

# Research Article

# Screening and Analysis of Biomarkers in the miRNA-mRNA Regulatory Network of Osteosarcoma

## Gang Han, Quanyi Guo 💿, Nan Li, Wenzhi Bi, Meng Xu, and Jinpeng Jia

Department of Orthopedics, The Fourth Medical Center of General Hospital of PLA, Beijing 100048, China

Correspondence should be addressed to Quanyi Guo; guoquanyi@pla304hosptal.org.cn

Received 10 January 2022; Revised 10 February 2022; Accepted 11 February 2022; Published 15 March 2022

Academic Editor: Bhagyaveni M.A

Copyright © 2022 Gang Han et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Osteosarcoma is a malignant disease, and few effective strategies can completely overcome the prognosis of these patients. This study attempted to reveal the key factors and related molecular mechanisms of osteosarcoma via excavating public microarray datasets. The data were obtained from the Gene Expression Omnibus (GEO) database; the differentially expressed miRNAs and differentially expressed genes were obtained in GSE69470 and GSE126851, respectively; the target of miRNAs were predicted with the miRDIP database; the functions of the factors were analyzed and visualized by the David database and R language, respectively. Moreover, the protein-protein interaction network and miRNA-mRNA network were performed with the STRING database and Cytoscape software to identify the hub nodes in GSE69470 and GSE12685. The results showed that 834 DEGs were found in GSE12685 and 37 miRNAs were found in GSE69470. Moreover, the target of 37 miRNAs were enriched in PI3K/AKT, P53, Wnt/ $\beta$ -catenin, and TGF- $\beta$  pathways and related with skeletal system development and cell growth. Besides, the miRNAs including miR-22-3p, miR-154-5p, miR-34a-5p, miR-93-5p, and miR-95-5p and the genes including LEF1, RUNX2, CSF1R, CDKN1A, and FBN1 were identified as the hub nodes via network analysis. In conclusion, this study suggested that the miRNAs including miR-22-3p, miR-154-5p, miR-34a-5p, miR-485-3p, miR-93-5p, and miR-9-5p and the genes including LEF1, RUNX2, CSF1R, CDKN1A, and FBN1 act as key factors in the progression of osteosarcoma.

# 1. Introduction

Osteosarcoma is a serious bone malignancy with a high tendency of aggressiveness and metastases, which has a high incidence in childhood [1, 2]. Statistically, more than 60% of patients with osteosarcoma are under 25 years old [3]. At present, surgery combined with neoadjuvant chemotherapy is the pillar for osteosarcoma treatment [4]. In recent ten years, the application of multiple radiotherapy and chemotherapy techniques and mature modern surgical skills have effectively changed the prognosis of patients [5]. With the current therapy strategies, a 5-year survival rate of patients has increased to 70% [6]. However, many patients still find it difficult to escape the bad outcomes in the long term. Although accumulating research has continually figured out the potential reasons of osteosarcoma development, the detailed mechanisms of osteosarcoma remain unclear, and deep research is urgently necessary.

MicroRNAs play important roles in the metabolic activities of cells via regulating the expression of proteins [7, 8]. Several reporters have indicated that the miRNA profile of osteosarcoma exhibits a high difference with the normal tissues, and some miRNAs have been identified as the hub nodes in regulating the malignant behavior of the tumor cells, such as rapid proliferation and high invasion [9]. For osteosarcoma, the miRNA profiles of patients also exhibit significant differences compared with normal people. Microarray analysis has been frequently used for revealing RNA profiles of diseases, which is useful for analyzing the related pathological mechanisms [10]. Some research has revealed the related molecular mechanism of some tumors via microarray analysis [11, 12].

This study aimed to analyze the miRNA profile and identify the key miRNAs of osteosarcoma via the GEO database, reveal the functions and potential regulation mechanisms of the miRNAs, and finally provide some novel references for osteosarcoma treatment.

