







## Research Article

# Rapid Determination of Pachymic Acid Content by Near-Infrared Spectroscopy

Jie Lu <sup>1</sup>, Changqin Li <sup>1,2</sup>, Lijun Liu <sup>1,2,3</sup>, Wangjing Chai <sup>1</sup>, Yabin Hou <sup>1,4</sup>  
and Changyang Ma <sup>1,2,5</sup>

<sup>1</sup>National R & D Center for Edible Fungus Processing Technology, Henan University, Kaifeng 475004, China

<sup>2</sup>Functional Food Engineering Technology Research Center of Henan Province, Kaifeng 475004, China

<sup>3</sup>Huaihe Hospital, Henan University, Kaifeng 475004, China

<sup>4</sup>Joint International Research Laboratory of Food & Medicine Resource Function, Kaifeng 475004, Henan, China

<sup>5</sup>Kaifeng Key Laboratory of Functional Components in Health Food, Kaifeng 475004, Henan, China

Correspondence should be addressed to Yabin Hou; [houyabin5916@sina.com](mailto:hoyabin5916@sina.com) and Changyang Ma; [changym@g.clemson.edu](mailto:changym@g.clemson.edu)

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In this paper, a rapid model for the determination of pachymic acid content in *Poria* was established by partial least squares (PLS) regression and near-infrared spectroscopy (NIR). During the research, a total of 108 batches of *Poria* samples from different producing regions were used, while their corresponding pachymic acid contents by high-performance liquid chromatography (HPLC) were adopted as reference. These samples were divided randomly into calibration sets for model establishment and validation sets for model validation. The test results from the calibration set showed that the best preprocessing method of the NIR spectra model was the standard normal variate (SNV) + second derivatives (SD), and the most suitable number of principal factors was 9. In this model, the coefficient of determination of the calibration set ( $r_c^2$ ) and validation set ( $r_v^2$ ) was 0.915 and 0.917, respectively. Meanwhile, the root mean square error of calibration (RMSEC) and the root mean square error of validation (RMSEP) with the calibration set were 0.051 mg/g and 0.054 mg/g, respectively. These results indicated this model could rapidly and reliably predict the pachymic acid content in *Poria* and increase the determination efficiency of pachymic acid in *Poria*. This is conducive to promote the development of industry.

## 1. Introduction

*Poria cocos*, one important medicinal fungus, has been used in China for a long time [1, 2]. Pachymic acid, a rich component in *Poria*, is the key active ingredient of *Poria*, with the effects of anti-inflammatory [3], antitumor [4], and hypoglycemic [5]. Usually, pachymic acid was used as the mark component to reflect the quality of *Poria* [6]. Nowadays, the common determination methods for pachymic acid include the colorimetric method and the HPLC method. For its high accuracy and stability, HPLC is usually taken as a reference method, but its analysis process is labor-intensive and time-consuming, which has hampered its wide applicability in the market. Moreover, the 2020 edition of the “Chinese Pharmacopoeia” has not included one quantitative

determination method for this component. Therefore, a method with high efficiency and throughput for the determination of pachymic acid content in *Poria* is necessary.

NIR spectroscopy is one rapid determination method widely used in the research field of traditional Chinese medicine recently for the advantages of fast speed [7, 8], high efficiency, high sensitivity, simple sample processing, and convenient instrument operation [9–11]. NIR spectroscopy has been widely used in food [12, 13], medicine [14–16], and other industries [17, 18]. Gao Hongbin et al. [19] used vector normalization and first derivative to process the NIR spectrum data of *Glycyrrhizae Radix et Rhizoma* Yinbian and established the corresponding rapid NIR analysis model for the determination of glycyrrhizic acid and glycyrrhizin with the PLS method. The correlation coefficients of the

models were 0.981 and 0.919, and the mean square errors of cross-validation were  $0.184 \mu\text{g/mL}$  and  $0.144 \mu\text{g/mL}$ , respectively. Ying Liang [20] used the PLS method combined with cross-validation methods with regression analysis and MSC+SD data processing methods to develop a NIR analysis model for rapid determination of naringin content. The model has good indexes and can rapidly evaluate the content of naringin in *Exocarpium Citri Grandis*.

