

### Research Article

## Examining the Optimal Amount of Moringa Leaf Extract to Improve the Morphological and Inner Quality of Cabbage (*Brassica oleracea* var. *capitata*)

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Moringa leaves contain a proper amount of antioxidants, amino acids, vitamins, hormones, macronutrients, and micronutrients. Therefore, it is applied as a natural, inexpensive, and simple-to-make biostimulant to boost nutritional value and growth parameters in a variety of plants. The main purpose of this study is to determine the optimal concentration of *Moringa oleifera* L. leaf extract (MLE) as an effective biostimulant to improve nutritional and physical quality in cabbage (*Brassica oleracea* var. *capitata*). To do this, three concentrations of MLE (6, 8, and 10%) were foliar sprayed to the plant leaves at a rate of 25 mL plant<sup>-1</sup> for every two weeks from transplantation till harvest, while control plants ( $\emptyset$ ) were sprayed with distilled water only. Our results show that MLE greatly increased cabbage growth, nutrient content, pigment content, and nutrient absorption, while a high concentration of 10% MLE could also significantly reduce nitrate content in cabbage leaves. In comparison to the control plants, sprays of 6%, 8%, and 10% MLE reduced nitrate content by 23%, 14%, and 12%, respectively. However, the lowest nitrate content was found for the plants sprayed with 6% MLE. Except for the dry matter, all the growth parameters, mineral content, and pigment content were significantly higher after spraying with 10% MLE, while better vitamin C and lower nitrate were found in the plants treated with 6% MLE. A Pearson correlation reveals that head weight has a positive correlation with head diameter, head height, chlorophyll a, and carotenoids at *p* levels of 0.01 and a positive correlation with chlorophyll b and vitamin C at *p* levels of 0.05. Vitamin C and dry matter, on the other hand, had a negative connection with nitrate content.

#### 1. Introduction

Cabbage (*Brassica oleracea* L. var. *capitata*) is a member of the *Brassicaceae family*, the biggest plant family. It is a coolseason vegetable with short roots that is cultivated for its enormous leafy head. It was initially cultivated in Western Europe [1]. According to the Food and Agriculture Organization of the United Nations (FAO), worldwide cabbage production in 2021 was 71.7 million tons, making it one of

the world's most widely grown vegetables, with Asia accounting for 78.2%, Europe 13%, Africa 5.7%, America 2.9%, and Oceania 0.2% [2]. In 2021, Hungary's total production of white cabbage was about 37 thousand tons [3]. The plant is grown for its large leaves. The physical properties of the cabbage cultivars, such as head size, shape, and colour, as well as the texture of the leaves, vary greatly amongst cultivars. Cabbage has been classified into three categories: white cabbage, red cabbage, and savoy cabbage [4]. The cabbage leaves are well-known for their high nutritional value as antioxidant phytochemicals [1, 5], macroand micronutrient content [6], and richness in vitamins C, E, and K [7]. Cabbage is one of the most important vegetables for healthy diets and can be cooked or consumed raw.

Approximately, 100 g of leaf sample contains 93 mL water, 15 g protein, 0.2 g fat, 4.0 g carbohydrate, 4 g calcium, and 0.5 g iron [8]. Cabbage also contains plenty of essential micronutrients which are crucial for human health and plant biological, chemical, and physiological functions such as copper (Cu), zinc (Zn), and iron (Fe) [9].

Leafy vegetables (particularly cabbage) require a high amount of N, P, and K, especially nitrogen fertilizer [10]. However, this may lead to various issues, like the excessive and careless use of chemical fertilizers, which raises the cost of planting and increases the risk of environmental degradation. Therefore, it is critical to enhance fertilization practices and fertilizer use efficiency to support agriculture's sustainable development.

Many research studies have recommended a variety of natural extracts to minimize fertilizer usage without reducing the quality and total yield of the products. Biostimulants, especially moringa leaf extract from the moringa tree (*Moringa oleifera* Lam.), are among the suggestions that have been practiced not only to reduce the amount of fertilizer used but also to improve the quality of many vegetables [11], such as lettuce (*Lactuca sativa* L.) [12–14], tomatoes (*Solanum lycopersicum*) [15], peppers (*Capsicum annuum*) [16], cabbage (*Brassica oleracea* var. *capitata*) [17], onions (*Allium cepa* L.) [18, 19], and squash (*Cucurbita pepo* L.) [20].

Moringa leaf extract is a currently introduced plant extract in Hungary as an active and helpful plant biostimulant to improve vegetable plant quality characteristics and reduce nitrate content in some leafy vegetables [14]. However, because it grows well in tropical and subtropical climates, the plant is most known in African and Asian countries [21, 22] rather than in Europe. Much scientific research in recent years has demonstrated that the plant contains significant nutritional value, antioxidants, minerals, and growth hormones. However, the content or nutritional value varies depending on the age of the leaf and the technique of extraction [23]. There is very limited research on the influence of moringa concentration on cabbage quality and yield. The purpose of this study was to see if moringa (Moringa oleifera Lam) leaf extract could alleviate nutritional value and physical parameters of cabbage (Brassica oleracea var. capitata) cultivated in an unheated plastic housing in Hungary and which concentration of extract may lead to an improvement in the quality of cabbage. Additionally, we aimed to increase the quality of fresh vegetables by using natural plant biostimulants while reducing the amount of artificial fertilizers used in vegetable cultivation. By doing this, consumers receive fresher, healthier food while also reducing environmental pollution, and farmers save money on fertilizer purchases.

#### 2. Materials and Methods

2.1. Experimental Setup and Layout. The design of our study was a complete randomized block design (CRBD), with three replications and 20 plants per replication. The cabbage seeds were first grown in a glasshouse, and once the plants had grown to four true leaves, they were transplanted to direct soil in an unheated plastic house from where some metrological data are shown in Table 1.

Two weeks after transplanting, seedlings were foliar sprayed with moringa leaf extract (MLE) at the concentrations of 6, 8, and 10%, whereas control plants ( $\emptyset$ ) received only distilled water.

The chemical components of soil samples were taken from 10 to 15 cm depth adjacent to plant roots to determine whether the plant needs to be fertilized. The experiment was carried out on lowland chernozem soil and the quality characteristics of the experimental soil are shown in Table 2.