#### 2. Materials and Methods

2.1. Data Source. The microarray datasets of GSE12685 and GSE69470 were obtained from the GEO database (https://www.ncbi.nlm.nih.gov). The raw data of the datasets were downloaded with the packages of the R language including GEOquery, dplyr, and Limma.

2.2. Identification of Differentially Expressed Genes. All datasets were analyzed with the GEO2R tools of GEO, and the expressed matrix files of the datasets were obtained. For GSE12685, 11 clinical pathological samples of osteosarcoma tumor and 2 differential normal human osteoblasts were used to identify the DEGs. For GSE69470, the DEGs were analyzed in human osteosarcoma cell lines and four normal human cell lines including chondrocyte cells, mesenchymal stem cells, osteoblasts, and skeletal muscle cells. The genes with |logFC| > 2 and P < 0.05 were considered as the differentially expressed genes (DEGs).

2.3. Screening of Potential Targets of the miRNAs. The common genes in the differentially expressed miRNAs in the matrix files were selected and then used for the next analysis. For GSE69470, the common differentially expressed miRNAs in the matrix files are the osteosarcoma tumor cells and differentially normal human osteoblast cells, osteosarcoma tumor cells, and mesenchymal stem cells (MSCs). The potential targets of common miRNAs were predicted with the miRDIP database (https://ophid.utoronto.ca/mirDIP/). The genes with a high minimum score (top 5%) were selected for the targets of the common miRNAs. Moreover, the common genes of the matrices were screed and visualized with VENNY (https://bioinfogp.cnb.csic.es/tools/venny/ index.html).

2.4. Kyoto Encyclopedia of Genes and Genomes (KEGG) Enrichment Analysis. A KEGG enrichment analysis was performed for investigating the related pathways of osteosarcoma. For GES12685, the DEGs were analyzed with KEGG enrichment. For GSE69470, the potential targets of differentially expressed miRNAs were used for KEGG enrichment analysis. The genes were annotated and analyzed by the David database (https://david.ncifcrf.gov/), and the related pathways in the results with a *P* value <0.05 were visualized with the R language.

2.5. Gene Ontology (GO) Enrichment Analysis. GO enrichment analysis was used to identify the molecular functions of genes. For GES12685, the DEGs were annotated by the David database, and then the related ENTREZID of genes were analyzed with the packages of the R language including clusterProfiler and org.Hs.eg.db. For the miRNAs, the targets of them were analyzed with GO enrichment. Finally, the related functions module with a P value <0.05 were visualized with the R language.

2.6. Network Analysis. The PPI network analysis was performed to identify the hub nodes of osteosarcoma. In brief, the common genes of the targets of the miRNAs in GSE69470 and the DEGs in GSE69470 were analyzed with the STRING database (https://cn.string-db.org/), and then visualized with Cytoscape software. For the miRNA-mRNA network, the connection of the differentially expressed miRNAs in GSE69470 and the DEGs in GES12685 were structured with Cytoscape. For the miRNA-mRNA network, the differentially expressed miRNAs in GSE69470 and their targets were visualized by Cytoscape software (3.4.0).

#### 3. Results

3.1. Identification of miRNAs of the Datasets. The datasets were used to analyze the differentially expressed miRNAs. For GSE12685, 11 clinical pathological samples of osteosarcoma tumor and 2 healthy samples of normal human osteoblast were used to identify the DEGs, and 424 downregulated genes and 410 upregulated genes were found in osteosarcoma (Figures 1(c) and 2(c)). For GSE69470, the differentially expressed miRNAs were analyzed in human osteosarcoma cell lines and four normal human cell lines including chondrocyte cells, mesenchymal stem cells, osteoblasts, and skeletal muscle cells. Compared with the expression profiles of chondrocyte, osteoblasts, and skeletal muscle cells, 45 downregulated genes and 10 upregulated genes were found in human osteosarcoma cell line (Figure 1). Compared with the expression profile of human mesenchymal stem cells, 43 downregulated genes and 16 upregulated genes were found in human osteosarcoma cell lines (Figure 1). Moreover, 37 common miRNAs were found in the matrices of GSE69470.