According to references, it can be concluded that the NIR spectrum majorly reflects the doubling- and combination-frequency absorption of the movement of some characteristic groups (such as C-H, O-H, C=O,  $-\text{CH}_2$ , and  $-\text{CH}_3$ ), and these groups have a fixed vibration frequency which can excite to generate resonance when irradiated by infrared rays [21]. The quantity and connection of these groups of pachymic acid could form a specific NIR spectrum in the instrument, and the amount of energy in the near-infrared light absorbed by the pachymic acid, which can be mined in the NIR spectrum by the *Poria* sample, could reflect the quantity of pachymic acid. In this study, NIR were used with PLS regression to establish a rapid quantitative analysis model to detect pachymic acid, which was expected to provide technical reference for quality control and market management of *Poria*.

## 2. Materials and Methods

**2.1. Materials and Reagents.** A total of 108 batches of *Poria* samples were collected in small cube shapes from the market in 2019. The collected regions covered the main production provinces and the Dabie Mountains of China, including Hunan, Anhui, Yunnan, Hubei, Sichuan, Guizhou, Chongqing, Fujian, Guangdong, Henan, Jiangxi, Zhejiang, and Guangxi Provinces.

**2.2. Main Instruments and Reagents.** Spot Light 400 NIR Spectrometer (PerkinElmer Inc., USA), Multiskan MK3 Microplate (Thermo Electron Corp., USA), and LC-20AT high performance liquid chromatograph (Shimadzu Corp., Japan) were used in this research.

During this research, acetonitrile (HPLC grade) and methanol (HPLC grade) were purchased from Longxi Science Co., Ltd., acetic acid (HPLC grade) was purchased from Tianjin Fuyu Fine Chemical Co., Ltd., purified water were from Hangzhou Wahaha Baili Food Co., Ltd., and pachymic acid (Batch No. 18062101) was purchased from Chengdu Pufei Biotechnology Co., Ltd.

### 2.3. Method for the Determination of Pachymic Acid Content

**2.3.1. Preparation of the Reference Solution.** Pachymic acid was accurately weighed and dissolved in methanol to prepare a  $0.49 \text{ mg/mL}$  reference substance solution, which was filtered through a  $0.22 \mu\text{m}$  microporous membrane and reserved for testing.

**2.3.2. Preparation of the Test Solution.** Each sample was crushed into powder of 80 mesh fineness and placed in a

TABLE 1: LC time program.

$t/\text{min}$	Acetonitrile (%)	0.04% acetic acid solution (%)
0	50	50
5	55	45
12	60	40
15	65	35
19	70	30
25	75	25
30	75	25
35	100	0

10 mL conical flask. The powder was dissolved with methanol to make a  $0.125 \text{ g/mL}$  solution and was disposed of by ultrasonic (1200 W) for 40 min. The supernatant was taken and reserved for testing after filtering through a  $0.22 \mu\text{m}$  microporous filter membrane.

**2.3.3. The HPLC Conditions.** The pachymic acid content of the reference solution and test solution was determined by HPLC with the chromatographic column being Thermo BDS Hypersil C18 ( $250 \times 4.6 \text{ mm}$ ,  $5 \mu\text{m}$ ). The LC time program is shown in Table 1. The flow rate was  $1.4 \text{ mL/min}$ , the column temperature was  $30^\circ\text{C}$ , the detection wavelength was  $210 \text{ nm}$ , and the sample size was  $20 \mu\text{L}$ . Based on this condition, the corresponding chromatogram is shown in Figure 1. The peak with the retention time of 26 min was the pachymic acid.

**2.3.4. Evaluation of HPLC Methods. Linear Relation Investigation.** In this part, the chromatograph and corresponding peak areas of 5 injections of reference solutions with different concentrations ( $0.0245$ ,  $0.049$ ,  $0.49$ ,  $0.98$ , and  $1.47 \text{ mg/mL}$ ) were recorded. The linear regression was established with the content of reference solutions ( $x$ ) and corresponding chromatographic peak areas ( $y$ ) and shown as the equation  $y = 3 \times 10^8 x + 4162.2$ . The linear regression coefficient of this equation was  $0.9999$ , indicating that the peak area of pachymic acid had a good linear correlation with the content in the range of  $0.0245\text{--}1.4700 \text{ mg/mL}$ .

**Reproducibility.** A total of 6 testing solutions prepared from the same sample were tested separately. The RSD of the test was  $0.48\%$ , indicating that this method had a good repeatability.

