

Review Article

Monoclonal Antibodies for Non-Hodgkin's Lymphoma: State of the Art and Perspectives

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Monoclonal antibodies have been the most successful therapeutics ever brought to cancer treatment by immune technologies. The use of monoclonal antibodies in B-cell Non-Hodgkin's lymphomas (NHL) represents the greatest example of these advances, as the introduction of the anti-CD20 antibody rituximab has had a dramatic impact on how we treat this group of diseases today. Despite this success, several questions about how to optimize the use of monoclonal antibodies in NHL remain open. The best administration schedules, as well as the optimal duration of rituximab treatment, have yet to be determined. A deeper knowledge of the mechanisms underlying resistance to rituximab is also necessary in order to improve the activity of this and of similar therapeutics. Finally, new antibodies and biological agents are entering the scene and their advantages over rituximab will have to be assessed. We will discuss these issues and present an overview of the most significant clinical studies with monoclonal antibodies for NHL treatment carried out to date.

1. Introduction

In 1975, Kohler and Milstein heralded a new era in antibody research with their discovery of hybridoma technology [1]. Mouse hybridomas were the first reliable source of monoclonal antibodies. Subsequently, the introduction of recombinant technologies, transgenic animals, and phage display technology has modernized selection, humanization and production of therapeutic antibodies. The use of mAbs in cancer treatment stems from the idea that these, because of their intrinsic specificity, could be used to selectively target cancer cells based on the expression of one or more antigens. In such approaches, antibodies could be used alone or be conjugated to toxins, radioactive moieties, or enzymes in order to achieve toxic concentrations of these agents in the cancerous tissues while sparing healthy organs.

Indeed, since their initial discovery, more than 20 mAbs have been approved by the US Food and Drug Administration (FDA) for the treatment of several conditions, including several types of cancers. This success has opened new therapeutic perspectives and prompted research efforts aimed to improve their activity, select for those patients who

will most benefit from them, and, potentially, to expand their therapeutic indications. The anti-CD20 mAb rituximab is one of the best examples of this new class of therapeutics, since it has rapidly become a key part of the pharmacological schemes used to treat Non-Hodgkin's lymphomas (NHLs). Moreover, due to its capacity to eliminate B lymphocytes, it has recently been applied in immune-mediated disorders [2].

Here, we will focus on the use of rituximab in the treatment of NHL, on the clinical issues associated with this therapeutic, and on the most recent advances in the field of lymphoma immunotherapy.

2. Tumor Antigens in NHL

When designing a therapeutic approach for NHL, cancer immunologists face the issue of selecting the best target antigen. Tumor antigens are traditionally divided in tumor-specific antigens (proteins that are uniquely expressed by cancer cells) and tumor-associated antigens (molecules that are expressed by cancer cells, although their expression is also found on normal cells) [3]. Ideally, an immune response

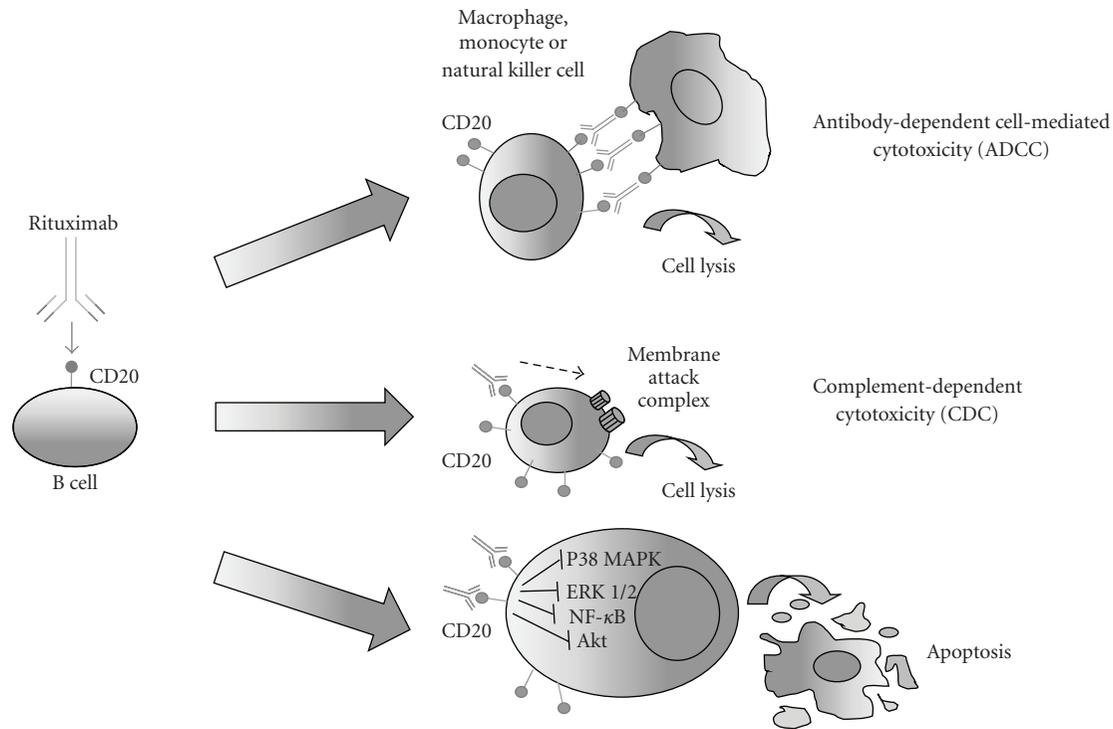


FIGURE 1: Schematic representation of the putative mechanisms mediating rituximab's anticancer activity in NHL cells. The anti-CD20 monoclonal antibody rituximab has several mechanisms of action, including antibody-dependent cellular cytotoxicity (ADCC), which involves recruitment of effector cells, mediated by Fc γ receptors; complement-dependent cytotoxicity (CDC); apoptosis induction.

against tumor antigens should destroy tumor cells without damaging normal cells. Thus, cancer-specific antigens would be the first choice. Unfortunately, true cancer-specific antigens, such as new proteins resulting from fusion oncogenes, are not frequent in NHL. Another important issue is to ensure that the chosen antigen does not mutate in a way that allows cancer cells to avoid destruction by the immune system [3].

The cell surface protein CD20 is a 33-kDa protein expressed by mature B cells and most malignant B cells, but not by pre-B cells or differentiated plasma cells [4–8]. In vitro studies have revealed that CD20 acts as a calcium ion channel [9, 10], and may also activate intracellular signaling through its ability to associate with the B-cell receptor (BCR) [11]. Interestingly, CD20's ability to induce cytosolic Ca²⁺ flux appears to be BCR dependent. Rituximab (Rituxan, Mabthera), is the first anti-CD20 monoclonal antibody approved by the Food and Drug Administration (FDA) (on November 26, 1994) for the treatment of relapsed or refractory, CD20+ follicular lymphoma (FL). It is a chimeric anti-CD20 antibody derived from the mouse mAb 2B8, targeting CD20 antigens, following replacement of the heavy and light chain constant regions with the corresponding regions of a human IgG1 mAb. Importantly, rituximab depletes both malignant and normal CD20⁺ B lymphocytes [4, 12, 13].

3. Rituximab's Mode of Action in Lymphoma Cells

Although the exact *in vivo* mechanisms of action for rituximab are not fully understood, the mechanisms of B-cell killing by this mAb have been exhaustively analyzed [14].

Briefly, the major mechanism of rituximab-induced B-cell depletion involves antibody-dependent cell-mediated cytotoxicity (ADCC) and complement dependent cytotoxicity (CDC) [15]. Additionally rituximab was reported to directly induce apoptosis, inhibit B-cell proliferation, and to enhance the cytotoxic activity of chemotherapeutic agents [16] (Figure 1).

Rituximab-induced CDC is triggered upon rituximab binding to B cells with consequent initiation of the complement cascade starting from C₁ activation. This mechanism causes osmotic lysis of neoplastic B cells [13, 14]. ADCC is triggered by the interaction between rituximab and the Fc receptor of natural killer (NK) cells [13, 14]. Once activated, NK cells release small proteins, including perforin and granzymes, which in turn form pores in the malignant B-cell membrane, and thus induce apoptosis or osmotic cell lysis. Finally, recent data demonstrate the novel role of rituximab as a signal-inducing antibody, and as a chemosensitizing agent, capable of negative regulation of major survival pathways [16].

Besides these mechanisms, rituximab's activity appears to be linked, at least in part, to its signaling via CD20. In this field, studies in B-NHL cell lines revealed several mechanisms involved in rituximab-mediated chemo/immunosensitization. Rituximab was shown to inhibit the p38 mitogen-activated protein kinase, nuclear factor- κ B (NF- κ B), extracellular signal-regulated kinase 1/2 (ERK 1/2), and Akt antiapoptotic survival pathways [17]. All of these effects result in upregulation of PTEN and of Raf kinase inhibitor protein (RKIP) [18], in the downregulation of antiapoptotic gene products, such as Bcl-2, Bcl-xL and Mcl-1, and, as a result, in chemo/immunosensitization [19]. In addition, treatment with rituximab inhibits the overexpressed transcription repressor Yin Yang 1 (YY1) [20]. YY1 downregulates Fas and DR5 expression and its inhibition leads to sensitization to Fas ligand and tumor necrosis factor-related apoptosis-inducing ligand- (TRAIL-) induced cell death [21].

Interestingly, recent studies also show that rituximab strongly affects BCR signaling [22]. Pretreatment of lymphoma cells or healthy B-cells with rituximab results in a time-dependent inhibition of the BCR-signaling cascade involving Lyn, Syk, PLC γ 2, Akt, ERK, and calcium flux. Such inhibitory effects by rituximab are associated with a decrease in raft-associated cholesterol, inhibition of BCR relocalization to lipid rafts, and BCR downregulation. Since BCR signaling appears to be crucial for healthy and malignant B cell survival and expansion [23–25], this mode of action of rituximab could actually have an important role in mediating its anticancer activity.