Soil quality was affected by pH, total water-soluble salt, humus, and accessible nutrients. That is why, adding fertilizer has been recommended for leafy plants in such type of soil, as indicated by Liu et al. [24]. Based on the experimental soil testing, the inorganic fertilizer called FERTICARE 24-8-16 (NPK) was applied to our plants at the beginning of the experiment with the amount of 200 kg N ha<sup>-1</sup> once.

2.2. Preparation of Moringa Leaf Extract and Spraying Treatment. Fresh green moringa leaves (approximately 40 days old) were harvested, washed, and cleaned from moringa trees grown in a glasshouse at the Agrar Campus of the University of Debrecen, Hungary. The leaves were then dried in a shaded area. According to Sowley et al. [25], the dried leaves were finely grounded with a grinder and then sieved with fine netting to create moringa leaf powder.

The extract was made from moringa leaf powder, as recommended by Makkar and Becker [26], who advocates making moringa leaf extract from powdered leaves of 20 g mixed with 225 mL of 80% ethanol every 3 hours to obtain 675 mL of extract. The extract was then maintained at 0°C until spraying began. The extract was diluted to 6, 8, and 10% before being sprayed onto the plant leaves every two weeks in the early morning with 25 mL per plant. We solely sprayed distilled water to treat the controlled plants. Some chemical contents of the extract were measured as it is shown in Table 3.

2.3. Growth and Physical Measurements. Major important physical and growth parameters (head weight, head height, stem length, and dry matter) were measured on the 5 best plants from each replication on the day of harvest. Also, the leaf samples of about 0.25 g were ground in 2 mL of 80% ethanol in combination with sand 0.1% or 0.1 CaCO<sub>3</sub> to assess the chlorophyll and carotenoid content ( $\mu$ g mL<sup>-1</sup>). The samples were filtered using filter papers, and the final volume was reduced to (25 mL) before being transferred to a spectrophotometer to calculate absorbency at the wavelengths of (A663, A646, and A470) using equations (1)–(3)

TABLE 1: Temperature, humidity, and light intensity averages under a plastic house during the spring experiments.

	Months	
March	April	May
Temperature (°C), m	ean	
11.3	15.6	21.2
Relative humidity (R	RH %), mean	
79.4	58.8	38.7
Light emission (W/n	1 <sup>2</sup> ), mean	
175.6	196.2	245.5

TABLE 2: Soil chemical content of the experimental location.

Soil content	Value
pH (KCl)	7.58
Plasticity index of Arany K <sub>A</sub>	45
Total water-soluble salt (m/m)	0.04
Calcium hydroxide (slaked lime) Ca (OH) <sub>2</sub> (m/m)	1.24
Organic carbon (humus content) (m/m)	2.89
Phosphorus pentoxide P <sub>2</sub> O <sub>5</sub> (mg/kg)	1365
Potassium oxide (k <sub>2</sub> O)	368
Nitrate NO <sub>3</sub> <sup>-</sup>	206

Source: Agricultural Laboratory Centre, University of Debrecen.

TABLE 3: Content of minerals in 100 g of moringa leaf extract (MLE) from powdered leaves.

Minerals	Values (mg)
Phosphorus (P)	100
Potassium (K)	350
Sulphur (S)	267
Calcium (Ca)	326
Magnesium (Mg)	86.8
Sodium (Na)	11.4
Iron (Fe)	2.24
Copper (Cu)	0.319
Zinc (Zn)	0.477
Boron (B)	0.581

Source: Yaseen and Takacs-Hajos [14].

[27–30]. In this investigation, a Japanese-made spectrophotometer named APEL PD-303 (APEL CO., LTD., Japan) was used and the measured data were assessed.

The cabbage samples were dried at 60°C for 4 days to analyse mineral contents such as Ca, K, Mg, P, and NO<sub>3</sub> which were measured using the ICP-OES (iCAP<sup>™</sup> 7400, Thermo Scientific) by European Virtual Institute for Speciation Analysis (EVISA, Germany) and approached according to Yaseen and Takacs-Hajos [31], where 0.5000 g of the properly prepared sample was measured in a highpressure teflon bomb. 5 mL of deionized water with cc.  $HNO_3$  and 3 mL of 30%  $H_2O_2$  were mixed. It was sealed and digested in the Ethos Plus Microwave Digestion System (Milestone) according to the manufacturer's technique (Application Note 076) (3 min 85°C, 9 min 145°C, 4 min 200°C, and 14 min 200°C). After the teflon bombs were cooled, the digested samples were diluted to 50 mL, homogenized, and filtered (MN 640W, Millipore) before element analysis.

2.3.1. Ascorbic Acid (Vitamin C) Measurement. The vitamin C level was evaluated by redox titration with iodine solution, as described by Ciancaglini et al., [32] ascorbic acid +  $I_2 \rightarrow 2$   $I^-$  + dehydroascorbic acid.

2.3.2. Nitrate Determination. The nitrate concentration in cabbage leaves was determined using a segmented flow analyzer AA II (Bran + Luebbe, Norderstedt, Germany) at 540 nm after reduction in a copper-covered cadmium column ( $NO_3^- + 2e^{-} \otimes NO_2^-$ ) to generate a diazo compound, as described by Kmecl et al. [33] and Louro et al. [34].

2.3.3. Statistical Analyses. The raw data or result data were analysed using SPSS version 25.0. The statistical significance was determined by one-way analysis of variance (ANOVA) in Duncan's multiple range test and the  $p \le 0.05$  was considered to be statistically significant. Also, some Pearson correlation was performed at p levels 0.01 and 0.05 to see if there was any correlation between the physical and growth parameters.

#### 3. Results and Discussion

#### 3.1. Effect of MLE on Morphological Parameters on Cabbage

3.1.1. Head Weight. Head weight is one of the most essential criteria for commercially growing plants. The cabbage heads were harvested at the peak of maturity on May 20, 2022 (82 days after transplanting). The sprayed plants outweighed the control in terms of head weight. The plants treated with 10% moringa leaf extract (MLE 10%) had the biggest or significantly increased plant head weight at 1527 g plant<sup>-1</sup>, whereas the lowest head weight was for the control plants  $(\emptyset)$  at 1169 g plant<sup>-1</sup> (Table 4). This might be because MLE contains different growth hormones such as zeatin which leads to improving the cell division; as a result, the final weight would be increased [35]. Other research demonstrates that it may relate to the mineral content in MLE; having a high amount of potassium and zinc can regulate the growth and development of fruits [36]. Many research investigations have indicated that spraying plants with various concentrations of MLE improves fruit weight and size, for example, heavier fruit is produced in tomatoes when spraying plants with MLE (25 mL per plant) [37]; higher fruit lengths, diameters, and weights were recorded in pepper plants treated with different concentrations of MLE [16].