3.2. Identification of the Targets of the Common miRNAs. To identify the key miRNAs in the development of osteosarcoma, the 37 common miRNAs including 30 downregulated miRNAs and 7 upregulated miRNAs were screened in the differentially expressed miRNAs of osteosarcoma cell lines compared with normal differential cells or mesenchymal stem cells (Figure 2(c)). 37 common miRNAs were screened in GSE69470, and 4037 potential targets were predicted by the miRDIP database. Moreover, 114 common genes were screened from the DEGs of GSE12685 and the targets of miRNAs in in GSE69470 (Figures 3(c) and 3(d)).

*3.3. Functional Modules Analysis.* To investigate the pathological mechanism of osteosarcoma, the targets of the screened miRNAs in GSE69470, the DEGs in GSE12685, and the common genes of them were analyzed with the KEGG enrichment. The DEGs in GSE12685 were associated with the extracellular matrix organization, regulation of vasculature development, skeletal system development, and so on (Figure 3(a)). For GSE69470, it was found that the targets of



FIGURE 1: Volcano plots of differentially expressed factors in GSE69470 and GSE126851. (a) The differentially expressed factors of human osteosarcoma cells and differentiated bone cells in GSE69470. (b) The differentially expressed factors of human osteosarcoma cells and mesenchymal stem cells in GSE69470. (c) The differentially expressed factor in GSE126851.

the screened miRNAs were involved in the skeletal system development, covalent chromatin modification, cell growth, and so on (Figure 3(b)). Moreover, it was found that the common genes were related with skeletal system development, regulation of cellular response to growth factor stimulus, regulation of cell growth, and so on (Figure 3(e)).

3.4. Pathways Analysis. To reveal the regulation mechanism of osteosarcoma, the related pathways of the factors in GSE12685 and GSE69470 were investigated with KEGG enrichment. The results showed that the DEGs in GSE12685 were related with the PI3K-AKT pathway, the Hippo pathway, the MAPK pathway, and microRNAs in cancer, P53 pathways, the Rap1 pathway, the TGF- $\beta$  pathway, and so on (Figure 4(a)). The targets of the screened miRNAs in GSE69470 were connected with pathways in cancer, the PI3K-AKT pathway, the MAPK pathway, and microRNAs in cancer, the Rap1 pathway, the P53 pathways, the TGF- $\beta$  pathway, and so on (Figure 4(b)). Moreover, it was also found that the common genes of the targets in GSE69470 and DEGs in GSE12685 were also involved in the PI3K/AKT pathway, the Hippo pathway, the MAPK pathway, and microRNAs in cancer, P53 pathways, the TGF- $\beta$ pathway, and the Rap1 pathway (Figure 4(c)).

3.5. Network Analysis. To reveal the molecular mechanism of osteosarcoma, the targets of the differentially expressed miRNAs in GSE69470, the DEGs in GSE12685, and their

common genes were analyzed with the STRING database, and then visualized by Cytoscape. The results showed that LEF1, RUNX2, CSF1R, PTPRD, VAV3, WIF1, BMP3, TFEC, FZD3, PTCH1, THBS1, CDKN1A, FBN1, TGFB1, and IGFBP3 had connections with miR-22-3p, miR-154-5p, miR-31-5p, miR-34a-5p, miR-485-3p, miR-93-5p, and miR-9-5p (Figure 5).

#### 4. Discussion

Although considerable research has focused on finding effective therapeutic strategies for osteosarcoma treatment in recent years, the prognosis of patients has been significantly improved [13]. In this study, the differentially expressed miRNAs in GSE69470 and the DEGs in GSE12685 were analyzed to reveal the pathological mechanism of osteosarcoma. miRNAs dysfunction has been widely confirmed as the critical reason for cancer development. For osteosarcoma, several reporters have indicated that miRNA disorders promote the malignant behaviors of the tumor cells [13]. In this study, a significant difference in the miRNA profiles of normal cell lines and tumor cell lines was also found in the datasets. Moreover, 834 DEGs were identified in GSE12685. Considering the difference of BM-MSCs and differentiated bone cells in miRNA profiles, this study identified 37 common differentially expressed miRNAs in osteosarcoma cells.