The relative importance of each mechanism of action of rituximab is likely to vary with the type of tumor and the type of treatments that are administered together with this mAb. CDC and ADCC appear to be important to target leukemia/lymphoma cells circulating in the bloodstream [26]. Conversely, an immunological mechanism of action seems to be less important in the presence of nodal and extranodal involvement.

4. Rituximab's Applications in Hematological Malignancies

We will discuss here the current therapeutic applications of rituximab in indolent NHL, diffuse large B cell non-Hodgkin lymphoma (DLBCL), and in B-cell chronic lymphocytic leukemia (B-CLL). Although trials may have had endpoint definitions that are not always identical, almost all defined complete response (CR) as the complete disappearance of the symptoms and signs of lymphoma (including bone marrow clearing for >28 days), and partial response (PR) as a >50% decrease in the size or number of the lymphomas lesions, without any evidence of progressive disease for >28 days. CR and PR together represent the objective response (OR) rate [4, 27].

4.1. Follicular and Low Grade Lymphoma. Until the early 90's, the first-line therapy in symptomatic low-grade NHL was chlorambucil and prednisone [42]. Subsequently, several

randomized trials showed the efficacy of rituximab in combination with other chemotherapeutic agents such as fludarabine (R-F), fludarabine, and cyclophosphamide (R-FC), fludarabine, cyclophosphamide and mitoxantrone (FCM-R), cyclophosphamide, vincristine, and prednisone (R-CVP), CVP plus mitoxantrone (R-CNOP), fludarabine, dexamethasone, and mitoxantrone (R-FND) as well as CHOP (R-CHOP) [28, 43–45] (Table 1). The clinical response rates of rituximab-containing regimens were encouraging, with an OR rates consistently around 95% and with a CR and PR rates ranging from 45% to 100%, and from 0% to 52%, respectively.

Importantly, clinical data on the benefit of rituximab combined with chemotherapy has also become available in patients with relapsed or refractory indolent B-cell NHL. Also here, the results are very encouraging, with OR of 81% for R-CVP, 97% for R-FC, 88% for R-CHOP, and 95% for FCM-R respectively [4, 46].

Finally, the efficacy of rituximab monotherapy in patients with relapsed or refractory CD20-positive low-grade or follicular lymphoma was examined in noncomparative multicentre trials [33–35, 47–55]. The overall response rates were 38%–48% after a 4-week therapy with rituximab, and 57% after 8 weeks of rituximab administration. CR rates ranging between 3 and 17% were recorded in these studies.

Remarkably, studies show that, in FL, sequential administration of standard chemotherapy followed by rituximab induces molecular clearance (as detected by PCR for the Bcl-2/IgH rearrangement) in more than 70% of the patients [42, 43, 56]. The actual clinical impact of achieving a molecular response in FL still has to be determined, since long-term remissions have been reported also in patients with persistently detectable Bcl-2/IgH rearrangement [57]. Moreover this rearrangement may occasionally be found in healthy peripheral blood lymphocytes [58]. In fact, a recent study by van Oers and coworkers suggests that BCL-2/IgH polymerase chain reaction status at the end of induction treatment would not be predictive for progression-free survival in relapsed/resistant FL [59]. Nonetheless, the above-mentioned studies support the efficacy of rituximab in FL, and indicate its potential for treating minimal residual disease in this type of disorder.

In summary, the current guidelines for the treatment of FL recommend that rituximab is administered in combination with standard chemotherapy in previously untreated stage III–IV FL, and at first relapse (at a dosage of 375 mg/m² on day 1 of each chemotherapy cycle, for up to eight doses). Rituximab is recommended as a monotherapy for stage III–IV chemoresistant FL, or at second (or subsequent) relapse after chemotherapy (375 mg/m² once weekly for four doses) (http://www.ema.europa.eu/docs/en_GB/document_library/Summary_of_opinion/human/000165/WC500097025.pdf).

4.2. DLBCL. After the disappointing results obtained with third-generation chemotherapy regimens in the United States, the CHOP regimen was reverted to as the standard of

TABLE 1: Principal clinical trials of chemotherapy plus Rituximab versus chemotherapy alone in NHL.

Lymphoma Subtype	Treatment	Patients (no.)	% Overall response rate (<i>P</i> value)*	Median Follow-up (mo.)	Reference
Follicular	CVP versus R-CVP	321	57 versus 81 (<.001)	53	Marcus et al. [28]
Follicular	CHOP versus R-CHOP	428	90 versus 96 (=,011)	18	Hiddemann et al. [29]
Follicular	CHOP versus R-CHOP	465	72.3 versus 85.1 (<.001)	39,4	van Oers et al. [30]
Follicular	FCM versus R-FCM	176	71 versus 95 (=,01)	26	Forstpointner et al. [31]
Follicular	MCP versus R-MCP	201	75 versus 92 (=,009)	47	Herold et al. [32]
relapsed/refractory low grade	R	37	46	13,4	Maloney et al. [33]
relapsed/refractory low grade	R	30	47	19	Feuring-Buske et al. [34]
relapsed/refractory low grade	R	166	48	19,5	McLaughlin et al. [35]
DLBCL	CHOP versus R-CHOP	399	63 versus 76 (=,005)*	24	Coiffier et al. [36]
DLBCL	CHOP versus R-CHOP	824	84 versus 93 (=,0001)**	34	Pfreundschuh et al. [37]
DLBCL	CHOP versus R-CHOP	632	57 versus 67 (=,05)**	42	Habermann et al. [38]
DLBCL	CHOP versus R-CHOP	122	75 versus 94 (=,0054)	18	Lenz et al. [39]
B-CLL	FC versus R-FC	552	58 versus 69.9 (=,0034)	25	Robak et al. [40]
B-CLL	FC versus R-FC	817	82.5 versus 87.2 (=,012)**	37,7	CLL8- German CLL Study Group*** [41]

* CR, CR-unconfirmed, partial response.

** CR rate.

*** 3-year OS.

therapy. The efforts to introduce rituximab in the treatment of this aggressive hematological disease led to two essential clinical trials: the Mabthera International trial (MinT) [37] and the Groupe d'Etude des lymphomes de l'Adulte study (GELA) [36]. The first one involved young, the latter elderly, DLBCL patients. In the multicenter study conducted by Coiffier and colleagues, therapy using rituximab combined with standard CHOP chemotherapy demonstrated a higher efficacy than CHOP alone, in terms of both event-free survival at 2 years (57% versus 38%, $P < .001$), overall survival at 2 years (70% versus 57%, $P < .01$), and CR rate (76% versus 63%, $P < .01$). Likewise, the MinT study showed an increased OS of the combined rituximab-adding regimen, compared to standard therapy, from 84% to 93%. These results led to FDA approval of rituximab in combination with CHOP chemotherapy for previously untreated patients with DLBCL. Whether or not all patients need rituximab has been questioned. Studies from France and the American National Cancer Institute suggested that the benefit of rituximab would be observed in patients with tumors overexpressing Bcl-2. On the other hand, a recent

report from the French group shows benefit in both Bcl-2-positive and Bcl-2-negative lymphomas using the method of competing risks [27, 60, 61]. Therefore, the question of whether molecular features should or will direct treatment decisions remains unanswered.

Finally, for recurrent DLBCL, the standard of care is salvage chemotherapy followed by high-dose chemotherapy with stem cell transplantation. Also in this setting, rituximab proved to be effective and has been incorporated into salvage chemotherapy regimens, since it may improve the overall response rate with ICE (ifosfamide, carboplatin, and etoposide) and DHAP (dexamethasone, high-dose cytarabine, and cisplatin) [62].

In summary rituximab is approved for previously untreated DLBCL patients in combination with CHOP chemotherapy and with salvage chemotherapy regimens in relapsed/refractory patients. The recommended rituximab dosage is 375 mg/m² on day 1 of each chemotherapy cycle, for up to eight doses (http://www.ema.europa.eu/docs/en_GB/document_library/Summary_of_opinion/human/000165/WC500097025.pdf).

4.3. Rituximab and Autologous Stem Cells Transplantation for Advanced Stage DLBCL. Young high-risk patients with DLBCL achieving a complete remission after a complete course of chemotherapy are likely to benefit from autologous stem cell transplantation (ASCT) [27, 63]. Several studies are assessing the role of rituximab as part of high-dose regimens (HDT) pre-ASCT in DLBCL because of its effectiveness, limited toxicity, and its ability to deplete B cells. In this field, in a 2-year study by Khouri and colleagues evaluating the efficacy and safety of high-dose rituximab in combination with high-dose BEAM and ASCT, the OS was 80% for the study group compared to 53% for the control group [64]. Superior survival rates have also been reported for patients who become PCR negative for BCL2/JH rearrangements in peripheral blood or bone marrow compared with those who remain positive.

4.4. Rituximab Maintenance Therapy for FL and DLBCL. Despite the fact that rituximab used in combination with chemotherapy has been shown to prolong the survival of patients with NHL, residual lymphoma cells (which then become responsible for disease relapses) frequently remain [65]. As a matter of fact, NHL relapses continue to be an important clinical issue. Therefore, several randomized trials have been conducted in order to analyze the benefit of rituximab maintenance treatment in NHL [66–68]. The studies that were done for FL adopted different schemes for induction (rituximab 375 mg/m² weekly × 4 in Ghielmini et al. and in Hainsworth et al.; CHOP or R-CHOP in van Oers et al.; fludarabine, cyclophosphamide, and mitoxantrone with or without rituximab in Forstpointner et al.) as well as for the maintenance treatment (375 mg/m² intravenously weekly for 4 weeks at six-month intervals in Hainsworth et al.; 375 mg/m² intravenously weekly for 4 weeks for Ghielmini et al.; 375 mg/m² rituximab intravenously once every 3 months in van Oers et al.; 2 further courses of 4-times-weekly doses of rituximab after 3 and 9 months in Forstpointner et al.). However, overall, they unequivocally show that rituximab maintenance increases event-free survival (EFS) and duration of response in indolent NHL. In Ghielmini et al., at a median followup of 35 months, the median EFS was 12 months in the no-maintenance group versus 23 months in the prolonged treatment arm ($P = .02$) [69]. The authors reported that the difference was particularly notable in chemotherapy-naïve patients (19 versus 36 months; $P = .009$) and in patients responding to induction treatment (16 versus 36 months; $P = .004$). In the study by van Oers et al., rituximab maintenance significantly improved EFS compared with observation (median, 3.7 years versus 1.3 years; $P < .00$), both after CHOP induction ($P < .001$) and R-CHOP ($P = .003$) [30]. The 5-year overall survival (OS) was 74% in the rituximab maintenance arm, and it was 64% in the observation arm ($P = .07$). Finally, also in the trial by Forstpointner and colleagues, response duration was significantly prolonged by rituximab maintenance, with the median not being reached in this evaluation versus an estimated median of 16 months in the observation group ($P = .001$) [31]. This beneficial effect was

also observed when analyzing FL ($P = .035$) and mantle cell lymphoma ($P = .049$) separately.