3.1.2. Head Diameter. Cabbage head diameter represents the head width or horizontal growth of the head size. This is regarded as a key physical feature in many vegetables, including lettuce, cabbage, and root vegetables. Yang et al. [38] indicate that head diameter has a strong positive regression to the head weight at  $r^2 = 0.92$ . According to our findings, the head diameter ranged from 14.60 to 16.10 cm head<sup>-1</sup>. Our study, which agrees with the previous one, demonstrates that head diameter is quadratically related to head weight at  $r^2 = 0.49$  (Figure 1).

TABLE 4	4: So	me o	t the	physical	parameters	were	altered	by	spraying	different	amounts	of moring	a lea	t extract.	
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Treatments	Head weight (g plant <sup>-1</sup> )	Head diameter (cm plant <sup>-1</sup> )	Head height (cm plant <sup>-1</sup> )	Stem length (cm plant <sup>-1</sup> )	Dry matter (%)
Control (Ø)	1169.00 <sup>b</sup>	14.60 <sup>b</sup>	13.60 <sup>b</sup>	6.90 <sup>ab</sup>	4.63 <sup>d</sup>
MLE 6%	1359.00 <sup>ab</sup>	15.70 <sup>a</sup>	14.30 <sup>ab</sup>	6.30 <sup>b</sup>	5.85 <sup>a</sup>
MLE 8%	$1411.00^{ab}$	$16.00^{\rm a}$	$14.40^{ab}$	6.60 <sup>ab</sup>	4.87 <sup>c</sup>
MLE 10%	1527.00 <sup>a</sup>	16.10 <sup>a</sup>	15.10 <sup>a</sup>	7.60 <sup>a</sup>	5.04 <sup>b</sup>

Mean values (n = 20) in each column followed by different letters are significantly different at  $p \le 0.05$  by Duncan's multiple range test.

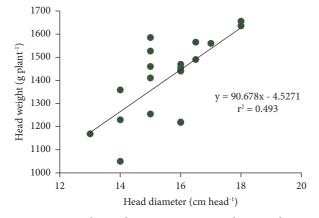


FIGURE 1: Scatter plots and regression equations between diameter and head weight.

The application of foliar moringa leaf extract had a significant impact on the enhancement of cabbage head diameter. Increasing the concentration of MLE leads to an increase in head diameter. Plants treated with 10% MLE had the largest head diameter at 16.10 cm head<sup>-1</sup>, followed by plants treated with 8 and 6% MLE at 16.0 and 15.70 cm head<sup>-1</sup>, respectively (Table 4).

Moringa aqueous extract contain cytokines, macro- and micronutrients, and other components that might be a reason for the significant improvement in head diameter. In addition to other substances that promote growth, such as phenols and ascorbic acid, MLE contains the cytokine zeatin [39, 40]. Additionally, MLE includes essential plant pigments such as carotenoids and chlorophyll, which are crucial for enhancing photosynthesis and thus boosting plant growth [41]. Abd-Elkafie et al. [42] observed maximum vegetative growth in coriander (*Coriandrum sativum* L.) sprayed with MLE.

3.1.3. Head Height. The above-ground growth of the cabbage head was measured as head height. According to the analysis of variance, spraying plants with MLE gradually improved the above-head height in cabbage. However, the only significant difference was identified when using a high concentration of 10% MLE, whereas 6 and 8% MLE did not affect head height (Table 4). Rayorath et al. [43] showed that higher concentrations of seaweed biostimulants (1 g L<sup>-1</sup>) are required to significantly boost Arabidopsis plant height, although lower concentrations (0.1 g L<sup>-1</sup>) can improve root weight. Hoque et al. [17] observed comparable results spraying MLE onto cabbage leaves 2 weeks and 4 weeks after transplanting, and 2 weeks after transplanting, and after every two weeks thereafter. Gibberellic acid and cytokines, two plant hormones, have given plant extract the value of a growth promoter and are most likely the cause of the cabbage plant's increased head height. Additionally, several physiological characteristics of the plant can be influenced by the micro- and macronutrients in MLE [44]. Culver [45] observed comparable outcomes regarding the enhancement of height in rape and cabbage plants that received foliar MLE spraying every two weeks.

3.1.4. Stem Length. Cabbage has a short stem that the leaves connect to, giving the head its round shape. Improving stem length produces rapid seed stalk and early maturity in the Brassicaceae family, resulting in a lower overall product quality. Many factors, including genetics, temperature, plant hormones [46-48], and agricultural techniques such as gibberellin A3 (GA3) spraying, can trigger steam development and bolting in cabbages [49]. Moringa leaf extract contains plenty of growth-promoting hormones (auxins, gibberellins, and cytokinins) [50, 51]; however, low concentration of 6-8% had no significant improvement in stem length, while by increasing the concentration of MLE to 10%, the stem length has a slight improvement but not significant (Table 4). This finding is intriguing since it demonstrates that applying MLE biostimulant has no negative effect on cabbage as bolting or early maturity.

3.1.5. Dry Matter Content. Dry matter is an important quality criterion in cabbages because plants with poor dry matter can have deformed leaves; as a result, ununiform shape occurs [52]. Many factors influence the nutritional quality including dry matter content in cabbages such as plant varieties, agricultural practices, and physiology of the plant [53, 54]. Cabbage plants treated with 6, 8, and 10% MLE significantly increased total dry matter content compared to untreated plants. The lowest dry matter content was found in the control ( $\emptyset$ ) plants at 4.63%, while plants foliar sprayed with 6% MLE had the higher dry matter content at 5.85% (Table 4).

#### 3.2. The Effect of MLE on the Development of Pigment Contents in Cabbage Leaves

3.2.1. Chlorophyll and Carotenoid Contents. Chlorophyll (Chl) and carotenoids are important phytochemical pigments because they play the main role in essential plant metabolisms such as photosynthesis process which relates to plant improvement and increasing the total yield [55]. Chlorophyll and carotenoids are light-harvesting pigments that play vital physiological functions in all photosynthetic organisms [56]. Many research studies had shown the positive role of MLE in improving chlorophyll and carotenoid content in different plants as in rice (*Oryza sativa* L.) under water stress conditions [57], peppers [16], fennel plants [58], SPAD value in lettuce [13, 14], and common bean (*Phaseolus vulgaris* L.) [59, 60].