FIGURE 2: Heat maps of differentially expressed factors in GSE69470 and GSE12685l. (a) The differentially expressed factors of human osteosarcoma cells and differentiated bone cells in GSE69470. (b) The differentially expressed factors of human osteosarcoma cells and mesenchymal stem cells in GSE69470. (c) The DEGs of GSE12685l. (d) The common differentially expressed miRNAs in the matrix files are the osteosarcoma tumor cells and differential normal human osteoblast cells and osteosarcoma tumor cells and mesenchymal stem cells (MSCs).

miRNAs are characterized with targeting 3'-UTRs of mRNA to repress the translation process of the related proteins [14]. In this study, 43 downregulated DEGs and 91 upregulated DEGs in GSE12685 were related with 7 upregulated miRNAs and 24 downregulated miRNAs in GSE69470, respectively. The upregulated genes including LEF1, RUNX2, CSF1R, PTPRD, VAV3, WIF1, BMP3, TFEC, FZD3, and PTCH1 were found to be upregulated in osteosarcoma. Moreover, LEF1, RUNX2, CSF1R, VAV3, and FZD3 have been reported to take part in the development of cancer. Reduced LEF1 could enhance the sensitivity of breast cancer cells to resensitizing docetaxel, and LEF1 downregulation could also inhibit the growth of osteosarcoma cells [15]. RUNX2 has been identified as an oncogene, which could activate the PI3K/AKT pathways to promote the development of breast cancer, and RUNX2 has been identified as a biomarker for osteosarcoma

development. Low-expressed CSF1R could impede the growth and metastasis of osteosarcoma cells though inactivating ERK signaling [16, 17]. Downregulated VAV3 could inhibit the angiogenesis of osteosarcoma, and inhibited FZD3 can also block the invasion and malignant growth of osteosarcoma cells [18, 19]. In this study, genes including THBS1, CDKN1A, FBN1, TGFB1, and IGFBP3 were found to be downregulated in tumor cell lines. However, only CDKN1A and FBN1 function as the inhibitors in caner progression. Decreased CDKN1A has been found in multiple tumor cells, and several studies have shown that IGFBP3 could effectively inhibit the malignant behaviors of cancer cells. For osteosarcoma, CDKN1A upregulation could inhibit the proliferation and cell cycle of the cells [20]. Reduced FBN1 is related with the high invasiveness and metastasis of osteosarcoma, and increased FBN1 could also inhibit the epithelial-mesenchymal transition of

#### Journal of Healthcare Engineering



FIGURE 3: The modular functions of DEGs in the progression of osteosarcoma. (a) Module gene GO enrichment analysis of DEGs in GSE126851. (b) Module gene GO enrichment analysis of the targets of the differentially expressed miRNAs in GSE69470. (c) The common genes of the downregulated DEGs in GSE126851 and the upregulated differentially expressed miRNAs in GSE69470. (d) The common genes of the upregulated DEGs in GSE126851 and the downregulated differentially expressed miRNAs in GSE69470. (e) Module gene GO enrichment analysis of the common genes.



FIGURE 4: The modular functions of DEGs in the progression of osteosarcoma. (a) KEGG enrichment analysis of DEGs in GSE126851. (b) KEGG enrichment analysis of the targets of the differentially expressed miRNAs in GSE69470. (c) KEGG enrichment analysis of the common genes.



FIGURE 5: Continued.



FIGURE 5: Network analysis of the differentially expressed factors in GSE69470 and GSE126851. (a) The PPI network of DEGs in GSE126851. (b) The PPI network of the targets of downregulated differentially expressed miRNAs in GSE69470. (c) The PPI network of the targets of upregulated differentially expressed miRNAs in GSE69470. (d) The miRNA-mRNA network of upregulated differentially expressed miRNAs and the related targets. (e) The miRNA-mRNA network of downregulated differentially expressed miRNAs and the related targets. (f) The miRNA-mRNA network of hub nodes (miRNA: yellow and target: green).

osteosarcoma cells [21]. Thus, this study suggests that the hub nodes LEF1, RUNX2, CSF1R, VAV3, FZD3, CDKN1A, and FBN1 are key factors in the progression of osteosarcoma.