**Precision.** In this part, one reference solution was injected 6 times continuously for the precision testing of this method. The RSD was  $0.31\%$ , which indicated that the instrument and method could determine the content of pachymic acid accurately.

**Stability.** Samples prepared according to 2.3.2 were injected at 0, 4, 8, 16, 20, and 24 h, respectively, for testing the stability of the testing solution. The results showed that the RSD of this method was  $0.28\%$ , indicating that pachymic acid in the test solution was basically stable within 24 h.

**Recovery.** In this part, 6 samples with a known content of pachymic acid were added with a certain amount of standard pachymic acid, respectively. The average recovery rate of this

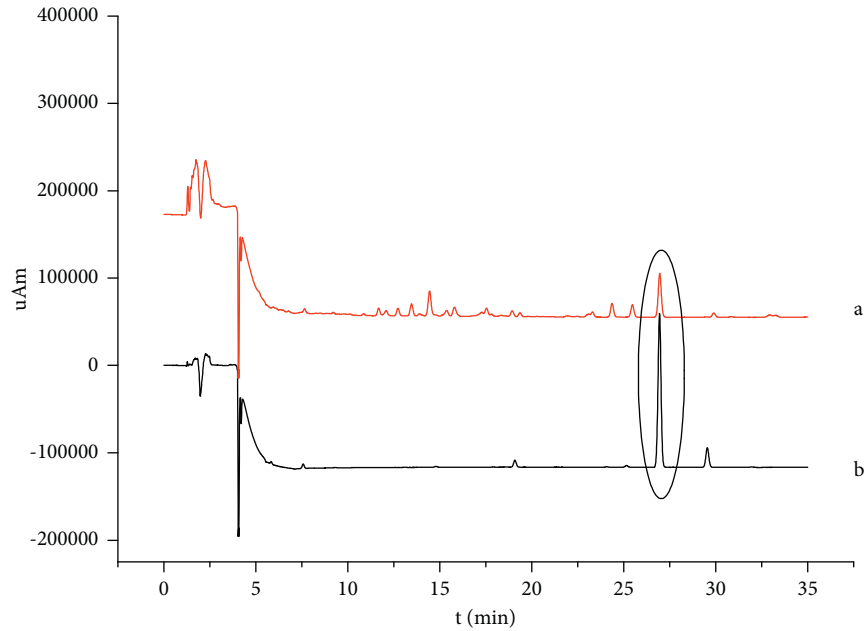


FIGURE 1: Chromatogram of (a) *Poria* sample and (b) reference solution.

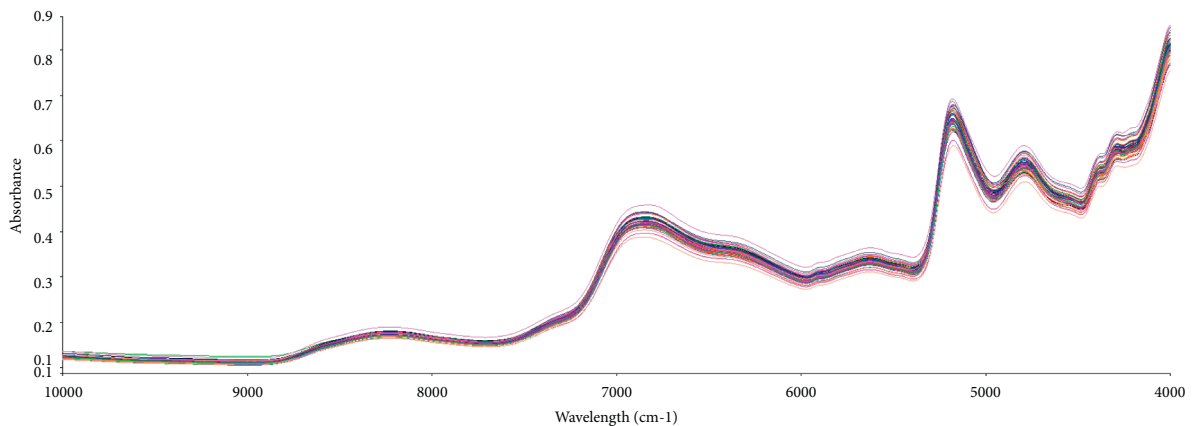


FIGURE 2: NIR spectrum of *Poria*.