Unlike in indolent NHL, rituximab maintenance therapy in DLBCL has failed to demonstrate benefit in the published clinical trials [38].

In conclusion, the current guidelines recommend the use of rituximab as a maintenance therapy only in relapsed or refractory follicular lymphoma responding to induction therapy with chemotherapy with or without rituximab. The recommended dosage of rituximab is 375 mg/m² once every 3 months until disease progression or for a maximum of 2 years (http://www.ema.europa.eu/docs/en_GB/document_library/Summary_of_opinion/human/000165/WC500097025.pdf).

4.5. B-cell Chronic Lymphocytic Leukemia (B-CLL). B-CLL is a heterogeneous disorder with a variable course (i.e., following diagnosis, survival ranges from months to decades) and risk factors such as age and performance status should be considered when selecting the most appropriate treatment option [70].

Rituximab monotherapy is generally not associated with sustained responses in B-CLL, possibly reflecting altered rituximab pharmacokinetics in patients with B-CLL [40, 70–73]. However, studies show that the addition of rituximab to fludarabine plus cyclophosphamide (FC) does improve clinical outcomes in B-CLL patients. The first study, known as CLL8, was conducted by the German CLL Study Group on 817 previously untreated B-CLL patients (ClinicalTrials.gov number, NCT00281918). The second trial, known as REACH, enrolled 552 patients with relapsed or refractory B-CLL following prior systemic therapy [40]. Both studies showed a benefit in terms of OS rates in the R-FC arm versus FC arm (86% versus 73% in the CLL8 trial and 54% versus 45% in the REACH). In addition, the benefit of adding rituximab to chemotherapy in B-CLL was shown by several other trials [40, 74–77].

Interestingly, since rituximab plus FC represents the standard treatment for B-CLL, clinical studies compared the conventional regimen to rituximab plus low-dose FC (i.e., FCR-Lite) or to sequential FC and rituximab [49], since these alternative regimens are expected to be associated with less grade 3 or 4 neutropenia than the conventional R-FC regimen [5, 50].

The current international guidelines recommend that chemoimmunotherapy regimens with R-FC are preferred as the first-line treatment for advanced CLL (stage II–IV) in patients without del(17p) who are aged <70 years or aged >70 years without significant comorbidities. Among patients with relapsed or refractory disease, those with a long response (i.e., >3 years) can be retreated with one of the first-line treatment options. Various chemoimmunotherapy options are suggested for patients with a short response (i.e., <2 years) (e.g., rituximab may be administered in combination with FC or with CHOP). The recommended dosage of rituximab is 375 mg/m² the day before starting chemotherapy, followed by 500 mg/m² on day 1 of cycles 2–6 (National Comprehensive Cancer Network. NCCN

clinical practice guidelines in oncology: non-Hodgkin's lymphoma).

5. Tolerability

Adverse events were reported in 84% of patients, receiving rituximab, during therapy or within the first 30 days following treatment [4, 35]. However, more than 95% of these events were described as mild to moderate in severity, of brief duration, and observed during the first infusion. The most common adverse effects were infusion-related reactions and lymphopenia. Ten percent of the patients reported severe fever, chills, infections, or other adverse effects. Serious adverse effects included severe infusion-related reactions, tumor lysis syndrome, mucocutaneous reactions, hypersensitivity reactions, cardiac arrhythmias, angina, and renal failure [4, 51].

These adverse events were less common during the subsequent rituximab administrations. One possible hematological adverse event is the reduction in peripheral B-lymphocyte counts, which can last for up to 6 months with a recovery period of 9 to 12 months [4, 35]. Nevertheless, the risk of serious opportunistic infections appears to be much lower than that reported with conventional therapy [4]. Interestingly, Bedognetti and coworkers have recently evaluated the impact of rituximab on the effectiveness of an anti-flu vaccine in patients who had previously been treated with this mAb [52]. Due to the fact that disease status might affect immune response, only NHL patients without evidence of disease, who had completed rituximab no less than 6 months before the accrual, were selected for this evaluation. The study showed that patients who had previously received rituximab had a significantly lower seroconversion rate in response to the vaccine. Remarkably, while peripheral CD27-naïve B cells were present, Bedognetti et al. found a profound depletion in CD27+B memory cells, which may well explain the defective induction of anti-flu immunity. Thus, concerns remain that patients who have been treated with the anti-CD20 mAb may be at risk for infections and that they may need careful monitoring.

6. Improving Rituximab Efficacy and Overcoming Resistance

Despite the expression of CD20 on their lymphoma cells, some patients exhibit primary resistance and do not respond well to this targeted antibody therapy. Moreover, an initially responsive lymphoma can subsequently become resistant to rituximab (secondary/acquired resistance). Several mechanisms have been reported that have the potential to contribute to reductions in rituximab efficacy. The identification of such mechanisms has allowed for the proposal of strategies to overcome these issues, and thus achieve better *in vivo* activity. Some of these mechanisms have been reviewed elsewhere [53]. Here, we will summarize some of the most recent and promising observations, and the related suggestions for therapeutic interventions.

(i) *Interfering with CD20 Downregulation/Shaving*. Initial *in vitro* observations suggested that CD20 would not be downregulated in the presence of anti-CD20 antibodies. Namely, the anti-CD20/CD20 complex was found to remain at the cell surface long enough to ensure cell killing by specific mechanisms. However, these observations may not be reproduced in *in vivo* settings. A recent report by Beers et al. showed that rituximab is able to induce CD20 internalization in a B-CLL mouse model [54]. Interestingly, these authors demonstrated that the degree of CD20 down-modulation correlates inversely with some types of NHL's susceptibility to rituximab. Namely, CLL and mantle cell lymphoma showed greater down-modulation of CD20 in response to rituximab than FL and DLBCL did, and were less responsive to treatment. Previous reports by Beum et al. described a "shaving reaction" in which mAb-CD20 complexes were "shaved" off CLL cells, by phagocytes, as the malignant cells circulated [78]. Whether the observed reduction in CD20 levels actually reflects shaving, or rather antigen masking by rituximab, remains unclear [79, 80]. Downregulation of CD20 access, irrespective of the underlying cause, appears to be an important mechanism affecting rituximab efficacy, as antigen loss by malignant cells will prevent rituximab activity. New anti-CD20 mAbs (tositumomab-like) may be able to induce considerably less CD20 down-modulation than rituximab, and thus possibly be more effective (see below) [54]. It is also of interest that CD20 expression on lymphoma cells can be increased with HDAC inhibitors, such as valproic acid and romidepsin [81]. These were shown to transactivate the CD20 gene through promoter hyperacetylation and Sp1 recruitment. In line with these premises, HDAC inhibitors potentiated the activity of rituximab both *in vitro* and *in vivo* in murine lymphoma models.

(ii) *Targeting CD20 Transcript Variants Associated with Resistance*. Henry and coworkers have recently identified an alternative CD20 transcript variant (Δ CD20) associated with resistance to rituximab [82]. This novel, alternatively spliced CD20 variant encodes for a truncated 130 amino acid protein lacking large parts of the four transmembrane domains, suggesting that Δ CD20 is a nonanchored membrane protein. Δ CD20 expression was detected in B-cell leukemias, B-cell lymphomas, and activated B cells, but not in healthy resting B cells. Finally, the authors went on to show that Δ CD20 is associated with resistance to rituximab, although the mechanism whereby this CD20 splice variant impairs the benefit of rituximab remains to be determined. The authors suggest that, given its selective expression in malignant (and activated) B-cells, Δ CD20 could become a therapeutic target, for instance for the development of antilymphoma vaccines. Whether this approach will prove effective remains to be assessed.