This study confirmed that the targets of the differentially expressed miRNAs were related with PI3K/AKT, AMPK, Wnt/ $\beta$ -catenin, TGF- $\beta$ , P53 pathways, and so on, and miR-22-3p, miR-154-5p, miR-31-5p, miR-34a-5p, miR-485-3p, miR-93-5p, and miR-9-5p were related with hub nodes. The study has indicated that miR-22-3p can inactivate the Wnt/  $\beta$ -catenin to inhibit the progression of via gastric cancer directly targeting BCL9 [22]. Increased miR-154-5p, miR-34a-5p, and miR-485-3p could also suppress the progression of osteosarcoma. [23]. The studies have indicated that both miR-34a-5p and LEF1 were related with the activity of Wnt/  $\beta$ -catenin pathways. Moreover, it has been verified that miR-34a-5p can directly target LEF1 to act as a cancer suppressor in esophageal squamous cells [24]. The miR-22-3p has been proved to suppress the activity of the PI3K/AKT pathways and thus inhibit the development of colorectal cancer [25]. CSF1R has also been found to boost the viability of T-cell lymphoma [26]. In this study, CSF1R was identified as a potential target of miR-22-3p, which suggests that the increased CSF1R induced by miR-22-3p downregulation is one of the reasons for osteosarcoma development. However, increased miR-31-5p has been found in osteosarcoma cells but has been proved to have inhibitor roles in osteosarcoma, which is inconsistent with the profile of GSE12685. Moreover, miR-93-5p may functions as an oncogene which takes part in the development of various tumors [27, 28]. The study showed that CDKN1A upregulation induced by miR-93-5p CDKN1A could block the TGF- $\beta$  and thus increase the therapeutic effects of chemotherapy on small cell lung cancer cells [29]. miRNA-9-5p has been found to upregulate and activate PI3K/AKT pathways in osteosarcoma [30].

Besides, miR-485-3p has been also proved to be involved in the cancer development via regulating TGF- $\beta$  pathways, and RUNX2 has been widely confirmed as the downstream factors of TGF- $\beta$ . In this study, RUNX2 was identified as the potential target of miR-485-3p.

#### **5. Conclusion**

In summary, this study suggests the miRNAs including miR-22-3p, miR-154-5p, miR-34a-5p, miR-485-3p, miR-93-5p, and miR-9-5p and the genes including LEF1, RUNX2, CSF1R, CDKN1A, and FBN1 are the key factors in the progression of osteosarcoma.

#### **Data Availability**

The data to support the findings of this study are available on reasonable request from the corresponding author.

## **Conflicts of Interest**

The authors have no conflicts of interest.

#### References

- Q. Huang, X. Liang, T. Ren et al., "The role of tumor-associated macrophages in osteosarcoma progression - therapeutic implications," *Cellular Oncology*, vol. 44, no. 3, pp. 525–539, 2021 Jun.
- [2] K.-H. Lu, E. W.-H. Lu, C.-W. Lin, J.-S. Yang, and S.-F. Yang, "New insights into molecular and cellular mechanisms of zoledronate in human osteosarcoma," *Pharmacology & Therapeutics*, vol. 214, Article ID 107611, 2020 Oct.
- [3] Y. Liu, N. Huang, S. Liao et al., "Current research progress in targeted anti-angiogenesis therapy for osteosarcoma," *Cell Proliferation*, vol. 54, no. 9, Article ID e13102, 2021 Sep.