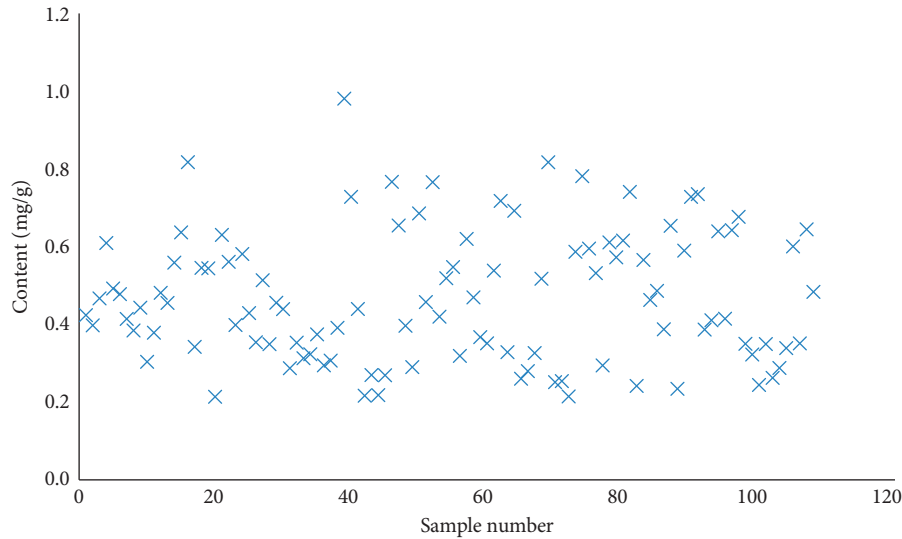
method for the pachymic acid was 101.55% and 2.46%, and the RSD value was 2.36%, which was eligible.

#### 2.4. Procedure of NIR Spectroscopy Analysis

**2.4.1. NIR Spectrum Acquisition and Analysis.** The *Poria* samples were crushed into powder, sieved through 80 mesh, and evenly spread on the bottom of the test dish (its diameter is 5 cm). The resolution was set to  $8\text{ cm}^{-1}$ , scanning was 64 times per average spectrum within the spectral range of  $4000\text{--}10000\text{ cm}^{-1}$ , the scanning type was interleaved mode, and the internal reference was automatically used as the background. Based on the determination parameters, the spectrum of each sample could be obtained (Figure 2).

**2.4.2. Establishment of the NIR Model.** PLS, artificial neural networks (ANN), support vector machines (SVM), and other nonlinear correction methods are used to establish

the model with their own advantages and disadvantages [22], and in this research, PLS was adopted to build the NIR analysis model with the software Spectrum Quant. In addition, 4 parameters including  $r_c^2$ , RMSEC, RMSEP, and  $r_v^2$  were used to evaluate the established model.  $r_c^2$  is used to evaluate the degree of linearity between the predicted and measured values of the samples. The closer  $r_c^2$  is to 1, the better the regression or prediction result is. RMSEC is mainly used to evaluate the feasibility and prediction ability of modeling methods. RMSEP is mainly used to evaluate the prediction ability of the model for external samples. The smaller the value is, the better the prediction ability of the model for external samples is.  $r_v^2$  is used to evaluate the linearity between the predicted value and the measured value of the model for the unknown sample during the external validation of the model [21]. Its value is approximately close to 1, indicating that the model predicts better results. Spectrum Quant was used for modeling in this study.

FIGURE 3: Scatter plot of pachymic acid content in *Poria* samples.TABLE 2: Pachymic acid content range in *Poria* samples from the calibration set and validation set.

	Size	Maximum value (mg/g)	Minimum value (mg/g)	Mean $\pm$ SD (mg/g)
Calibration set	91	0.981	0.214	$0.472 \pm 0.166$
Validation set	17	0.735	0.242	$0.429 \pm 0.144$

TABLE 3: Test results for the ranges of NIR spectroscopy.

Spectral range ( $\text{cm}^{-1}$ )	$r_c^2$	RMSEC (mg/g)	RMSEP (mg/g)
4200–5390	0.504	0.124	0.131
5390–6070	0.524	0.121	0.127
6070–7500	0.832	0.072	0.079
4000–10000	0.915	0.051	0.054

TABLE 4: The results for the data preprocessing methods.