- (iii) *Preventing NK Cell-Mediated ADCC Exhaustion.* NK cell-mediated ADCC can be exhausted. Studies showed that NK cells can engage and kill 3-4 target cells in 16 hours. Thereafter, cells become exhausted, possibly due to a reduction in the available levels of perforin and granzyme B [83]. Indeed, incubation of NK cells with rituximab-coated target cells leads to CD16 (FcγRIIIa) downregulation and to upregulation of CD107a, a marker for degranulation and exhaustion [84, 85]. Finally, NK cell-mediated target cell killing was shown to become less efficient in the presence of high burdens of rituximab-opsonized lymphoma cells [86]. Remarkably, IL-2 treatment can restore NK cell-mediated ADCC. In line with this concept, Berdeja and coworkers found that systemic interleukin-2 and adoptive transfer of lymphokine-activated killer cells improves antibody-dependent cellular cytotoxicity in patients with relapsed B-cell lymphoma treated with rituximab [86]. In this context, recent studies showed that also complement components, such as C3b, can inhibit NK-cell mediated killing of mAb-opsonized lymphoma cells [55]. Importantly, C3 depletion by cobra venom factor, or the related drug (HC3-1496), appears to effectively overcome this mechanism and improve the activity of rituximab in lymphoma-bearing mice. Thus, overall, strategies aimed to improve NK cell activity could help enhance the efficacy of rituximab and should therefore be further investigated.
- (iv) *Enhancing CDC.* Studies showed that also complement can be depleted upon rituximab infusion in B-CLL patients [79]. Kennedy et al. found that fresh frozen plasma would then restore rituximab efficacy. More studies on this approach should be performed in order to confirm its viability. Another approach to enhance rituximab-induced CDC has recently been proposed by Wang and colleagues [87]. These authors observed that many tumors, including lymphomas, upregulate the expression of CD46, an inhibitory complement receptor. As a means to overcome this issue, they identify a recombinant adenovirus type 35 fiber knob protein (Ad35K⁺⁺) which, when incubated with lymphoma cells, leads to CD46 downregulation and cooperates with rituximab in inducing CDC. In xenograft models with human lymphoma cells, preinjection of Ad35K⁺⁺ dramatically increased the efficacy of rituximab, suggesting that the Ad35K⁺⁺-based approach has potential implications in mAb therapy of NHL. Finally, Sato and colleagues have recently reported the identification of a novel CDC-enhancing variant of rituximab (113F) [88]. Compared to rituximab, 113F appeared to mediate highly enhanced CDC against primary CD20-expressing lymphoma cells *in vitro*. Moreover, these authors were able to establish a human tumor-bearing NOD/Shi-scid-IL-2Rγ(null) mouse model, in which human complement functions as the CDC mediator. Using this model, the authors demonstrated that 113F exerted significantly more potent antitumor effects than rituximab.
- (v) *Improving Phagocytosis Through CD47 Blockade.* Chao and colleagues have recently shown that multiple B-cell NHL subtypes, including DLBCL, FL, and B-CLL, exhibit increased levels of CD47, a transmembrane protein which activates SIRP1a in phagocytic cells [89]. This results in initiation of a signal transduction cascade which leads to phagocytosis inhibition. These authors demonstrate that CD47 overexpression correlates with worse prognosis. Blocking anti-CD47 antibodies promote phagocytosis of NHL cells and cooperate with rituximab both *in vitro* and *in vivo* in murine NHL xenotransplant models. Again, whether this approach will prove useful in humans remains to be assessed.
- (vi) *Topical IFN-α Delivery.* Finally, Xuan and colleagues have proposed an approach to target IFN-α molecules to lymphoma sites by constructing a fusion protein consisting of IFN-α and an anti-CD20 mAb [90]. IFN-α has potent immunostimulatory properties and antiproliferative effects in some B-cell NHLs, but its systemic administration is frequently associated to significant toxicity. The CD20-IFN-α fusion proteins showed efficient anticancer activity against an aggressive rituximab-resistant human CD20+ murine lymphoma (38C13-huCD20) and a human B-cell lymphoma (Daudi). Further experimentation with this administration method is warranted to assess its applicability in patients.
- (vii) *Rituximab Mutants with Proapoptotic Activity.* In order to improve rituximab anticancer activity, Li and colleagues modulated the binding property of this mAb by introducing several point mutations in its complementarity-determining regions [91]. These authors found that the CDC potency of such CD20 mAbs was independent of the off-rate. However, they were able to identify a rituximab triple mutant (H57DE/H102YK/L93NR) with an extremely potent apoptosis-inducing activity. This triple mutant efficiently initiated both caspase-dependent and-independent apoptosis, and exhibited potent *in vivo* activity even in a rituximab-resistant lymphoma model. These modified versions of rituximab hold promise as new therapeutic agents for B-cell lymphomas, although their efficacy in patients still has to be assessed.
- (viii) *Combining Rituximab with Other mAbs.* Rituximab activity in NHL as a single agent is limited, especially when administered to pretreated patients. However, combining rituximab with chemotherapy does achieve significantly better outcomes than chemotherapy alone. In addition, strategies to use two mAbs have also been proposed. Combinations such as anti-CD20 plus anti-CD22, anti-CD20 plus

anti-HLA-DR, anti-CD20 plus anti-TRAIL-R1, anti-CD20 plus anti-CD80 have been evaluated preclinically and/or clinically, showing enhanced antitumor activity both *in vitro* and *in vivo* [92–94]. An interesting approach to achieve the benefit of multiple targeting in NHL consists of the generation of multivalent antibodies using the so-named Dock-and-Lock (DNL) method, which enables site-specific self-assembly of two modular components with each other, resulting in a covalent structure with retained bioactivity [95]. Using this approach, Rossi and colleagues generated bispecific anti-CD20/CD22 hexavalent antibodies with promising antilymphoma activity *in vitro* and *in vivo* [96, 97]. Interestingly, in a recent study, these authors were able to correlate the strong direct cytotoxicity of the anti-CD20/CD22 hexavalent antibodies, compared to their bivalent parental antibodies, with their increased ability to upregulate PTEN, phospho-p38, and cyclin-dependent kinase inhibitors, such as p21, p27 and Kip2 [98].

7. Other mAbs for NHL

In addition to the above-mentioned strategies aiming to improve rituximab activity, numerous research efforts have led to new mAbs directed against different target antigens and to the development of radioimmunoconjugates. The most promising newer therapeutics are listed below.

- (i) Epratuzumab: a humanized IgG1 anti-CD22 antibody. It induces ADCC and CDC in preclinical studies. Phase I/II studies demonstrated objective responses in relapsed/refractory FL (24%) [99], and in DLBCL (15%) [100], without dose-limiting toxic effects.
- (ii) Galiximab: a primatised anti-CD80 (IgG1 λ) mAb with human constant regions and primate (cynomolgus macaque) variable regions [101]. CD80 is a costimulatory molecule involved in regulating T-cell activation. It is transiently expressed on the surface of activated B cells, dendritic cells, and T cells of healthy individuals [102]. Additionally, a variety of lymphoid malignancies constitutively express CD80, making this antigen a suitable target [103]. A phase-I/II study showed that GALIXIMAB is able to enhance rituximab antitumor activity in previously untreated NHL patients, with a response reported in 70% of patients [104].
- (iii) Alemtuzumab (Campath): a humanized monoclonal antibody against CD52 (an antigen expressed by normal and malignant B- and T-lymphocytes, monocytes, and NK cells). It is indicated for the treatment of patients with B-CLL refractory to fludarabine (ORR of 56%) [105], for advanced-stage mycosis fungoides/Sezary syndrome [106], and for relapsed or refractory peripheral T-cell lymphomas [107, 108]. Notably, although clinically effective, this mAb

induces a dramatic decrease in CD4+ and CD8+ T lymphocytes and thus strongly increases the risk of infections.

- (iv) Apolizumab (Hu1D10): a humanized anti-HLA-DR antibody that induces CDC, ADCC, and apoptosis. HLA class II antigens are expressed at the surface of professional antigen presenting cells, including B cells. They are involved in antigen presentation and in promoting cell proliferation. Thus, mAbs against HLA-DR inhibit B-cell proliferation and induce apoptosis through activation of the extrinsic apoptotic pathway. Recently, this type of approach has shown promising results in B-cell malignancies [109]. Single agent therapy APOLIZUMAB in previously untreated B-CLL patients showed an ORR of 83% [110]. Moreover, the combination of APOLIZUMAB and rituximab in relapsed/refractory B-cell lymphoma and B-CLL showed an ORR of 42% [111].
- (v) *Radioimmunotherapy*: this type of treatment involves the administration of an antibody linked to a radioisotope. This approach permits the targeting of the radioactive isotopes to cancer tissues and is especially interesting as it allows for killing neighboring cancer cells that either are inaccessible to the antibody or express insufficient antigen for the antibody to bind in adequate quantities. Two anti-CD20 radioimmunoconjugates are approved for use in patients with relapsed or refractory follicular or low-grade lymphoma:
 - (1) Yttrium-90: labelled ibritumomab tiuxetan (zevalin),
 - (2) iodine-131: labelled tositumomab (bexxar).

These therapeutics hold great promise for the treatment of NHL and their usefulness has recently been confirmed by several clinical trials [112–122].

About 80% of patients with follicular or low-grade lymphomas respond to treatment with Zevalin, with 20 to 30% achieving a CR. Interestingly, the duration of response appears to exceed 3 years in about 25% of patients [123]. The benefit of adding a radioisotope to the antibody was confirmed in a study enrolling patients with indolent NHL that were refractory to rituximab. In this study, Zevalin showed a 74% response rate and 15% of CR [115]. Additionally, as compared to rituximab, Zevalin produces higher response rates among patients with follicular or low-grade lymphoma who have not previously received antibody-based treatments (ORR 80% versus 56%, $P = .002$; CR 30% versus 16%, $P = .04$ [115]). Finally Zevalin also appears to be effective against some diffuse large B-cell lymphomas, and mantle-cell lymphomas, when used in sequence with chemotherapy (ORR of 53% versus 19%; OS 22.4, versus 4.6, resp.) [117].

Similar results are obtained with Bexxar. In particular in patients with NHL refractory to standard chemotherapy, treatment with Bexxar resulted in CR in 20% of patients [124]. Additionally, in one study, 95% of patients with NHL had responses to 131I-labeled tositumomab used as

initial treatment, with 75% demonstrating CR [125]. Finally, the results of a recently completed study (ClinicalTrials.gov number, NCT00006721), comparing CHOP followed by 131I-labeled tositumomab to rituximab plus CHOP for the initial treatment of FL, is predicted to redefine standard therapy for this disorder.

Importantly, there are other radiolabeled immunotherapeutics for NHL that are currently under evaluation [112, 114, 126–130]. These include

- (a) LL₂ anti-CD22, conjugated to either ¹³¹I or ⁹⁰Y; Lym-I;
- (b) anti-HLA-DR, conjugated to ⁹⁰Y or ⁶⁷Cu;
- (c) rituximab, conjugated to ²¹¹At, ¹⁸⁶Re, or ²²⁷Th;
- (d) anti-CD19 mAb conjugated to ⁹⁰Y.

8. Conclusions and Perspectives

Combining rituximab with chemotherapy has proven to be an effective treatment for both indolent and aggressive forms of NHL. The same type of treatment can be used in patients with B-CLL, although its efficacy in this disorder appears to be lower. In addition, it has also been demonstrated that using rituximab alone as a maintenance therapy improves the prognosis and extends disease-free survival in FL. Although a standard scheme for rituximab maintenance therapy has not been established yet, it is currently under investigation and the ongoing studies will establish the most effective regimen.

For patients in which treatment with rituximab has not given the expected results, autologous stem cell transplantations have shown promise. It has been demonstrated that using a cycle of rituximab in association with stem cell transplantations and after it as maintenance therapy, yields better results than transplant alone.