Similar to the previous investigations, our findings show that cabbages treated with MLE had considerably higher levels of chlorophyll a, chlorophyll b, and carotenoids than control plants ( $\emptyset$ ). This could be due to moringa leaf extract inducing cytokinin, which leads to increased leaf area and higher chlorophyll levels in plant leaves [61]. Plants treated with 6, 8, and 10% MLE improved chlorophyll a and carotenoids significantly, while only plants treated with 10% MLE improved chlorophyll b significantly (Table 5).

3.3. Effects of MLE on Nutritional Quality and Mineral Content of Vitamin C. Vitamin C, commonly known as ascorbic acid, is a six-carbon molecule related to glucose. Its primary biological function is to act as a reducing agent in a variety of essential hydroxylation processes in the human body. The recommended daily allowance (RDA) of vitamin C for an adult is 60 mg per day, although this amount is debatable [32]. Another benefit of ascorbic acid is that it protects the body from several disorders caused by faulty collagen assemblies, such as skin lesions, unstable blood vessels, bleeding gums, osteoporosis, hemorrhages, and anemia [32, 62]. Our results show that plants treated with MLE could significantly improve the concentration of vitamin C. Among the treatments, MLE 6% could have a significant result of 33.10 (mg 100  $g^{-1}$ ) followed by the plants treated with 10% and 8% of moringa leaf extract (Table 6).

It has been stated that moringa leaf extract is a valuable product in hormones such as zeatin, a plant hormone cytokine [50, 63]. This could be the reason for increasing antioxidants and, as a result, ensuring plant vitality [64, 65]. Our findings agree with those of Toscano et al. [66], who discovered a significant increase in vitamin C content in baby leaf kale plants treated with MLE with the amount of 200 mg  $L^{-1}$ . In a study conducted by Yaseen and Takacs-Hajos [14], vitamin C levels in lettuce leaves of the Great Lakes variety were dramatically increased when 6% MLE was applied.

3.3.1. Nitrate Content. Nitrate or nitrate salts are the main components of fertilizers that have been used largely to boost crop development and increase vegetable yield in recent decades. The level of human nitrate exposure has risen considerably in recent years, which could be attributed to the enrichment of the biosphere with reactive nitrogen, as well as an increase in vegetable and preserved animal product consumption [67]. Recent research shows that the physiological role of nitrate or nitrite is responsible for their harmful function in our food and water supplies. About 80% of nitrate comes from vegetables we eat, followed by drinking water (10–20% or higher in various Asian and African countries), preserved meat (5%), and other sources.

The primary two disadvantages of higher nitrate intake in our meals are increasing methemoglobinemia (MetHb) disorder in infants and the possibility of an increase in cancer disease in adults [68].

Among vegetables, lettuce, spinach, celery, and beetroot have the highest nitrate contents (above 1000 mg kg<sup>-1</sup>), whereas potatoes, cabbage, and spring greens contain about 100–1000 mg kg<sup>-1</sup>, and tomatoes have the lowest concentrations lower than 100 mg kg<sup>-1</sup> [69]. According to studies, the ADI (acceptable daily intake) must be 3.65 mg kg<sup>-1</sup> body weight [70]; nevertheless, the estimated dietary intake of nitrate in various European countries ranges from 31 to 185 mg/day<sup>-1</sup>.

Our research studies show that spraying of MLE with the concentration of 6, 8, and 10% had a significant reduction in nitrate content compared to the control plants; however, by increasing the concentration of MLE by 10%, the nitrate content was gradually increased (Table 6).

This could be because a larger concentration of MLE allows the plant to absorb more nitrogen [71]. Furthermore, moringa leaf extract contains a high concentration of nitrate and other essential nutrients [72], which allows the plant to accumulate a higher concentration of nitrate in the leaves. Sardar et al. [73] discovered that spraying stevia (*Stevia rebaudiana* Bertoni) with 20% MLE could considerably enhance N, P, K, Ca, Mg, Na, and Zn accumulation in plant leaves, whereas Yaseen and Takacs-Hajos [14] initiate nitrate reduction in lettuce leaves by spraying with low concentration (6%) of moringa leaf extract.

3.3.2. Mineral Content in Cabbage Leaves. Moringa leaf extract contains a high concentration of macro- and micronutrients (Table 3), which provides additional nutrition to the treated plants. Our mineral enhancement findings are consistent with previous research on vegetable crops. Table 7 shows significant increases for plants treated with 6, 8, and 10% MLE. Increasing MLE concentrations, except for nitrogen, has resulted in enhanced nutritional content such as Ca, K, Mg, and P. This could be because moringa leaf extract reduces nitrate levels in plant leaves [13, 66]. Various studies on other vegetables suggest that treatment with MLE can increase macro and micronutrient levels in many plants.

This could be because MLE promotes nutrient uptake by increasing root membrane permeability and nutrient mobility [50], as well as the extract's nutrient content. High concentrations of 10, 20, and 30% MLE could improve the leaf's N, P, K, Ca, Mg, Na, and Zn concentrations in stevia (*Stevia rebaudiana* Bertoni) [73], nutrient improvement in basil plant (*Ocimum basilicum*) under salt stress [74]; also, N, P, K, Ca, Mg, and Fe improvement was found in pea seeds (*Pisum sativum*) through foliar sprayed with 1, 2, 3, and 4% MLE [75].

3.4. Correlation of the Vegetation Growth of Cabbage to Some Growth Measurements. Table 8 clearly shows that all vegetative parameters have a favourable correlation to growth

Treatments	Chlorophyll <i>a</i> ( $\mu$ g mL <sup>-1</sup> )	Chlorophyll <i>b</i> ( $\mu$ g mL <sup>-1</sup> )	Carotenoids ( $\mu g m L^{-1}$ )
Control (Ø)	13.09 <sup>c</sup>	4.23 <sup>b</sup>	4.86 <sup>c</sup>
MLE 6%	14.96 <sup>b</sup>	$4.80^{ab}$	5.98 <sup>b</sup>
MLE 8%	16.61 <sup>ab</sup>	$5.00^{\mathrm{ab}}$	$6.07^{b}$
MLE 10%	17.95 <sup>a</sup>	5.64 <sup>a</sup>	7.29 <sup>a</sup>

TABLE 5: Chlorophyll a, chlorophyll b, and carotenoid content in cabbage fresh leaves.