Methods	$r_c^2$	RMSEC (mg/g)	RMSEP (mg/g)
MSC	0.209	0.156	0.163
SNV	0.208	0.156	0.164
FD	0.504	0.124	0.129
SD	0.922	0.049	0.052
MSC + SD	0.915	0.051	0.054
SNV + SD	0.915	0.051	0.054
MSC + FD	0.582	0.113	0.120
SNV + FD	0.590	0.112	0.118

### 3. Results and Analysis

**3.1. The Content Value of Pachymic Acid.** Figure 3 is a scatter plot about the pachymic acid content of 108 batches of *Poria*.

A total of 91 *Poria* samples were randomly selected from 108 samples according to the sequence of each sample by using the random function in Excel for composing the calibration set, and the remaining samples were included in the validation set. At the same time, the maximum and minimum values of pachymic acid content from the calibration set should cover the validation set. The information characteristics of the samples from the two sets are shown in Table 2.

**3.2. Selection of Spectral Interval of NIR Analysis Model.** The absorption effect of substances varies in different NIR spectra, and it is necessary to determine the proper spectral range for the PA determination [23]. In this study, 4 potential spectral intervals were adopted, and the evaluation results of the corresponding analysis model are shown in Table 3.

According to the results, the spectral range is  $4000\text{--}10000\text{ cm}^{-1}$ , the full spectral range with the largest  $r_c^2$ ,

the smallest RMSEC and RMSEP, as well as the proper ratio of latter two indices were selected for modeling.

**3.3. Selection of Spectral Pretreatment Methods for NIR Analysis Models.** During the data analysis of NIR spectra, some random factors, such as sample particle size and light interference, could cause the baseline drift, affecting the reliability and stability of model building [24–26]. Single and combination data preprocessing methods were used for the modeling and the results are listed in Table 4.

MSC mainly eliminates the scattering effect caused by uneven sample size and particle size. SNV is mainly used to eliminate the influence of particle size, surface scattering light, and optical path change on the near-infrared spectrum. Derivatives can effectively eliminate baseline and other background interference [21]. It can be seen from Table 4 that SD, SNV + SD, and MSC + SD are the better preprocessing methods for the spectral information of the NIR analysis model. The  $r_c^2$  of the three pretreatment methods is

TABLE 5: The test results for the number of principal components.

No.	Method, principal factor number	$r_c^2$	RMSEC (mg/g)	RMSEP (mg/g)
1	SNV + SD, 10	0.949	0.040	0.042
2	SNV + SD, 9	0.915	0.051	0.054
3	SNV + SD, 8	0.873	0.062	0.065
4	MSC + SD, 10	0.949	0.040	0.042
5	MSC + SD, 9	0.915	0.051	0.054
6	MSC + SD, 8	0.873	0.062	0.065
7	SD, 10	0.941	0.043	0.046
8	SD, 9	0.922	0.049	0.052
9	SD, 8	0.853	0.067	0.069

TABLE 6: The parameters of the optimal NIR detection model.

Project	Parameters
Spectral pretreatment method	SNV + SD
Model parameters	Principal factor number of PLS
	9
	Spectral range
	4000–10000 $\text{cm}^{-1}$
	$r_c^2$
	0.915
The evaluation results	RMSEC (mg/g)
	0.051
	RMSEP (mg/g)
	0.054
	$r_v^2$
	0.917

greater than 0.9, and RMSEC and RMSEP meet the basic requirements of model building. Among them, the values of the three indexes ( $r_c^2$ , RMSEC, and RMSEP) of SNV + SD and MSC + SD pretreatment methods have a slight difference, so these three pretreatment methods are taken into consideration.

**3.4. Determination of PLS Principal Factor Number in the NIR Analysis Model.** When PLS was used to establish the NIR analysis model, the number of PLS principal factors was also a key factor affecting the quality of the model. Either too many or too less principal factors would cause underfitting or overfitting problems, respectively, increasing the prediction error [27]. In this part, the corresponding results of the models with specific data preprocessing method and principle factors are listed in Table 5.

It can be seen from Table 5 that all alternative data preprocessing methods with more principal factors could cause higher  $r_c^2$  and smaller RMSEC and RMSEP, and the models with over 9 principal factors would have a suitable  $r_c^2$  higher than 0.90. So 9 principal factors were the basic requirements for the modeling.