Radiolabeled antibodies may be effective in rituximab-resistant and chemotherapy-resistant disease, but their clinical use is still limited when compared to that of unlabeled mAbs. Recent data suggest that sequential radioimmunotherapy after chemotherapy may have significant clinical value. Additionally, novel monoclonal antibodies are under development. If these will prove to be more effective than rituximab will have to be assessed by randomized comparative trials.

Overall, the results obtained with antibody-based therapeutics in NHL are clearly highly promising. They herald the advent of therapeutic strategies based on targeted agents that will likely be more effective and, at the same time, less toxic than traditional chemotherapy-based treatments.

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References

- [1] G. Kohler and C. Milstein, "Continuous cultures of fused cells secreting antibody of predefined specificity," *Nature*, vol. 256, no. 5517, pp. 495–497, 1975.
- [2] N. Cooper and D. M. Arnold, "The effect of rituximab on humoral and cell mediated immunity and infection in the treatment of autoimmune diseases," *British Journal of Haematology*, vol. 149, no. 1, pp. 3–13, 2010.
- [3] O. J. Finn, "Cancer immunology," *New England Journal of Medicine*, vol. 358, no. 25, pp. 2704–2715, 2008.
- [4] G. L. Plosker and D. P. Figgitt, "Rituximab: a review of its use in non-Hodgkin's lymphoma and chronic lymphocytic leukaemia," *Drugs*, vol. 63, no. 8, pp. 803–843, 2003.
- [5] A. Demidem, T. Lam, S. Alas, K. Hariharan, N. Hanna, and B. Bonavida, "Chimeric anti-CD20 (idec-C2B8) monoclonal antibody sensitizes a B cell lymphoma cell line to cell killing by cytotoxic drugs," *Cancer Biotherapy and Radiopharmaceuticals*, vol. 12, no. 3, pp. 177–186, 1997.
- [6] L. M. Nadler, J. Ritz, and R. Hardy, "A unique cell surface antigen identifying lymphoid malignancies of B cell origin," *Journal of Clinical Investigation*, vol. 67, no. 1, pp. 134–140, 1981.
- [7] M. Petryk and M. L. Grossbard, "Rituximab therapy of B-cell neoplasms," *Clinical Lymphoma*, vol. 1, no. 3, pp. 186–194, 2000.
- [8] N. M. Almasri, R. E. Duque, J. Iturraspe, E. Everett, and R. C. Braylan, "Reduced expression of CD20 antigen as a characteristic marker for chronic lymphocytic leukemia," *American Journal of Hematology*, vol. 40, no. 4, pp. 259–263, 1992.
- [9] J. P. Deans, H. Li, and M. J. Polyak, "CD20-mediated apoptosis: signalling through lipid rafts," *Immunology*, vol. 107, no. 2, pp. 176–182, 2002.
- [10] J. K. Bubien, L. J. Zhou, P. D. Bell, R. A. Frizzell, and T. F. Tedder, "Transfection of the CD20 cell surface molecule into ectopic cell types generates a Ca conductance found constitutively in B lymphocytes," *Journal of Cell Biology*, vol. 121, no. 5, pp. 1121–1132, 1993.
- [11] C. A. Walshe, S. A. Beers, R. R. French et al., "Induction of cytosolic calcium flux by CD20 is dependent upon B cell antigen receptor signaling," *Journal of Biological Chemistry*, vol. 283, no. 25, pp. 16971–16984, 2008.
- [12] A. J. Grillo-López, E. Hedrick, M. Rashford, and M. Benyunes, "Rituximab: ongoing and future clinical development," *Seminars in Oncology*, vol. 29, no. 1, pp. 105–112, 2002.
- [13] S. V. Onrust, H. M. Lamb, and J. A. Balfour, "Rituximab," *Drugs*, vol. 58, no. 1, pp. 79–88, 1999.
- [14] M. J. Glennie, R. R. French, M. S. Cragg, and R. P. Taylor, "Mechanisms of killing by anti-CD20 monoclonal antibodies," *Molecular Immunology*, vol. 44, no. 16, pp. 3823–3837, 2007.
- [15] J. L. Teeling, W. J. M. Mackus, L. J. J. M. Wiegman et al., "The biological activity of human CD20 monoclonal antibodies is linked to unique epitopes on CD20," *Journal of Immunology*, vol. 177, no. 1, pp. 362–371, 2006.
- [16] A. R. Jazirehi and B. Bonavida, "Cellular and molecular signal transduction pathways modulated by rituximab (rituxan, anti-CD20 mAb) in non-Hodgkin's lymphoma: implications in chemosensitization and therapeutic intervention," *Oncogene*, vol. 24, no. 13, pp. 2121–2143, 2005.

- [17] B. Bonavida, "Rituximab-induced inhibition of antiapoptotic cell survival pathways: implications in chemo/immuno-resistance, rituximab unresponsiveness, prognostic and novel therapeutic interventions," *Oncogene*, vol. 26, no. 25, pp. 3629–3636, 2007.
- [18] A. R. Jazirehi, M. I. Vega, D. Chatterjee, L. Goodglick, and B. Bonavida, "Inhibition of the Raf-MEK1/2-ERK1/2 signaling pathway, BCL-2 down-regulation, and chemosensitization of non-Hodgkin's lymphoma B cells by rituximab," *Cancer Research*, vol. 64, no. 19, pp. 7117–7126, 2004.
- [19] S. Alas and B. Bonavida, "Rituximab inactivates signal transducer and activation of transcription 3 (STAT3) activity in B-non-Hodgkin's lymphoma through inhibition of the interleukin 10 autocrine/paracrine loop and results in down-regulation of Bcl-2 and sensitization to cytotoxic drugs," *Cancer Research*, vol. 61, no. 13, pp. 5137–5144, 2001.
- [20] M. I. Vega, A. R. Jazirehi, S. Huerta-Yepez, and B. Bonavida, "Rituximab-induced inhibition of YY1 and Bcl-xL expression in ramos non-Hodgkin's lymphoma cell line via inhibition of NF-kappaB activity: role of YY1 and Bcl-xL in Fas resistance and chemoresistance, respectively," *Journal of Immunology*, vol. 175, no. 4, pp. 2174–2183, 2005.
- [21] S. Huerta-Yepez, M. Vega, A. Jazirehi et al., "Nitric oxide sensitizes prostate carcinoma cell lines to TRAIL-mediated apoptosis via inactivation of NF-kB and inhibition of Bcl-expression," *Oncogene*, vol. 23, no. 29, pp. 4993–5003, 2004.
- [22] S. Kheirallah, P. Caron, E. Gross et al., "Rituximab inhibits B-cell receptor signaling," *Blood*, vol. 115, no. 5, pp. 985–994, 2010.
- [23] L. Chen, S. Monti, P. Juszczynski et al., "SYK-dependent tonic B-cell receptor signaling is a rational treatment target in diffuse large B-cell lymphoma," *Blood*, vol. 111, no. 4, pp. 2230–2237, 2008.
- [24] L. Leseux, S. M. Hamdi, T. Al Saati et al., "Syk-dependent mTOR activation in follicular lymphoma cells," *Blood*, vol. 108, no. 13, pp. 4156–4162, 2006.
- [25] R. M. Young, I. R. Hardy, R. L. Clarke et al., "Mouse models of non-hodgkin lymphoma reveal Syk as an important therapeutic target," *Blood*, vol. 113, no. 11, pp. 2508–2516, 2009.
- [26] M. R. Smith, "Rituximab (monoclonal anti-CD20 antibody): mechanisms of action and resistance," *Oncogene*, vol. 22, no. 47, pp. 7359–7368, 2003.
- [27] J. O. Armitage, "How I treat patients with diffuse large B-cell lymphoma," *Blood*, vol. 110, no. 1, pp. 29–36, 2007.
- [28] R. Marcus, K. Imrie, P. Solal-Celigny et al., "Phase III study of R-CVP compared with cyclophosphamide, vincristine, and prednisone alone in patients with previously untreated advanced follicular lymphoma," *Journal of Clinical Oncology*, vol. 26, no. 28, pp. 4579–4586, 2008.
- [29] W. Hiddemann, M. Kneba, M. Dreyling et al., "Front-line therapy with rituximab added to the combination of cyclophosphamide, doxorubicin, vincristine, and prednisone (CHOP) significantly improves the outcome for patients with advanced-stage follicular lymphoma compared with therapy with CHOP alone: results of a prospective randomized study of the German Low-Grade Lymphoma Study Group," *Blood*, vol. 106, no. 12, pp. 3725–3732, 2005.
- [30] M. H. J. Van Oers, R. Klasa, R. E. Marcus et al., "Rituximab maintenance improves clinical outcome of relapsed/resistant follicular non-Hodgkin lymphoma in patients both with and without rituximab during induction: results of a prospective randomized phase 3 intergroup trial," *Blood*, vol. 108, no. 10, pp. 3295–3301, 2006.
- [31] R. Forstpointner, M. Unterhalt, M. Dreyling et al., "Maintenance therapy with rituximab leads to a significant prolongation of response duration after salvage therapy with a combination of rituximab, fludarabine, cyclophosphamide, and mitoxantrone (R-FCM) in patients with recurring and refractory follicular and mantle cell lymphomas: results of a prospective randomized study of the German Low Grade Lymphoma Study Group (GLSG)," *Blood*, vol. 108, no. 13, pp. 4003–4008, 2006.
- [32] M. Herold, A. Haas, S. Srock et al., "Rituximab added to first-line mitoxantrone, chlorambucil, and prednisolone chemotherapy followed by interferon maintenance prolongs survival in patients with advanced follicular lymphoma: an East German study group hematology and oncology study," *Journal of Clinical Oncology*, vol. 25, no. 15, pp. 1986–1992, 2007.
- [33] D. G. Maloney, A. J. Grillo-López, C. A. White et al., "IDEC-C2B8 (rituximab) anti-CD20 monoclonal antibody therapy in patients with relapsed low-grade non-Hodgkin's lymphoma," *Blood*, vol. 90, no. 6, pp. 2188–2195, 1997.
- [34] M. Feuring-Buske, M. Kneba, M. Unterhalt et al., "IDEC-C2B8 (Rituximab) anti-CD20 antibody treatment in relapsed advanced-stage follicular lymphomas: results of a phase-II study of the German low-grade lymphoma study group," *Annals of Hematology*, vol. 79, no. 9, pp. 493–500, 2000.
- [35] P. McLaughlin, A. J. Grillo-López, B. K. Link et al., "Rituximab chimeric anti-CD20 monoclonal antibody therapy for relapsed indolent lymphoma: half of patients respond to a four-dose treatment program," *Journal of Clinical Oncology*, vol. 16, no. 8, pp. 2825–2833, 1998.
- [36] B. Coiffier, E. Lepage, J. Brière et al., "Chop chemotherapy plus rituximab compared with chop alone in elderly patients with diffuse large-B-cell lymphoma," *New England Journal of Medicine*, vol. 346, no. 4, pp. 235–242, 2002.
- [37] M. Pfreundschuh, L. Trümper, A. Österborg et al., "CHOP-like chemotherapy plus rituximab versus CHOP-like chemotherapy alone in young patients with good-prognosis diffuse large-B-cell lymphoma: a randomised controlled trial by the MabThera International Trial (MInT) Group," *Lancet Oncology*, vol. 7, no. 5, pp. 379–391, 2006.
- [38] T. M. Habermann, E. A. Weller, V. A. Morrison et al., "Rituximab-CHOP versus CHOP alone or with maintenance rituximab in older patients with diffuse large B-cell lymphoma," *Journal of Clinical Oncology*, vol. 24, no. 19, pp. 3121–3127, 2006.
- [39] G. Lenz, M. Dreyling, E. Hoster et al., "Immunochemotherapy with rituximab and cyclophosphamide, doxorubicin, vincristine, and prednisone significantly improves response and time to treatment failure, but not long-term outcome in patients with previously untreated mantle cell lymphoma: results of a prospective randomized trial of the German Low Grade Lymphoma Study Group (GLSG)," *Journal of Clinical Oncology*, vol. 23, no. 9, pp. 1984–1992, 2005.
- [40] T. Robak, A. Dmoszynska, P. Solal-Céligny et al., "Rituximab plus fludarabine and cyclophosphamide prolongs progression-free survival compared with fludarabine and cyclophosphamide alone in previously treated chronic lymphocytic leukemia," *Journal of Clinical Oncology*, vol. 28, no. 10, pp. 1756–1765, 2010.
- [41] CLL8- German CLL Study Group.
- [42] E. Kimby, M. Björkholm, G. Gahrton et al., "Chlorambucil/prednisone vs. CHOP in symptomatic low-grade non-Hodgkin's lymphomas: a randomized trial from the