- [4] J. Li, Y. Lu, G. Chen et al., "Cryoablation-aided joint retention surgery for epiphysis involvement in osteosarcoma compared with endoprosthetic replacement," *The Bone & Joint Journal*, vol. 103-B, no. 8, pp. 1421–1427, 2021 Aug.
- [5] M. Heng, A. Gupta, P. W. Chung et al., "The role of chemotherapy and radiotherapy in localized extraskeletal osteosarcoma," *European Journal of Cancer*, vol. 125, pp. 130–141, 2020 Jan.
- [6] R. Liu, Y. Hu, T. Liu, and Y. Wang, "Profiles of immune cell infiltration and immune-related genes in the tumor microenvironment of osteosarcoma cancer," *BMC Cancer*, vol. 21, no. 1, Article ID 1345, 2021 Dec 18.
- [7] M. A. Lopez-Ramirez, A. Reijerkerk, H. E. de Vries, and I. A. Romero, "Regulation of brain endothelial barrier function by microRNAs in health and neuroinflammation," *The FASEB Journal*, vol. 30, no. 8, pp. 2662–2672, 2016 Aug.
- [8] N. Bruno, J. M. ter Maaten, E. S. Ovchinnikova et al., "MicroRNAs relate to early worsening of renal function in patients with acute heart failure," *International Journal of Cardiology*, vol. 203, pp. 564–569, 2016 Jan 15.
- [9] Y. Wen, B. Li, M. He, S. Teng, Y. Sun, and G. Wang, "circHIPK3 promotes proliferation and migration and invasion via regulation of miR-637/HDAC4 signaling in osteosarcoma cells," *Oncology Reports*, vol. 45, no. 1, pp. 169–179, 2021 Jan.
- [10] Z. Jamali, M. Taheri-Anganeh, Z. Shabaninejad et al., "Autophagy regulation by microRNAs: novel insights into osteosarcoma therapy," *IUBMB Life*, vol. 72, no. 7, pp. 1306–1321, 2020 Jul.
- [11] Y. Sun, L. Wang, C. Li et al., "Construction of an integrated human osteosarcoma database, HOsDb, based on literature mining, microarray analysis, and database retrieval," *BMC Cancer*, vol. 20, no. 1, p. 390, 2020 May 6.
- [12] X. Zhou, Y. Fan, W. Ye, B. Jia, Y. Yang, and Y. Liu, "Identification of the novel target genes for osteosarcoma therapy based on comprehensive bioinformatic analysis," *DNA and Cell Biology*, vol. 39, no. 7, pp. 1172–1180, 2020 Jul.
- [13] L. Raimondi, A. De Luca, A. Gallo et al., "Osteosarcoma cellderived exosomes affect tumor microenvironment by specific packaging of microRNAs," *Carcinogenesis*, vol. 41, no. 5, pp. 666–677, 2020 Jul 10.
- [14] S. A. Lozano Calderón, C. Garbutt, J. Kim et al., "Clinical and molecular analysis of pathologic fracture-associated osteosarcoma: MicroRNA profile is different and correlates with prognosis," *Clinical Orthopaedics and Related Research*, vol. 477, no. 9, pp. 2114–2126, 2019 Sep.
- [15] M. Prieto-Vila, I. Shimomura, A. Kogure et al., "Quercetin inhibits Lef1 and resensitizes docetaxel-resistant breast cancer cells," *Molecules*, vol. 25, no. 11, p. 2576, 2020 Jun 1.
- [16] H. Lu, T. Jiang, K. Ren et al., "RUNX2 plays an oncogenic role in esophageal carcinoma by activating the PI3K/AKT and ERK signaling pathways," *Cellular Physiology and Biochemistry*, vol. 49, no. 1, pp. 217–225, 2018.
- [17] B. A. Smeester, N. J. Slipek, E. J. Pomeroy et al., "PLX3397 treatment inhibits constitutive CSF1R-induced oncogenic ERK signaling, reduces tumor growth, and metastatic burden in osteosarcoma," *Bone*, vol. 136, Article ID 115353, 2020 Jul.
- [18] Y. Xiao, C. Li, H. Wang, and Y. Liu, "LINC00265 targets miR-382-5p to regulate SAT1, VAV3 and angiogenesis in osteosarcoma," *Aging*, vol. 12, no. 20, pp. 20212–20225, 2020 Aug 14.
- [19] S. Zhu, Y. Liu, X. Wang, J. Wang, and G. Xi, "IncRNA SNHG10 promotes the proliferation and invasion of

osteosarcoma via wnt/ $\beta$ -catenin signaling," *Molecular Therapy - Nucleic Acids*, vol. 22, pp. 957–970, 2020 Oct 15.