#### 4. Validation and Evaluation of the Model

On the basis of the former parts of this research, 6 aforementioned models (No. 1, 2, 4, 5, 7, and 8 in Table 5) with  $r_c^2$  over 0.90 were further verified by using validation set samples in this part, and it was found that the maximum  $r_v^2$  of model 2 is 0.917. From this, the optimal NIR analysis model (No. 2 model in Table 5) for predicting the content of

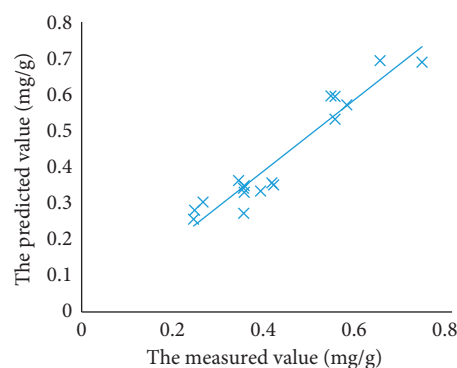


FIGURE 4: Comparison between predicted values and experimental values.

pachymic acid of *Poria* was obtained, and its parameters are shown in Table 6.

The distribution of the predicted and experimental values was scattered intuitively in a line graph (Figure 4) [28]. At the same time, SPSS statistical software (SPSS Inc., Chicago, USA) was used to analyze the difference between the two values by comparing the ratio of them with 1, and there was no significant difference found between them ( $P = 0.512$ ). This result indicated that the NIR analysis model had good prediction accuracy for pachymic acid of *Poria*.

#### 5. Conclusion

There are already many national standards by using NIR for qualitative and quantitative detection now [29, 30]. NIR has also been widely recognized in the field of traditional Chinese medicine. It is widely used in the production process of traditional Chinese medicines, including qualitative identification of the types and authenticity of traditional Chinese medicines [31], quantitative determination of effective components, and monitoring of the drug production process [32, 33]. In this study, different NIR analysis models for pachymic acid were evaluated with different parameters by  $r_c^2$ , RMSEC, RMSEP, and  $r_v^2$  values. The results showed that the NIR analysis model with pretreatment method of standard normal variate (SNV) + second derivative (SD), the spectral interval of 4000–10000  $\text{cm}^{-1}$ , and 9 PLS major factors could predict the pachymic acid content in *Poria* exactly. Besides, there was no significant difference

between the predicted value and the measured value of the prediction set ( $P = 0.512$ ).

According to Williams et al.'s results, this NIR model is more precise than the standard for most applications and almost reaches the standard for quality assurance ( $r^2 \geq 0.92$ ) [34]. Hence, many improvements still need to be made to this model in further study, in order to promote its applicability and practicability.

The pachymic acid content of *Poria* was successively determined by NIR combined with stoichiometry, expanding the applicability of NIR. Compared with other analytical methods, the NIR method has the advantages of simple pretreatment, fast analytical speed, high multisample processing capacity, and low cost, which is beneficial to the development promotion of the *Poria* industry [35].

### Data Availability

The data used to support the findings of this study are included within the article.

### Ethical Approval

Not applicable.

### Conflicts of Interest

The authors declare that they have no conflicts of interest regarding the publication of this paper.

### Authors' Contributions

Jie Lu and Changqin Li contributed equally to this work.