Mean values (n = 20) in each column followed by different letters are significantly different at  $p \le 0.05$  by Duncan's multiple range test.

TABLE 6: Ascorbic acid (vitamin C) and nitrate content in cabbage fresh leaves.

Treatments	Vitamin C (mg $100 \text{ g}^{-1}$ )	$NO_{3}^{-} (mg \ kg^{-1})$
Control Ø	28.56 <sup>c</sup>	527.00 <sup>a</sup>
MLE 6%	33.10 <sup>a</sup>	428.66 <sup>b</sup>
MLE 8%	29.90 <sup>b</sup>	462.33 <sup>b</sup>
MLE 10%	30.46 <sup>b</sup>	470.33 <sup>b</sup>

Mean values (n = 20) in each column followed by different letters are significantly different at  $p \le 0.05$  by Duncan's multiple range test.

TABLE 7: Mineral elements and nitrate in cabbage samples treated with moringa leaf extract (MLE).

Treatments	Ca (m/m) %	K (m/m) %	Mg (m/m) %	P (m/m) %	N (m/m) %
Control (Ø)	0.48 <sup>c</sup>	3.62 <sup>c</sup>	0.17 <sup>b</sup>	$0.54^{\mathrm{b}}$	2.30 <sup>c</sup>
MLE 6%	$0.56^{\mathrm{b}}$	3.95 <sup>b</sup>	0.21 <sup>a</sup>	$0.58^{a}$	2.68 <sup>b</sup>
MLE 8%	$0.60^{\mathrm{ab}}$	4.16 <sup>b</sup>	0.22 <sup>a</sup>	0.59 <sup>a</sup>	2.71 <sup>a</sup>
MLE 10%	0.64 <sup>a</sup>	4.46 <sup>a</sup>	0.26 <sup>a</sup>	0.61 <sup>a</sup>	2.62 <sup>b</sup>

Mean values (n = 20) in each column followed by different letters are significantly different at  $p \le 0.05$  by Duncan's multiple range test.

TABLE 8: Pearson correlation between vegetative parameters and growth measurements in cabbage (Brassica oleracea L. var. capitata).

Parameters	Head weight	Stem length	Head diameter	Head height	Dry matter	Chl a	Chl b	Carotenoids	Vitamin C	Nitrate
Head weight	1									
Stem length	0.414	1								
Head diameter	0.702**	0.28	1							
Head height	0.671**	0.178	0.798**	1						
Dry matter	0.319	-0.143	0.241	0.229	1					
Chl a	0.750**	0.334	$0.637^{*}$	0.465	0.123	1				
Chl b	0.635*	0.142	0.498	0.38	0.089	0.925**	1			
Carotenoids	0.758**	0.333	0.663*	0.516	0.136	0.990**	0.903**	1		
Vitamin C	0.635*	-0.197	0.516	0.432	0.870**	0.442	0.414	0.44	1	
Nitrate	0.311	0.554	0.252	0.31	-0.462	0.458	0.459	0.472	-0.308	1

\*\*Correlation is significant at the 0.01 level (2-tailed). \*Correlation is significant at the 0.05 level (2-tailed).

measurements. The only negative correlation detected was between vitamin C and nitrate level, which accords with the findings of Koh et al. [76], who discovered a negative correlation between nitrate, ascorbic acid, and vitamin C in spinach (*Spinacia oleracea* L.) leaves. This finding will be advantageous because a research study has revealed that ascorbic acid and flavonoids can reduce the harmful effects of nitrites [77].

Head weight correlated significantly with head diameter, head height, chlorophyll a, and carotenoids at the p level 0.01, while chlorophyll b and vitamin C correlated significantly at the p level 0.05. However, no significant association was observed across stem length and head height and growth factors. Head diameter, on the other hand, was found to have a significantly positive correlation with head height at p level 0.01, as well as a positive correlation with chlorophyll a and carotenoids at p level 0.05. The dry matter has a significant positive correlation with vitamin C at p level 0.01, whereas a negative correlation with nitrate. Chlorophyll a and chlorophyll b had a substantial positive association with carotenoids among the growth parameters, while chlorophyll a had a positive correlation to chlorophyll b at p level 0.01. Our result shows that cabbage plants with greater chlorophyll content can produce better physical parameters in terms of growth performance and delaying flowering or bolting.

#### 4. Conclusion

We can conclude that MLE is an effective plant biostimulant to be used to improve nutritional and physical parameters in cabbage (*Brassica oleracea* var. *capitata*). This study suggests that foliar application of 10% MLE to cabbage had a significant positive effect on morphological parameters, mineral absorption, nutrient content, and bioactive compounds without improving stem length, resulting in a significant increase in the gross and marketable yield of the vegetable crop and nitrate content in cabbage leaves. However, by increasing the percentage of MLE from 6% to 10%, the nitrate reduction is decreasing by about half. Nitrate had a negative correlation with vitamin C and dry matter. These findings imply that plants with higher pigment content have higher morphological and nutrient value.

#### **Data Availability**

The data used to support the study are available from the corresponding author upon request.

#### **Conflicts of Interest**

The authors declare that they have no conflicts of interest.

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#### References

- J. Singh, A. K. Upadhyay, A. Bahadur, B. Singh, K. P. Singh, and M. Rai, "Antioxidant phytochemicals in cabbage (Brassica oleracea L. var. capitata)," *Scientia Horticulturae*, vol. 108, no. 3, pp. 233–237, 2006.
- [2] Food and Agriculture Organization (Fao), Production Quantities Of Cabbages By Country, FAO, Rome, Italy, 2023.
- [3] Statista, "Production volume of white cabbage in Hungary from 2010 to 2021(in tons)," 2023, https://www.statista.com/ statistics/1301698/hungary-production-volume-of-whitecabbage/.
- [4] M. Nieuwhof, *Cole Crops: Botany, Cultivation and Utilization*, Leonard Hill, London, UK, 1969.
- [5] A. Sharma, I. J. Rathore, A. Ali et al., "Major diseases and pathogen ecology of cabbage," *The Pharma Innovation Journal*, vol. 7, no. 7, pp. 574–578, 2018.
- [6] M. Turan, M. Ekinci, E. Yildirim et al., "Plant growthpromoting rhizobacteria improved growth, nutrient, and hormone content of cabbage (Brassica oleracea) seedlings," *Turkish Journal of Agriculture and Forestry*, vol. 38, no. 3, pp. 327–333, 2014.
- [7] E. Ibukunoluwa Moyin-Jesu, "Use of different organic fertilizers on soil fertility improvement, growth and head yield parameters of cabbage (Brassica oleraceae L)," *International Journal of Recycling of Organic Waste in Agriculture*, vol. 4, no. 4, pp. 291–298, 2015.
- [8] M. Moamogwe, "Adaptation trial of introduced cabbage cultivars ARP training reports between 1993–1995," AVRDC Africa Regulation Program. Arusha Tanzania, vol. 4, no. 1, pp. 27–29, 1995.