- Lymphoma Group of Central Sweden," *Annals of Oncology*, vol. 5, no. 2, pp. 67–71, 1994.
- [43] M. S. Czuczman, R. Weaver, B. Alkuzweny, J. Berlfein, and A. J. Grillo-López, "Prolonged clinical and molecular remission in patients with low-grade or follicular non-Hodgkin's lymphoma treated with rituximab plus CHOP chemotherapy: 9-year follow-up," *Journal of Clinical Oncology*, vol. 22, no. 23, pp. 4659–4664, 2004.
- [44] M. S. Czuczman, A. Koryzna, A. Mohr et al., "Rituximab in combination with fludarabine chemotherapy in low-grade or follicular lymphoma," *Journal of Clinical Oncology*, vol. 23, no. 4, pp. 694–704, 2005.
- [45] W. Hiddemann, C. Buske, M. Dreyling et al., "Treatment strategies in follicular lymphomas: current status and future perspectives," *Journal of Clinical Oncology*, vol. 23, no. 26, pp. 6394–6399, 2005.
- [46] E. Domingo-Domenech, E. González-Borco, C. Estany, A. Sureda, J. Besalduch, and A. F. De Sevilla, "Combined treatment with anti CD20 (rituximab) and CHOP in relapsed advanced-stage follicular lymphomas," *Haematologica*, vol. 87, no. 11, pp. 1229–1230, 2002.
- [47] T. A. Davis, A. J. Grillo-Lopez, C. A. White et al., "Rituximab anti-CD20 monoclonal antibody therapy in non-Hodgkin's lymphoma: safety and efficacy of re-treatment," *Journal of Clinical Oncology*, vol. 18, no. 17, pp. 3135–3143, 2000.
- [48] T. A. Davis, C. A. White, A. J. Grillo-López et al., "Single-agent monoclonal antibody efficacy in bulky non-Hodgkin's lymphoma: results of a phase II trial of rituximab," *Journal of Clinical Oncology*, vol. 17, no. 6, pp. 1851–1857, 1999.
- [49] N. Lamanna, J. G. Jurcic, A. Noy et al., "Sequential therapy with fludarabine, high-dose cyclophosphamide, and rituximab in previously untreated patients with chronic lymphocytic leukemia produces high-quality responses: molecular remissions predict for durable complete responses," *Journal of Clinical Oncology*, vol. 27, no. 4, pp. 491–497, 2009.
- [50] T. Ahmadi and S. J. Schuster, "Variations on the fludarabine, cyclophosphamide, and rituximab combination in chronic lymphocytic leukemia therapy: what have we learned?" *Journal of Clinical Oncology*, vol. 27, no. 4, pp. 479–480, 2009.
- [51] L. Kunkel, A. Wong, T. Maneatis et al., "Optimizing the use of rituximab for treatment of B-cell non-Hodgkin's lymphoma: a benefit-risk update," *Seminars in Oncology*, vol. 27, no. 6, pp. 53–61, 2000.
- [52] D. Bedognetti, G. Zoppoli, C. Massucco et al., "Patients treated with rituximab-containing immunochemotherapy have a significant and prolonged lack of humoral response to influenza vaccine associated with a persistent depletion of b memory cells," in *Blood (ASH Annual Meeting Abstracts)*, November 2009.
- [53] B. D. Cheson and J. P. Leonard, "Monoclonal antibody therapy for B-cell non-Hodgkin's lymphoma," *New England Journal of Medicine*, vol. 359, no. 6, pp. 613–626, 2008.
- [54] S. A. Beers, R. R. French, H. T. C. Chan et al., "Antigenic modulation limits the efficacy of anti-CD20 antibodies: implications for antibody selection," *Blood*, vol. 115, no. 25, pp. 5191–5201, 2010.
- [55] S. Y. Wang, S. Veeramani, E. Racila et al., "Depletion of the C3 component of complement enhances the ability of rituximab-coated target cells to activate human NK cells and improves the efficacy of monoclonal antibody therapy in an in vivo model," *Blood*, vol. 114, no. 26, pp. 5322–5330, 2009.
- [56] A. Rambaldi, M. Lazzari, C. Manzoni et al., "Monitoring of minimal residual disease after CHOP and rituximab in previously untreated patients with follicular lymphoma," *Blood*, vol. 99, no. 3, pp. 856–862, 2002.
- [57] C. G. A. Price, J. Meerabux, S. Murtagh et al., "The significance of circulating cells carrying t(14;18) in long remission from follicular lymphoma," *Journal of Clinical Oncology*, vol. 9, no. 9, pp. 1527–1532, 1991.
- [58] J. Limpens, R. Stad, C. Vos et al., "Lymphoma-associated translocation t(14;18) in blood B cells of normal individuals," *Blood*, vol. 85, no. 9, pp. 2528–2536, 1995.
- [59] M. H. van Oers, E. Tönnissen, M. Van Glabbeke et al., "BCL-2/IgH polymerase chain reaction status at the end of induction treatment is not predictive for progression-free survival in relapsed/resistant follicular lymphoma: results of a prospective randomized EORTC 20981 phase III intergroup study," *Journal of Clinical Oncology*, vol. 28, no. 13, pp. 2246–2252, 2010.
- [60] N. Mounier, J. Briere, C. Gisselbrecht et al., "Rituximab plus CHOP (R-CHOP) overcomes bcl-2-associated resistance to chemotherapy in elderly patients with diffuse large B-cell lymphoma (DLBCL)," *Blood*, vol. 101, no. 11, pp. 4279–4284, 2003.
- [61] N. Mounier, J. Briere, C. Gisselbrecht, F. Reyes, P. Gaulard, and B. Coiffier, "Estimating the impact of rituximab on bcl-2-associated resistance to CHOP in elderly patients with diffuse large B-cell lymphoma," *Haematologica*, vol. 91, no. 5, pp. 715–716, 2006.
- [62] T. Kewalramani, A. D. Zelenetz, E. E. Hedrick et al., "High-dose chemoradiotherapy and autologous stem cell transplantation for patients with primary refractory aggressive non-Hodgkin lymphoma: an intention-to-treat analysis," *Blood*, vol. 96, no. 7, pp. 2399–2404, 2000.
- [63] C. Haioun, E. Lepage, C. Gisselbrecht et al., "Survival benefit of high-dose therapy in poor-risk aggressive non-Hodgkin's lymphoma: final analysis of the prospective LNH87-2 protocol—a groupe d'étude des lymphomes de l'adulte study," *Journal of Clinical Oncology*, vol. 18, no. 16, pp. 3025–3030, 2000.
- [64] A. Nademanee and S. J. Forman, "Role of Hematopoietic Stem Cell Transplantation for Advanced-Stage Diffuse Large Cell B-Cell Lymphoma-B," *Seminars in Hematology*, vol. 43, no. 4, pp. 240–250, 2006.
- [65] N. L. Berinstein, "Principles of maintenance therapy," *Leukemia Research*, vol. 30, no. 1, pp. S3–S10, 2006.
- [66] J. D. Hainsworth, S. Litchy, H. A. Burris et al., "Rituximab as first-line and maintenance therapy for patients with indolent non-Hodgkin's lymphoma," *Journal of Clinical Oncology*, vol. 20, no. 20, pp. 4261–4267, 2002.
- [67] J. D. Hainsworth, S. Litchy, D. W. Shaffer, L. Van Lackey, M. Grimaldi, and F. A. Greco, "Maximizing therapeutic benefit of rituximab: maintenance therapy versus re-treatment at progression in patients with indolent non-Hodgkin's lymphoma—a randomized phase II trial of the Minnie Pearl Cancer Research Network," *Journal of Clinical Oncology*, vol. 23, no. 6, pp. 1088–1095, 2005.
- [68] C. Buske and W. Hiddemann, "Rituximab maintenance therapy in indolent NHL: a clinical review," *Leukemia Research*, vol. 30, no. 1, pp. S11–S15, 2006.
- [69] M. Ghielmini, S. F. H. Schmitz, S. B. Cogliatti et al., "Prolonged treatment with rituximab in patients with follicular lymphoma significantly increases event-free survival and response duration compared with the standard weekly \times 4 schedule," *Blood*, vol. 103, no. 12, pp. 4416–4423, 2004.