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### References

- [1] G. Ma, W. Yang, L. Zhao, F. Pei, D. Fang, and Q. Hu, "A critical review on the health promoting effects of mushrooms nutraceuticals," *Food Science and Human Wellness*, vol. 7, no. 2, pp. 125–133, 2018.
- [2] Z. Yin, Z. Liang, C. Li, J. Wang, C. Ma, and W. Kang, "Immunomodulatory effects of polysaccharides from edible fungus: a review," *Food Science and Human Wellness*, vol. 10, no. 4, pp. 393–400, 2021.
- [3] E. M. Giner, S. Máñez, M. C. Recio, R. M. Giner, M. Cerdá-Nicolás, and J. L. Ríos, "In vivo studies on the anti-inflammatory activity of pachymic and dehydrotumulosic acids," *Planta Medica*, vol. 66, no. 3, pp. 221–227, 2000.
- [4] C. Lu, J. Ma, and D. Cai, "Pachymic acid inhibits the tumorigenicity of gastric cancer cells by the mitochondrial pathway," *Anti-Cancer Drugs*, vol. 28, no. 2, pp. 170–179, 2017.
- [5] F. S. A. Saadeldien, Y. Niu, H. Wang et al., "Natural products: regulating glucose metabolism and improving insulin resistance," *Food science and human wellness*, vol. 9, no. 3, pp. 214–228, 2020.
- [6] S. S. Tian, X. M. Zhao, and Y. Liu, "Studies on quality standards of *Poria*," *China Journal of Chinese Materia Medica*, vol. 45, no. 8, pp. 1734–1744, 2020.
- [7] S. Geng, Y. Chen, and Y. Jin, "Rapid quantitative determination of multi-indicators in *Lonicera japonica* flos by near-infrared spectroscopy," *Chinese Journal of Modern Applied Pharmacy*, vol. 33, no. 2, pp. 144–150, 2016.
- [8] C. J. Lai, R. R. Zhou, and Y. Yu, "Rapid identification of geographical origins and determination of polysaccharides contents in *Ganoderma lucidum* based on near infrared spectroscopy and chemometrics," *China Journal of Chinese Materia Medica*, vol. 43, no. 16, pp. 3243–3248, 2018.
- [9] Y. Yang, X. Liu, W. Li et al., "Rapid measurement of epimedin A, epimedin B, epimedin C, icariin, and moisture in *Herba Epimedii* using near infrared spectroscopy," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 171, pp. 351–360, 2017.
- [10] T. Y. Di, X. J. Gao, and X. Zhang, "Comparative identification studies on wild and cultivated *Glycyrrhiza uralensis* produced Ningxia based on index components and near-infrared spectroscopy," *China Journal of Chinese Materia Medica*, vol. 42, no. 03, pp. 536–541, 2017.
- [11] C. Gu, X. K. Yue, and W. F. Du, "Establishment of NIRS quantitative model for synchronous quantitative detection of contents of three alkaloids in *corydalis rhizoma*," *Chinese Journal of Experimental Traditional Medical*, vol. 23, no. 3, pp. 60–65, 2017.
- [12] M. Zareef, Q. Chen, M. M. Hassan et al., "An overview on the applications of typical non-linear algorithms coupled with NIR spectroscopy in food analysis," *Food Engineering Reviews*, vol. 12, no. 2, pp. 173–190, 2020.
- [13] M. A. Berhow, M. Singh, M. J. Bowman, N. P. J. Price, S. F. Vaughn, and S. X. Liu, "Quantitative NIR determination of isoflavone and saponin content of ground soybeans," *Food Chemistry*, vol. 317, no. 1, pp. 126373–126373.9, 2020.
- [14] Y. Yang, L. Wang, Y. Wu et al., "On-line monitoring of extraction process of *Flos Lonicerae Japonicae* using near infrared spectroscopy combined with synergy interval PLS and genetic algorithm," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 182, pp. 73–80, 2017.
- [15] C.-H. Zhang, Y.-H. Yun, W. Fan, Y.-Z. Liang, Y. Yu, and W.-X. Tang, "Rapid analysis of polysaccharides contents in *Glycyrrhiza* by near infrared spectroscopy and chemometrics," *International Journal of Biological Macromolecules*, vol. 79, pp. 983–987, 2015.
- [16] Y. Xie, R. R. Zhou, and H. L. Xie, "Application of near infrared spectroscopy for rapid determination the geographical regions and polysaccharides contents of *Lentinula edodes*," *International Journal of Biological Macromolecules*, vol. 122, pp. 1115–1119, 2018.
- [17] J. T. C. Rocha, L. M. S. L. Oliveira, J. C. M. Dias et al., "Sulfur determination in Brazilian petroleum fractions by mid-infrared and near-infrared spectroscopy and partial least squares associated with variable selection methods," *Energy and Fuels*, vol. 30, no. 1, pp. 698–705, 2016.
- [18] Y. P. Wang, C. K. Lee, and H. Y. Dai, "Effect of wetting on the determination of soil organic matter content using visible and near-infrared spectrometer," *Geoderma*, vol. 376, pp. 173–190, 2020.
- [19] H. B. Gao, H. Liu, and H. H. Xia, "Near-infrared spectroscopy in rapid determination of glycyrrhizic acid and liquiritin in *Glycyrrhizae Radix et Rhizoma Yinbian*," *Academic Journal of*