- [9] S. M. Nkosi and N. M. Msimango, "Screening of zinc, copper and iron in lettuce and Chinese cabbage cultivated in Durban, South Africa, towards human health risk assessment," *South African Journal of Science*, vol. 118, no. 11/12, pp. 1–5, 2022.
- [10] S. K. Pradhan, A. M. Nerg, A. Sjöblom, J. K. Holopainen, and H. Heinonen-Tanski, "Use of human urine fertilizer in cultivation of cabbage (Brassica oleracea)—impacts on chemical, microbial, and flavor quality," *Journal of Agricultural and Food Chemistry*, vol. 55, no. 21, pp. 8657–8663, 2007.
- [11] R. S. El-Serafy, A. N. A. El-Sheshtawy, U. A. Abd El-Razek et al., "Growth, yield, quality, and phytochemical behavior of three cultivars of quinoa in response to moringa and azolla extracts under organic farming conditions," *Agronomy*, vol. 11, no. 11, pp. 1–25, 2021.
- [12] N. Admane, G. Cavallo, C. Hadjila et al., "Biostimulant formulations and moringa oleifera extracts to improve yield, quality, and storability of hydroponic lettuce," *Molecules*, vol. 28, no. 1, p. 373, 2023.
- [13] A. A. Yaseen and M. T. Hájos, "The potential role of moringa leaf extract as bio-stimulant to improve some quality parameters of different lettuce (Lactuca sativa L.) genotypes," *Sarhad Journal of Agriculture*, vol. 37, no. 4, pp. 1107–1119, 2021.
- [14] A. A. Yaseen and M. Takacs-Hajos, "Evaluation of moringa (Moringa oleifera Lam.) leaf extract on bioactive compounds of lettuce (Lactuca sativa L.) grown under glasshouse environment," *Journal of King Saud University Science*, vol. 34, no. 4, Article ID 101916, 2022.
- [15] I. Ahmad, J. M. Dole, M. A. Khan, M. Qasim, T. Ahmad, and A. S. Khan, "Preliminary and regional reports," *Hort Technology*, vol. 26, no. 3, pp. 327–337, 2016.
- [16] H. Hala, E. Abou, and A. Nabila, "Effect of Moringa oleifera leaf extract (MLE) on pepper seed germination, seedlings improvement, growth, fruit yield and its quality," *Middle East Journal of Agriculture Research*, vol. 6, no. 2, pp. 448–463, 2017.
- [17] T. S. Hoque, M. S. Rana, S. A. Zahan, I. Jahan, and M. A. Abedin, "Moringa leaf extract as a bio-stimulant on growth, yield and nutritional improvement in cabbage," *Asian Journal of Medical and Biological Research*, vol. 6, no. 2, pp. 196–203, 2020.
- [18] A. A. Yaseen and M. Takácsné Hájos, "Effect of Moringa leaf extract and set size on the bulb weight, diameter and yield of onions (Allium cepa L.)," *Acta Agraria Debreceniensis*, vol. 2, pp. 127–131, 2020.
- [19] R. Mohammed, M. Olorukooba, M. Akinyaju, and E. Kambai, "Evaluation of different concentrations and frequency of foliar application of moringa extract on growth and yield of onion, Allium cepa lam," *Agrosearch*, vol. 13, no. 3, pp. 196–205, 2014.
- [20] T. A. Abd El-Mageed, W. M. Semida, and M. M. Rady, "Moringa leaf extract as biostimulant improves water use efficiency, physio-biochemical attributes of squash plants under deficit irrigation," *Agricultural Water Management*, vol. 193, pp. 46–54, 2017.
- [21] H. I. Muhammad, M. Z. Asmawi, and N. A. K. Khan, "A review on promising phytochemical, nutritional and glycemic control studies on Moringa oleifera Lam. in tropical and subtropical regions," *Asian Pacific Journal of Tropical Biomedicine*, vol. 6, no. 10, pp. 896–902, 2016.
- [22] V. Pakade, E. Cukrowska, and L. Chimuka, "Metal and flavonol contents of Moringa oleifera grown in South Africa," *South African Journal of Science*, vol. 109, no. 3/4, pp. 1–7, 2013.