- [70] K. A. Foon and M. J. Hallek, "Changing paradigms in the treatment of chronic lymphocytic leukemia," *Leukemia*, vol. 24, no. 3, pp. 500–511, 2010.
- [71] H. Schulz, S. K. Klein, U. Rehwald et al., "Phase 2 study of a combined immunochemotherapy using rituximab and fludarabine in patients with chronic lymphocytic leukemia," *Blood*, vol. 100, no. 9, pp. 3115–3120, 2002.
- [72] N. Lamanna, M. Kalaycio, P. Maslak et al., "Pentostatin, cyclophosphamide, and rituximab is an active, well-tolerated regimen for patients with previously treated chronic lymphocytic leukemia," *Journal of Clinical Oncology*, vol. 24, no. 10, pp. 1575–1581, 2006.
- [73] T. Robak, P. Smolewski, B. Cebula, O. Grzybowska-Izdorczyk, and J. Z. Błoński, "Rituximab plus cladribine with or without cyclophosphamide in patients with relapsed or refractory chronic lymphocytic leukemia," *European Journal of Haematology*, vol. 79, no. 2, pp. 107–113, 2007.
- [74] M. Hallek, G. Fingerle-Rowson, and A.-M. Fink, "First-line treatment with fludarabine (f), cyclophosphamide (c), and rituximab (r) (fcr) improves overall survival (os) in previously untreated patients (pts) with advanced chronic lymphocytic leukemia (cll): results of a randomized phase iii trial on behalf of an international group of investigators and the german cll study group," *Blood*, vol. 114, 2009.
- [75] M. J. Keating, S. O'Brien, M. Albitar et al., "Early results of a chemoimmunotherapy regimen of fludarabine, cyclophosphamide, and rituximab as initial therapy for chronic lymphocytic leukemia," *Journal of Clinical Oncology*, vol. 23, no. 18, pp. 4079–4088, 2005.
- [76] C. S. Tam, S. O'Brien, W. Wierda et al., "Long-term results of the fludarabine, cyclophosphamide, and rituximab regimen as initial therapy of chronic lymphocytic leukemia," *Blood*, vol. 112, no. 4, pp. 975–980, 2008.
- [77] W. Wierda, S. O'Brien, S. Wen et al., "Chemoimmunotherapy with fludarabine, cyclophosphamide, and rituximab for relapsed and refractory chronic lymphocytic leukemia," *Journal of Clinical Oncology*, vol. 23, no. 18, pp. 4070–4078, 2005.
- [78] P. V. Beum, M. A. Lindorfer, and R. P. Taylor, "Within peripheral blood mononuclear cells, antibody-dependent cellular cytotoxicity of rituximab-opsonized Daudi cells is promoted by NK cells and inhibited by monocytes due to shaving," *Journal of Immunology*, vol. 181, no. 4, pp. 2916–2924, 2008.
- [79] A. D. Kennedy, P. V. Beum, M. D. Solga et al., "Rituximab infusion promotes rapid complement depletion and acute CD20 loss in chronic lymphocytic leukemia," *Journal of Immunology*, vol. 172, no. 5, pp. 3280–3288, 2004.
- [80] P. V. Beum, A. D. Kennedy, and R. P. Taylor, "Three new assays for rituximab based on its immunological activity or antigenic properties: analyses of sera and plasmas of RTX-treated patients with chronic lymphocytic leukemia and other B cell lymphomas," *Journal of Immunological Methods*, vol. 289, no. 1-2, pp. 97–109, 2004.
- [81] R. Shimizu, J. Kikuchi, T. Wada, K. Ozawa, Y. Kano, and Y. Furukawa, "HDAC inhibitors augment cytotoxic activity of rituximab by upregulating CD20 expression on lymphoma cells," *Leukemia*, vol. 24, no. 10, pp. 1760–1768, 2010.
- [82] C. Henry, M. Deschamps, P. S. Rohrlach et al., "Identification of an alternative CD20 transcript variant in B-cell malignancies coding for a novel protein associated to rituximab resistance," *Blood*, vol. 115, no. 12, pp. 2420–2429, 2010.
- [83] R. Bhat and C. Watzl, "Serial killing of tumor cells by human natural killer cells—enhancement by therapeutic antibodies," *PLoS ONE*, vol. 2, no. 3, article no. e326, 2007.
- [84] J. A. Bowles and G. J. Weiner, "CD16 polymorphisms and NK activation induced by monoclonal antibody-coated target cells," *Journal of Immunological Methods*, vol. 304, no. 1-2, pp. 88–99, 2005.
- [85] L. Fischer, O. Penack, C. Gentilini et al., "The anti-lymphoma effect of antibody-mediated immunotherapy is based on an increased degranulation of peripheral blood natural killer (NK) cells," *Experimental Hematology*, vol. 34, no. 6, pp. 753–759, 2006.
- [86] J. G. Berdeja, A. Hess, D. M. Lucas et al., "Systemic interleukin-2 and adoptive transfer of lymphokine-activated killer cells improves antibody-dependent cellular cytotoxicity in patients with relapsed B-cell lymphoma treated with rituximab," *Clinical Cancer Research*, vol. 13, no. 8, pp. 2392–2399, 2007.
- [87] H. Wang, Y. Liu, Z. Y. Li, X. Fan, A. Hemminki, and A. Lieber, "A recombinant adenovirus type 35 fiber knob protein sensitizes lymphoma cells to rituximab therapy," *Blood*, vol. 115, no. 3, pp. 592–600, 2010.
- [88] F. Sato, A. Ito, T. Ishida et al., "A complement-dependent cytotoxicity-enhancing anti-CD20 antibody mediating potent antitumor activity in the humanized NOD/Shi-scid, IL-2R γ ^{null} mouse lymphoma model," *Cancer Immunology, Immunotherapy*, vol. 59, no. 12, pp. 1791–1800, 2010.
- [89] M. P. Chao, A. A. Alizadeh, C. Tang et al., "Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma," *Cell*, vol. 142, no. 5, pp. 699–713, 2010.
- [90] C. Xuan, K. K. Steward, J. M. Timmerman, and S. L. Morrison, "Targeted delivery of interferon-alpha via fusion to anti-CD20 results in potent antitumor activity against B-cell lymphoma," *Blood*, vol. 115, no. 14, pp. 2864–2871, 2010.
- [91] B. Li, L. Zhao, H. Guo et al., "Characterization of a rituximab variant with potent antitumor activity against rituximab-resistant B-cell lymphoma," *Blood*, vol. 114, no. 24, pp. 5007–5015, 2009.
- [92] R. Stein, Z. Qu, S. Chen et al., "Characterization of a new humanized anti-CD20 monoclonal antibody, IMMU-106, and its use in combination with the humanized anti-CD22 antibody, epratuzumab, for the therapy of non-Hodgkin's lymphoma," *Clinical Cancer Research*, vol. 10, no. 8, pp. 2868–2878, 2004.
- [93] S. Maddipatla, F. J. Hernandez-Ilizaliturri, J. Knight, and M. S. Czuczman, "Augmented antitumor activity against b-cell lymphoma by a combination of monoclonal antibodies targeting trail-r1 and cd20," *Clinical Cancer Research*, vol. 13, no. 15, pp. 4556–4564, 2007.
- [94] E. Tobin, G. L. DeNardo, N. Zhang, A. L. Epstein, C. Liu, and S. DeNardo, "Combination immunotherapy with anti-CD20 and anti-HLA-DR monoclonal antibodies induces synergistic anti-lymphoma effects in human lymphoma cell lines," *Leukemia and Lymphoma*, vol. 48, no. 5, pp. 944–956, 2007.
- [95] E. A. Rossi, D. M. Goldenberg, T. M. Cardillo, W. J. McBride, R. M. Sharkey, and C. H. Chang, "Stably tethered multifunctional structures of defined composition made by the dock and lock method for use in cancer targeting," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 18, pp. 6841–6846, 2006.
- [96] E. A. Rossi, D. M. Goldenberg, T. M. Cardillo, R. Stein, and C. H. Chang, "Hexavalent bispecific antibodies represent a

