.
- [26] X. Wang, P. Zhou, J. Cheng, Z. Chen, and X. Liu, "Use of straw mushrooms (Volvariella volvacea) for the enhancement of physicochemical, nutritional and sensory profiles of Cantonese sausages," *Meat Science*, vol. 146, pp. 18–25, 2018.
- [27] D. Li, D. Wang, Y. Fang et al., "A novel phase change coolant promoted quality attributes and glutamate accumulation in postharvest shiitake mushrooms involved in energy metabolism," *Food Chemistry*, vol. 351, Article ID 129227, 2021.
- [28] T. Ming, J. Li, P. Huo, Y. Wei, and X. Chen, "Analysis of free amino acids in russula griseocarnosa harvested at different stages of maturity using iTRAQ-LC-MS/MS," *Food analytical methods*, vol. 7, no. 9, pp. 1816–1823, 2014.
- [29] GB 5009 124-2016, "National Food Safety Standards Determination of Amino Acids in Food," 2016.
- [30] GB/T 30987-2020, "Determination of Free Amino Acids in Plant," 2020.

- [31] J. Lv, L. Wei, Y. Yang et al., "Amino acid substitutions in the neuraminidase protein of an H9N2 avian influenza virus affect its airborne transmission in chickens," *Veterinary Research*, vol. 46, no. 44, pp. 44–10, 2015.
- [32] X. Yu, M. J. Chen, and C. H. Li, "Effects of culture substrates on nutritional and flavor components of Volvariella volvacea," *Mycosystema*, vol. 37, no. 12, pp. 1731–1740, 2018.
- [33] J. Y. Duan, Z. Y. Li, and J. Li, "Comparison of nutritional and flavor characteristics between four edible fungi and four fruits and vegetables based on components and characteristics of free amino acids," *Mycosystema*, vol. 39, no. 6, pp. 1077–1089, 2020.
- [34] M. Lu, H. M. An, and X. H. Zhao, "Analysis of amino acids in *Rosa sterilis* and *Rosa roxburghii* fruits," *Food Science*, vol. 36, no. 14, pp. 118–121, 2015.
- [35] Y. Ma, X. X. Yuan, L. N. Liu et al., "Multielement principal component analysis and origin traceability of rice based on ICP-ms/MS," *Journal of Food Quality*, vol. 2021, pp. 1–12, Article ID 5536241, 2021.
- [36] Y. Zhou and H. Yang, "Effects of calcium ion on gel properties and gelation of tilapia (*Oreochromis niloticus*) protein isolates processed with pH shift method," *Food Chemistry*, vol. 277, pp. 327–335, 2019.
- [37] M. Golebiowski, A. Ostachowska, and M. Paszkiewicz, "Fatty acids and amino acids of entomopatho- genic fungus *Conidiobolus coronatus* grow on minimal and rich media," *Chemical Papers*, vol. 70, pp. 1360–1369, 2016.