- [23] P. Nobossé, E. N. Fombang, and C. M. F. Mbofung, "Effects of age and extraction solvent on phytochemical content and antioxidant activity of fresh Moringa oleifera L. leaves," *Food Science and Nutrition*, vol. 6, no. 8, pp. 2188–2198, 2018.
- [24] C. W. Liu, Y. Sung, B. C. Chen, and H. Y. Lai, "Effects of nitrogen fertilizers on the growth and nitrate content of lettuce (Lactuca sativa L.)," *International Journal of Envi*ronmental Research and Public Health, vol. 11, no. 4, pp. 4427–4440, 2014.
- [25] E. N. K. Sowley, F. Kankam, and J. Adomako, "Management of root-knot nematode (Meloidogyne spp.) on sweet pepper (Capsicum annuum L.) with moringa (Moringa oleifera Lam.) leaf powder," *Archives of Phytopathology and Plant Protection*, vol. 47, no. 13, pp. 1531–1538, 2014.
- [26] H. P. S. Makkar and K. Becker, "Nutrients and antiquality factors in different morphological parts of the Moringa oleifera tree," *Practice Nursing*, vol. 20, no. 2, p. 60, 2009.
- [27] H. K. Lichtenthaler and C. Buschmann, "Chlorophylls and carotenoids: measurement and characterization by UV-VIS spectroscopy," *Current Protocols in Food Analytical Chemistry*, vol. 1, no. 1, pp. F4–F2, 2001.
- [28] K. A. Ali, S. S. Noraldeen, and A. A. Yaseen, "An evaluation study for chlorophyll estimation techniques," *Sarhad Journal* of Agriculture, vol. 37, no. 4, pp. 1458–1465, 2021.
- [29] S. I. Vica, V. Laslo, S. Pantea, and G. E. Bandici, "Chlorophyll and carotenoids pigments from Mistletoe (Viscum album) leaves using different solvents," *Analele Universității din Oradea-Fascicula Biologie*, vol. 17, no. 2, pp. 213–218, 2010.
- [30] Ş. Dere, T. Güneş, and R. Sivaci, "Spectrophotometric determination of chlorophyll-A, B and total caretenoid contents of some algae species using different solvents," *Turkish Journal* of Botany, vol. 22, no. 1, pp. 13–16, 1998.
- [31] A. A. Yaseen and M. Takacs-Hajos, "The effect of plant biostimulants on the macronutrient content and ion ratio of several lettuce (Lactuca sativa L.) cultivars grown in a plastic house," *South African Journal of Botany*, vol. 147, pp. 223–230, 2022.
- [32] P. Ciancaglini, H. L. Santos, K. R. P. Daghastanli, and G. Thedei, "Using a classical method of vitamin C quantification as a tool for discussion of its role in the body," *Biochemistry and Molecular Biology Education*, vol. 29, no. 3, pp. 110–114, 2001.
- [33] V. Kmecl, T. Knap, and D. Žnidarčič, "Evaluation of the nitrate and nitrite content of vegetables commonly grown in Slovenia," *Italian Journal of Agronomy*, vol. 12, no. 2, pp. 79–84, 2017.
- [34] A. Louro, D. Báez, M. I. García, and L. Cárdenas, "Nitrous oxide emissions from forage maize production on a Humic Cambisol fertilized with mineral fertilizer or slurries in Galicia, Spain," *Geoderma Regional*, vol. 5, pp. 54–63, 2015.
- [35] N. Teribia, V. Tijero, and S. Munné-Bosch, "Linking hormonal profiles with variations in sugar and anthocyanin contents during the natural development and ripening of sweet cherries," *New Biotech*, vol. 33, no. 6, pp. 824–833, 2016.
- [36] M. Nasir, A. S. Khan, S. A. Basra, and A. U. Malik, "Foliar application of moringa leaf extract, potassium and zinc influence yield and fruit quality of "Kinnow" Mandarin," *Scientia Horticulturae*, vol. 210, pp. 227–235, 2016.
- [37] T. S. Hoque, M. A. Abedin, M. G. Kibria, I. Jahan, and M. A. Hossain, "Application of moringa leaf extract improves growth and yield of Tomato (Solanum lycopersicum) and Indian Spinach (Basella alba)," *Plant Science Today*, vol. 9, no. 1, pp. 137–143, 2021.

- [38] C. Yang, T. X. Liu, and J. H. Everitt, "Estimating cabbage physical parameters using remote sensing technology," *Crop Protection*, vol. 27, no. 1, pp. 25–35, 2008.
- [39] N. Foidle, H. Makkar, and Becker, "The potential of moringa oleifera for agricultural and industrial uses," in *The Miracle Tree: The Multipurpose Attributes of Moringa*, L. Fuglie, Ed., pp. 45–76, CTA, Wageningen, Netherlands, 2001.
- [40] L. Fuglie, "New uses of Moringa studied in Nicaragua," ECHO Development Notes, vol. 68, pp. 1–25, 2000.
- [41] M. A. Iqbal, "Role of moringa, Brassica and sorghum water extracts in increasing crops growth and yield: a review," *American-Eurasian Journal of Agricultural and Environmental*, vol. 14, no. 11, pp. 1150–1158, 2014.
- [42] O. Abd-ElKafie, M. Kasem, S. Abd-ElBaki, and M. Sidky, "Improving the vegetative growth and chemical contents of coriander (coriandrum sativum L.) plant by using moringa leaf extract," *Journal of Plant Production*, vol. 7, no. 12, pp. 1357–1363, 2016.
- [43] P. Rayorath, W. Khan, R. Palanisamy et al., "Extracts of the brown seaweed Ascophyllum nodosum induce gibberellic acid (GA3)-independent amylase activity in barley," *Journal* of Plant Growth Regulation, vol. 27, no. 4, pp. 370–379, 2008.
- [44] H. P. S. Makkar, G. Francis, and K. Becker, "Bioactivity of phytochemicals in some lesser-known plants and their effects and potential applications in livestock and aquaculture production systems," *Animal*, vol. 1, no. 9, pp. 1371–1391, 2007.
- [45] M. Culver, "Effect of Moringa oleifera leaf aqueous extract on growth and yield of rape and cabbage," *African Journal of Biotechnology*, vol. 11, no. 73, pp. 13796–13800, 2012.
- [46] E. Kou, X. Huang, Y. Zhu et al., "Crosstalk between auxin and gibberellin during stalk elongation in flowering Chinese cabbage," *Scientific Reports*, vol. 11, no. 1, p. 3976, 2021.
- [47] S. R. Dahanayake and N. W. Galwey, "Effects of interactions between low-temperature treatments, gibberellin (GA3) and photoperiod on flowering and stem height of spring rape (Brassica napus var. annua)," *Annals of Botany*, vol. 84, no. 3, pp. 321–327, 1999.
- [48] S. W. Song, Y. L. Lei, X. M. Huang, W. Su, R. Y. Chen, and Y. W. Hao, "Crosstalk of cold and gibberellin effects on bolting and flowering in flowering Chinese cabbage," *Journal* of *Integrative Agriculture*, vol. 18, no. 5, pp. 992–1000, 2019.
- [49] H. Guan, X. Huang, Y. Zhu et al., "Identification of della genes and key stage for ga sensitivity in bolting and flowering of flowering Chinese cabbage," *International Journal of Molecular Sciences*, vol. 22, p. 12092, 2021.
- [50] M. A. Mona, "The potential of Moringa oleifera extract as a biostimulant in enhancing the growth, biochemical and hormonal contents in rocket (Eruca vesicaria subsp. sativa) plants," *International Journal of Plant Physiology and Biochemistry*, vol. 5, no. 3, pp. 42–49, 2013.
- [51] A. Yaseen and M. T. Hájos, "Study on moringa tree (Moringa oleifera lam.) leaf extract in organic vegetable production: a review," *Research on Crops*, vol. 21, no. 2, pp. 402–414, 2020.
- [52] C. Avalhães, P. Rd, R. Lm, R. De, and C. Ma, "Omission of macronutrients of the growth and nutritional status of plants of cabbage grown in nutrient solution," *Bioscience Journal*, vol. 25, no. 5, pp. 21–28, 2009.
- [53] R. Pokluda, "Nutritional quality of Chinese cabbage from integrated culture," *Horticultural Science*, vol. 35, no. 4, pp. 145–150, 2008.
- [54] D. L. da Silva, R. de Mello Prado, L. F. L. Tenesaca, J. L. F. da Silva, and B. H. Mattiuz, "Silicon attenuates calcium deficiency by increasing ascorbic acid content, growth and

quality of cabbage leaves," *Scientific Reports*, vol. 11, no. 1, pp. 1770–1779, 2021.