- [20] X. Zhao, Y. Yang, J. Xu, Y. Luo, Y. Xin, and Y. Wang, "Downregulation of microRNA-95-3p suppresses cell growth of osteosarcoma via CDKN1A/p21 expression," *Oncology Reports*, vol. 39, no. 1, pp. 289–297, 2018 Jan.
- [21] W. Liu, P. Liu, H. Gao, X. Wang, and M. Yan, "Long noncoding RNA PGM5-AS1 promotes epithelial-mesenchymal transition, invasion and metastasis of osteosarcoma cells by impairing miR-140-5p-mediated FBN1 inhibition," *Molecular Oncology*, vol. 14, no. 10, pp. 2660–2677, 2020 Oct.
- [22] B. Guan, J. Ma, Z. Yang, F. Yu, and J. Yao, "LncRNA NCK1-AS1 exerts oncogenic property in gastric cancer by targeting the miR-22-3p/BCL9 axis to activate the Wnt/β-catenin signaling," *Environmental Toxicology*, vol. 36, no. 8, pp. 1640–1653, 2021.
- [23] Y. Xue, Y. Guo, N. Liu et al., "Upregulation of miRNA-154-5p prevents the tumorigenesis of osteosarcoma," *Biomedicine & Pharmacotherapy*, vol. 124, Article ID 109884, 2020 Apr.
- [24] X. Wang, Y. Zhao, Q. Lu et al., "MiR-34a-5p inhibits proliferation, migration, invasion and epithelial-mesenchymal transition in esophageal squamous cell carcinoma by targeting LEF1 and inactivation of the hippo-YAP1/TAZ signaling pathway," *Journal of Cancer*, vol. 11, no. 10, pp. 3072–3081, 2020 Mar 4.
- [25] Y. Wang and C. Lin, "Exosomes miR-22-3p derived from mesenchymal stem cells suppress colorectal cancer cell proliferation and invasion by regulating RAP2B and PI3K/AKT pathway," *Journal of oncology*, vol. 2021, Article ID 3874478, 2021 Jun 21.
- [26] C. Murga-Zamalloa, D. C. M. Rolland, A. Polk et al., "Colonystimulating factor 1 receptor (CSF1R) activates AKT/mTOR signaling and promotes T-cell lymphoma viability," *Clinical Cancer Research*, vol. 26, no. 3, pp. 690–703, 2020.
- [27] Y. Yang, B. Jia, X. Zhao, Y. Wang, and W. Ye, "miR-93-5p may be an important oncogene in prostate cancer by bioinformatics analysis," *Journal of Cellular Biochemistry*, vol. 120, no. 6, pp. 10463–10483, 2019.
- [28] J. Li, Z.-P. Chu, H. Han et al., "Suppression of miR-93-5p inhibits high-risk HPV-positive cervical cancer progression via targeting of BTG3," *Human Cell*, vol. 32, no. 2, pp. 160–171, 2019 Apr.
- [29] W. Huang, Y. Yang, J. Wu et al., "Circular RNA cESRP1 sensitises small cell lung cancer cells to chemotherapy by sponging miR-93-5p to inhibit TGF-β signalling," *Cell Death* & Differentiation, vol. 27, no. 5, pp. 1709–1727, 2020 May.
- [30] P. Yao, Y. Ni, and C. Liu, "Long non-coding RNA 691 regulated PTEN/PI3K/AKT signaling pathway in osteosarcoma through miRNA-9-5p," *OncoTargets and Therapy*, vol. 13, pp. 4597–4606, 2020.