- Second Military Medical University*, vol. 41, no. 8, pp. 921–925, 2020.
- [20] Y. Liang, H. B. Gao, and X. D. Li, “Content of naringin in *Exocarpium citrigrandis* was rapidly determined by near-infrared spectroscopy,” *Guangzhou Chemistry*, vol. 45, no. 04, pp. 23–29, 2020.
- [21] X. Chu, *Practical Manual of Near Infrared Spectroscopy Analysis Technology*, China Machine Press, Beijing, China, 2016.
- [22] S. Wold, M. Sjöström, and L. Eriksson, “PLS-regression: a basic tool of chemometrics,” *Chemometrics and Intelligent Laboratory Systems*, vol. 58, no. 2, pp. 109–130, 2001.
- [23] X. H. Lv, J. L. Jiang, and J. Yang, “Detection of capsaicin content by near-infrared spectroscopy combined with optimal wavelengths,” *Journal of Zhejiang University (Agriculture & Life Sciences)*, vol. 45, no. 6, pp. 760–766, 2019.
- [24] G. Ding, B. Li, Y. Han et al., “A rapid integrated bioactivity evaluation system based on near-infrared spectroscopy for quality control of *Flos Chrysanthemi*,” *Journal of Pharmaceutical and Biomedical Analysis*, vol. 131, pp. 391–399, 2016.
- [25] X. L. Chu, H. F. Yuan, and W. Z. Lu, “Progress and application of spectral data pretreatment and wavelength selection methods in NIR analytical technique,” *Progress in Chemistry*, vol. 4, pp. 528–542, 2020.
- [26] L. M. Wu, *Study on Rapid Detection for the Quality of Some Herbal Medicines and Chinese Patent Medicines by Near Infrared Spectroscopy*, Southwest University, Chongqing, China, 2013.
- [27] W. M. Zhang, W. He, and Y. J. Wu, “Determination of the content of compound aspirin/dicyanidone by near-infrared spectroscopy combined with partial least square method,” *Yunnan Chemical Technology*, vol. 46, no. 8, pp. 84–86, 2019.
- [28] J. J. Diao, W. Li, and L. Li, “Rapid detection of alcohol in wine by near infrared optical fiber spectroscopy,” *Food and Drug*, vol. 23, no. 1, pp. 6–10, 2021.
- [29] General Administration of Quality Supervision, Inspection and Quarantine, Standardization Management Committee, *Inspection of Grain, Oil and Corn, Determination of Starch Content, Near Infrared Method: GB/T 25219—2010*, China Standards Publishing House, Beijing, China, 2010.
- [30] Association of Official Analytical Chemists, *AOAC Official Method 997.06 Protein (Crude) in Wheat Whole Grain Analysis Near-Infrared Spectroscopic Method First Action 1997 Final Action*, Association of Official Analytical Chemists, Rockville, MD, USA, 2005.
- [31] T. Yuan, J. Wang, and W. Zhe, “The review of application and standards of near infrared spectrometry,” *Chinese Agricultural Science Bulletin*, vol. 29, no. 20, pp. 190–196, 2013.
- [32] Y. J. Kim, H. J. Lee, and H. S. Shin, “Near infrared reflectance spectroscopy as a rapid and non-destructive analysis tool for curcuminoids in turmeric,” *Journal of Phytochemical Analysis*, vol. 104, no. 1, pp. 445–452, 2020.
- [33] L. B. Schlegel, M. Schubert-Zsilavec, and M. Abdel-Tawab, “Quantification of active ingredients in semi-solid pharmaceutical formulations by near infrared spectroscopy,” *Journal of Pharmaceutical and Biomedical Analysis*, vol. 142, no. 5, pp. 178–189, 2017.
- [34] P. Williams and J. Antoniszyn, *Near Infrared Technology: Getting the Best Out of Light*, African Sun Media, Stellenbosch, South Africa, 2019.
- [35] T. J. Barzee, L. Cao, Z. Pan, and R. Zhang, “Fungi for future foods,” *Journal of Future Foods*, vol. 1, no. 1, pp. 25–37, 2021.