- new class of anticancer therapeutics—1. Properties of anti-CD20/CD22 antibodies in lymphoma,” *Blood*, vol. 113, no. 24, pp. 6161–6171, 2009.
- [97] E. A. Rossi, D. M. Goldenberg, T. M. Cardillo, R. Stein, Y. Wang, and C. H. Chang, “Novel designs of multivalent anti-CD20 humanized antibodies as improved lymphoma therapeutics,” *Cancer Research*, vol. 68, no. 20, pp. 8384–8392, 2008.
- [98] P. Gupta, D. M. Goldenberg, E. A. Rossi, and C.-H. Chang, “Multiple signaling pathways induced by hexavalent, monospecific, anti-CD20 and hexavalent, bispecific, anti-CD20/CD22 humanized antibodies correlate with enhanced toxicity to B-cell lymphomas and leukemias,” *Blood*, vol. 116, no. 17, pp. 3258–3267, 2010.
- [99] J. P. Leonard, M. Coleman, J. C. Ketas et al., “Phase I/II trial of epratuzumab (humanized anti-CD22 antibody) in indolent non-Hodgkin’s lymphoma,” *Journal of Clinical Oncology*, vol. 21, no. 16, pp. 3051–3059, 2003.
- [100] J. P. Leonard, M. Coleman, J. C. Ketas et al., “Epratuzumab, a humanized anti-CD22 antibody, in aggressive non-Hodgkin’s lymphoma: phase I/II clinical trial results,” *Clinical Cancer Research*, vol. 10, no. 16, pp. 5327–5334, 2004.
- [101] A. Younes, K. Hariharan, R. S. Allen, and B. R. Leigh, “Initial trials of anti-CD80 monoclonal antibody (galiximab) therapy for patients with relapsed or refractory follicular lymphoma,” *Clinical Lymphoma*, vol. 3, no. 4, pp. 257–259, 2003.
- [102] F. A. Vyth-Dreese, T. A. M. Dellemijn, D. Majoor, and D. De Jong, “Localization in situ of the co-stimulatory molecules B7.1, B7.2, CD40 and their ligands in normal human lymphoid tissue,” *European Journal of Immunology*, vol. 25, no. 11, pp. 3023–3029, 1995.
- [103] J. M. Munro, A. S. Freedman, J. C. Aster et al., “In vivo expression of the B7 costimulatory molecule by subsets of antigen-presenting cells and the malignant cells of Hodgkin’s disease,” *Blood*, vol. 83, no. 3, pp. 793–798, 1994.
- [104] J. P. Leonard, J. W. Friedberg, A. Younes et al., “A phase I/II study of galiximab (an anti-CD80 monoclonal antibody) in combination with rituximab for relapsed or refractory, follicular lymphoma,” *Annals of Oncology*, vol. 18, no. 7, pp. 1216–1223, 2007.
- [105] M. J. Keating, I. Flinn, V. Jain et al., “Therapeutic role of alemtuzumab (Campath-1H) in patients who have failed fludarabine: results of a large international study,” *Blood*, vol. 99, no. 10, pp. 3554–3561, 2002.
- [106] J. Lundin, H. Hagberg, R. Repp et al., “Phase 2 study of alemtuzumab (anti-CD52 monoclonal antibody) in patients with advanced mycosis fungoides/Sézary syndrome,” *Blood*, vol. 101, no. 11, pp. 4267–4272, 2003.
- [107] G. Enblad, H. Hagberg, M. Erlanson et al., “A pilot study of alemtuzumab (anti-CD52 monoclonal antibody) therapy for patients with relapsed or chemotherapy-refractory peripheral T-cell lymphomas,” *Blood*, vol. 103, no. 8, pp. 2920–2924, 2004.
- [108] A. Gallamini, F. Zaja, C. Patti et al., “Alemtuzumab (Campath-1H) and CHOP chemotherapy as first-line treatment of peripheral T-cell lymphoma: results of a GITIL (Gruppo Italiano Terapie Innovative nei Linfomi) prospective multicenter trial,” *Blood*, vol. 110, no. 7, pp. 2316–2323, 2007.
- [109] R. Stein, Z. Qu, S. Chen, D. Solis, H. J. Hansen, and D. M. Goldenberg, “Characterization of a humanized IgG4 anti-HLA-DR monoclonal antibody that lacks effector cell functions but retains direct antilymphoma activity and increases the potency of rituximab,” *Blood*, vol. 108, no. 8, pp. 2736–2744, 2006.
- [110] P. Hillmen, A. B. Skotnicki, T. Robak et al., “Alemtuzumab compared with chlorambucil as first-line therapy for chronic lymphocytic leukemia,” *Journal of Clinical Oncology*, vol. 25, no. 35, pp. 5616–5623, 2007.
- [111] K. Dunleavy, T. White, N. Grant et al., “Phase I study of combination rituximab with apolizumab in relapsed/refractory B-cell lymphoma and chronic lymphocytic leukemia,” vol. 23, 2005, ASCO Meeting Abstract 6607.
- [112] G. L. DeNardo, S. J. DeNardo, D. S. Goldstein et al., “Maximum-tolerated dose, toxicity, and efficacy of I-Lym-1 antibody for fractionated radioimmunotherapy of non-Hodgkin’s lymphoma,” *Journal of Clinical Oncology*, vol. 16, no. 10, pp. 3246–3256, 1998.
- [113] R. Schnell, M. Dietlein, K. Schomäcker et al., “Yttrium-90 ibritumomab tiuxetan-induced complete remission in a patient with classical lymphocyte-rich Hodgkin’s lymphoma,” *Onkologie*, vol. 31, no. 1-2, pp. 49–51, 2008.
- [114] D. Ma, M. R. McDevitt, E. Barendswaard et al., “Radioimmunotherapy for model B cell malignancies using Y-labeled anti-CD19 and anti-CD20 monoclonal antibodies,” *Leukemia*, vol. 16, no. 1, pp. 60–66, 2002.
- [115] T. E. Witzig, L. I. Gordon, F. Cabanillas et al., “Randomized controlled trial of yttrium-90-labeled ibritumomab tiuxetan radioimmunotherapy versus rituximab immunotherapy for patients with relapsed or refractory low-grade, follicular, or transformed B-cell non-Hodgkin’s lymphoma,” *Journal of Clinical Oncology*, vol. 20, no. 10, pp. 2453–2463, 2002.
- [116] T. E. Witzig, A. Molina, L. I. Gordon et al., “Long-term responses in patients with recurring or refractory B-cell non-Hodgkin lymphoma treated with yttrium 90 ibritumomab tiuxetan,” *Cancer*, vol. 109, no. 9, pp. 1804–1810, 2007.
- [117] F. Morschhauser, T. Illidge, D. Huglo et al., “Efficacy and safety of yttrium-90 ibritumomab tiuxetan in patients with relapsed or refractory diffuse large B-cell lymphoma not appropriate for autologous stem-cell transplantation,” *Blood*, vol. 110, no. 1, pp. 54–58, 2007.
- [118] P. L. Zinzani, M. Tani, S. Fanti et al., “A phase II trial of CHOP chemotherapy followed by yttrium 90 ibritumomab tiuxetan (Zevalin) for previously untreated elderly diffuse large B-cell lymphoma patients,” *Annals of Oncology*, vol. 19, no. 4, pp. 769–773, 2008.
- [119] S. J. Horning, A. Younes, V. Jain et al., “Efficacy and safety of tositumomab and iodine-131 tositumomab (Bexxar) in B-cell lymphoma, progressive after rituximab,” *Journal of Clinical Oncology*, vol. 23, no. 4, pp. 712–719, 2005.
- [120] M. S. Kaminski, M. Tuck, J. Estes et al., “¹³¹I-tositumomab therapy as initial treatment for follicular lymphoma,” *New England Journal of Medicine*, vol. 352, no. 5, pp. 441–449, 2005.
- [121] O. W. Press, J. M. Unger, R. M. Brazier et al., “Phase II trial of CHOP chemotherapy followed by tositumomab/iodine I-131 tositumomab for previously untreated follicular non-Hodgkin’s lymphoma: five-year follow-up of Southwest Oncology Group protocol S9911,” *Journal of Clinical Oncology*, vol. 24, no. 25, pp. 4143–4149, 2006.
- [122] A. K. Gopal, J. M. Pagel, J. R. Fromm, S. Wilbur, and O. W. Press, “¹³¹I anti-CD45 radioimmunotherapy effectively targets and treats T-cell non-Hodgkin lymphoma,” *Blood*, vol. 113, no. 23, pp. 5905–5910, 2009.
- [123] T. Witzig, C. Emmanouilides, A. Molina, L. Gordon, I. Gaston, and I. Flinn, “Yttrium-90 ibritumomab tiuxetan radioimmunotherapy (RIT) induces durable complete

- responses (CR/CRu) in patients with relapsed or refractory B-cell non-Hodgkin's lymphoma (NHL)," *Proceedings of the American Society of Clinical Oncology*, vol. 22, p. 597, 2003.
- [124] M. S. Kaminski, A. D. Zelenetz, O. W. Press et al., "Pivotal study of iodine I 131 tositumomab for chemotherapy-refractory low-grade or transformed low-grade B-cell non-Hodgkin's lymphomas," *Journal of Clinical Oncology*, vol. 19, no. 19, pp. 3918–3928, 2001.
- [125] M. S. Kaminski, M. Tuck, J. Estes et al., "¹³¹I-tositumomab therapy as initial treatment for follicular lymphoma," *New England Journal of Medicine*, vol. 352, no. 5, pp. 441–449, 2005.
- [126] M. E. Juweid, E. Stadtmauer, G. Hajjar et al., "Pharmacokinetics, dosimetry, and initial therapeutic results with I- and In-/Y-labeled humanized LL2 anti-CD22 monoclonal antibody in patients with relapsed, refractory non-Hodgkin's lymphoma," *Clinical Cancer Research*, vol. 5, no. 10, pp. 3292s–3303s, 1999.
- [127] R. O'Donnell, G. DeNardo, D. Kukis et al., "⁶⁷copper-2-iminothiolane-6-[p-(bromoacetamido) benzyl]-teta-lym-1 for radioimmunotherapy of non-hodgkin's lymphoma," *Clinical Cancer Research*, vol. 5, no. 10, p. 3330s, 1999.
- [128] R. T. O'Donnell, G. L. DeNardo, D. L. Kukis et al., "A clinical trial of radioimmunotherapy with Cu-2IT-BAT-Lym-1 for non-Hodgkin's lymphoma," *Journal of Nuclear Medicine*, vol. 40, no. 12, pp. 2014–2020, 1999.
- [129] E. Aurlien, R. H. Larsen, G. Kvalheim, and Ø. S. Bruland, "Demonstration of highly specific toxicity of the α -emitting radioimmunoconjugate ²¹¹At-rituximab against non-Hodgkin's lymphoma cells," *British Journal of Cancer*, vol. 83, no. 10, pp. 1375–1379, 2000.
- [130] S. Knop, A. Jakob, L. Kanz, H. Hebart, R. Bares, and B. Dohmen, "¹⁸⁶Rhenium-labeled anti-CD20 antibody radioimmunotherapy followed by autologous peripheral blood stem cell transplantation in patients with relapsed or refractory non-Hodgkin lymphoma," *Blood*, vol. 103, no. 3, p. 1175, 2004.



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