- [55] P. Swapnil, M. Meena, S. K. Singh, U. P. Dhuldhaj, Harish, and A. Marwal, "Vital roles of carotenoids in plants and humans to deteriorate stress with its structure, biosynthesis, metabolic engineering and functional aspects," *Current Plant Biology*, vol. 26, Article ID 100203, 2021.
- [56] N. Mezzomo and S. R. S. Ferreira, "Carotenoids functionality, sources, and processing by supercritical technology: a review," *Journal of Chemistry*, vol. 2016, Article ID 3164312, 16 pages, 2016.
- [57] S. Khan, A. Basit, M. B. Hafeez et al., "Moringa leaf extract improves biochemical attributes, yield and grain quality of rice (Oryza sativa L.) under drought stress," *PLoS One*, vol. 16, no. 7, 2021.
- [58] R. S. El-Serafy and A. A. El-Sheshtawy, "Effect of nitrogen fixing bacteria and moringa leaf extract on fruit yield, estragole content and total phenols of organic fennel," *Scientia Horticulturae*, vol. 265, Article ID 109209, 2020.
- [59] M. M. Rady and G. F. Mohamed, "Modulation of salt stress effects on the growth, physio-chemical attributes and yields of Phaseolus vulgaris L. plants by the combined application of salicylic acid and Moringa oleifera leaf extract," *Scientia Horticulturae*, vol. 193, pp. 105–113, 2015.
- [60] F. Zulfiqar, A. Casadesús, H. Brockman, and S. Munné-Bosch, "An overview of plant-based natural biostimulants for sustainable horticulture with a particular focus on moringa leaf extracts," *Plant Science*, vol. 295, Article ID 110194, 2020.
- [61] M. M. Rady, G. F. Mohamed, A. M. Abdalla, and Y. H. M. Ahmed, "Integrated application of salicylic acid and Moringa oleifera leaf extract alleviates the salt-induced adverse effects in common bean plants," *Journal of Agricultural Technologies*, vol. 11, no. 7, pp. 1595–1614, 2015.
- [62] L. Stryer, *Molecular Motors. Biochemistry*, WH Freeman and Company, New York, NY, USA, 4th edition, 1995.
- [63] B. M. L. Price, "The moringa tree," in ECHO Technical Note, pp. 1–19, ECHO, Sydney, Australia, 2007.
- [64] A. Yasmeen, S. M. A. Basra, R. Ahmad, and A. Wahid, "Performance of late sown wheat in response to foliar application of moringa oleifera lam. Leaf extract," *Chilean Journal of Agricultural Research*, vol. 72, no. 1, pp. 92–97, 2012.
- [65] H. U. Rehman, Q. Nawaz, S. M. A. Basra, I. Afzal, A. Yasmeen, and F. Ul-Hassan, "Seed priming influence on early crop growth, phenological development and yield performance of linola (Linum usitatissimum L.)," *Journal of Integrative Agriculture*, vol. 13, no. 5, pp. 990–996, 2014.
- [66] S. Toscano, A. Ferrante, F. Branca, and D. Romano, "Enhancing the quality of two species of baby leaves sprayed with moringa leaf extract as biostimulant," *Agronomy*, vol. 11, no. 7, p. 1399, 2021.
- [67] N. S. Bryan and H. van Grinsven, "The role of nitrate in human health," *Advances in Agronomy*, vol. 119, pp. 153–182, 2013.
- [68] J. L'hirondel, Nitrate and Man: Toxic, Harmless or Beneficial?, CABI, New York, NY, USA, 2002.
- [69] S. D. Gangolli, B. P. Van Den, V. J. Feron et al., "Nitrate, nitrite and N-nitroso compounds," *European Journal of Pharmacology: Environmental Toxicology and Pharmacology*, vol. 292, no. 1, pp. 1–38, 1994.
- [70] W. Derave and Y. Taes, "Beware of the pickle: health effects of nitrate intake," *Journal of Applied Physiology*, vol. 107, no. 5, p. 1677, 2009.

- [71] M. A. Mona, "The potential of Moringa oleifera extract as a biostimulant in enhancing the growth, biochemical and hormonal contents in rocket (Eruca vesicaria subsp. sativa) plants," *International Journal of Plant Physiology and Biochemistry*, vol. 5, no. 3, pp. 42–49, 2013.
- [72] Y. Arif, A. Bajguz, and S. Hayat, "Moringa oleifera extract as a natural plant biostimulant moringa oleifera extract as a natural plant biostimulant," *Journal of Plant Growth Regulation*, pp. 1–6, 2022.
- [73] H. Sardar, A. Nisar, M. A. Anjum et al., "Foliar spray of moringa leaf extract improves growth and concentration of pigment, minerals and stevioside in stevia (Stevia rebaudiana Bertoni)," *Industrial Crops and Products*, vol. 166, Article ID 113485, 2021.
- [74] M. A. Alkuwayti, F. El-Sherif, Y. K. Yap, and S. Khattab, "Foliar application of Moringa oleifera leaves extract altered stress-responsive gene expression and enhanced bioactive compounds composition in Ocimum basilicum," *South African Journal of Botany*, vol. 129, pp. 291–298, 2020.
- [75] A. R. M. A. Merwad, "Using Moringa oleifera extract as biostimulant enhancing the growth, yield and nutrients accumulation of pea plants," *Journal of Plant Nutrition*, vol. 41, no. 4, pp. 425–431, 2018.
- [76] E. Koh, S. Charoenprasert, and A. E. Mitchell, "Effect of organic and conventional cropping systems on ascorbic acid, vitamin C, flavonoids, nitrate, and oxalate in 27 varieties of spinach (Spinacia oleracea L.)," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 12, pp. 3144–3150, 2012.
- [77] R. Walker, "Nitrates, nitrites and N-nitrosocompounds: a review of the occurrence in food and diet and the toxicological implications," *Food Additives and Contaminants*, vol. 7, no. 6, pp. 717–768, 1990.